SCIENTIFIC REPORTS

natureresearch

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OPEN Genome-wide association study and polygenic risk score analysis of esketamine treatment response

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To elucidate the genetic underpinnings of the antidepressant efficacy of S-ketamine (esketamine) nasal spray in major depressive disorder (MDD), we performed a genome-wide association study (GWAS) in cohorts of European ancestry (n = 527). This analysis was followed by a polygenic risk score approach to test for associations between genetic loading for psychiatric conditions, symptom profiles and esketamine efficacy. We identified a genome-wide significant locus in *IRAK3* ($p = 3.57 \times 10^{-8}$, rs11465988, $\beta = -51.6$, SE = 9.2) and a genome-wide significant gene-level association in *NME7* $(p = 1.73 \times 10^{-6})$ for esketamine efficacy (i.e. percentage change in symptom severity score compared to baseline). Additionally, the strongest association with esketamine efficacy identified in the polygenic score analysis was from the genetic loading for depressive symptoms (p = 0.001, standardized coefficient $\beta = -3.1$, SE = 0.9), which did not reach study-wide significance. Pathways relevant to neuronal and synaptic function, immune signaling, and glucocorticoid receptor/stress response showed enrichment among the suggestive GWAS signals.

Esketamine nasal spray has been shown to have rapidly-acting antidepressant effects in patients with treatment resistant depression (TRD) and in patients with major depressive disorder (MDD) at imminent risk for suicide¹⁻⁸. Predictors for conventional oral antidepressant treatment outcome including sociodemographic, symptom profiles, genetics, and clinical comorbidities were systematically reviewed by Perlman et al.⁹ In a small clinical study assessing the antidepressant efficacy of ketamine, a racemate consisting of two enantiomers, R- and S-ketamine, it was recently reported that body mass index (BMI) was associated with the remission rate, with greater BMI being associated with greater remission rate¹⁰. BMI and clinical comorbidities are influenced by both genetic and environmental factors. Genetic loading of such traits provides an objective way of measuring the relationship between these and other predictors with antidepressant treatment response.

In studies assessing individual genetic factors, the brain-derived neurotrophic factor (BDNF) Val66Met allele was reported to impair basal and ketamine-stimulated synaptogenesis in prefrontal cortex in vitro¹¹, and a significant genetic association between Val66Met and ketamine treatment outcome at 4 h post treatment was reported in a candidate gene study of small sample size¹². A more recent study further suggested that the BNDF Val66Met polymorphism may influence the improvement in suicide ideation following ketamine infusion in a sample of depressed participants from Taiwan¹³. In general, however, genetic associations with MDD disease susceptibility outcome reported in relatively small candidate gene studies have proven difficult to replicate in studies of larger samples¹⁴. Therefore, studies of genetic effects influencing antidepressant treatment outcome may particularly benefit from the use of genome-wide association analysis (GWAS) approaches in clinical trials of larger patient samples. Here, we assessed the genetic contributions to esketamine treatment response from patients with TRD who participated in two Phase III trials testing the efficacy and safety of esketamine, using both a genome-wide association analysis and a polygenic risk score (PRS) approach.

Results

Esketamine treatment response outcome was assessed at the 4 week study endpoint using one continuous variable (percent change from baseline in the Montgomery-Asberg Depression Rating Scale (MADRS) score) and two dichotomized variables (responder status, defined by a reduction of \geq 50% on the MADRS, and remission status,

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	Remitters (n = 255)	Non-remitters (n = 272)	p-value				
Mean (SD)							
Age*	50.6 (13.8)	53.4 (13.5)	0.424				
Baseline BMI*	28.1 (5.6)	28.3 (5.8)	0.742				
Baseline MADRS score*	29.7 (4.7)	33.0 (4.7)	6.36E-13				
N (%)							
Gender, female	153 (60.0)	175 (64.3)	0.349				
Study			7.34E-05				
TRANSFORM-3	10 (3.9)	39 (14.3)					
SUSTAIN-2	245 (96.1)	233 (85.7)					
Concomitant antidepressant medications		0.782					
DULOXETINE	90 (35.3)	87 (32.0)					
ESCITALOPRAM	78 (30.6)	80 (29.4)					
SERTRALINE	42 (16.5)	52 (19.1)					
VENLAFAXINE XR	45 (17.6)	52 (19.1)					
None		1 (0.4)					

 Table 1. Characteristics of study participants comparing remitters from non-remitters. *p-value reported is based on type III test statistics controlling for study.

rsID	Chr	pos	A1	A2	FRQ	INFO	Beta/OR	SE	р	Func.refGene	Gene.refGene	GeneDetail.refGene
Percentage change of MADRS from baseline												
rs11465988	12	66641813	С	Т	0.9898	0.51	-51.6	9.2	3.57E-08	Exonic	IRAK3	
rs17767394	12	66636086	С	A	0.9843	0.78	-32.7	6	8.68E-08	Intronic	IRAK3	
rs4739050	8	64034747	G	A	0.3376	1.02	7.5	1.4	6.06E-08	Intergenic	TTPA;YTHDF3-AS1	dist = 36135; dist = 45537
rs151184257	4	105714757	A	G	0.9888	0.61	-40.5	7.9	4.51E-07	Intergenic	CXXC4-AS1;TET2	dist = 96008; dist = 352275
rs115141868	2	70816605	A	С	0.9898	0.48	-46.8	9.3	7.65E-07	Intergenic	TGFA;ADD2	dist = 35458; dist = 72611
Response status												
rs10957273	8	6.4E+07	Т	C	0.3028	1	0.3	0.2	8.07E-07	Intergenic	TTPA;YTHDF3-AS1	dist = 30479; dist = 51193

Table 2. Variants with association p-value less than 1×10^{-6} in GWAS. Note that beta coefficient is reported for percentage change of MADRS score from baseline and OR is reported for responder status.

defined by achieving a final MADRS score of < 12). The demographic and clinical characteristics of study participants are summarized in Table 1 and Supplemental Table 1. Participants of the randomized TRANSFORM-3 study were recruited from an elderly population and had a lower remission rate than participants of the openlabelled SUSTAIN-2 study. Gender and concomitant medication proportions were comparable between remitters and non-remitters. After controlling for study, the baseline demographic characteristics (age and baseline BMI) were comparable between remitters and non-remitters. As expected from the clinical literature, remitters had lower baseline depression symptom severity score than non-remitters.

The genome-wide association analysis revealed one genome-wide significant association between an exonic synonymous variant (rs11465988, $p = 3.57 \times 10^{-8}$) in the interleukin 1 receptor associated kinase 3 (*IRAK3*) gene and the percent change in MADRS score (Table 2, Fig. 1 for Manhattan plot, Fig. 2A for regional plot and Supplemental Fig. 1A for QQ plot, Genomic Control lambda (λ) = 0.986). SNPs (e.g. rs115989442, rs144324167, rs79138866, rs116371327, rs150373274, and rs144520864) in linkage disequilibrium (r²=0.64) with rs11465988 are part of the regions engaging in intra-chromosomal loop (Fig. 2B, Supplemental Table 2) and could potentially be regulatory elements. rs144520864 is in fact located in a region with an annotated enhancer. An additional regional plot using rs17767394 as index SNP is also shown as Supplemental Fig. 2.

The other two GWAS for responder and remission status, respectively, did not yield any genome-wide significant finding (Supplemental Figs. 3A,B for Manhattan plots; Supplemental Figs. 1B,C for QQ plots, $\lambda = 1.028$ and 0.997, respectively). Nevertheless, a suggestive signal that merits comment was identified in chromosome 8 (rs4739050, nominal $p = 6.06 \times 10^{-8}$, $\beta = 7.5$, SE = 1.4 for percentage change in MADRS score; rs10957273, nominal $p = 8.07 \times 10^{-7}$, OR = 0.3, SE = 0.2 for responder status) from both the continuous endpoint GWAS and the responder status GWAS. Rs4739050 is an expression quantitative trait locus (eQTL) for gamma-glutamyl hydrolase (*GGH*) based on eQTLGen ($p_{eQTL} = 4.24 \times 10^{-9}$). A full list of suggestive associations with p-values less than 1×10^{-4} is provided in Supplemental Table 3.

Gene-level association analysis revealed one significant gene NME/NM23 family member 7 (*NME7*, $p = 1.73 \times 10^{-6}$, Supplemental Fig. 4A) for the percentage change in MADRS score. In the percent change in MADRS score GWAS, a pathway enrichment analysis revealed suggestive enrichments of genes involved in the negative regulation of glucocorticoid metabolic process (nominal $p = 3.53 \times 10^{-5}$) and neuronal action potential





(nominal p = 0.0001). Pathway enrichment analysis also revealed suggestive (p-values listed are nominal) enrichments of genes involved in synaptic vesicle clustering ($p = 4.33 \times 10^{-5}$), negative regulation of glucocorticoid metabolic process ($p = 5.48 \times 10^{-5}$), regulation of synaptic vesicle clustering ($p = 6.13 \times 10^{-5}$), anterior posterior axon guidance (p = 0.0002), and netrin mediated repulsion signals (p = 0.0002) in the responder status GWAS, and in the negative regulation of extrinsic apoptotic signaling pathway ($p = 4.04 \times 10^{-5}$), NF- κ B canonical pathway ($p = 5.90 \times 10^{-5}$), stress pathway (p = 0.0002), and TNFR1 induced proapoptotic signaling (p = 0.0003) in the remission status GWAS (Supplemental Table 4). We did not identify an association between the change in MADRS score and the *BDNF* Val66Met polymorphism in the current study (p > 0.05).

After applying corrections for multiple testing, none of the associations between esketamine's antidepressant efficacy and the PRS genetic loading for psychiatric conditions or symptom profiles was significant at the study-wide level (which required p < 0.0004 for significance). In Table 3 and Supplemental Fig. 5 we list suggestive associations observed in these analyses, however, along with their nominal p-values. Thus we observed suggestive (i.e., p-values listed are nominal) negative correlations between the depressive symptom PRS¹⁵ (p = 0.001, Table 3 and Supplemental Fig. 5) and the esketamine treatment response outcome as measured by percentage change from baseline in the MADRS score at the end of four week treatment period (Table 3). In addition, the depressive symptom PRS (p = 0.004) displayed suggestive positive correlations with esketamine responder status. Lastly, depressive symptoms PRS (p = 0.002) and insomnia¹⁶ PRS (p = 0.003) exhibited suggestive positive correlations with esketamine remission status.

Discussion

In this investigation of genetic associations with the antidepressant outcome to esketamine treatment, two findings remained significant after applying corrections for multiple testing. From the GWAS, a genome-wide significant association was identified with the percent change in MADRS score in an exonic SNP in *IRAK3*. *IRAK3* encodes a member of the interleukin-1 receptor-associated kinase protein family that is primarily expressed in monocytes and macrophages, where it functions as a negative regulator of Toll-like receptor signaling. In addition, the gene-level association analysis revealed one significant gene *NME7* for the percentage change in MADRS score. NME7 is a γ -tubulin ring complex component that regulates the microtubule-nucleating activity of this complex¹⁷.

A suggestive signal observed in both the continuous endpoint GWAS and the responder status GWAS was an expression quantitative trait loci (eQTL) for *GGH*, an enzyme that regulates intracellular folate concentrations. Folate deficiency has been linked to oxidative stress¹⁸. Meta-analysis showed that individuals with depression had lower folate levels than those without depression¹⁹. Folic acid administration was also shown to ameliorate depression-like behavior in rats subjected to chronic unpredictable mild stress, a putative rodent depression model²⁰.

The previously reported association between the Val66Met *BDNF* variant and antidepressant response to IV ketamine was not replicated in the current study. A significant methodological difference between studies, however, was that in the previous trial that reported this association the antidepressant response was assessed 4 h post ketamine infusion¹², whereas in the current study, the antidepressant outcome was assessed after 4 weeks of repeated esketamine nasal spray administration. While the importance of this timing difference in detecting an association with the Val66Met *BDNF* variant remains unclear, it is noteworthy that hypotheses generated from relatively small candidate gene studies in MDD have typically proven difficult to replicate in larger samples¹⁴, emphasizing the importance of studying larger sample sizes to detect reliable genetic signals. Nevertheless, the findings from the current study also warrant replication in larger sample sizes given the relatively modest number of participants included.



Figure 2. Genome-wide significant locus *IRAK3*. (**A**) Regional association plot; (**B**) circos plot. For the regional association plot generated via LocusZoom⁵² v1.4 (https://locuszoom.sph.umich.edu/), SNPs in genomic risk loci are color-coded as a function of their r² to the index SNP rs11465988 in the locus, while SNPs with missing LD information are shown in grey. For the circos plot generated via FUMA⁵¹ v1.3.5e (https://fuma.ctglab.nl/), the outer most layer is Manhattan plot and the middle layer highlights genomic risk loci (as defined by FUMA⁵¹ using minimum P-value of lead SNPs of 1×10^{-5} and default values for other parameters) in blue, while the inner most layer highlights eQTLs and/or chromatin interactions. Only SNPs with p < 0.05 are displayed in the outer ring. SNPs in genomic risk loci are color-coded as a function of their maximum r² to the one of the independent significant SNPs in the locus. The rsID of the top SNPs in each risk locus are displayed in the most outer layer. For the inner most layer, if the gene is mapped only by chromatin interactions or only by eQTLs, it is colored orange or green, respectively. It is colored red when the gene is mapped by both.

It has been shown previously that apoptotic biochemical cascades can exert local actions on the functions and structural dynamics of growth cones and synapses²¹. In this context, it is of interest that several apoptotic signaling pathways were identified as suggestive enriched gene sets. In preclinical models ketamine enhances structural plasticity in mouse mesencephalic and human iPSC-derived dopaminergic neurons via AMPARdriven BDNF and rapamycin kinase (mTOR) signaling²². In a rat traumatic brain injury (TBI) model, posttraumatic administration of a sub-anesthetic dose of ketamine exerts neuroprotection via attenuating inflammation and autophagy²³. There have been conflicting reports as to whether ketamine induces apoptosis, which might reflect a dependence on dose and developmental period. It was reported that ketamine induced apoptosis in human uroepithelial SV-HUC-1 cells²⁴ and in the neonatal rat brain²⁵, while a study in chronic unpredictable stress model of depression suggested an anti-apoptotic and antidepressant effects of ketamine²⁶. In the exposure range encompassing concentrations at which esketamine nasal spray has been tested for antidepressant effects in humans, no evidence of neuronal toxicity was identified in experimental animals²⁷. Notably, in a putative rodent depression model involving chronic mild stress that produces dendritic atrophy in the medial prefrontal cortex, a single ketamine administration restored synaptic density and function toward normative levels²⁸. Such changes in synaptic plasticity are hypothesized to underlie the relatively long-lasting antidepressant effects of ketamine and esketamine following single or pulsed intermittent doses²⁹, and the pathophysiology of MDD is associated with regional atrophy in the medial prefrontal cortex and other anatomically related structures³⁰. In clinical studies the antidepressant response to ketamine has been predicted by peripheral blood evidence of low-grade inflammation at baseline or by the enhancement of stimulus-evoked somatosensory cortical responses (a putative in vivo measure of long term potentiation effects mediated via changes in synaptic plasticity) at 4 h post-administration^{31,32}. Pathways relevant to neuronal and synaptic function, immune signaling, and glucocorticoid receptor/stress response showed enrichment among the GWAS suggestive signals. These findings are consistent with the hypotheses that inflammation and synaptic plasticity play a role in differentiating esketamine responders from non-responders.

This study suggests that PRS for psychopathology/symptom profiles may influence the antidepressant treatment outcome for esketamine. Although the PRS for depressive symptoms¹⁵ did not reach study-wide significance, the suggestive associations were consistent across multiple p-value thresholds ($p_{\rm T}$) used to construct PRS and across three esketamine efficacy endpoints. In addition, the condition of "depression"33 PRS (PGC2_ MDD+UKB in Table 3) constructed by the summary statistics from the GWAS meta-analysis by Howard et al. (2019) (without the 23andMe cohort³⁴) showed suggestive associations that did not reach significance for the remission endpoint. The GWAS meta-analyses for depressive symptoms¹⁵ and that for the condition, "depression"³³ differ in two respects. First, while both studies included PGC phase 1 samples³⁵, the GERA³⁶ samples (7,231 cases and 49,316 controls), and the UK Biobank (UKB) samples, the "depression" GWAS meta-analysis³³ additionally included samples from other cohorts, e.g. iPSYCH, deCODE, GenScot, and the incremental core samples from PGC MDD Working Group phase 2 analysis³⁶. Second, the phenotypic definition differed in that "depressive symptoms" in the UKB cohort from the Okbay et al. (2016) study employed a continuous phenotype measure by combining responses to two mental health questionnaire (MHQ) questions deployed to UKB participants, which asked about the frequency in the past two weeks with which the respondent experienced feelings of lack of enthusiasm/interest and depression/hopelessness, whereas the Howard et al. (2019) study used a "broad depression" phenotype³⁷, e.g. self-reported past help-seeking for problems with "nerves, anxiety, tension or depression.

Finally, genetic loading of BMI was not associated with esketamine remission status, in contrast to the previous report of ketamine¹⁰. In a correlation analysis of the SUSTAIN-2 clinical data, baseline BMI was also not associated with remission status, either in the entire sample irrespective of race (p = 0.365, n = 667) or in the subsample with European ancestry (p = 0.742, Supplemental Table 1).

Methods

PsychArray genotyping data were generated using blood DNA samples collected from the SUSTAIN-2⁸ (NCT02497287, n = 598) and TRANSFORM-3² (NCT02422186, n = 95; only participants with age of onset less than 55 years were included) phase III pivotal clinical studies. All subjects genotyped were of European ancestry. The clinical studies were carried out in accordance with the ethical principles outlined in the Declaration of Helsinki, Good Clinical Practices guidelines, and applicable regulatory requirements. The study protocols were approved by the local, regional, or central Institutional Review Board (IRB) or Independent Ethics Committee (IEC) overseeing the respective clinical sites: Sterling Institutional Review Board, IRB-UConn Health, Human Research Protection Program (US); Comité de Etica e Investigación del Sanatorio Profesor, Comité de ética en Investigación de Winsett Rethman S.A. de C.V., Comité de Etica en Investigación del Hospital La Mision SA de CV (Mexico); Comité de Etica en Investigación (CEI-INAPSI), CEI Fundación Rusculleda, Comité de Etica en Investigacion Burzaco, Comité de Ética IPEM, Comité Institucional de Ética en Investigación en Salud CIEIS Hospital Italiano (Aregentina); Comite de Etica em Pesquisa da UNIFESP/EPM, Comite de Etica em Pesquisa do Complexo Hospitar HUOC/PROCAPE, Comite de Etica em Pesquisas do Hospital Pro-Cardiaco Rua Voluntarios da Patria (Brasil); Western Institutional Review Board, Oxford Health NHS Foundation Trust, Derbyshire Healthcare NHS Foundation Trust, South London and Maudsley NHS Foundation Trust, Northamptonshire Healthcare NHS Foundation Trust, Ashgate Medical Practice Ethics Committee, Oxfordshire Research Ethics Committee A (UK); Alfred Health Human Ethics Committee, Bellberry Limited (Australia); Regionala Etikprövningsnämnden i Lund, Komisja Bioetyczna przy Kujawsko-Pomorskiej OIL (Sweden); CPP ile de France VIII (France); Lithuanian Bioethics Committee (Lithuania); Comitato Etico per la sperimentazione clinica della Provincia di Vicenza (CESC-VI) (Italy); Naisten lasten ja psykiatrian eettinen toimikunta (Finland); Ethics Committee for Clinical Trials, AZ St.-Jan Brugge (Belgium); der Stadt Wien gemäß KAG, Ethik-Kommission der

Threshold	r ² _{PRS}	r ² _{Full}	r ² _{Null}	Standardized coefficient	Standard error	р	Number of SNP	Base GWAS	References	
Percentage change of MADRS from baseline										
0.05	0.017351	0.152098	0.134747	- 3.06	0.94	1.20E-03	13,443	Depressive symptoms	Okbay et al., 2016	
0.001	0.0111608	0.145908	0.134747	-2.50	0.96	9.54E-03	1,000	ADHD	Demontis et al., 2019	
0.001	0.00761273	0.14236	0.134747	2.02	0.94	3.25E-02	245	Anxiety	Otowa et al., 2016	
0.001	0.00699865	0.141746	0.134747	2.04	0.99	4.03E-02	2,742	PGC2_SCZ	Ripke et al., 2014	
0.3	0.00638174	0.141129	0.134747	-1.95	0.99	5.03E-02	44,314	Insomnia	Hammerschlag et al., 2017	
0.001	0.00521661	0.139964	0.134747	1.82	1.03	7.69E-02	1,362	PGC2_BIP	Stahl et al., 2019	
0.001	0.00354911	0.138297	0.134747	-1.42	0.97	1.45E-01	1,917	PGC2_MDD+UKB	Howard et al., 2019	
0.5	0.00295958	0.137707	0.134747	-3.77	2.83	1.83E-01	60,258	SA_in_MDD_BIP_ SCZ	Mullins et al., 2019	
0.001	0.00229691	0.137044	0.134747	-1.36	1.16	2.41E-01	471	SA_in_MDD	Mullins et al., 2019	
0.001	0.00101084	0.135758	0.134747	0.76	0.98	4.37E-01	5,295	EA	Lee et al., 2018	
0.05	0.000785382	0.135533	0.134747	-0.72	1.06	4.93E-01	12,344	ASD	Grove et al., 2019	
0.05	0.000728554	0.135476	0.134747	0.65	0.98	5.09E-01	10,729	SWB	Okbay et al., 2016	
0.05	0.000634123	0.135382	0.134747	-0.58	0.95	5.38E-01	18,460	СР	Lee et al., 2018	
0.05	0.0006288	0.135376	0.134747	-0.59	0.97	5.40E-01	13,743	Neuroticism	Okbay et al., 2016	
0.001	0.00050388	0.135251	0.134747	-0.53	0.96	5.83E-01	6,764	BMI	Yengo et al., 2018	
Response stat	tus									
0.05	0.0218424	0.34341	0.321568	0.43	0.15	4.39E-03	13,443	Depressive symptoms	Okbay et al., 2016	
1	0.018916	0.340484	0.321568	0.41	0.15	7.83E-03	77,733	ADHD	Demontis et al., 2019	
0.4	0.0182367	0.339804	0.321568	-0.64	0.25	9.57E-03	56,270	PGC2_SCZ	Ripke et al., 2014	
0.001	0.0103815	0.331949	0.321568	-0.28	0.14	5.02E-02	245	Anxiety	Otowa et al., 2016	
0.5	0.00654157	0.328109	0.321568	0.69	0.44	1.16E-01	60,258	SA_in_MDD_BIP_ SCZ	Mullins et al., 2019	
0.2	0.0050513	0.326619	0.321568	0.20	0.15	1.67E-01	34,449	Insomnia	Hammerschlag et al., 2017	
0.001	0.00454651	0.326114	0.321568	-0.19	0.14	1.89E-01	5,295	EA	Lee et al., 2018	
0.05	0.0041483	0.325716	0.321568	-0.19	0.15	2.10E-01	10,729	SWB	Okbay et al., 2016	
0.001	0.00366392	0.325232	0.321568	-0.16	0.14	2.39E-01	733	ASD	Grove et al., 2019	
0.4	0.00251489	0.324083	0.321568	0.34	0.35	3.30E-01	52,541	SA_in_MDD	Mullins et al., 2019	
0.1	0.00228732	0.323855	0.321568	-0.15	0.16	3.53E-01	24,185	PGC2_MDD+UKB	Howard et al., 2019	
0.001	0.00184734	0.323415	0.321568	0.12	0.14	4.02E-01	6,764	BMI	Yengo et al., 2018	
0.05	0.000794085	0.322362	0.321568	-0.13	0.24	5.83E-01	15,002	PGC2_BIP	Stahl et al., 2019	
0.3	0.000469917	0.322038	0.321568	-0.06	0.15	6.73E-01	47,523	СР	Lee et al., 2018	
0.001	0.000213117	0.321781	0.321568	-0.04	0.14	7.76E-01	1,108	Neuroticism	Okbay et al., 2016	
Remission sta	atus									
0.05	0.020447	0.218933	0.198486	0.30	0.10	2.29E-03	13,443	Depressive symptoms	Okbay et al., 2016	
1	0.019018	0.217504	0.198486	0.31	0.10	3.25E-03	79,083	Insomnia	Hammerschlag et al., 2017	
0.001	0.0131928	0.211679	0.198486	-0.26	0.11	1.41E-02	1,362	PGC2_BIP	Stahl et al., 2019	
0.001	0.0127337	0.211219	0.198486	-0.25	0.10	1.61E-02	2,742	PGC2_SCZ	Ripke et al., 2014	
0.001	0.0103469	0.208833	0.198486	0.22	0.10	2.93E-02	1,917	PGC2_MDD+UKB	Howard et al., 2019	
0.05	0.00709676	0.205582	0.198486	-0.18	0.10	7.04E-02	6,160	Anxiety	Otowa et al., 2016	
0.1	0.00668848	0.205174	0.198486	-0.18	0.10	7.90E-02	20,193	ASD	Grove et al., 2019	
0.5	0.00611421	0.2046	0.198486	0.48	0.29	9.31E-02	60,258	SA_in_MDD_BIP_ SCZ	Mullins et al., 2019	
0.001	0.00482744	0.203313	0.198486	0.15	0.10	1.35E-01	1,000	ADHD	Demontis et al.,2019	
0.05	0.00292355	0.201409	0.198486	0.11	0.10	2.45E-01	13,743	Neuroticism	Okbay et al., 2016	
0.5	0.00220535	0.200691	0.198486	0.10	0.10	3.12E-01	61,486	СР	Lee et al., 2018	
0.001	0.00119781	0.199684	0.198486	-0.07	0.10	4.56E-01	5,295	EA	Lee et al., 2018	
0.001	0.00113909	0.199625	0.198486	0.09	0.12	4.67E-01	471	SA_in_MDD	Mullins et al., 2019	
1	0.00091008	0.199396	0.198486	0.07	0.10	5.16E-01	67,436	SWB	Okbay et al., 2016	
0.3	0.000424952	0.198911	0.198486	-0.04	0.10	6.57E-01	40,520	BMI	Yengo et al.,2018	

Table 3. Polygenic Risk Score association with esketamine treatment outcome. *MDD* major depressive disorder, *BIP* bipolar disorder, *SCZ* schizophrenia, *ADHD* attention deficit/hyperactivity disorder, *ASD* autism, *SWB* subjective well-being, *CP* cognitive performance, *EA* education attainment, *UKB* UK Biobank, *PGC* Psychiatric Genomic Consortium. *For the reported standardized coefficient in this table, only PRS was scaled while the dependent variable was kept in its original scale.

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Landesärztekammer Brandenburg, Ethik-Kommission der Ärztekammer Westfalen-Lippe und der Medizinischen Fakultät der Westfälischen Wilhelms-Universität Münster (Germany); Uludag University Medical Faculty Clinical Research Ethics Committee, Dicle University Medical Faculty Clinical Research Ethical Committee (Turkey). All participants provided written informed consent before enrollment.

The analysis was composed of TRD patients who received esketamine combined with a newly initiated oral antidepressant treatment (SSRI or SNRI) either in an open labelled (for SUSTAIN-2) or in a randomized (for TRANSFORM-3) fashion. A total of 527 samples were included in the final analysis. Treatment response endpoints were defined as follow: (1) a quantitative trait using percentage of change of MADRS score at the end of study compared to baseline; (2) response defined as $\geq 50\%$ improvement from baseline in the MADRS Score; (3) remission defined as MADRS ≤ 12 at study endpoint. Additional details of these clinical studies are provided in the Supplemental Text or could also be found in https://clinicaltrials.gov/.

Genotypes were imputed based on the 1000 Genome Project³⁸ Phase I reference panel. A SNP-wise genomewide association analysis was performed using PLINK^{39,40}. In addition, a gene-wise genome-wide association followed by pathway enrichment analysis was performed using MAGMA⁴¹. In all analyses the models corrected for gender, study ID, baseline symptom severity, and 5 principal components representing the population substructure. Detailed methods are described in the Supplemental Text.

Polygenic risk scores (PRS) were constructed based on well-powered genome-wide association studies (GWAS) of 15 PRS phenotypes, of which six were constructed for psychiatric conditions (depression³³, bipolar disorder⁴², schizophrenia⁴³, autism⁴⁴, ADHD⁴⁵, anxiety⁴⁶) and seven psychiatric characteristics (history of suicide attempt⁴⁷ among depressive subjects or among schizophrenia, bipolar, and depressive subjects), depressive symptoms¹⁵, subjective well-being¹⁵, neuroticism¹⁵, insomnia¹⁶, education attainment⁴⁸, and cognitive performance⁴⁸), and BMI⁴⁹. To correct the resulting p-values for performing comparisons in multiple PRS phenotypes and at 8 p-value thresholds assessed (i.e., 5e–08, 0.001, 0.05, 0.1,...0.5, 1), the association p-value < 0.05/ (15×8) ~ 0.0004 (for 15 phenotypes and 8 P_T bins) between PRS and any esketamine treatment response outcome was considered to be study-wide significant. To balance Type 2 error, nonsignificant associations that reached nominal *p* < 0.005 were considered "suggestive". The PRS analysis was performed using PRSice-2⁵⁰. All p-values reported in this study were uncorrected p-values.

Received: 27 March 2020; Accepted: 1 July 2020 Published online: 28 July 2020

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Acknowledgements

We are grateful to the study volunteers for participating in the research studies and to the clinicians and support staff for enabling patient recruitment and blood sample collection. Informed consent was obtained from all participants. We thank the staff at Covance and the Neuroscience Biomarkers team at Janssen for managing the EDTA blood samples, the staff at HD Bio for DNA extraction and plating, and the staff at Illumina for genotyping Janssen DNA samples. The work was funded by Janssen Research & Development, LLC.

Author contributions

Q.S.L. conceived and initiated the project, performed the analysis, and wrote the first draft of the manuscript. All authors contributed to the data analysis review, discussions, and contributed to the final manuscript.

Competing interests

All authors are employees of Janssen Research & Development, LLC and equity shareholders in Johnson & Johnson, the parent company of the Janssen companies. Drs. Li, Wajs, Ochs-Ross, Singh, and Drevets declare that, except for income received from their primary employer, no financial support or compensation has been received from any individual or corporate entity over the past three years for research or professional service, and there are no other personal financial holdings that could be perceived as constituting a potential conflict of interest.

Additional information

Supplementary information is available for this paper at https://doi.org/10.1038/s41598-020-69291-6.

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