

# Safety of human embryonic stem cells in patients with terminal/ incurable conditions- a retrospective analysis

Geeta Shroff<sup>1</sup>, J.K Barthakur<sup>2</sup>

<sup>1</sup>Nutech Mediworld, New Delhi, India; <sup>2</sup>Retired Additional Secretary, Ministry of Home Affairs, Government of India

## KEY WORDS

Human Embryonic Stem Cells  
Transplantation  
Stem cell therapy

## Corresponding Author:

Geeta Shroff  
Tel : +91 11 26565548  
E-mail : geetashroff@hotmail.com

## ABSTRACT

**Background:** Human embryonic stem cells (hESCs) are pluripotent cells that have the potential to self-renew and differentiate into all types of human cells.

**Purpose:** The present study was aimed at establishing the safety of hESC therapy in patients with terminal/ incurable conditions.

**Methods:** This was a single cohort study conducted at Nutech Mediworld, New Delhi. The patients suffering from various degenerative diseases were included in the study from year 2002 to 2004. hESCs (0.25 mL) were injected under skin in the abdominal wall. The safety of hESC therapy was evaluated by assessing the AEs experienced by patients during the study. Any disabling symptom/ sign, teratoma or antigen-antibody reaction that a patient suffered post transplantation of hESCs was considered as an AE.

**Results:** A total of four, six and twenty three patients received hESC therapy in the year 2002, 2003 and 2004 respectively. Pain and fever were the most common AEs observed during the study. Other AEs included headache, mild pain in the abdomen, swelling of legs (edema), urinary tract infection (UTI), rash/ erythema, pain at the lower back and limbs and body ache. All the AEs reported were mild in nature and resolved within one or two days with symptomatic medication and rest. No serious AEs were reported. The improvement in specific parameters of the patients was observed after the therapy.

**Conclusion:** hESCs used in the present study are safe for use in humans afflicted with incurable/terminal conditions. Future, prospective controlled studies to substantiate the present study are ongoing.

doi : 10.5214/ans.0972.7531.220303



## Introduction

Human embryonic stem cells (hESCs) are self-renewing cells with a potential to differentiate into all types of human cells.<sup>1</sup> These cells have the potential for cell replacement and regeneration therapies for human diseases. hESCs were derived and characterized as early as 1982 from fresh or frozen cleavage stage donated human embryos produced by *in vitro* fertilization (IVF).<sup>2</sup> The viable cell lines were obtained from the inner cell mass or blastocyst. hESCs have also been derived and established from single blastomeres of the 4 or 8 celled embryo and 16 celled morula.<sup>3-7</sup> Since then, a plethora of research has been done using hESCs for various diseases like diabetes, liver disorders, auto-immune disorders, immune disorders, Parkinson's disease, Alzheimer's disease, age related macular degeneration and spinal cord injury.<sup>8-14</sup>

Despite huge potential in curing chronic and terminal conditions, hESCs have not been used extensively in humans. This is largely due to the ethical consideration in procuring the hESC lines and also lack of knowledge for the use of hESCs. Further, hESC cell lines have shown chromosomal and genomic instability, with acquisition of loss of heterozygosity or copy-number variation in cancer-related genes.<sup>15,16</sup> hESCs have also been associated with teratoma formation and fear of being immunologically rejected.<sup>17</sup> These challenges have hindered the use of hESCs to their full potential.

A Phase 1 human clinical trial using hESCs was approved by FDA in 2009 popularly referred to as the Geron trial. Though the initial results of the trial were promising, it was left mid-way due to financial constraints.<sup>18</sup> Further, there have been safety concerns and challenges in the use of hESCs. Most of the hESCs being used have been exposed to xeno-products during isolation and propagation. As a result, these could carry a risk of xenogenetic pathogen cross transfer and other unknown substances capable of eliciting a detrimental immune response in transplanted hosts. The cells used in Geron trial also contained animal components such as B27 supplement or Matrigel.<sup>18</sup> Recently, Asterias Biotechnology Inc. of Menlo Park has bought the rights of Geron to conduct clinical trial with hESC in humans; that has been approved by FDA.<sup>19</sup> Advanced Cell technology (ACT), Inc. is also focusing on developing hESC based therapies for various disorders and has got promising initial results in patients with macular degeneration.<sup>20</sup>

We used an in-house developed patented technology to culture and maintain hESCs in our GMP, GLP and GTP certified laboratory. The hESCs were obtained from a one-time harvest made at the pre-blastocyst stage. The cell line thus developed is created from a single expendable fertilized ovum 24-48 hours after fertilization when the conceptus is assumed to have reached 4-16 celled stage. Further, we have not used any animal product or exposed our cell lines to any animal product. We have developed a simplified cell culture system free of exogenous cells and supplements of animal origin for expansion of hESCs in a substantially undifferentiated state. In this article, we present the safety and efficacy data of our cell lines.

## Methods

### Study Characteristics

This was a single cohort study to establish safety and efficacy of hESCs in terminally ill patients carried out at Nutech Mediworld, New Delhi. The study included the patients enrolled in different cohorts in 2002 and 2004. The first patient was admitted on 31 March 2002. The patients were included in the study after an informed consent. The consent process involved a detailed discussion about the hESC therapy with the patient accompanied by a family member or caretaker. All the patients were informed that the treatment protocol being followed is being developed and is not yet finalized. The patients were also made aware of the adverse events (AEs) that might occur due to hESC therapy. All the information regarding the patients, a detailed report of the therapy and outcomes on patients and commencement of therapy was given to Government of India. We followed the guidelines of biomedical research (year 2000) on human participants in India.<sup>20</sup> During the entire procedure, an anesthetist was present and safety and sterile measures for hESC transplantation were followed. The transplantation was done at a medical centre registered under Delhi Government.

### Study Population

Patients suffering from various degenerative diseases including Parkinson's disease, spinal cord injuries, autosomal recessive disorders, motor sensory neuropathy with diplopia (vasculitis), mild diffuse cerebral atrophy, Huntington's chorea, liver metastasis, diabetic foot (amputation), diabetes mellitus, psoriasis, chronic renal failure secondary to lupus nephritis, systemic lupus erythematosus (SLE), Duchene's muscular dystrophy (DMD), cirrhosis, mental retardation with microcephaly, hypothalamic astrocytoma, post traumatic paraplegia, developmental delay, colitis, and acute cauda equine lesions received hESC therapy. Patients who were pregnant, lactating or confirmed to have received other forms of cell therapy within 12 months of the treatment were not included.

### Cell Culture and Differentiation

hESCs were obtained from a single, spare, expendable, pre implantation stage fertilized ovum taken during natural *in vitro* fertilization (IVF) process with due consent. These cells are cultured and maintained as per our proprietary in-house technology in a GMP, GLP and GTP certified laboratory. The technology has now been patented (United States Granted Patent No US 8592, 208, 52). The detailed cell culture and differentiation techniques have been elaborated elsewhere. The cell lines have been cultured and maintained in animal products free conditions making them suitable for clinical cell therapy. A detailed composition of hESCs used in this study their derivatives, methods of use, and methods of preparation are available at <http://patentscope.wipo.int/search/en/WO2007141657>.

A quality check was performed on the stored cell batches which included integrity, viability and microbial contamination. The cells were characterized and the transplanted cells were octamer-binding transcription factor 4 positive (OCT4 +ve); Stage-specific embryonic antigen 3 (SSEA3)+ve; NANOG +ve; SOX +ve;  $\beta$  actin +ve;  $\beta$ -human chorionic gonadotropin ( $\beta$ -HCG) +ve; alkaline phosphatase +ve; CD 34 +ve; Nestin +ve, GATA +ve; GAF +ve; NeuN +ve; and transfer gene (TRA) -ve. The characterization was done by fluorescence-activated cell sorting (FACS), polymerase chain reaction (PCR) and immuno-

fluorescence (Nikon Ellipse E200; BD Acuri, Biorad T 100 Thermal cycle) (Unpublished data, Paper under submission).

### Study Procedure

The cells were transplanted through simple injections of hESC suspension under skin in the abdominal wall. A single injection contained 0.25 mL of hESC suspension (1 mL contains approximately 4 million cells).

### Variables for Analysis

**Safety Evaluation:** The safety of hESC therapy was evaluated by assessing the AEs experienced by patients during the study. Any disabling symptom/sign that a patient suffered after the test dose was given was considered as an AE. The medical staff of Nutech Mediworld carefully examined the patients for any AEs keeping the ones related to hESCs in mind. These AEs included teratomas, antigen-antibody reactions and any other sign and symptom.

**Efficacy Evaluation:** The efficacy of hESC therapy was evaluated based on the improvement seen in the patients.

### Data Analysis

No formal sample size was calculated for this study. Each case was assessed at admission or soon after admission to determine the pre-therapy status of the case. The safety analyses were performed on safety population (patients who took at least one dose of hESC). The efficacy was assessed based on the improvement seen in the patients. No statistical assumptions were made.

## Results

### Study Patients

In the year 2002, 4 patients started hESC therapy. The first injection of hESCs was given to a patient with cortico basal degeneration on 31 Mar 2002. In year 2003, six patients received hESC therapy. The details of these patients are shown in Table 1. In the year 2004, 23 patients with different chronic or terminal conditions were administered hESC therapy. Of these 23 patients, 4 had SCI, 3 had Duchenne's muscular dystrophy, 2 had Huntington's Chorea, 1 had Parkinsonism, 1 had Psoriasis, 1 had cauda equina Syndrome, 1 had postastrocytoma brain damage with seizures and slow learning, 1 had systemic lupus erythematosus, 1 had developmental delay, 1 had dementia, 1 had Alzheimer's disease, 1 had alcoholic cirrhosis with portal hypertension, 1 had genetic disorder, 2 had post cerebral vascular accident (CVA), 1 had uncontrolled diabetes with right bundle branch block and 1 had multiple sclerosis.

### Safety Evaluation

No serious AEs were reported during the study period. All the AEs were mild in nature and resolved within one or two days with symptomatic medication and rest. Pain and fever were the most common AEs observed during the study. Headache, mild pain in the abdomen, swelling of legs (edema), urinary tract infection (UTI), rash/erythema, pain at the lower back and limbs and body ache were the other common AEs observed during the study. Table 2 lists the AEs observed during the study period in our patients.

### Efficacy Evaluation

The improvement in patients after the therapy is tabulated in Table 1. Since this was not an efficacy study, we did not analyze

Table 1: Characteristics of Patients, hESC Therapy Schedule and Condition Before and After the Treatment (Year 2000 and 2003)

Code	Age (yr)/ Gender	Diagnosis	Presenting Condition	hESC Therapy Schedule	Condition after Therapy
<b>Year 2002</b>					
80001	56/M	Cortico Basal degeneration	Rigidity Stammering and slurring Tightly clenched fists Inability to blink Inability to swallow Inability to walk	2 injections 31 Mar 2002 1 Dec 2002	Rigidity decreased Speech improved No clenched fists Could swallow and chew food Could walk a few steps
80002	64/M	Parkinsonism	Pain in lower back and lower limbs Pain before micturition Tremors in the upper limbs Inability to speak properly Inability to walk properly	3 injections 26 Apr 2002 8 Sep 2002 26 Oct 2002	Lesser pain in lower back and lower limbs Micturition normal No tremors Speech improved Ability to walk improved Medication decreased
80003	53/M	Becker's Muscular Dystrophy	Slurring of speech Inability to walk Inability to lift arms Difficulty in breathing Weak hand grip	2 injections 29 Sep 2002 17 Nov 2002	Speech improved Able to walk without support Weakness in limbs decreased Breathing pattern improved Hand grip improved
80004	37/F	Post traumatic encephalomalacia and gliosis with quadriplegia (Right>Left)	Poor gait Inability to walk without support Scoliosis Poor ability to perform everyday activities Poor speech Poor finger movements Rigid neck Impaired cognition	Single injection 16 Nov 2002	Better gait Able to walk without support Scoliosis improved More independent in performing everyday activities Slight improvement in finger movements Better neck movements Improved cognition
<b>Year 2003</b>					
80005	65/M	Diabetes Mellitus with renal and cardiac involvement	Chest pain Breathlessness Pain in the right shoulder Serum creatinine-2.4 mg LVEF-48%	Single injection 31 Jan 2003	Decreased chest pain Less pain in right shoulder Serum creatinine- 1.8 mg LVEF-57.3%
80006	72/F	Non-healing burn wound	Non-healing ulcer after a burn Chest pain Breathlessness	Single injection 12 Feb 2003	Skin covered the ulcer Decreased chest pain
80007	66/M	Type II Diabetes Mellitus with Hypertension and Parkinsonism	High blood sugar levels Abnormal kidney function Tremors	Single injection 5 May 2003	Lower blood sugar levels, insulin dosage reduced to half Normal kidney function Decreased tremors
80008	35/F	Primary ovarian failure	No ovulation despite medication	Single injection 11 Jun 2003	Delivered a normal healthy baby
80009	28/F	Becker's muscular dystrophy with cerebral atrophy	Drooling Low IQ-35	Single injection 15 Nov 2003	Mental condition improved IQ-100
80010	65/F	Atrophy with spino cerebellar ataxia	Catheterization for micturition Difficulty in swallowing Tremors and nodding of head Inability to walk Slurring of speech Poor balance Hypertension	Single injection 30 Dec 2003	No catheterization Able to swallow food Decreased tremors and nodding of head Able to walk with walker Speech improved Better balance

**Table 2: Adverse Events (AEs) Observed in Patients during the Study Period**

Sr. No.	Adverse Event (AE)	n (%)
<b>Year 2002 and 2003 (N = 10)</b>		
1	Pain (Legs, Abdomen, Chest)	5 (50)
2	Fever	4 (40)
2	Urinary Tract Infection	3 (30)
3	Headache	2 (20)
4	Loose motion	2 (20)
5	Constipation	2 (20)
7	Cough	2 (20)
8	Dyspnea	1 (10)
9	Breathlessness	1 (10)
<b>Year 2004 (N = 23)</b>		
1	Fever	4 (17.4)
2	Cough	3 (13)
2	Lack of sleep	3 (13)
3	Headache	2 (8.7)
5	Pain in the legs	2 (8.7)
6	Loose motions	2 (8.7)
7	Vomiting	2 (8.7)
8	Backache	2 (8.7)
9	Restlessness	1 (4.4)
10	Burning pain in the area of ulceration (in psoriatic cases)	1 (4.4)
11	Pain in the lower abdomen	1 (4.4)
12	Breathlessness	1 (4.4)
13	Constipation	1 (4.4)
14	Swelling	1 (4.4)
15	Knee pain	1 (4.4)
16	Loss of appetite	1 (4.4)

the efficacy data. However, we observed an improvement in specific parameters of all our patients during the study, Figure 1.

### Discussion

hESCs have an inexhaustible potential to differentiate into different cell types making it a promising treatment option for many debilitating conditions. The first embryonic stem cells (ESCs) were derived from mice (mESCs) in the year 1981.<sup>21</sup> Thomson *et al* isolated the first hESCs derived from human blastocysts about two decades later.<sup>2</sup> Since then, studies have been ongoing to prove and utilize their therapeutic prowess. However, ethical considerations and safety regarding the use of hESCs have limited their widespread use.

Adult stem cells are ethically preferable but are lineage restricted and have a limited capacity of self-renewal, which is the essence of stem cell therapy. Further, the sources of human adult stem cells are limited and their isolation is a challenge and can



**Fig. 1:** Images of Two Patients Before and After Receiving hESC Therapy.

be painful for the patient. Human induced pluripotent stem cells (iPSCs) derived from various somatic cells have generated a tremendous interest for their use in stem cell therapy and regenerative medicine. Though effective, iPSCs might cause genetic and epigenetic abnormalities that could take place during reprogramming or maintenance of iPSCs in subsequent cell culture.<sup>22-24</sup> The potential tumorigenicity and immunogenicity associated with iPSC-based cell therapy is of significant concern.<sup>25-27</sup> hESCs have an edge over adult stem cells and iPSCs as they display low immunogenicity and could be transplanted with minimal immunosuppression.<sup>28-29</sup>

Maintaining an undifferentiated stem cell state during large scale expansion (without spontaneous differentiation) is an uphill task that has hindered their widespread use. Another issue is direct or indirect exposure of cells to animal products while culturing that results in high risk of graft rejection and transfer of non-human pathogens to the recipient. Some researchers have been able to maintain and expand undifferentiated hESC on human feeder layers or feeder free matrices but the scale of expansion is low.<sup>30,31</sup> Further, majority of the methods adopted for culturing hESCs could result in genetically unstable and aberrant cell lines.<sup>32</sup>

Most of the hESCs used till date have been derived from inner cell mass (ICM) of blastocyst embryos before implantation.<sup>2,33,34</sup> hESC lines are obtained by enzymatic dispersion of the ICM



and culturing under particular conditions. This blastocyst is the 256-celled stage of human embryo that has developed from 2-celled, 4-celled, 8-celled stage and so on. The blastocyst continues to mature for an additional 24 hours and is ready to implant into the uterine wall. These preimplantation embryos are able to develop in synthetic culture media for several days and are highly adaptable.<sup>35</sup> Though pluripotency at this blastocyst stage has been studied extensively using various marker and characterization studies, there is a paucity of studies about cells at the initial stages of developments. In our study, we used hESCs that are generated in a culture from a one-time harvest made at the pre-blastocyst stage. Expendable, fertilized ovum was taken after a natural IVF cycle and informed consent was sought from the donor. The cell line thus developed is created from a single fertilized ovum 24–48 hr after fertilization when the conceptus is assumed to have reached the 4–16 cell stage. All the media used in the culture are free from animal contaminants and cells of animal origin. The composition of the present therapy is simple to prepare and cost effective. The ready to inject form is easily transportable, scalable and has a good shelf life. The evidence for the use of hESCs at our facility has been gathered over a number of years and was accepted as a written evidence to House of Lords, Regenerative Medicine, Science and Technology Committee report.<sup>36</sup>

Stem cells have a unique functioning when transplanted. Previous studies suggest that various factors like chemokines, cytokines, and other growth factors released from the site of injury attractant the transplanted stem cells. These cells then migrate to the damage site due to up regulation of selectins and integrins on their surface, a process called “homing”.<sup>37–40</sup> After homing at the injured site, the stem cells help in “rescue” and “replacement” of the injured cells. We assume that the hESC in our study also acted in a similar way and reached the site of injury after transplantation. Once there, a trigger of factors led to their differentiation into the cell type of injured area and helped in recovery/regeneration.

When we tested our cell line in patients with differing chronic and life threatening conditions, we got promising results. The studies from 2002 to 2004 were done to establish the safety of these cell lines and develop a protocol for using the therapy. During these years, we did not observe any serious AE associated with the use of our cell line. Fever and generalized pain were the most common AEs that we observed after hESC injections. All the AEs were mild in nature and were observed majorly not due to hESC transplant *per se*, but were a direct consequence of the patient’s illness and were part of the normal course of the disease. Though not the primary objective, efficacy of the hESC therapy was promising and we observed a continued benefit in our patients. Our cell line is a mixture of “neuronal” and “non-neuronal” cells. The non-neuronal cell lines include progenitor cells for hematopoietic stem cells progenitors, insulin producing stem cells, mesenchymal stem cell, epithelial stem cells, hepatocyte stem cell, and cardiac stem cells. The presence of these two directed cell lines makes them appropriate for usage in a wide variety of conditions. In this study, our patients suffered from different non-curable diseases like cortico-basal degeneration, SCI, cerebral palsy, corticovisual impairment, Parkinsonism etc. but the hESCs seemed to have beneficial effects for all of them. Further, all our patients had come to us after trying all the conventional treatments without any improvement.

Now, the protocol of using hESC therapy in patients with different chronic and life threatening conditions has been fully developed and we have used our therapy in over 1300 patients over last decade. Till date, we have not observed any serious AE in our patients. As largely concerned about the usage of hESCs, we have not observed any teratoma formation in our patients till date. We did not give steroids or immunosuppressant to our patients. None of the patient had an immune response. Our staff is trained to observe any antigenic/anaphylactic response in the patients.

In conclusion, our hESC cell line is safe for use in humans afflicted with incurable conditions. We did not observe any serious AE in our patients. No teratoma formation or immune response was observed. We also observed clinical benefits of these cell lines in all our patients. Future, prospective controlled studies to substantiate the present study are ongoing.

#### Authorship Contributions

**Geeta Shroff:** Drafted the manuscript, carried out the experiments, conceived of the study, and participated in its design and coordination and helped to draft the manuscript, **J.K Barthakur:** participated in the design of the study and performed the statistical analysis, **Geeta Shroff and J.K Barthakur:** Read and approved the final manuscript.

#### Acknowledgements

The authors acknowledge Staff of Nutech Mediworld and the patients of this study. The authors also acknowledge Knowledge Isotopes Pvt. Ltd (<http://www.knowledgeisotopes.com>) for writing support.

#### Competing Interests

The authors declare that they have no competing interests.

#### Disclaimer

This is a case/isolated study conducted by Nutech Mediworld to test the efficacy of ESC on humans.

This article complies with International Committee of Medical Journal editor’s uniform requirements for manuscript.

Conflict of Interests: None: Source of funding: None.

Received Date : 16 November 2014; Revised Date : 20 February 2015;

Accepted Date : 18 March 2015

#### References

1. Heins, N, Englund MC, Sjöblom C, et al. Derivation, characterization, and differentiation of human embryonic stem cells. *Stem Cells*. 2004; 22(3): 367–76.
2. Thomson JA, Joseph IE, Sander SS et al., Embryonic stem cell lines derived from human blastocysts. *Science*. 1998; 282(5391): 1145–7.
3. Chung Y, Irina K, Sandy B, et al. Human embryonic stem cell lines generated without embryo destruction. *Cell Stem Cell*. 2008; 2(2): 113–7.
4. Geens M, Mateizel I, Sermon K et al., Human embryonic stem cell lines derived from single blastomeres of two 4-cell stage embryos. *Hum Reprod*. 2009; 24(11): 2709–17.
5. Klimanskaya I, Young C, Sandy B et al. Human embryonic stem cell lines derived from single blastomeres. *Nature* 2006; 444(7118): 481–5.
6. Strelchenko N, Verlinsky O, Kukharenko V et al. Morula-derived human embryonic stem cells. *Reprod Biomed Online*. 2004; 9(6): 623–9.
7. Strelchenko N, Verlinsky Y. Embryonic stem cells from morula. *Methods Enzymol*. 2006; 418: 93–108.

8. Agarwal S, Holton KL, Lanza R. Efficient differentiation of functional hepatocytes from human embryonic stem cells. *Stem Cells*. 2008; 26(5): 1117–27.
9. Cai J, Zhao Y, Liu Y et al. Directed differentiation of human embryonic stem cells into functional hepatic cells. *Hepatology*. 2007; 45(5): 1229–39.
10. Cloutier F, Siegenthaler MM, Nistor G, et al. Transplantation of human embryonic stem cell-derived oligodendrocyte progenitors into rat spinal cord injuries does not cause harm. *Regen Med*. 2006; 1(4): 469–79.
11. Idelson M, Alper R, Obolensky A et al. Directed differentiation of human embryonic stem cells into functional retinal pigment epithelium cells. *Cell Stem Cell*. 2009. 5(4): 396–408.
12. Kroon E, Martinson LA, Kadoya K et al. Pancreatic endoderm derived from human embryonic stem cells generates glucose-responsive insulin-secreting cells in vivo. *Nat Biotechnol*. 2008. 26(4): 443–52.
13. Senju S, Hirata S, Motomura Y et al. Pluripotent stem cells as source of dendritic cells for immune therapy. *Int J Hematol*. 2010. 91(3): 392–400.
14. Wong SS, Bernstein HS. Cardiac regeneration using human embryonic stem cells: producing cells for future therapy. *Regen Med*. 2010; 5(5): 763–75.
15. Lefort N, Feyeux M, Bas C et al. Human embryonic stem cells reveal recurrent genomic instability at 20q11.21. *Nat Biotechnol*. 2008; 26(12): 1364–6.
16. Närvä E, Autio R, Rahkonen N et al. High-resolution DNA analysis of human embryonic stem cell lines reveals culture-induced copy number changes and loss of heterozygosity. *Nat Biotechnol*. 2010; 28(4): 371–7.
17. Bradley JA, Bolton EM, Pedersen RA. Stem cell medicine encounters the immune system. *Nat Rev Immunol*. 2002; 2(11): 859–71.
18. Lukovic D, Stojkovic M, Moreno-Manzano V et al. Perspectives and future directions of human pluripotent stem cell-based therapies: lessons from Geron's clinical trial for spinal cord injury. *Stem Cells Dev*. 2014. 23(1): 1–4.
19. R, L., Stem cell trial for spinal cord injuries cleared by FDA. Available from: <http://www.bizjournals.com/sanfrancisco/blog/biotech/2014/08/embryonic-stem-cells-asterias-geron-spinal-cord.html>; accessed on 17th September, 2014.
20. ICMR. Ethical guidelines for biomedical research on human participants. Available at: [http://icmr.nic.in/ethical\\_guidelines.pdf](http://icmr.nic.in/ethical_guidelines.pdf); assessed on 7 November, 2014.
21. Evans MJ, Kaufman MH. Establishment in culture of pluripotential cells from mouse embryos. *Nature*. 1981; 292(5819): 154–6.
22. Hussein SM, Batada NN, Vuoristo S et al., Copy number variation and selection during reprogramming to pluripotency. *Nature*. 2011; 471(7336): 58–62.
23. Lister R, Pelizzola M, Kida YS et al. Hotspots of aberrant epigenomic reprogramming in human induced pluripotent stem cells. *Nature*. 2011; 471(7336): 68–73.
24. Mummery C. Induced pluripotent stem cells—a cautionary note. *N Engl J Med*, 2011. 364(22): p. 2160–2.
25. Miura K, Okada Y, Aoi T et al. Variation in the safety of induced pluripotent stem cell lines. *Nat Biotechnol*. 2009; 27(8): 743–5.
26. Tsujia O, Miuraa K, Okada Y et al. Therapeutic potential of appropriately evaluated safe-induced pluripotent stem cells for spinal cord injury. *Proc Natl Acad Sci U S A*, 2010. 107(28): 12704–9.
27. Zhao T, Zhang ZN, Rong Z et al. Immunogenicity of induced pluripotent stem cells. *Nature*, 2011; 474(7350): 212–5.
28. Drukker M, Katz G, Urbach A et al. Characterization of the expression of MHC proteins in human embryonic stem cells. *Proc Natl Acad Sci U S A* 2002; 99(15): 9864–9.
29. Li L, Baroja ML, Majumdar A et al. Human embryonic stem cells possess immune-privileged properties. *Stem Cells*. 2004; 22(4): 448–56.
30. Li Y, Powell S, Brunette E et al., Expansion of human embryonic stem cells in defined serum-free medium devoid of animal-derived products. *Biotechnol Bioeng*. 2005; 91(6): 688–98.
31. Yoo SJ, Yoon BS, Kim JM et al. Efficient culture system for human embryonic stem cells using autologous human embryonic stem cell-derived feeder cells. *Exp Mol Med*. 2005; 37(5): 399–407.
32. Catalina P, Montes R, Ligeró G et al. Human ESCs predisposition to karyotypic instability: Is a matter of culture adaptation or differential vulnerability among hESC lines due to inherent properties? *Mol Cancer* .2008; 7: 76.
33. Reubinoff BE, Pera MF, Fong CY et al. Embryonic stem cell lines from human blastocysts: somatic differentiation in vitro. *Nat Biotechnol*. 2000; 18(4): 399–404.
34. Cowan CA, Klimanskaya I, McMahon J et al. Derivation of embryonic stem-cell lines from human blastocysts. *N Engl J Med*. 2004; 350(13): 1353–6.
35. Cockburn K, Rossant J. Making the blastocyst: lessons from the mouse. *J Clin Invest*. 2010; 120(4): 995–1003.
36. House of Lords SATSC. Regenerative Medicine. Available from <http://www.parliament.uk/documents/lords-committees/science-technology/RegenerativeMedicine/RegenMed.pdf> ; accessed on 3 November 2014, 2012.
37. Borlongan CV, Glover LE, Tajiri N et al. The great migration of bone marrow-derived stem cells toward the ischemic brain: therapeutic implications for stroke and other neurological disorders. *Prog Neurobiol*. 2011; 95(2): 213–28.
38. Ezzat T, Dhar DK, Malago M et al. Dynamic tracking of stem cells in an acute liver failure model. *World J Gastroenterol*. 2012; 18(6): 507–16.
39. Kang SK, Shin IS, Ko MS et al., Journey of mesenchymal stem cells for homing: strategies to enhance efficacy and safety of stem cell therapy. *Stem Cells Int* 2012; 2012: 342968.
40. Sohni A, Verfaillie CM. Mesenchymal stem cells migration homing and tracking. *Stem Cells Int*. 2013; 2013: 130763.

Serial no.	Comment	Response
<b>Review Round 1</b>		
1.	That patients were not charged of any fees.	No fee was charged for the therapy to the patients treated between the years 2002-2004.
2.	Please mention if pregnant or lactating patients received any therapies.	No, pregnant or lactating patients were not given any therapy
3.	Embryonic Stem cell usage is under the restricted category of research and cannot be used in injections in human subjects. Hence authors are requested to provide Stem Cell Research committee approval.	We are providing the approval letter from the Institutional Ethics Committee (IEC) with the revised draft.
4.	Any usage of cells in human subjects has to be conducted as a trial which involves an animal study before permission is given to conduct in humans but this stage has to be allowed by regulations and until date regulations are slow to react. Big companies like Geron etc (which has been mentioned in the article) have gone into a trial after a rigorous study by FDA.	At the time (in year 2000), when we started hESC research in our facility; stem cell research in India was in its infancy. We recruited our first patient in 2002 and established safety of hESC therapy from 2002 to 2004. Till this time also, no guidelines were available for stem cell research. The only guidelines that were available are from Indian Council of Medical Research (ICMR) on their website and are issued every year (available at <a href="http://icmr.nic.in/ethical_guidelines.pdf">http://icmr.nic.in/ethical_guidelines.pdf</a> ). Few points to consider here are: For every patient that we recruited, we informed and reported to regulatory authority. Detailed reports of outcome of every patient after the therapy were also submitted . The institute has IEC/ICSCRT and has applied for registration to NAC-SC etc. The guidelines state that tissues for transplantation can be obtained from embryos after spontaneous or induced abortion. Further, informed consent should be obtained from the mother whose embryo is sought. This was done. Lastly, an IEC is in place since 2003. The Institutional Committee for Stem Cell Research and Therapy (ICSCRT) sent annual report to the National Apex Body (ICMR).
5.	For any research, there has to be a quantitative value of improvement. The 2 tables that were shown here were of qualitative nature. They were of no significant value.	We agree that we have not quantified the improvement. Please note that, all the patients who visited us had chronic level of injury. The patients came after undergoing several other treatments that did not show any benefits. However, after undergoing hESC therapy, all patients suffering from different medical conditions showed improvement in their health. These are medical observations by qualified doctors. The objective of this was only to assess the safety of hESC therapy. Hence, we did not assess the quantitative improvement.
<b>Review Round 2</b>		
6.	The informed consent is written very very badly and there are glaring mistakes and misinterpretations. This has to be changed by the author.	The consent is very old and we agree that it is not up to the mark. We have changed our consent form. Further, we are also taking video consents for our patients.
7.	The pictures are sent separately and have to be a part of the paper. This needs to be looked at by the author.	We would modify the paper and send the pictures embedded in the paper.
8.	As this involves a contentious issue, the committees have been very careful to grant permission to such a study unless it goes through a full blown trial such as the Phase I through IV. This involves millions of dollars and not just going to the patient with no backup data.	The backup data is present. All the data from these patients and others also was validated by independent CROs. These included Moody's International, QSA and GVK biosciences.
9.	This study is not under the scope of DCGI in India so it cannot be a drug and ICMR does not take responsibility for these cells on human subjects.	We agree to this. However, we would like to bring it to your notice that we started this study, we made a project proposal and approached DST. DST guided us to ICMR and then we submitted our project to them. Since then, we have been submitting all our reports to ICMR. Recently, in Dec 2014, we presented our work and submitted report to the expert committee meeting.
10.	This is under the restrictive category of research. Restrictive and prohibited categories are banned in India.	Restrictive research with minimal manipulation is allowed under 2012 guidelines (Section 7.1, page 14).

Disclaimer: This is a pure experimental study and the study does not come under the purview of accepted Medical Treatment for the patient as stipulated by regulatory agencies world wide who control the use of Stem Cells on Human subjects.