



Association of extended myositis panel results, clinical features, and diagnoses: a single-center retrospective observational study

Shamma Ahmad Al Nokhatha¹ · Eman Alfares² · Luke Corcoran¹ · Niall Conlon² · Richard Conway¹

Received: 6 July 2021 / Accepted: 21 September 2021 / Published online: 4 October 2021
© The Author(s) 2021

Abstract

Myositis-specific antibodies (MSA) and myositis-associated antibodies (MAA) are a feature of the idiopathic inflammatory myopathies (IIM), but are also seen in other rheumatic diseases, and in individuals with no clinical symptoms. The aim of this study was to assess the clinical utility of MSA and MAA and in particular the clinical relevance of weakly positive results. We included all patients at our institution who had at least one positive result on the Immunoblot EUROLINE myositis panel over a 6-year period (2015–2020). Associations with clinical features and final diagnosis were evaluated. Eighty-seven of 225 (39%) myositis panel tests met the inclusion criteria. There were 52 strong positives and 35 weak positives for one or more MSA/MAAs. Among the strong positive group, 15% (8/52) were diagnosed with IIM, 34.6% (18/52) with interstitial lung disease, 7.7% (4/52) with anti-synthetase syndrome, 25% (13/52) with connective tissue disease, and others accounted for 25% (13/52). In weak-positive cases, only 14% (5/35) had connective tissue disease and none had IIM. 60% (21/35) of weak-positive cases were not associated with a specific rheumatic disease. A significant number of positive myositis panel results, particularly weak positives, are not associated with IIM or CTD.

Keywords Myositis · Autoimmune · Antibodies · Inflammatory

Introduction

The idiopathic inflammatory myopathies (IIM) are a heterogeneous group of autoimmune rheumatic diseases characterized by proximal muscle weakness and frequent involvement of other organ systems [1]. The prevalence of IIM can be estimated between 2.4 and 33.8 per 100,000 persons [2].

Historically, the Bohan and Peter criteria were used for IIM, until 2017 when the European League Against Rheumatism and American College of Rheumatology (EULAR/ACR) proposed new classification criteria [3, 4]. These new classification criteria reflect the advances of medicine in the last 40 years as well as providing higher performance (sensitivity/specificity, 93%/88% with biopsies, 87%/82% without biopsies). The new criteria are based primarily on clinical history, examination, and biopsy results. Only one antibody, Anti-Jo-1, is included. The criteria are in the form of a calculator which gives a probability score of the patient having myositis. A classification tree is then used to help determine the subcategory (polymyositis (PM), dermatomyositis (DM), inclusion body myositis, and juvenile dermatomyositis) [4].

However, autoantibodies have been reported in more than 80% of patients with IIM. These autoantibodies can be myositis-specific antibodies (MSA), or myositis-associated antibodies (MAA) which are also seen in a host of other connective tissue diseases (CTD). MSA have a 90% diagnostic specificity, while MAA are noted in up to 50% of myositis patients. These antibodies can help anticipate the clinical course and disease prognosis [5, 6].

✉ Richard Conway
drrichardconway@gmail.com
Shamma Ahmad Al Nokhatha
shamma.alnokhatha@gmail.com
Eman Alfares
Eman.alfaris@yahoo.com
Luke Corcoran
lukepcorcoran@gmail.com
Niall Conlon
NiaConlon@stjames.ie

¹ Department of Rheumatology, St. James's Hospital, James's Street, Dublin, Ireland

² Department of Immunology, St. James's Hospital, James's Street, Dublin, Ireland

MSA include anti-ARS (aminoacyl-tRNA synthetases) antibodies; (histidyl (Jo-1), threonyl (PL-7), alanyl (PL-12), glycylic (EJ), isoleucyl (OJ), asparaginylic (KS), tyrosyl (Ha), and phenylalanyl (Zo)), anti-Mi2 (nucleosome-remodeling deacetylase complex), anti-SRP (signal recognition particle), anti-TIF1 (transcription intermediary factor 1) and anti-NXP-2 (nuclear matrix protein 2), anti-MDA5 (melanoma differentiation-associated protein 5), and anti-SAE (small ubiquitin-like modifier activating enzyme). MAA include anti-PM-Scl, U1RNP, Ku, and Ro52 [7–9].

Autoantibodies are a feature of the subclinical phase of systemic rheumatic diseases and can be present for many years before the onset of clinical symptoms [10, 11]. MSA and MAA are associated with IIM; however, only anti Jo-1 is included in the EULAR/ACR criteria. Weak-positive MSA/MAA are frequently seen and of uncertain clinical significance. Therefore, the aim of the study is to assess the clinical utility of MSA and MAA and in particular the clinical relevance of weakly positive results.

Materials and methods

Study design and setting

This study is a single-center retrospective observational study, performed over a 6-year period (2015–2020). All patients who had an extended myositis antibody panel in this period were assessed for eligibility. Those over age 18 with at least one positive MSA/MAA were included and patients who were followed up in other institutions were excluded. IIM patients with positive MSA/MAA were compared to weak-positive MSA/MAA patients. The study was approved by the St. James' Hospital (SJH)/Tallaght University Hospital (TUH) Joint Research Ethics Committee under protocol number 2020–04 List 15, in May 2020.

Determination/procedure

Myositis antibody testing was performed using the Immunoblot EUROLINE myositis panel, according to the manufacturer's specifications. This assay allows the detection of human IgG autoantibodies to a range of different antigens. This includes 12 MSA (Mi-2a, Mi-2b, TIF1, MDA5, NXP2, SAE1, SRP, Jo-1, PL-7, PL-12, EJ, and OJ), in addition to 4 MAA (Ku, PM-Scl100, PM-Scl75, and Ro/SSA-52). Our immunology lab reports PM-Scl100 and PM-Scl75 separately. Some consider both anti-PM-Scl100 and anti-PM-Scl75 antibodies as one, since they target two closely related isoforms of the same protein. For the purpose of this study, we have included those who were positive for PM-Scl75 and/or PM-Scl100 under the one result. The same applies for Mi-2a and Mi-2b [12]. Anti-nuclear antibody (ANA)

screening by indirect immunofluorescence (IIF) on HEp-2 cells is performed in tandem with each myositis panel to improve specificity, as some myositis antibodies have a distinct ANA staining pattern [13]. The assay was performed according to the manufacturer's recommendations, using a screening dilution of 1:80. Comments are on the presence or absence of antibodies, in addition to the pattern.

Measurement

Immunoblot strips were analyzed using the EuroBlotOne Analyzer/Euroline Scan. This assay provides a semi-qualitative result based on signal intensity of each measured antibody. Results are reported as: negative, weak positive, and strong positive. According to the manufacturer's recommendations, an antibody is considered negative if the signal is < 11. Low positivity is a signal between 11 and 25, and strong positivity beyond 25. The turnover time for the assay is 21 days.

Clinical features

Clinical features were defined as follows. Interstitial lung disease was diagnosed by a respiratory physician. Other features were identified by a rheumatologist and/or immunologist. Arthritis was defined as swelling and tenderness of one or more joints, arthralgia as joint pain with no evidence of arthritis, myositis as muscle weakness supported by relevant investigations, Raynaud's phenomenon as recurrent events of sharply demarcated pallor and/or cyanosis of the skin of the digits with or without reactive hyperaemia, and cutaneous manifestations as Gottron's papules or sign, heliotrope rash, photosensitive rash, calcinosis, digital ulceration, psoriasis, livedo reticularis, or sclerodactyly. Malignancy was defined as any cancer within 5 years of the index study.

Statistical analysis

Statistical analysis was performed using SPSS v26. Descriptive statistics were reported, with results given as frequency and percentages. Categorical variables were compared using Chi-square tests. $p \leq 0.05$ was considered statistically significant throughout.

Results

Patients and demographics

A total of 225 myositis panels were performed in the 6-year study period. 87/225 (39%) patients had positive myositis panel results and met the inclusion criteria, 39% were male and 61% female, with a mean (SD) age of 58 (+16) years.

Of the positive results, 60% (52/87) were strong positive for and 40% (35/87) weak positive for one or more MSA/MAAs. Full demographic data are shown seen in Table 1 (strong positive cohort) and Table 2 (weak-positive cohort).

Clinical features

Tables 1 and 2 summarize the clinical features, ANA results, medication, and outcome of included cases. A creatine kinase (CK) level was performed in 52% of patients, with a median result of 69 (IQR 44.5–277, $p=0.57$). Respiratory medicine accounted for the highest number of test requests (33%, 29/87), followed by rheumatology and immunology (24%, 21/87 each).

Strong-positive MSA/MAA

Anti-PL12 was the most frequent strong positive MSA and anti-Ro52 the most common strong positive MAA (Table 3). The most frequently observed clinical features were arthralgia in 38% (20/52), ILD in 35% (18/52), and cutaneous manifestations in 29% (15/52). Arthritis was seen in 15% (8/52), Raynaud's phenomenon in 15% (8/52), myositis in 13% (7/52), and malignancy in 12% (6/52). Thirteen percent (8/52) were diagnosed with dermatomyositis and 8% (4/52) with anti-synthetase syndrome.

Weak-positive MSA/MAA

Anti-Mi2 was the most frequent weak-positive MSA and anti-Ro52 the most frequent weak-positive MAA (Table 3). The most common clinical manifestations were ILD in 34% (12/35), cutaneous manifestations in 20% (7/35), and arthralgia in 17% (6/35), with Raynaud's phenomenon and arthritis in 11% each (4/35) and myositis and malignancy in 3% (1/35) each. No patients were diagnosed with IIM or anti-synthetase syndrome.

Clinical correlates of positive MSA/MAA

A statistically significant association between arthralgia and a positive myositis panel was identified ($p=0.033$) (Table 4). There were numerical differences for presentations of ILD ($p=0.975$), myositis ($p=0.093$), and cutaneous ($p=0.140$) manifestations, but these did not reach statistical significance. A diagnosis of IIM was associated with a strong positive panel ($p=0.008$). Symptom duration < 1 year was associated with a weakly positive panel ($p=0.022$).

Details of clinical features and diagnosis by individual MSA and MAA are shown in Supplementary Tables 1–7. There was no evident difference between single MSA/MAA positivity and positivity for more than one MSA/MAA and clinical features or diagnosis.

Discussion

Our study shows that those with a strong positive myositis panel were more likely to be diagnosed with an IIM and were more likely to present with arthralgia. There were no diagnoses of IIM in the weakly positive myositis panel group.

A review of the literature shows variations of clinical presentation and serology across different populations. It is felt that genetic factors and environmental triggers may be responsible for this disparity [14]. For example, a study of a Greek population found that the most frequently detected MAA was anti-Ro-52 (30%), while the most frequently detected MSA was anti-Jo-1 (22%) [15]. In our total population, only 3% tested positive for anti-Jo-1.

Our study shows the association of MSA and MAA with IIM, ILD, and CTD are much higher at the strong positive antibody level when compared with the weak positive. However, the diagnostic yield of MSA was generally lower than previously reported studies [16, 17]. This may be because of a relatively short follow-up in our population compared to other published studies or may be due to testing in patients with a lower pre-test probability.

The American thoracic society/European respiratory society/Japanese respiratory society/Latin American thoracic society diagnostic guidelines recommend serial antibody testing in ILD to identify seroconversion and differentiate idiopathic pulmonary fibrosis (IPF) from CTD-ILD. In our study, 34% of all patients were diagnosed with ILD and respiratory having the highest number of requests. This shows the value of MSA testing in ILD as it may present with no or minimal symptoms suggestive of CTD [18]. As CTD-ILD confers a better prognosis and different treatment approach than IPF, it is of paramount importance to detect this subset at an early stage [19].

In our study, MSA were detected in many other inflammatory and non-inflammatory diseases. This finding is in contrast to the majority of prior studies. For instance, Vulsete et al. reported positive MSA in half of patients with IIM compared to only 3.5% of patients with systemic inflammatory diseases and none in healthy controls [20]. This could suggest that MSA sensitivity and specificity vary from one testing lab to another [15, 16]. It may also be the case that there are differences in the populations being tested, with resultant variation in the pre-test probability.

We perform ANA in conjunction with the myositis panel to improve diagnostic performance [13]. 83% of weakly positive myositis panels in our cohort were ANA negative compared to 46% of strong positive panels (~93% correctly matched the non-ANA staining in the positive panel). A false-positive test should be considered if the autoantibody staining/pattern does not correlate with the ANA result and

Table 1 Strong-positive myositis panel characteristics

ANA	Age/gender	MAA	MSA	ILD	Arthritis	Arthralgia	Myositis	Raynaud	Cutaneous	Malignancy	Final diag-nosis	Treatment	Outcome
Inflammatory myositis													
1	+ S 43M	Ro52		+	+	+	+	+	+		Dermatomy- ositis Hidradenitis suppurati- va	Prednisolone+HCQ	Remission/ stable
2	+ S 76F	NXP2					+		+		Dermatomy- ositis Myasthenia gravis	Prednisolone+IVIG+ Azathioprine+pyridostigmine	Remission/ stable
3	+ S 45F	Ro52					+		+	+	Para- neoplastic dermato- myositis, stage 4 high-grade serous ovarian carcinoma	Predniso- lone+MMF+IVIG+chem- otherapy	Worsening
4	- 54F		MDA5		+				+		Amyopathic dermato- myositis	Prednisolone+MTX	Remission/ stable
5	+ S 42F		SAE1				+		+		Dermatomy- ositis	Prednisolone+MTX	Remission/ stable
6	- 77M		Mi2b						+		Dermatomy- ositis	Topical corticosteroid	Remission/ stable
7	- 55M	PMscl100/75 Ro52			+				+		Dermatomy- ositis	Prednisolone+MTX	Remission/ stable
8	+ H 62F	PMscl100/75							+		Dermatomy- ositis sine myositis	MTX	Remission/ stable
Interstitial lung disease													
9	- 66F	Ro52	PL12	+							IPF	Prednisolone	Died
10	+ S 55M		SAE1/OJ	+							IPF	No medication	Lost follow-up
11	+ 68M	Ro52		+							IPF	Prednisolone	Remission/ stable
12	+ S 72F	PMscl100/75		+							IPF	Pirfenidone	Remission/ stable
13	- 83M		PL12	+							IPF	No medication	Remission/ stable

Table 1 (continued)

ANA	Age/gender	MAA	MSA	ILD	Arthritis	Arthralgia	Myositis	Raynaud	Cutaneous	Malignancy	Final diag- nosis	Treatment	Outcome
14	-	73M	EJ	+							IPF	No medication	Remission/ stable
15	-	78M	Ro52	+							IPF	Pirfenidone	Remission/ stable
16	C	46M	Ro52	+	+			+			IPAF Pyoderma gangreno- sum	Prednisolone Adalimumab	Remission/ stable
17	+S	53M	Ro52	+			+				IPAF	Prednisolone + MMF	Remission/ stable
18	-	72F	PL12	+						+	IPAF	Under evaluation	Remission/ stable
19	+H	85F	PMscl100/75	+							IPAF	Prednisolone	Remission/ stable
20	C	71F	PL7	+	+		+				Anti- synthetase syndrome	Prednisolone Cyclophosphamide then Aza- thioprine	Remission/ stable
21	C	62M	PL7	+			+				Anti- synthetase syndrome	Prednisolone + Rituximab	Remission/ stable
22	-	43M	JO-1	+				+			Anti- synthetase syndrome	No medication	Remission/ stable
23	-	66F	JO-1	+							Anti- synthetase syndrome	Prednisolone + MMF then rituximab	Remission/ stable
24	-	73M	SAE1/SRP	+						+	Progressive pulmonary fibrosis (post COVID, ARDS and recurrent aspiration) Esopha- geal Ca T1N2M0 s/p esophagec- tomy	Antibiotics + supportive care	Remission/ stable

Table 1 (continued)

ANA	Age/gender	MAA	MSA	ILD	Arthritis	Arthralgia	Myositis	Raynaud	Cutaneous	Malignancy	Final diag- nosis	Treatment	Outcome
25	C	66M	Ro52	+	+	+					RA-ILD	Prednisolone+Rituximab	Remission/ stable
26	-	74F	Ro52	+	+	+					Sjogren -ILD	Prednisolone+AZA+HCQ	Remission/ stable
Connective tissue disease													
26	+S	60F	Ro52								SLE	HCQ	Lost follow-up
27	+H	75F	Ro52		+	+					Sjogren	HCQ	Remission/ stable
28	+S	69F	Ro52								Sjogren	HCQ	Remission/ stable
29	+S	53F	Ro52		+	+		+	+	+	Sjogren Breast cancer	No medication Surgery+Radiotherapy+Hormonal	Remission/ stable
30	+S	18F	Ro52		+	+					Sjogren	HCQ	Remission/ stable
31	-	33M	Ro52		+	+					Sjogren with neuropsy- chiatry manifesta- tion	AZA	Remission/ stable
32	+	73F	Ro52		+	+					Sjogren	HCQ	Remission/ stable
34	+	66F	Ku/Ro52		+	+		+	+		Undifferenti- ated CTD	No medication	Lost follow-up
35	+S	19 F	U1snRNP		+	+		+	+		Undifferenti- ated CTD	Prednisolone MTX+HCQ	Remission/ stable
36	-	70M	Ro52					+	+	+	Undifferenti- ated CTD query para- neoplastic on back- ground melanoma and eosin- ophilia	Nifedipine	Remission/ stable
37	+S	48F	U1snRNP/ Ro52		+	+			+		MCTD Autoimmune hepatitis	Prednisolone+AZA+HCQ	Remission/ stable

Table 1 (continued)

ANA	Age/gender	MAA	MSA	ILD	Arthritis	Arthralgia	Myositis	Raynaud	Cutaneous	Malignancy	Final diag- nosis	Treatment	Outcome
38	+Ce 52F		SRP					+	+		Limited cutaneous scleroderma	Nifedipine	Remission/stable
39	+ 54F	PMscl100/75						+	+		Scleroderma Scleroderma renal crisis	HCQ and ramipril	Remission/stable
Others													
40	- 72M		NXP2			+					Polymyalgia rheumatica	Prednisolone	Remission/stable
41	+S 61M	ku	Mi2b			+					large vessel vasculitis	Prednisolone Tocilizumab	Remission/stable
42	- 35F		PL12			+			+		PsA	MTX	Remission/stable
43	+S 49F		Mi2b								PBC	Ursodeoxycholic acid	Remission/stable
44	+N 50F	Ro52									Liver cir-rhosis	No medication	Remission/stable
45	+S 53 F	Ro52									Autoimmune limbic encephalitis	IV methyl-pred+IVIG+plasma exchange+cyclophosphamide	Died
46	+N 46F	Ro52							+		Fibromyalgia	No medication	Remission/stable
47	- 37F	PMscl100/75 Ro52							+		Fibromyalgia	No treatment	Remission/stable
48	C 45F	Ku/Ro52									Chronic spontaneous urticaria	Anti-histamine Levothyroxine	Remission/stable
49	- 71F	Ro52							+		High grade serous ovarian carcinoma with metastasis	Surgery and chemotherapy	Remission/stable
50	+H 62F	Ro52									Uterine fibroid	No treatment	Remission/stable

Table 1 (continued)

ANA	Age/gender	MAA	MSA	ILD	Arthritis	Arthralgia	Myositis	Raynaud	Cutaneous	Malignancy	Final diag- nosis	Treatment	Outcome
51	+H 64F	Ro52			+						Rheumatoid arthritis	MTX	Remission/ stable
52	- 73F	PMscl100/75					+				Extranodal NK/T lym- phoma	-	Died

S speckled, H homogenous, C cytoplasmic, Ce centromere, N nucleolar

Table 2 Weak-positive myositis panel characteristics

Age/gender	ANA	MAA	MSA	ILD	Arthritis	Arthralgia	Myositis	Raynaud	Cutaneous	Malignancy	Final diagnosis	Medications	Outcome
1 19M	-	PmScl 100/75	EJ OJ		+	+					IBD-related spondyloar- thropathy	Adalimumab	Remission/stable
2 29F	-		TIF1								MDR TB and neuropathy Intrauterine fibroid	Antibiotic Pregabalin	Remission/stable
3 32M	-		Mi2b					+			Psoriasis	No medication	Remission/stable
4 63F	-		Mi2b	+							Asymptomatic idiopathic bi-apical fibrosis	No medication	Remission/stable
5 57M	-		SRP	+							Sarcoidosis	Nintedanib	Lost follow-up
6 54M	-		Mi2		+	+		+			Bilateral interstitial pulmonary fibrosis	Prednisolone + MTX	Remission/stable
7 79F	-		SRP	+							IPF query RA related	Prednisolone	Died
8 52F	C		Mi2b								Fatty liver along with hepatosplenomegaly Hypothyroidism	No medication	Remission/stable
9 54M	-	Ro52					+				NSCLC-adenocarcinoma T2N1M0 + antiphos- pholipid syndrome and VTE history	Prednisolone + chemo- therapy	Remission/stable
10 59F	C	Ro52						+			Idiopathic livedo vs erythema ab igne	No medication	Remission/stable
11 71M	+S	U1snRNP	PL12				+				Poorly controlled Myas- themia Gravis	IVIG + steroid + pyri- dostigmine	Remission/stable
12 43F	+H	Ro52	PL7		+			+			Coeliac disease Hypothy- roidism	HCQ + MMF	Remission/stable
13 56M	-	PmScl 100/75 Ro52		+				+			Scleroderma/pulmonary fibrosis	Prednisolone + Rituxi- mab + MTX	Remission/stable
14 73F	-		MDA5/SAE1								IgA deficiency	-	Remission/stable
15 66M	+H	PmScl 100/75			+						Degenerative lumbosa- cral spine	No medications	Remission/stable
16 41F	-	PmScl 100/75						+			No Unclear diagno- sis—paroxysms of inflammation cause unclear	Supportive	Remission/stable
17 67F	+H	Ro52	Mi2a/b SRP								Raynaud phenomenon	-	Remission/stable
18 18F	-										MGUS	-	Remission/stable
19 61M	-		Mi2a SRP	+							Chilblains likely second- ary to anorexia nervosa	-	Remission/stable
20 32F	-	Ro52			+			+			Idiopathic pulmonary fibrosis	Nintedanib	Remission/stable
21 78F	-		SAE1								Peripheral SpA Autoimmune hepatitis	Certilizumab	Remission/stable

Table 2 (continued)

Age/gender	ANA	MAA	MSA	ILD	Arthritis	Arthralgia	Myositis	Raynaud	Cutaneous	Malignancy	Final diagnosis	Medications	Outcome
22 33F	-	PmScl 100/75			+				+		Livedo-reticularis and previous peritacheal vasculitis rash in LL		Remission/stable
23 53F	+H	Ro52		+				+			Diffuse systemic sclerosis	Steroid + MMF + Rituximab + Nintendinib	Remission/stable
24 62F	-		Mi2a								Pontine stroke and under workup for MS	Clopidogrel	Remission/stable
25 72M	-		Mi2b								AML and organizing pneumonia	Chemo + steroid taper for OP	Remission/stable
26 89M	-		Mi2a	+							UIP-ILD / IPF		Remission/stable
27 60F	-		Mi2b SAE1 SRP	+							IPF query RA related		Died
28 64F	-	Ro52				+			+		Discoid lupus	Was on steroid, HCQ + MMF	Remission/stable
29 60F	-	Ro52	MDA5	+							IPF query RA related	o2	Remission/stable
30 69M	-		Mi2b								COPD and asthma	Inhalers + on/off steroid	Remission/stable
31 76M	-	Ku									Hospital Acquired Pneumonia with parapneumonic effusions		Remission/stable
32 64M	-		PL-12	+							IPF		Remission/stable
33 79M	-		SAE1/PL-7								IPF	Nintedanib	Remission/stable
34 67M	-		NXP2	+							IPAF-ILD secondary to CTD	Steroid +	Remission/stable
35 46F	+S	U1snRNP Ro52							+		MCTD	HCQ	Remission/stable

S speckled, H homogenous, C cytoplasmic

Table 3 The results of the antibodies for both positive and weakly positive

Antibody	Positive	Weakly positive
MSA		
Anti-PL-12	4	2
Anti-SAE1	3	3
Anti-Mi2	3	12
Anti-NXP2	2	1
Anti-Jo	2	1
Anti-SRP	2	5
Anti-PL7	2	2
Anti-EJ	2	1
Anti-OJ	2	1
Anti-MDA5	1	2
MAA		
Anti-Ro52	29	10
Anti-PMScI	7	5
Anti-Ku	3	–
Anti-U1RNP	2	2

clinical context [9]. However, some MSA exhibit negative ANA testing due to cytoplasmic localisation, and as such

negative ANA does not necessarily imply autoantibody negativity in IIM.

This study was not without its limitations. Our power to detect significant differences was impacted by a relatively small sample size and low number of IIM diagnoses. This highlights the need for larger collaborative studies to evaluate these rare conditions. This was a single-center study and our findings require confirmation in other settings to confirm external validity. Given the significant mortality and morbidity burden of IIM, early and accurate diagnosis should be a primary goal in all cases. Based on the above, we have proposed an algorithm to guide the interpretation of myositis antibody panel results, Fig. 1. This highlights our findings and suggests that weak-positive panels should be repeated to confirm the result.

The current EULAR/ACR guidelines suggest that clinical assessment and biopsy are the core components of the diagnostic approach to IIM. Our expanding knowledge of the importance of MSA/MAA suggests a key adjunctive role in diagnosis. Our study found that positive panels are more likely to be associated with IIM; however, a significant number of cases had no clinical features suggestive of CTD or IIM. A combined clinical and serological framework may be useful in IIM diagnosis.

Table 4 Chi-square analysis between weak-positive and positive myositis panel

	Type				<i>p</i> value
	Weak-positive myositis panel		Positive myositis panel		
	Count	Column <i>N</i> %	Count	Column <i>N</i> %	
ILD	12	34.3	18	34.6	0.975
Arthritis	4	11.4	8	15.4	0.600
Arthralgia	6	17.1	20	38.5	0.033*
Myositis	1	2.9	7	13.5	0.093
Raynaud	4	11.4	8	15.4	0.600
Cutaneous	7	20.0	18	34.6	0.140
Malignancy	1	2.9	6	11.5	0.144
Final diagnosis					
Inflammatory myositis	0	0.0	8	15.4	0.008*
Interstitial lung disease	12	34.3	18	34.6	
Connective tissue disease	5	14.3	14	26.9	
Others	18	51.4	12	23.1	
Management					
Corticosteroid	3	8.6	5	9.6	0.115
Corticosteroid + immunosuppression	7	20.0	17	32.7	
Immunosuppression	3	8.6	12	23.1	
No treatment	11	31.4	9	17.3	
Others	11	31.4	9	17.3	
Outcome					
Died	2	5.7	3	5.8	0.773
Remission/stable	32	91.4	45	86.5	
Worsening	0	0.0	1	1.9	
Lost follow-up	1	2.9	3	5.8	
Duration					
= < 1 year	23	65.7	22	42.3	0.022*
2 years	6	17.1	14	26.9	
3 years	6	17.1	5	9.6	
4 years	0	0.0	8	15.4	
5 years	0	0.0	3	5.8	

**p* < 0.05

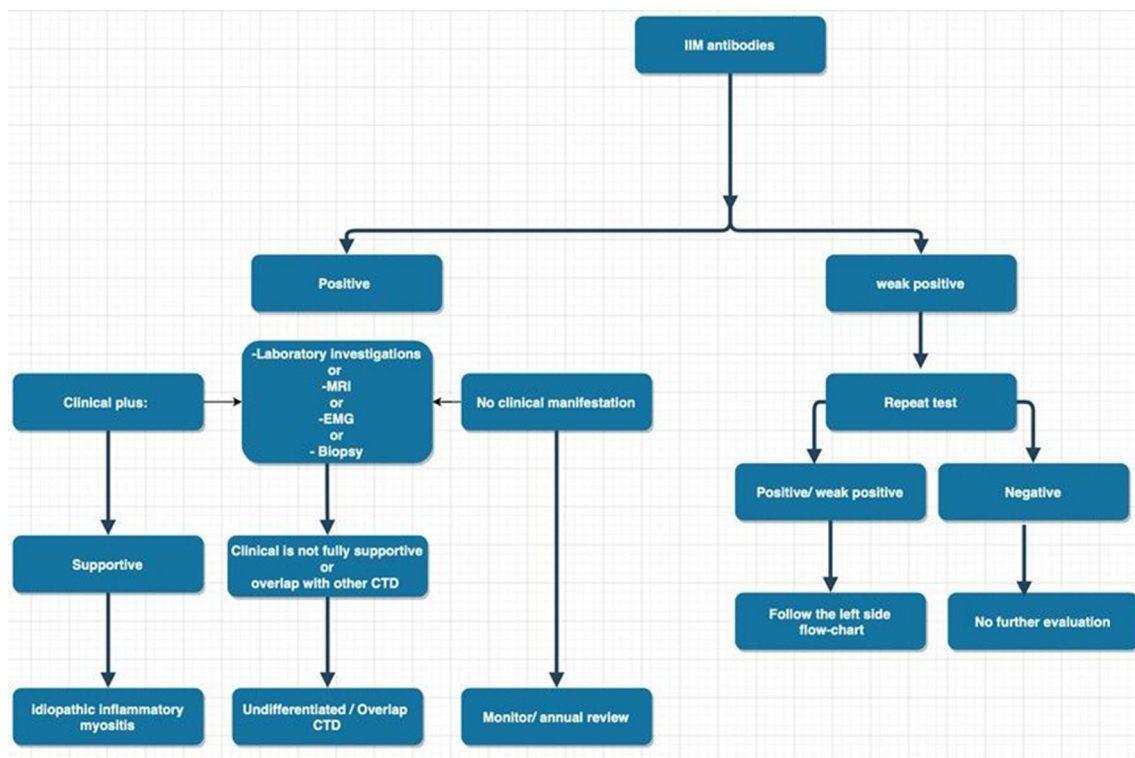


Fig. 1 A proposed algorithm to guide interpretation of myositis antibody panel results

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1007/s00296-021-05012-0>.

Author contributions SAN, NC, and RC: substantial contributions to the conception or design of the work and the acquisition, analysis, and interpretation of data for the work; and drafting the work or revising it critically for important intellectual content; and final approval of the version to be published; and agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. EA and LC: substantial contributions to the acquisition, analysis, and interpretation of data for the work; and drafting the work or revising it critically for important intellectual content; and final approval of the version to be published; and agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Funding Open Access funding provided by the IReL Consortium. None.

Availability of data and materials Available from authors on request.

Code availability Not applicable.

Declarations

Conflict of interest The authors declare that they have no conflict of interest.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

1. Firestein G, Budd R, Gabriel S, McInnes I, O'Dell J (2017) Kelley's textbook of rheumatology. Elsevier, Philadelphia
2. Meyer A, Meyer N, Schaeffer M, Gottenberg JE, Geny B, Sibilia J (2015) Incidence and prevalence of inflammatory myopathies: a systematic review. *Rheumatology (Oxford)* 54:50–63. <https://doi.org/10.1093/rheumatology/keu289>
3. Bohan A, Peter JB (1975) Polymyositis and dermatomyositis (first of two parts). *N Engl J Med* 292:344–347. <https://doi.org/10.1056/nejm197502132920706>
4. Lundberg IE, Tjärnlund A, Bottai M, Werth VP, Pilkington C, Visser M, Alfredsson L, Amato AA, Barohn RJ, Liang MH, Singh JA, Aggarwal R, Arnardottir S, Chinoy H, Cooper RG, Dankó K, Dimachkie MM, Feldman BM, Torre IG, Gordon P, Hayashi T,

- Katz JD, Kohsaka H, Lachenbruch PA, Lang BA, Li Y, Oddis CV, Olesinska M, Reed AM, Rutkowska-Sak L, Sanner H, Selva-O'Callaghan A, Song YW, Vencovsky J, Ytterberg SR, Miller FW, Rider LG (2017) 2017 European League Against Rheumatism/American College of Rheumatology classification criteria for adult and juvenile idiopathic inflammatory myopathies and their major subgroups. *Ann Rheum Dis* 76:1955–1964. <https://doi.org/10.1136/annrheumdis-2017-211468>
5. Cruellas MG, Viana Vdos S, Levy-Neto M, Souza FH, Shinjo SK (2013) Myositis-specific and myositis-associated autoantibody profiles and their clinical associations in a large series of patients with polymyositis and dermatomyositis. *Clinics (Sao Paulo)* 68:909–914. [https://doi.org/10.6061/clinics/2013\(07\)04](https://doi.org/10.6061/clinics/2013(07)04)
 6. Ghirardello A, Borella E, Beggio M, Franceschini F, Fredi M, Doria A (2014) Myositis autoantibodies and clinical phenotypes. *Auto Immun Highlights* 5:69–75. <https://doi.org/10.1007/s13317-014-0060-4>
 7. Chino H, Sekine A, Baba T, Iwasawa T, Okudela K, Takemura T, Itoh H, Sato S, Suzuki Y, Ogura T (2016) Radiological and pathological correlation in anti-MDA5 antibody-positive interstitial lung disease: rapidly progressive perlobular opacities and diffuse alveolar damage. *Intern Med (Tokyo, Japan)*. 55:2241–2246. <https://doi.org/10.2169/internalmedicine.55.5774>
 8. Mahler M, Miller FW, Fritzler MJ (2014) Idiopathic inflammatory myopathies and the anti-synthetase syndrome: a comprehensive review. *Autoimmun Rev* 13:367–371. <https://doi.org/10.1016/j.autrev.2014.01.022>
 9. Palterer B, Vitiello G, Carraresi A, Giudizi MG, Cammelli D, Parronchi P (2018) Bench to bedside review of myositis autoantibodies. *Clin Mol Allergy* 16:5. <https://doi.org/10.1186/s12948-018-0084-9>
 10. Ma WT, Chang C, Gershwin ME, Lian ZX (2017) Development of autoantibodies precedes clinical manifestations of autoimmune diseases: a comprehensive review. *J Autoimmun* 83:95–112. <https://doi.org/10.1016/j.jaut.2017.07.003>
 11. Vulsteke JB, Blockmans D, Moons V, Vijgen S, Bossuyt X, De Langhe E (2020) Detection of anti-Mi-2 autoantibodies before dermatomyositis-specific manifestations. *Rheumatology (Oxford)* 59:e60–e62. <https://doi.org/10.1093/rheumatology/keaa055>
 12. Lecouffe-Desprets M, Hémond C, Néel A, Toquet C, Masseur A, Hamidou M, Josien R, Martin JC (2018) Clinical contribution of myositis-related antibodies detected by immunoblot to idiopathic inflammatory myositis: a one-year retrospective study. *Autoimmunity* 51:89–95. <https://doi.org/10.1080/08916934.2018.1441830>
 13. Damoiseaux J, Andrade LEC, Carballo OG, Conrad K, Francescantonio PLC, Fritzler MJ, Garcia de la Torre I, Herold M, Klotz W, Cruvinel WM, Mimori T, von Muhlen C, Satoh M, Chan EK (2019) Clinical relevance of HEp-2 indirect immunofluorescent patterns: the International Consensus on ANA patterns (ICAP) perspective. *Ann Rheum Dis* 78:879–889. <https://doi.org/10.1136/annrheumdis-2018-214436>
 14. Alenzi FM (2020) Myositis specific autoantibodies: a clinical perspective. *Open Access Rheumatol* 12:9–14. <https://doi.org/10.2147/oarr.S231195>
 15. Zampeli E, Venetsanopoulou A, Argyropoulou OD, Mavragani CP, Tektonidou MG, Vlachoyiannopoulos PG, Tzioufas AG, Skopouli FN, Moutsopoulos HM (2019) Myositis autoantibody profiles and their clinical associations in Greek patients with inflammatory myopathies. *Clin Rheumatol* 38:125–132. <https://doi.org/10.1007/s10067-018-4267-z>
 16. Platteel ACM, Wevers BA, Lim J, Bakker JA, Bontkes HJ, Curvers J, Damoiseaux J, Heron M, de Kort G, Limper M, van Lochem EG, Mulder AHL, Saris CGJ, van der Valk H, van der Kooij AJ, van Leeuwen EMM, Veltkamp M, Schreurs MWJ, Meek B, Hamann D (2019) Frequencies and clinical associations of myositis-related antibodies in The Netherlands: a one-year survey of all Dutch patients. *J Transl Autoimmun* 2:100013. <https://doi.org/10.1016/j.jtauto.2019.100013>
 17. González-Bello Y, García-Valladares I, Reyes-Pérez IV, García-Cerda D, Medrano-Ramírez G, Navarro-Zarza JE, Andrade-Ortega L, Maradiaga-Ceceña M, Cardenas-Anaya A, Nava-Zavala AH, Orozco-Barocio G, Vázquez-Del Mercado M, Rojo-Mejía A, Loyo E, Gottschalk P, Iglesias-Gamarra A, Vega K, Rojas C, Mantilla R, Gómez G, García-Kutzbach A, Fritzler MJ, García-De La Torre I (2020) Myositis-specific antibodies and myositis-associated antibodies in patients with idiopathic inflammatory myopathies from the PANLAR myositis study group. *J Clin Rheumatol*. <https://doi.org/10.1097/rhu.0000000000001350>
 18. De Sadeleer LJ, De Langhe E, Bodart N, Vigneron A, Bossuyt X, Wuyts WA (2018) Prevalence of myositis-specific antibodies in idiopathic interstitial pneumonias. *Lung* 196:329–333. <https://doi.org/10.1007/s00408-018-0108-8>
 19. Yoshimura K, Kono M, Enomoto Y, Nishimoto K, Oyama Y, Yasui H, Hozumi H, Karayama M, Suzuki Y, Furuhashi K, Enomoto N, Fujisawa T, Nakamura Y, Inui N, Sumikawa H, Johkoh T, Colby TV, Sugimura H, Suda T (2018) Distinctive characteristics and prognostic significance of interstitial pneumonia with autoimmune features in patients with chronic fibrosing interstitial pneumonia. *Respir Med* 137:167–175. <https://doi.org/10.1016/j.rmed.2018.02.024>
 20. Vulsteke J-B, Bossuyt X, Dillaerts D, Poesen K, Claeys K, Lenaerts J, Westhovens R, Blockmans D, Haes PD, Langhe ED (2017) FRI0393 Prevalence of myositis-specific antibodies in idiopathic inflammatory myopathy compared to disease and healthy controls. *Ann Rheum Dis* 76:636–637. <https://doi.org/10.1136/annrheumdis-2017-eular.5143>

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.