Review

The effects of long non-coding ribonucleic acids on various cellular components in rheumatoid arthritis

Yilong Fang^{1,2,*}, Jiajie Tu^{1,2,*}, Dafei Han^{1,2}, Yawei Guo^{1,2}, Wenming Hong^{1,2} and Wei Wei^{1,2}

Abstract

RA is a chronic, autoimmune-mediated inflammatory pathology. Long non-coding RNAs (IncRNAs) are a novel group of non-coding RNAs with a length of >200 nucleotides. There are reports emerging that suggest that IncRNAs participate in establishing and sustaining autoimmune diseases, including RA. In this review article, we highlight the functions of IncRNAs in different cell types in RA. Our review indicates that IncRNAs affect various cellular components and are novel candidates that could constitute promising targets for the diagnosis and treatment of RA.

Key words: RA, IncRNA, FLS, T cells, PBMCs

Rheumatology key messages

- LncRNAs are involved in RA via regulating different cells, including fibroblast-like synoviocytes, T cells and peripheral blood mononuclear cells.
- LncRNAs are promising hotspots for the diagnosis and treatment of RA.
- To identify individual IncRNAs and their specific roles in RA is essential for future study.

Introduction

More than 90% of human genomic DNA can be transcribed into RNA, but only 2% of this RNA represents mRNA that can be translated into protein post-transcriptionally. The remaining 98% of the genome that do not encode protein are referred to as non-coding RNAs [1]. Long non-coding RNAs (IncRNAs) are an accumulation of transcripts with a length >200 nucleotides [2]. Until recently, IncRNAs have not been the focus of intensive research, since IncRNA was considered to be a by-product of RNA polymerase II transcription. This 'noise' product produced by gene transcription was supposed to have no biological function associated with it [3]. Nevertheless, recent studies have confirmed that IncRNAs play a role in many basic biological processes,

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*Yilong Fang and Jiajie Tu contributed equally to this study.

including nucleation, splicing, RNA degradation and translation [4]. Furthermore, IncRNAs can contribute to gene regulation through epigenetic modifications such as methylation, ubiquitination and chromatin remodelling [5]. More importantly, numerous studies have shown that IncRNAs play a key role in the occurrence of the pathological progression of autoimmune diseases, including RA.

RA is a relatively common chronic inflammatory autoimmune disorder that is characteristic of systemic inflammation, peripheral joint destruction and erosion of bone [6]. Moreover, it seriously affects the health of patients, limiting quality of life and leading to severe disabilities. Currently the specific underlying pathological mechanisms that cause RA are still unclear. It is generally believed that several factors, including recurrent infection, cold stimulation, fatigue and environmental factors, play a significant role in the aetiology of RA. Recently, accumulating studies have demonstrated that IncRNAs could play a prominent role in a multitude of biological processes that participate in the pathogenesis of RA [7]. Here we summarize the functions of IncRNAs in several essential cellular compartments that have been identified to be involved in the pathogenesis of RA, including fibroblast-like synoviocytes (FLS), T lymphocytes (T cells) and peripheral blood mononuclear cells (PBMCs).

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¹Institute of Clinical Pharmacology and ²Key Laboratory of Anti-Inflammatory and Immune Medicine, Ministry of Education, Anhui Collaborative Innovation Center of Anti-Inflammatory and Immune Medicine, Anhui Medical University, Hefei, China

Correspondence to: Wei Wei, Institute of Clinical Pharmacology, Anhui Medical University, 81 Meishan Road, Shushan District, Hefei City, Anhui Province, China. E-mail: wwei@ahmu.edu.cn

IncRNAs in different cell types of RA

IncRNAs in FLS of RA

FLS, which are also referred to as synovial fibroblasts or type B synoviocytes, are mesenchymal cells of the synovial joints that have major regulatory functions in joint health. During RA pathology, this cellular compartment can abundantly proliferate and invade cartilage and bone [8], participating in the pathogenesis of RA, and it has been identified as a promising target for RA treatment [9].

Maternally expressed gene 3 (MEG3) is a long noncoding RNA gene locus that acts as tumour suppressor. A loss of this RNA has been defined as an important marker of various human cancers [10-12]. Interestingly, it has been described in synovial tissues and cells of adjuvant arthritis rat models that nucleotide oligomerization domain-like receptor 5 (NLRC5), a member of the NODlike receptor family that participates in inflammatory and immune responses [13], is increased dramatically while the expression of MEG3 is markedly decreased [14]. Furthermore, in adjuvant arthritis rat models the MEG3 promoter was methylated in FLS. The methylation inhibitor 5-aza-2-deoxycytidine suppressed MEG3 promoter methylation significantly and simultaneously downregulated NLRC5 expression. Interference with the expression of MEG3 may result in decreased levels of NLRC5 and inflammatory cytokines. Therefore strong expression of MEG3 and downregulation of NLRC5 could reduce the migrative and invasive abilities of RA FLS and it provides a new perspective that MEG3 may potentially alleviate RA symptoms by targeting NLRC5.

The IncRNA lowly expressed in rheumatoid fibroblastlike synoviocytes (LERFS) has a lower expression in synovial tissues and cells of RA patients than in healthy controls. Furthermore, LERFS suppress the ability of FLS to migrate, invade and proliferate by interacting with heterogeneous nuclear ribonucleoprotein Q (hnRNP Q) [15]. LERFS-hnRNP Q complex binds to the mRNA of RhoA, Rac1 and CDC42, a small GTPase protein that could regulate of FLS motility and proliferation, to target their mRNA, inhibiting its stability and translation, and to negatively regulate its protein expression in healthy controls. In contrast, the decrease in LERFS levels leads to a decrease of LERFS-hnRNP Q complex in RA FLS, which decreases the integration between hnRNP Q and target mRNA, strengthening its stability and translation and upregulating its protein expression. In conclusion, IncRNA LERFS negatively regulate FLS aggression and proliferation, with the downregulated expression of LERFS leading to synovial proliferation and joint destruction of RA.

Previous reports have demonstrated that Tanshinone IIA (Tan IIA) is used to promote apoptosis of RA FLS, thereby hindering the progression of RA [16]. Expression of IncRNA growth arrest-specific 5 (GAS5) is downregulated in RA FLS, however, the GAS5 level is upregulated after treatment with Tan IIA. Apoptosis is significantly repressed after knocking down GAS5 by small interfering RNA (siRNA) transfection [17]. Meanwhile, the knockdown of Gas5 downregulates caspase-3 and caspase-9 levels and activates the phosphoinositide 3-kinase (PI3K)/AKT signalling pathway. Therefore these results show that the pro-apoptotic effect of Tan IIA on RA FLS is achieved by upregulating IncRNA GAS5.

Urothelial carcinoma associated 1 (UCA1) is a novel IncRNA that resides in the chromosome 19p13.12, with three exons and two introns [18]. A series of reports have demonstrated that UCA1 plays a key role in the pathologic processes of diverse disorders. Yan *et al.* [19] analysed the effect of the activity of IncRNAs on FLS and the pathologic processes of RA. They found that UCA1 shows an enhanced expression in normal FLS compared with that in RA FLS. Down-regulated UCA1 in normal FLS increases cell viability, while upregulated UCA1 in RA FLS reduces cell viability. Furthermore, they also showed that UCA1 induces the apoptosis of FLS by regulating caspase-3 and Wnt6. These results demonstrate that UCA1 could affect the pathological progression of RA by promoting FLS apoptosis.

The association of IncRNA-NR024118 with the inhibition of shikonin on inflammatory response in FLS has been demonstrated [20]. An injury of soft tissue and bone in shikonin-treated arthritis mice can be detected. Shikonin dramatically induces IncRNA-NR024118 and suppressor of cytokine signalling (SOCS3) expression and suppresses secretion and expression of the pro-inflammatory mediators IL-8, IL-6 and various MMPs. In addition, shikonin inhibits the proliferation of FLS and enhances acetylation of histone H3 at the promoter of IncRNA-NR024118 in a dose- and time-dependent manner in RA FLS. Upregulation and siRNA silencing of IncRNA-NR024118 has been shown to enhance and reduce, respectively, the expression of SOCS3. These results suggest that the inhibitory effects of shikonin on the inflammatory response in RA FLS is mediated via affecting IncRNA-NR024118.

The IncRNA ZFAS1 expression level in synovial tissue has been compared between healthy controls and RA patients [21]. The expression of ZFAS1 is enhanced in RA FLS compared with FLS from healthy donors. ZFAS1 siRNA interference markedly suppresses the migratory and invasive phenotype of RA FLS while ZFAS1 overexpression boosts the migration and invasive phenotype of FLS. Importantly, the upregulation of microRNA 27a (miR-27a) almost abolishes the invasive phenotype of RA FLS. However, in a pull-down experiment it was shown that miR-27a allows ZFAS1 to exert its function. These findings indicate that IncRNA ZFAS1 promotes the migration and invasion of FLS via suppression of miR-27a in RA.

Mo *et al.* [22] recently demonstrated that IncRNA GAPLINC (gastric adenocarcinoma associated, positive CD44 regulator, long intergenic non-coding RNA) affects the functional role of RA FLS. Moreover, GAPLINC is expressed in RA FLS, compared with healthy FLS isolated from trauma patients. In order to verify whether the increased GAPLINC can exert its function in RA FLS, the RA FLS proliferation has been validated by a Cell Counting Kit-8 (CCK-8) assay using an siRNA-mediated

GAPLINC knockdown in RA FLS. The CCK-8 results show that GAPLINC knockdown decreases RA FLS proliferation. Moreover, the migratory and invasive abilities of RA FLS are suppressed by GAPLINC silencing. Inflammatory cytokines or proteinases such as IL-6, IL-8 and MMP-9, which are expressed in FLS and intensify the inflammatory response, are also suppressed in GAPLINC knockdown RA FLS. Further studies have illustrated that GAPLINC downregulation leads to an enhanced expression of two anti-inflammatory miRNAs, miR-382-5p and miR-575. These results indicate that GAPLINC could enhance RA FLS tumour-like features, including proliferation, migration and invasion, by downregulating miR-382-5p and miR-575 [23-25].

Quercetin is a ubiquitous dietary antioxidant that has been proven to be helpful in the treatment of arthritis [26, 27]. Pan et al. [28] analysed the mechanisms of guercetin promoting the apoptosis of RA FLS. RA FLS viability is considerably reduced in the wake of elevated concentrations of guercetin, and FLS viability is time-dependently decreased after treatment with the same concentration of quercetin. A PCR array has been used to screen and mark the IncRNAs, and it has been shown that IncRNA MALAT1 (Metastasis Associated Lung Adenocarcinoma Transcript 1) was notably upregulated. Furthermore, knockdown of MALAT1 significantly suppresses the apoptosis of the RA FLS and the protein expression of caspase-3, caspase-9 and Bax. In contrast, Bcl-2 expression is enhanced and the PI3K/AKT pathway is activated in FLS following treatment with siRNA targeting MALAT1. Thus the upregulation of IncRNA MALAT1 is a requirement for the guercetininduced apoptosis of FLS in RA.

Long non-coding interleukin-7 receptor (Inc-IL7R) has been authenticated recently. This RNA is considerably upregulated by stimulation with lipopolysaccharide (LPS) and is able to impair LPS-induced inflammatory responses in the human monocyte line THP1 and human umbilical vein endothelial cells (HUVECs) [29]. Researchers showed that Inc-IL7R overexpression enhances FLS proliferation [30]. Further work illustrated that Inc-IL7R can drive cell cycle progression and suppress cell apoptosis, thus enhancing FLS proliferation. Importantly, the function of Inc-IL7R on proliferation, cell cycle and apoptosis is reversed by knocking down Inc-IL7R. It was hypothesized and later confirmed that Inc-IL7R forms a close contact with enhancer of zeste homologue 2 (EZH2), which is a central constituent of polycomb repressive complex 2 (PRC2) to modulate gene expression by affecting histone modification, and Inc-IL7R serves an essential role in participating in suppression induced by PRC2, which incorporates cyclin-dependent kinase inhibitor 1A (p21) and cyclin-dependent kinase inhibitor 2A (p16). EZH2 knockdown almost abolishes the ability of FLS to proliferate, stalls the cell cycle and boosts cell apoptosis. These results demonstrate that Inc-IL7R could enhance FLS growth by interacting with EZH2.

Collectively, activated FLS is a key feature of RA. FLS proliferate excessively and play an important role in

inflammatory responses, autoimmunity and joint destruction in RA [6]. A number of IncRNAs affect many aspects of FLS (Fig. 1), including viability, apoptosis, proliferation, migration and invasion, to aggravate or alleviate RA via various mechanisms.

IncRNAs in PBMCs of RA

PBMCs are key constituents in host defence responses and are used to identify new disease vectors/variants and therapeutic responses [31]. Furthermore, PBMCs are easier to obtain when compared with synovial tissues. Therefore specific lncRNAs in PBMCs may be used as a general marker for clinical diagnosis of RA [32].

To investigate the expression of IncRNA between RA patients and a normal group, Luo et al. [33] initially used human IncRNA microarrays that included 30 586 IncRNA and 26 109 coding transcripts to profile the patterns of gene expression of PBMCs in RA patients, revealing the potential function of IncRNA in the pathological progression of RA. There were 5045 IncRNA differentially expressed in RA patients (2410 up and 2635 down). The most dramatically upregulated IncRNA was ENST00 000583574 (257.7 times) and the most remarkably downregulated was ENST00000559539 (4726.8 times). Most differentially expressed IncRNAs are derived from the intergenic regions (42%), natural antisense (19%) and intronic antisense to the protein-coding locus (15%). Moreover, 135 potential IncRNA-mRNA pairs were verified for 85 abnormally expressed IncRNA and 109 abnormally expressed mRNA. This is the first report to study the relationship between PBMCs and IncRNA in RA.

Another group also utilized microarray technology to analyse the differentially expressed IncRNAs in PBMCs in RA. The bioinformatic pathway and gene ontology analyses found that the IncRNAs could regulate the aberrantly expressed mRNAs, which played a significant role in the pathological progression of RA [34]. For instance, the increased cellular components, including MHC class II protein complex, cell periphery and trans-Golgi network membrane, and the concentrated biological processes, including system development, acute-phase response and cell-cell adhesion, were associated with upregulated mRNAs. The downregulated mRNAs participated in the binding of nucleic acid, a heterocyclic compound. The increased cellular components, including nucleus and intracellular components, and the biological processes, including nucleus and cellular compound metabolic processes, were associated with downregulated mRNAs. Quantitative PCR showed that ENST00000456270 expression was prominently upregulated in PBMCs in RA patients. Moreover, the increasing level of ENST00000456270 was associated with the serum levels of IL-6, TNF- α , and the Simplified Disease Activity Index in RA patients, which demonstrated that ENST00000456270 may serve as a biomarker for assessing and diagnosing RA patients.

Zhang *et al.* [35] used quantitative RT-PCR to detect the expression of IncRNAs in PBMCs of 65 RA patients and 54 control patients. Meanwhile, three single-nucleotide polymorphisms (SNPs) of Inc0640 and Inc5150 were





(A) UCA1 affects the pathogenesis of RA by reducing viability and inducing apoptosis of FLS. LncRNA-NR024118 is involved in the inhibition of shikonin on inflammation in FLS. MALAT1 is important to quercetin-induced apoptosis of FLS in RA. GAS5 increases after the Tan IIA treatment in RA FLS and GAS5 upregulates caspase-3 and caspase-9 levels and silences the PI3K/AKT pathway. (B) ZFAS1 promotes the migration and invasion of FLS by inhibiting miR-27a to aggravate RA. GAPLINC enhances inflammatory cytokine or proteinase expression. GAPLINC promotes RA FLS tumour-like features, including proliferation, migration and invasion by downregulating miR-382-5p and miR-575. Lnc-IL7R enhances RA FLS proliferation and suppresses its apoptosis to aggravate RA by interacting with EZH2. (C) In RA FLS, MEG3 promoter is methylated to decrease MEG3 expression, thus the increasing expression of NLRC5 participating in inflammatory and immune responses. Downregulated LERFS leads to a decrease of LERFS-hnRNP Q complex in RA FLS.

genotyped. Compared with the healthy controls, the expression of Inc0640 in PBMCs from RA patients was markedly increased, while the expression of Inc5150 was decreased. The expression of Inc0640 and Inc5150 in patients with RA was significantly correlated with CRP and Inc5150 expression was closely related to the ESR. There was a statistical correlation between the TT genotype of rs13039216 in the Inc0640 gene and RA risk reduction. And the risk of variation in the recessive model rs13039216 was reduced. In addition, compared with RF-negative patients, the G allele frequency of the Inc5150 gene rs141561256 polymorphism was notably decreased in RF-positive RA patients. These results show the Inc0640 and Inc5150 participate in RA pathological progression.

The genomic region of TRAF1-C5 is relatively large and contains several candidate genes associated with validated susceptibility genes of RA that include major histocompatibility complex, class II, DR Beta-1 (HLA-DRB1). lymphoid tyrosine phosphatase (PTPN22) and signal transducer and activator of transcription 4 (STAT4). Messemaker et al. [36] found that the TRAF1-C5 region was related to RA. TRAF1 served as a target in the treatment of RA that could negatively regulate TNF-a signalling, while the C5 expression level increased in the joints of RA patients [37, 38]. The C5T1IncRNA-regulated gene transcription level could be identified in the same genomic region. The expression of C5T1IncRNA was positively associated with C5 mRNA and PBMCs in numerous tissues. C5T1IncRNA induction was determined in LPS-stimulated PBMCs in a dose-dependent manner. Furthermore, the expression of C5T1IncRNA was strongly linked to C5 in PBMCs stimulated by LPS. Knocking down C5T1IncRNA reduced C5 mRNA levels in hepatocytes and in FLS. However, levels of TRAF1 mRNA were not influenced [36]. Therefore C5T1IncRNA, which has been considered as a novel IncRNA, is completely located in the relevant area and is expressed in several cells associated with RA to regulate the transcript levels of C5.

In addition to its role in apoptosis, p53, a tumour suppressor protein, also has anti-inflammatory effects [39]. LincRNA-p21 plays a key role in regulating the responses of cells to p53 [40]. Spurlock et al. [41] attempted to investigate the correlation of p53, lincRNA-p21 and nuclear factor (NF)-kB activity in the context of RA and MTX treatment by analysing blood samples from RA sufferers. They found that RA patients had a lower basal expression level of lincRNA-p21 and a higher expression level of phosphorylated p65 (RelA), the marker of NF-kB activation. Compared with RA sufferers who were treated with lowdose MTX, RA sufferers who did not receive MTX therapy showed lower expression levels of lincRNA-p21 and higher expression levels of ReIA. In PBMC lines and primary cell culture, MTX induced expression of lincRNAp21 by the activation of DNA-dependent protein kinase catalytic subunit and lincRNA-p21 directly inhibited NFκB activity.

To summarize, IncRNAs regulate the aberrantly expressed mRNAs in PBMCs, which participate in the pathological progression of RA (Fig. 2). Meanwhile, differentially expressed IncRNAs in PBMCs from RA patients could be used as targets for potential diagnosis and treatment of RA.

IncRNAs in T lymphocytes (T cells) of RA

T cells from RA patients show several aberrant functions, for instance, premature aging, self-reactivity and T cell subset disorder [42–44]. Employing soluble cytotoxic T lymphocyte antigen 4 recombinant proteins to inhibit T cell activity could relieve the inflammation of RA sufferers [45].



Fig. 2 Regulatory roles and expression of IncRNAs in PBMCs of RA

The expression of C5T1IncRNA is positively associated with C5 in PBMCs of RA. Upregulating C5 promotes complement activation to aggravate inflammatory response. Furthermore, C5T1IncRNA induces LPS-stimulated inflammatory response in PBMCs in a dose-dependent manner [36]. RA patients have a lower basal expression level of lincRNA-p21 and a higher expression level of phosphorylated p65 (ReIA), the marker of NF- κ B activation. MTX induces expression of lincRNA-p21 by the activation of DNA PKcs and lincRNA-p21 directly inhibits NF- κ B activity [41].

A multitude of SNPs in the protein tyrosine phosphatase non-receptor type 2 (PTPN2) locus are connected with many autoimmune diseases, including juvenile idiopathic arthritis [46], type 1 diabetes [47] and RA [48]. It has been reported that knockout PTPN2 mice die from grievous inflammatory diseases at 3-5 weeks after birth [49] and PTPN2 was identified as a negative regulator of T cell receptor signalling to prevent autoimmune diseases [50]. PTPN2 could restrain the differentiation of CD8⁺ T cells after antigen cross-presentation [51]. Houtman et al. [52] investigated the autoimmune disease and link with the PTPN2 locus. The results revealed that autoimmune disease-associated SNPs in the PTPN2 locus are highly linked to differential DNA methylation at four CpG sites downstream of PTPN2 and expression of IncRNA LINC01882 and RP11-973H7.1 occur primarily in T cells in RA patients. To investigate the function of LINC01882 in T cells, LINC01882 was silenced in Jurkat T cells and it could be shown that there were six genes upregulated and six genes downregulated. The most significantly upregulated gene was BZRAP1, whose role in immune cells was not identified. MAP2K4, mediating the response of cells to cytokine signalling and stress, and ZEB1, encoding the transcription factor that inhibits the expression of IL-2 in T cells, were downregulated by LINC01882 knockdown models [53]. Taken together, these results demonstrate that LINC01882 participated in the activation of T cells of RA.

In another study, Lu *et al.* [54] evaluated whether several abnormally expressed IncRNAs in T cells from RA patients could facilitate the inflammatory response. They identified that the expression of IncRNA LOC100652951 and LOC100506036 was considerably increased in T cells from patients with RA. RA sufferers had a lower expression level of LOC100652951 and female RA patients had a

lower expression level of LOC100506036 after the use of biologic agents. Furthermore, the expression level of LOC100506036, rather than LOC100652951, was notably enhanced in Jurkat T cells activated by phorbol 12-myristate 13-acetate and ionomycin. Silencing LOC100506036 decreased the production of IFN- γ and nuclear factor of activated T cells in Jurkat T cells. In addition, knockdown of LOC100506036 repressed sphingomyelin phosphodiesterase 1 (SMPD1), which may further regulate secretion of IL-2 and cytotoxic granules [55-57], T cell apoptosis and differentiation [58, 59]. Together, the expression of IncRNA LOC100652951 and LOC100506036 were considerably increased in T cells from patients with RA, and the latter could regulate numerous genes including SMPD1 and nuclear factor of activated T cells along with promoting RA inflammatory responses.

T cells play a central role in the pathogenesis of RA and promote RA bone injury by directly contacting other pathogenic cells or secreting mediators. The above-mentioned lncRNAs regulate inflammation in RA via activation, apoptosis and differentiation of T cells (Fig. 3). They are likely to manage RA persistently by regulating lncRNAs in T cells in the joints.

IncRNAs in other cellular components of RA

Monocytes are the core participants in the coordination of complex immune responses [60]. Yang *et al.* [61] established the relation between 17 kb IncRNA non-coding transcripts in T cells (NTT) and monocytes. It was found that NTT was expressed in primary monocytes and in macrophages derived from monocytes, and was adjusted by C/EBP, a crucial transcription factor for monocytes. Together, C/EBP/NTT was overexpressed in PBMCs from patients initially diagnosed as RA without any treatment, however, these genes were downregulated in the

Fig. 3 Regulatory roles and expression of IncRNAs in T cells of RA



BZRAP1, whose role in immune cells is not identified, is remarkably upregulated after knocking down LINC01882. MAP2K4, mediating the response of cells to cytokine signalling and stress, and ZEB1, coding the transcription factor that inhibits the expression of IL-2 in T cells, are downregulated by LINC01882 knockdown [53]. The expression of LINC01882, IncRNA LOC100652951 and LOC100506036 were remarkably increased in T cells from RA patients, LOC100506036 regulates IFN-γ, nuclear factor and SMPD1 to activate T cells [55-57]. wake of treatment. Moreover, IncRNA NTT was found to regulate inflammation in monocytes and its activation was closely associated with the differentiation of monocytes and macrophages and was involved in RA pathological progression.

As a member of the IncRNAs, IncRNA-AF085935 was related to HCV in the serum of patients with RA. Compared with healthy donors, serum IL-10, IL-17 and IncRNA-AF085935 in were significantly increased in patients with RA associated with HCV. The serum levels of IL-10 and HCV RNA were remarkably increased after corticosteroid treatment, while the serum levels of IL-17 were notably decreased, with negligible levels of IncRNA-AF085935 observed. Therefore IncRNA-AF085935 plays a role in detecting early RA associated with HCV and provides a novel strategy in the treatment of RA associated with HCV [62].

LncRNA growth arrest-specific 5 (GAS5), a glucocorticoid receptor inhibitor in the manner of its RNA 'glucocorticoid response element', was accumulated in the wake of starvation-induced cellular growth arrest [63]. In the whole blood of patients with multiple sclerosis and clinically isolated syndrome with the earliest clinical manifestations of multiple sclerosis, GAS5 levels were significantly increased [64]. In addition, the levels of GAS5 in CD4 T cells and whole blood were slightly affected by viral infection. The abundance of GAS5 in whole blood was decreased in sufferers with bacterial sepsis [65]. GAS5 levels were not sensitive to fasting in immune organs, including the spleen and thymus, in contrast to the metabolic organs such as the liver, fat and skeletal muscles [66]. Together, GAS5 regulating glucocorticoid receptor transcriptional activity through its decoy RNA 'glucocorticoid response element' plays a role in the immunological function and pathological/pathophysiological regulation of autoimmunity, inflammation and infectious diseases.

Remarkably upregulated expression of IncRNA HOX transcript antisense RNA (HOTAIR) was detected in PBMCs and serum exosomes of patients with RA, which could accelerate the migration of macrophages. In addition, Song et al. [67] confirmed upregulated HOTAIR suppressed MMP-2 and MMP-13 expression in osteoclasts and RA synoviocytes, and its expression dramatically decreased in LPS-treated RA chondrocytes. The cell proliferation and inflammatory factors IL-17 and IL-23 were remarkably repressed in HOTAIR-overexpressed chondrocytes [68]. Another study showed that the overexpression of miR-138 partly reversed the effects on cell proliferation and inflammation by upregulating HOTAIR in LPS-induced chondrocytes [69]. In addition, the activation of NF-κB treated by LPS was notably repressed by the overexpression of HOTAIR, which is a significant actor in the activation of NF-kB in macrophages [70]. In conclusion, HOTAIR alleviates RA by inhibiting the miR-138 and NF- κ B pathway.

More recently, Shaker *et al.* [71] found that the mRNA expression levels of HOTAIR and IncRNA-Cox2, related to the activation or suppression of immunomodulator gene expression, were markedly upregulated in serum from RA patients compared with healthy subjects. Meanwhile,

LncRNA	Expression	Sample	Pathway	Functional role	Related gene	Reference
IncRNA NTT	Up	THP-1 cells (human monocytic laukaemia cell line)	C/EBP regulating	Regulating inflammation in monocytes	C/EBP	[61]
IncRNA- AF085935	Up	Serum of RA patients associated with HCV	IL-10, IL-17 expression increasing	Detecting early RA associated with HCV	HCV RNA	[62]
GAS5	Down	CD4 T cells and B cells obtained from RA and SLE patients	Regulated GR transcriptional activity through its decoy RNA 'GRF'	Regulating effect on autoimmunity, inflammation and infectious	GRE	[63]
HOTAIR	Чр	Serum exosome and blood mononuclear cells in RA	Repressing the activation of MMP-13 and MMP-2; in- hibiting miB-138 and NE-2	Alleviating RA	MMP-13 MMP-2	[67, 69, 70]
H19	Up	Synovial rissue in RA and OA patients; isolated synovial	None	Be a tumour markerRelating to TIMP-2 in synovial fibroblasts	PI3k ERK-1/2	[73]
HOTAIR and IncRNA-Cox2	Up	Serum from RA patients	None	Discriminating RA patients from healthy folks	IL-6 and MMP-9	[71]
GR: glucocorticoid re	sceptor; GRE: gl	ucocorticoid response element.				

TABLE 1 LncRNA expression traits in other cellular components

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HOTAIR and IncRNA-Cox2 could distinguish between RA and healthy control groups using the receiver operating characteristics curve. Thus HOTAIR and IncRNA-Cox2 in serum are able to serve as non-invasive biomarkers for diagnosing RA.

The expression of H19 RNA is at a basal level in all tissues except skeletal muscle after birth. However, its expression reoccurs in diverse tumours, so H19 RNA may be a tumour biomarker [72]. H19 expression in synovial tissue from patients with RA was upregulated compared with normal/joint trauma controls. Positive signals of antisense probe for H19 were observed in the stroma, diffuse infiltrates and lining layer regions of RA synovium by in situ hybridization. What's more, quantitative RT-PCR was used to confirm the expression of H19 in both isolated macrophages and synovial fibroblasts. Meanwhile, it was encouraging to find that the expression of H19 in RA synovial fibroblasts was dramatically upregulated in the context of starvation, either in the presence or absence of stimulation with IL-1 β , TNF- α or platelet-derived growth factor BB (PDGF-BB). Moreover, the expression of H19 was downregulated or upregulated by the inhibitors of MAP kinase ERK-1/2 and the phosphatidylinositol 3-kinase after stimulation of IL-1β or PDGF-BB, respectively [73]. Thus H19 RNA may be a biomarker for embryonal dedifferentiation of adult synovial tissue and inflammatory response in RA. The function and related mechanism of IncRNAs in RA is quite versatile (Table 1). If we can comprehend explicitly the mechanism of action of IncRNA in RA, it will provide new ideas for the diagnosis and treatment of RA and other autoimmune-related diseases.

IncRNAs as targets for diagnosis and treatment of RA

In recent years, studies of IncRNAs in RA have focussed on functional changes and related molecular mechanisms of IncRNAs in various dysregulated cells of RA patients. The ultimate objective is to seek the potential application of IncRNAs in clinical diagnosis and therapy of RA in the future.

Diverse bioinformatic and high-throughput technologies have been used to investigate the role of IncRNAs in RA, such as the above-mentioned microarray analysis and gene ontology terminology. Luo et al. [33] used human IncRNAs microarrays that included 30 586 IncRNAs and 26 109 coding transcripts to evaluate the differential expression of IncRNAs in PBMCs of RA patients and healthy controls. There were 5045 (2410 up and 2635 down) differentially expressed IncRNAs in PBMCs of RA patients. The largest upregluated one was ENST00000583574 (257.7 times) and the largest downregulated one was ENST00000559539 (4726.8 times). Gene ontology analysis also predicted that a large number of differentially expressed IncRNAs are closely associated with inflammation and immune response [33], suggesting that these abnormally expressed IncRNAs in PBMCs may be used to indicate the progress or prognosis of RA with high sensitivity and specificity [74]. In addition, Spurlock et al. [41] found that MTX can induce lincRNA-p21 by activating DNA PKcs. The specific response of lincRNA-p21 to MTX reminded us that it is possible to use the encapsulation technique to deliver IncRNA to specific tissues, such as a joint cavity, for RA treatment. A previous study also showed that nanofibres encapsulating siRNA-treated RA mice have achieved promising results [75]. It is known that IncRNAs show a relatively low conservation between species, various structures and complex molecular regulatory mechanisms. Currently it is difficult to find strong evidence of IncRNAs to replace or replenish conventional clinical biomarkers or therapy in RA. However, it is believed that IncRNAs will have wider use in RA with a deeper understanding of the specific role of individual IncRNAs in various essential cells of RA (Fig. 4).





The differential expression of IncRNAs in PBMCs could be used as potential diagnostic or prognostic biomarkers for RA. LncRNAs may also serve as potential therapeutic targets and an encapsulation technique can be used to deliver IncRNAs to a specific tissue, such as a joint cavity, to treat RA.

Conclusion and future perspectives

RA is a chronic and common autoimmune disorder and its aetiology is still elusive. There are still knowledge gaps. although the underlying pathogenesis has been extensively described [6]. There have been reports of a significant role for IncRNAs in RA. However, the functional role of IncRNAs in RA is largely unexplored. It is a challenge to identify individual IncRNAs and their specific roles in RA pathological progression. Structure is the basis of function, and function cannot be separated from its specific molecular structure. Due to the relatively long length, IncRNAs possess the characteristics of being large molecules, with high molecular weight and complex spatial structure, with lower sequence conservation of secondary structures [76, 77]. Thus identifying the spatial structure of IncRNAs and their interactions is essential for understanding their function and mechanisms. In addition, in order to carry out much deeper research on IncRNAs in different cellular components in RA, cutting-edge technology should be applied. The application of single-cell sequencing could increase the precision of research on IncRNAs, which helps us to better understand the specific role of IncRNAs at the single cell level [78]. Finally, in spite of a large number of IncRNAs serving as biomarkers for assessing and diagnosing RA patients, researchers need to study more clinical samples and further validate the role of IncRNAs in RA patients.

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