



# Determination of the iron(IV) local spin states of the Q intermediate of soluble methane monooxygenase by K $\beta$ X-ray emission spectroscopy

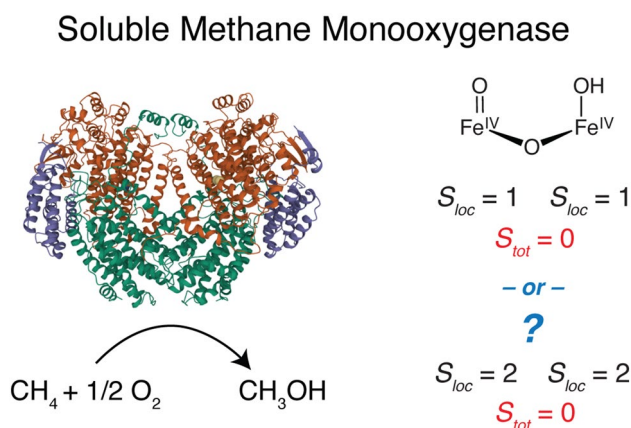
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Received: 13 April 2022 / Accepted: 7 August 2022 / Published online: 21 August 2022  
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## Abstract

Soluble methane monooxygenase (sMMO) facilitates the conversion of methane to methanol at a non-heme Fe<sup>IV</sup><sub>2</sub> intermediate MMOH<sub>Q</sub>, which is formed in the active site of the sMMO hydroxylase component (MMOH) during the catalytic cycle. Other biological systems also employ high-valent Fe<sup>IV</sup> sites in catalysis; however, MMOH<sub>Q</sub> is unique as Nature's only identified Fe<sup>IV</sup><sub>2</sub> intermediate. Previous <sup>57</sup>Fe Mössbauer spectroscopic studies have shown that MMOH<sub>Q</sub> employs antiferromagnetic coupling of the two Fe<sup>IV</sup> sites to yield a diamagnetic cluster. Unfortunately, this lack of net spin prevents the determination of the local spin state (*S*<sub>loc</sub>) of each of the irons by most spectroscopic techniques. Here, we use Fe K $\beta$  X-ray emission spectroscopy (XES) to characterize the local spin states of the key intermediates of the sMMO catalytic cycle, including MMOH<sub>Q</sub> trapped by rapid-freeze-quench techniques. A pure XES spectrum of MMOH<sub>Q</sub> is obtained by subtraction of the contributions from other reaction cycle intermediates with the aid of Mössbauer quantification. Comparisons of the MMOH<sub>Q</sub> spectrum with those of known *S*<sub>loc</sub> = 1 and *S*<sub>loc</sub> = 2 Fe<sup>IV</sup> sites in chemical and biological models reveal that MMOH<sub>Q</sub> possesses *S*<sub>loc</sub> = 2 iron sites. This experimental determination of the local spin state will help guide future computational and mechanistic studies of sMMO catalysis.

## Graphical abstract



**Keywords** Methane monooxygenase · Non-heme iron · Spin state · X-ray emission spectroscopy · Rapid freeze-quench

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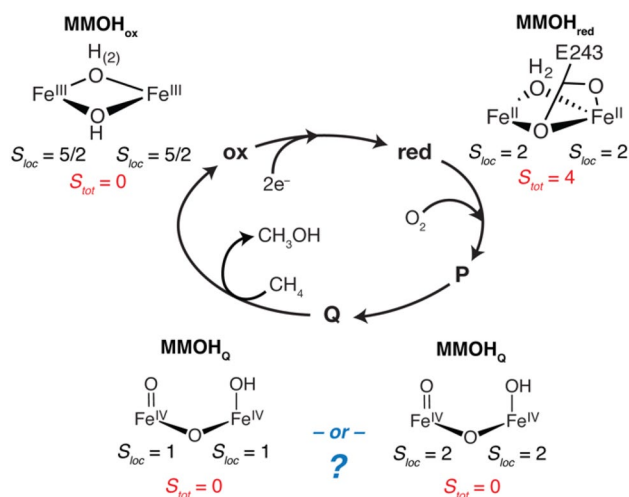
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## Introduction

The conversion of methane to methanol is of significant interest for technologies aimed at gas-to-liquid fuel conversion, and for the sequestration of a greenhouse gas, alongside the desire to understand the fundamental chemistry of strong C–H bond activation. Nature facilitates this challenging chemical reaction with two different methane monooxygenase enzymes [1]. The most common type is the copper-containing particulate methane monooxygenase (pMMO) [2–4], which is a topic of intense research, in particular for the assignment of the active site and its structure [5–11]. In copper limited environments [2, 12], methanotrophic bacteria express the iron-containing soluble methane monooxygenase (sMMO), which has a well characterized non-heme carboxylate-bridged di-iron active site [13, 14].

The active site of sMMO is located within the hydroxylase protein (MMOH) and forms various catalytic intermediates that have been spectroscopically identified [13, 15–25]. An abbreviated catalytic mechanism of MMOH is shown in Fig. 1. For the critical intermediate of interest,  $\text{MMOH}_\text{Q}$ , the O–O bond of the preceding peroxo intermediate ( $\text{MMOH}_\text{P}$ ) has been cleaved and the resultant intermediate is able to activate the strong 105 kcal/mol C–H bond of methane, allowing for its subsequent conversion to methanol. To our knowledge,  $\text{MMOH}_\text{Q}$  is the only di-iron(IV) intermediate that has been identified in Nature [17, 19, 20].

The exact geometric structure of the  $\text{MMOH}_\text{Q}$  intermediate has been hotly debated for over two decades [14, 17, 18, 21, 23–27]. Spectroscopic characterizations of  $\text{MMOH}_\text{Q}$



**Fig. 1** Abbreviated catalytic scheme of sMMO with corresponding iron oxidation states, local spin states ( $S_{\text{loc}}$ ) and cluster ground spin state ( $S_{\text{tot}}$ ) for  $\text{MMOH}_{\text{ox}}$ ,  $\text{MMOH}_{\text{red}}$ , and  $\text{MMOH}_\text{Q}$ . The structure of  $\text{MMOH}_\text{Q}$  is drawn as the proposed ‘open-core’ structure [23, 24], although a closed  $\text{Fe}_2(\mu\text{-O})_2$  core is also proposed [21, 26]

have included vibrational studies (transient resonant Raman and nuclear resonance vibrational scattering, NRVS), which have also been used to argue for a “closed-core” bis- $\mu$ -oxo structure [21, 25]. However, high-resolution Fe K-edge X-ray absorption (HERFD XAS) characterization and pre-edge analysis of  $\text{MMOH}_\text{Q}$  in comparison with biomimetic models support an “open core” structure [23]. Recent HERFD extended X-ray absorption fine structure (EXAFS) measurements of RFQ samples of sMMO were fit with a long Fe–Fe distance of  $\sim 3.3$  Å and showed no evidence for the initially reported 2.46 Å di-iron distance that would have supported a closed-core structure [24]. The short Fe–Fe distance in the previous 1997 EXAFS report [18] was shown to arise from metallic iron background scattering contributions, which are suppressed in the HERFD EXAFS [24]. Because the RFQ samples in the HERFD EXAFS study contained primarily  $\text{MMOH}_\text{Q}$  ( $\sim 50\%$ ) and  $\text{MMOH}_{\text{red}}$  (40%), the fitted distance is weighted average of these contributions. As the di-iron distance in  $\text{MMOH}_{\text{red}}$  is established to be  $\sim 3.2$  Å [28], the longer fitted distance of the RFQ samples therefore suggests a longer di-iron distance for  $\text{MMOH}_\text{Q}$  of  $\sim 3.4$  Å, consistent with an open core structure [24]. Further, a recent quantum mechanics-molecular mechanics (QM/MM) computational study of  $\text{MMOH}_\text{Q}$  has shown that the computed spectroscopic signatures of an open core structure are consistent with the Mössbauer, Fe HERFD pre-edge XAS, EXAFS di-iron distance measurements, and resonance Raman [29]. These QM/MM computational studies, however, have not yet been extended to the recently reported NRVS spectrum of  $\text{MMOH}_\text{Q}$  [25].

While the geometric structure of  $\text{MMOH}_\text{Q}$  remains controversial, its electronic structure is also key to understanding catalytic activity and predicting spectroscopic signatures. The initial spectroscopic characterization and identification of  $\text{MMOH}_\text{Q}$  included its characteristic optical absorption at 430 nm and a single observed Mössbauer quadruple doublet in the case of  $\text{MMOH}_\text{Q}$  trapped using the enzyme from *Methylosinus trichosporium* OB3b [16, 19, 20]. The single Mössbauer doublet from two irons was interpreted to indicate that the iron sites have similar coordination environments, thereby yielding overlapping doublets. The isomer shift of the observed doublet is entirely consistent with  $\text{Fe}^{\text{IV}}$  oxidation state, however, the Mössbauer characterization of  $\text{MMOH}_\text{Q}$  remains inconclusive for the assigned local spin states of the iron sites. Low temperature and applied magnetic field Mössbauer spectroscopy of  $\text{MMOH}_\text{Q}$  determined that the total ground spin state of the cluster is  $S_{\text{tot}} = 0$ , meaning that the  $\text{Fe}^{\text{IV}}$  ions are antiferromagnetically coupled, where iron sites must have local spin states ( $S_{\text{loc}}$ ) of intermediate-spin ( $S_{\text{loc}} = 1$ ) or high-spin ( $S_{\text{loc}} = 2$ ) [20]. As originally stated by Münck, the isomer shift and quadrupole splittings of  $\text{MMOH}_\text{Q}$  are consistent with both  $S_{\text{loc}} = 1$  and  $S_{\text{loc}} = 2$  local spin states; the applied field measurement

only serves to measure the coupling strength between the two sites [20]. Non-diamagnetic cluster spin states allow for clear local spin-state assignments by Mössbauer spectroscopy. For other mononuclear Fe<sup>IV</sup> synthetic chemical and biological centers, applied field Mössbauer measurement has been essential to differentiate between intermediate and high-spin iron [30–34]. For example, in the case of the  $S_{\text{tot}} = 1/2$  Fe<sup>III</sup>–Fe<sup>IV</sup> cluster intermediate (X) of ribonucleotide reductase (RNR) [35], the paramagnetic nature of the cluster allowed iron hyperfine and local spin states to be determined by applied field measurements.

Attempts to understand or model both the local and cluster spin states of MMOH<sub>Q</sub> through biomimetic chemistry have also been challenging [33, 36–39]. Different local and cluster spin states have been identified in these models, including those with  $S_{\text{loc}} = 1$  or 2 and ferro- or antiferromagnetic coupling of the iron sites. A sampling of both mono and di-nuclear biomimetic models and their Mössbauer parameters are listed in Table 1. The observed Mössbauer isomer shifts and quadrupole splittings for both  $S_{\text{loc}} = 1$  and 2 Fe<sup>IV</sup> sites span a fairly wide range around what is observed for MMOH<sub>Q</sub>, making a direct assignment of a local spin state from these parameters alone prohibitive.

Perhaps the primary support for the now more commonly accepted  $S_{\text{loc}} = 2$  spin state of MMOH<sub>Q</sub> arises in part from analogy to other non-heme iron chemistry [39]. Enzymatic non-heme Fe<sup>IV</sup> sites favor local high-spin states, in part due to their weak-field ligand environments [30], similar to that

present in MMOH. Furthermore, the enhanced reactivity of high-spin  $S = 2$  Fe<sup>IV</sup> sites to perform hydrogen atom transfer reactions, as seen in other forms of non-heme biological catalysis and biomimetic chemistry [31, 33, 34, 38, 48, 49], is often used to infer a spin-state assignment for MMOH<sub>Q</sub>. Cryo-reduction experiments of MMOH<sub>Q</sub> have also revealed an electronic structure similar in character to RNR-X, suggesting locally high-spin Fe<sup>IV</sup> sites [50]. Previous computational studies of MMOH<sub>Q</sub> also favor  $S_{\text{loc}} = 2$  iron spin states [29, 51–53]. Ultimately, these chemical analogies or other studies have fallen short of a direct experimental characterization of the local spin states in MMOH<sub>Q</sub>, motivating an investigation of the electronic structure by means of an alternative spectroscopic approach.

For 3d transition metals, K $\beta$  (3p → 1s) X-ray emission spectroscopy (XES) has been well demonstrated to be excellent reporter of electronic structure due to 3p–3d exchange in the final state [54–58]. K $\beta$  mainlines have previously been demonstrated to both report and correlate to oxidation state and spin state. As a marker of oxidation state, the maximum of the K $\beta$  mainline, the K $\beta_{1,3}$  feature, generally shifts to higher energy with increased oxidation-state [54]. On the other hand, the intensity of the K $\beta'$  feature, at lower energy, correlates with the number of unpaired electrons, thereby allowing for spin-state assignments. For instance, for ferrous complexes, the presence or absence of the K $\beta'$  has been used as a marker of high-spin ( $S = 2$ ) or low-spin ( $S = 0$ ) electronic configurations [57–60].

**Table 1** Mössbauer parameters of various Fe<sup>IV</sup> sites

	$S_{\text{loc}}$	$S_{\text{tot}}$	$\delta$ (mm/s)	$\Delta E_Q$ (mm/s)	References
MMOH <sub>Q</sub>		0	0.17	0.53	[20]
TauD J	2	2	0.31	– 0.88	[40]
RnR X (Fe <sup>III</sup> – <b>Fe<sup>IV</sup></b> ) <sup>a</sup>	5/2, 2	1/2	0.26	– 0.6	[35]
[L(OH)Fe-( $\mu$ -O) <b>Fe(O)L</b> ] <sup>3+</sup> ; L = TPA* <sup>a,b</sup>	1, 1	0	– 0.03	+ 0.92	[41]
[L( <b>OH</b> )Fe-( $\mu$ -O)Fe(O)L] <sup>3+</sup> ; L = TPA* <sup>a,b</sup>	1, 1	0	0.0	+ 1.96	[41]
[Fe <sup>IV</sup> <sub>2</sub> ( $\mu$ -O) <sub>2</sub> (TPA*) <sub>2</sub> ] <sup>4+</sup>	1, 1	0	– 0.04	2.09	[36]
[(O)(L)Fe <sup>IV</sup> –O–Fe <sup>IV</sup> (O)(L)] <sup>2+</sup> ; L = TPA* <sup>b</sup>	2, 2	0	0.14	0.52	[39]
[(O)(L)Fe <sup>IV</sup> –O–Fe <sup>IV</sup> (O)(L)] <sup>2+</sup> ; L = TPA* <sup>b</sup>	1, 2	3	– 0.02, 0.14	– 1.17, 0.82	[39]
[(TMC)Fe(O)NCMe] <sup>2+</sup> <sup>c</sup>	1	1	0.17	+ 1.24	[42]
[(N4Py)Fe(O)] <sup>2+</sup> <sup>d</sup>	1	1	– 0.04	0.93	[43]
[Fe <sup>IV</sup> (O)(2PyN2Q)] <sup>2+</sup> <sup>e</sup>	1	1	0.03	0.56	[44]
[Fe <sup>IV</sup> (O)(TMG <sub>3</sub> tren)] <sup>2+</sup> <sup>f</sup>	2	2	0.24	– 0.29	[45]
[Fe <sup>IV</sup> (O)( <i>t</i> Bu <sub>3</sub> TACN)] <sup>2+</sup> <sup>g</sup>	2	2	0.11	+ 0.96	[46]
Aqueous Fe <sup>IV</sup> = O	2	2	0.38	– 0.33	[47]

<sup>a</sup>In the case of asymmetric di-iron cluster, the parameters are detailed for the iron in bold

<sup>b</sup>TPA\* = tris(4-methoxy-3,5-dimethylpyridyl-2-methyl)amine

<sup>c</sup>TMC = 1,4,8,11-tetramethyl-1,4,8,11-tetraazacyclotetradecane

<sup>d</sup>N4Py = *N,N*-bis(2-pyridylmethyl)bis(2-pyridyl)methylamine

<sup>e</sup>2PyN2Q = 1,1-di(pyridin-2-yl)-*N,N*-bis(quinolin-2-ylmethyl)methanamine

<sup>f</sup>TMG<sub>3</sub>tren = tris[2-(*N*-tetramethylguanidyl)ethyl]amine

<sup>g</sup>*t*Bu<sub>3</sub>TACN = 1,4,7-tri-*tert*-butyl-1,4,7-triazacyclononane

While these rule-of-thumb trends may hold true in idealized systematic studies, covalency is shown to have profound influences on the shape and energies of K $\beta$  mainlines [56, 58]. Increasing metal–ligand covalency delocalizes metal 3d orbital character onto the ligands and thus decreases the metal 3p–3d exchange integrals, which determine the final state splitting in K $\beta$  spectra [56]. Ultimately, this relationship predicts that expected shifts of the K $\beta_{1,3}$  to higher energy with increased oxidation-state may be counteracted by increased metal–ligand covalency, making some interpretations across an oxidation state series difficult. However, these challenges generally do not hinder the ability of K $\beta$  XES to probe local spin states and distinguish between spin states in samples of known oxidation-state and similar covalency [54, 56–58, 61, 62]. Generally, Fe K $\beta$  XES is not sensitive to the magnetic spin coupling ( $J$ ) between the iron centers in bimetallic or larger clusters; therefore, the bulk XES measurement reports on the average local spin state of all iron sites [63–66]. This makes the approach attractive to possibly probe the local spin states in diamagnetic clusters such as MMOH<sub>Q</sub> and distinguish between its local spin-state assignments.

## Experimental

The sMMO proteins <sup>57</sup>Fe-enriched MMOH and MMOB were purified according to protocols described recently in the literature [22, 67]. The Fe K $\beta$  mainline XES spectra were collected on frozen solution samples of MMOH<sub>red</sub>, MMOH<sub>ox</sub> and freeze-quenched samples (MMOH-RFQ) to trap MMOH<sub>Q</sub> of the same samples described in a previously published HERFD EXAFS study [24]. The MMOH-RFQ samples were prepared as previously described by freezing the reaction mixture in precooled sample cell of super-cooled liquid nitrogen (-199 °C) [24].

<sup>57</sup>Fe Mössbauer spectroscopy was used to quantitate the three components, MMOH<sub>red</sub>, MMOH<sub>ox</sub> and MMOH<sub>Q</sub>, in the MMOH-RFQ sample. As previously reported [24], the frozen solution samples had 46% MMOH<sub>Q</sub>, 39.5% MMOH<sub>red</sub>, and 14.5% MMOH<sub>ox</sub>.

The  $S = 1$ , Fe<sup>IV</sup> = O complex ([Fe<sup>IV</sup>(O)(2PyN2Q)](PF<sub>6</sub>)<sub>2</sub> (2PyN2Q = 1,1-di(pyridin-2-yl)-*N,N*-bis(quinolin-2-ylmethyl)methanamine)) was synthesized and isolated following published procedures [44, 68]. Solid samples for XES data collection were prepared in 1.0 mm Al spacers and sealed with 38  $\mu$ m thick Kapton tape.

The Fe K $\beta$  XES data were collected at beam line ID-26 of the European Synchrotron Radiation Facility (ESRF) operating at 6 GeV and 200 mA. All samples were measured in a liquid helium cryostat operating at 20 K. The energy of the incident beam was selected using either a Si(111) or Si(311) double crystal monochromator, each of which was calibrated by setting the first inflection of an iron foil to 7111.2 eV. The

incident monochromator was then set to an excitation energy of 7800 eV to non-resonantly excite the sample. The XES spectra were collected with a 1 m radius Johann spectrometer equipped with five Ge(620) spherically bent analyzer crystals and calibrated by the scanning of elastic scattering lines. A collected reference K $\beta$  XES spectrum of Fe<sub>2</sub>O<sub>3</sub> is displayed in Fig. S1. The measured resolution of the elastic scattering line was ~1.5 eV (fwhm) which includes the contribution of the upstream Si(111) monochromator. For the synthetic solid sample, an avalanche photodiode (APD) detector was used, while the more dilute protein samples required the use of an energy resolving Ketek detector to further improve the S/N of the emission spectra. For the XES collection of the protein, the incident beam was attenuated to a maximum estimated flux of  $4 \times 10^{12}$  photons/s within a spot size of 1.2 mm ( $w$ )  $\times$  0.1 mm ( $v$ ) to further reduce radiation damage. Maximum sample exposure dwell times were determined through the evaluation of repeated fast X-ray near edge absorption (XANES) scans (5–60 s) to determine total acceptable photon doses [24]. Collection of the protein XES was completed in small segments (~12 eV wide, 50–61 points) of the entire XES spectra (7.025–7.08 keV) with 2 eV overlap per energy segment (10–11 data points). Each segment was collected on a fresh sample spot, limiting the total sample spot exposure time. The maximum dwell times per data point were: MMOH<sub>red</sub>, 1.25 secs; MMOH<sub>ox</sub> 0.40 secs; MMOH<sub>Q</sub>, 0.15 secs. This limited the sample spot exposure times to a maximum of 76.25 secs (MMOH<sub>red</sub>), 24.4 secs (MMOH<sub>ox</sub>) and 15.25 secs (MMOH<sub>Q</sub>). This collection process allowed for improved signal-to-noise to be acquired at each data point, and to mitigate beam induced damage. All data segments were splined together through the averaging of their overlapping components. A sample of the process is shown in Fig. S2.

All K $\beta$  XES spectra were normalized to a unit area of 1 over the energy range of 7025 to 7080 eV. The XES spectrum of pure MMOH<sub>Q</sub> was calculated by subtracting the MMOH<sub>red</sub> and MMOH<sub>ox</sub> components from MMOH-RFQ at the ratios determined by Mössbauer and renormalizing to a unit area of 1. The first moments reported are determined over the described energy ranges by the following equation,

$$M_1 = \frac{\sum_i (E_i I_i)}{\sum_i I_i}$$

where  $E_i$  and  $I_i$  are the energy and intensities, respectively, at data point  $i$ .

## Results

Samples of MMOH<sub>red</sub>, MMOH<sub>ox</sub> and rapid-freeze-quenched (RFQ) samples of MMOH<sub>Q</sub> were prepared as previously described [24]. The Fe K $\beta$  XES spectra of the three prepared

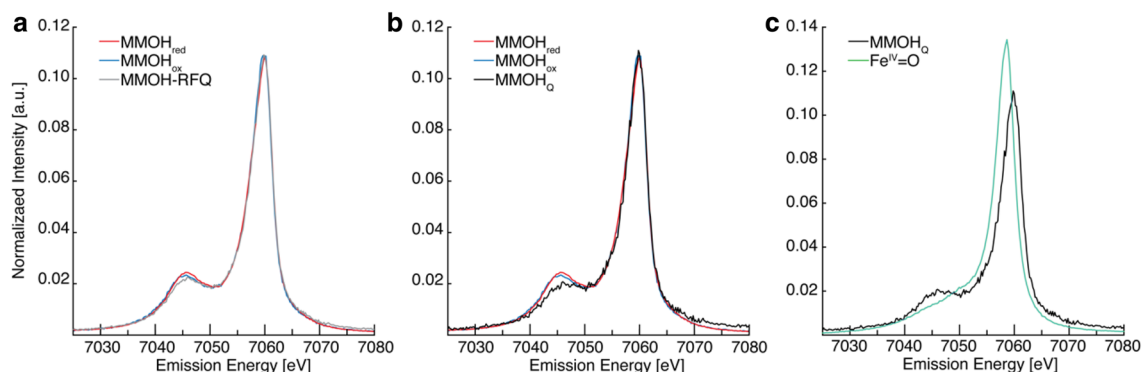
samples of sMMO ( $\text{MMOH}_{\text{red/ox/RFQ}}$ ) are shown in Fig. 2a. No significant differences in the  $\text{K}\beta_{1,3}$  maxima are apparent. One does, however, observe slight intensity differences in the  $\text{K}\beta'$  region (at  $\sim 7045$  eV) following a trend of  $\text{MMOH}_{\text{red}} > \text{MMOH}_{\text{ox}} > \text{MMOH-RFQ}$ . As the RFQ sample is a mixture of all three different components and therefore three oxidation states, an assignment of the convoluted spectrum is challenging. The individual components of the RFQ samples were quantitated by  $^{57}\text{Fe}$  Mössbauer spectroscopy, with an estimated 46% yield of  $\text{MMOH}_Q$  [24]. The  $^{57}\text{Fe}$  Mössbauer quantification of each component in the  $\text{MMOH-RFQ}$  sample allows for the ‘pure’  $\text{MMOH}_Q$  spectrum to be calculated as previously established in the HERFD XAS studies of sMMO and other non-heme di-iron proteins [23, 69].

The ‘pure’  $\text{MMOH}_Q$  mainline spectrum is shown in Fig. 2b, overlaid with  $\text{MMOH}_{\text{red}}$  and  $\text{MMOH}_{\text{ox}}$ . Here, one can clearly see that the  $\text{K}\beta_{1,3}$  peak position does not shift for the various oxidation states  $\text{Fe}^{\text{II}}_2/\text{Fe}^{\text{III}}_2/\text{Fe}^{\text{IV}}_2$  of sMMO. Often, the first moment of the  $\text{K}\beta_{1,3}$  is used rather than the peak maximum to determine energy shifts because the first moment has been shown to have more sensitivity to small changes [54, 70–72]. However, the first moment analysis of the three spectra, Table 2, does not reveal significant energy

shifts either, even though it was previously shown that all three of these samples exhibit clearly different XAS pre-edge and rising edge energies (see Figure S5 in Ref. [24]), consistent with their anticipated oxidation states and previous HERFD-XAS characterization [23].

While the  $\text{K}\beta_{1,3}$  may not exhibit clear differences with oxidation-state changes, the  $\text{K}\beta'$  intensity of the three states clearly varies, Fig. 2b. The  $\text{K}\beta'$  feature of  $\text{MMOH}_{\text{red}}$  appears slightly more intense than  $\text{MMOH}_{\text{ox}}$ . This is in agreement with the subtle intensity differences observed in other high-spin  $\text{Fe}^{\text{II}}$  versus  $\text{Fe}^{\text{III}}$  mainlines, such as  $[\text{Fe}(\text{H}_2\text{O})_6]^{2+/3+}$  [58].  $\text{MMOH}_Q$  has a clear  $\text{K}\beta'$  feature, indicating that there are unpaired d electrons ( $S_{\text{loc}} > 0$ ). The first moment of the  $\text{K}\beta'$  of  $\text{MMOH}_Q$  feature is at approximately the same energy as the more reduced iron species. In fact, the  $\text{K}\beta'$  features for each of the three  $\text{MMOH}$  states appear at approximately the same energies, with only the intensities of the  $\text{K}\beta'$  features varying, making the  $\Delta(\text{K}\beta_{1,3} - \text{K}\beta')$  splitting approximately the same for each species, Table 2.

The mainline of  $\text{MMOH}_Q$  may be the result of two local  $\text{Fe}^{\text{IV}}$  spin states: high-spin  $S = 2$ , or intermediate-spin  $S = 1$ . To determine the spin state, the mainline is directly compared to the  $\text{Fe}^{\text{IV}} S = 1$  mainline from an  $\text{Fe}^{\text{IV}} = \text{O}$  complex  $[\text{Fe}(\text{O})2\text{PyN}2\text{Q}]^{2+}$ , Scheme 1. Two immediate



**Fig. 2** **a** Fe  $\text{K}\beta$  mainline XES of  $\text{MMOH}_{\text{red}}$ ,  $\text{MMOH}_{\text{ox}}$  and  $\text{MMOH-RFQ}$  samples of sMMO. **b** The  $\text{MMOH}_Q$  spectrum is determined by subtraction of the  $\text{MMOH}_{\text{red}}$  and  $\text{MMOH}_{\text{ox}}$  components from the  $\text{MMOH-RFQ}$  spectrum. **c** The  $\text{MMOH}_Q$  XES is compared to that of an  $S = 1$   $\text{Fe}^{\text{IV}} = \text{O}$

**Table 2** Key Fe  $\text{K}\beta$  mainline parameters

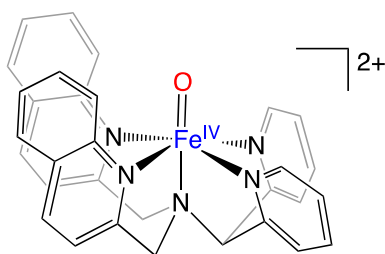
	$\text{K}\beta$ First Moment <sup>a</sup>	$\text{K}\beta_{1,3}$ First Moment <sup>b</sup>	$\text{K}\beta'$ First Moment <sup>c</sup>	$\Delta(\text{K}\beta_{1,3} - \text{K}\beta')$
$\text{MMOH}_{\text{red}}$	7054.94	7058.96	7043.59	15.37
$\text{MMOH}_{\text{ox}}$	7055.00	7059.00	7043.75	15.25
RFQ	7055.20	7059.05	7043.59	15.46
$\text{MMOH}_Q$	7055.45	7059.12	7043.43	15.69
$\text{Fe}(\text{IV}) = \text{O}$	7055.38	7058.16	7044.10	14.06

All values have energy units of eV

<sup>a</sup>Determined over entire measured  $\text{K}\beta$  mainline spectrum, 7025–7080 eV

<sup>b</sup>Determined over the energy range of 7051–7070 eV

<sup>c</sup>Determined over the energy range of 7025–7051 eV



**Scheme 1**  $\text{Fe}^{\text{IV}}=\text{O}; [\text{Fe}(\text{O})(2\text{PyN}2\text{Q})]^{2+}$

differences are observed in the spectra plotted in Fig. 2c. First, the  $\text{K}\beta_{1,3}$  energy for the  $\text{Fe}^{\text{IV}}=\text{O}$   $S=1$  model complex is clearly shifted to lower energy by approximately 1 eV compared to  $\text{MMOH}_Q$  and second, the  $\text{K}\beta'$  region for the  $\text{Fe}^{\text{IV}}=\text{O}$  model is significantly less intense than that of  $\text{MMOH}_Q$ . Furthermore, the same first moment analysis of the  $\text{K}\beta'$  of the  $\text{Fe}^{\text{IV}}=\text{O}$  mainline reveals an +1 eV compared to  $\text{MMOH}_Q$ , decreasing the  $\Delta(\text{K}\beta_{1,3}-\text{K}\beta')$  splitting by  $\sim 2$  eV, comparatively.

With respect to the  $\text{K}\beta'$  intensity of  $\text{MMOH}_Q$ , one may refer to the fingerprinting analyses performed for ferrous ion sites [57–60]. The presence of the  $\text{K}\beta'$  in the high-spin  $S=2$  and its absence in low-spin  $S=0$  centers is used as a spin-state diagnostic tool. For  $\text{MMOH}_Q$ , the clearly greater intensity of its  $\text{K}\beta'$  feature and larger  $\Delta(\text{K}\beta_{1,3}-\text{K}\beta')$  splitting compared to that of the established  $S=1$   $\text{Fe}^{\text{IV}}=\text{O}$  in Fig. 2c strongly suggests *higher* local spin states for the  $\text{Fe}^{\text{IV}}$  ions of  $\text{MMOH}_Q$ .

For the  $\Delta(\text{K}\beta_{1,3}-\text{K}\beta')$  splitting, it has been previously demonstrated that the energetic splitting is proportional to the nominal spin of the metal center, but these energy shifts may be further modulated by covalency [56]. However, the quantification of the covalency in the various states of  $\text{MMOH}$  is difficult to estimate. One may naturally consider a potential di-iron(IV) open core structure that contains terminal oxos to be more covalent than the  $\text{MMOH}_{\text{ox}}$  and  $\text{MMOH}_{\text{red}}$  states. However, the previous Mn  $\text{K}\beta$  mainline characterization of the step-wise deprotonation of high-spin  $\text{Mn}^{\text{IV}}$   $S=2$   $\text{Mn}_2(\mu\text{-OH})_2$  dimers to the  $\text{Mn}_2(\mu\text{-O})$  cores exhibited only minimal shifts ( $\sim 0.5$  eV) in the  $\text{K}\beta_{1,3}$  energies [64]. This suggests that when comparing potential oxo vs hydroxo ligation to the iron sites, the covalency changes will result in only minor energetic perturbations and that local iron spin state should be the dominant contribution to the  $\text{K}\beta$  mainline energies. Our analysis and comparison of the sMMO  $\text{K}\beta$  mainline lines clearly supports the conclusion that the local spin states of the iron sites of  $\text{MMOH}_Q$  are  $S=2$ . Furthermore, the  $\text{K}\beta$  mainline analysis of  $\text{MMOH}_Q$  as an  $S_{\text{loc}}=2$  is very consistent with the  $S_{\text{loc}}=2$  and  $S_{\text{loc}}=5/2$  spin states of the  $\text{MMOH}_{\text{red}}$  and  $\text{MMOH}_{\text{ox}}$ , respectively [13, 73].

## Discussion

While previous Mössbauer experiments of  $\text{MMOH}_Q$  provide evidence for isomer shifts in clear agreement with  $\text{Fe}^{\text{IV}}$  ions, the antiferromagnetic coupling and  $S_{\text{tot}}=0$  cluster spin state do not yield conclusive information of the local spin states of the iron sites. Here, we have used Fe  $\text{K}\beta$  XES to probe the local spin state of  $\text{MMOH}_Q$ . The  $\Delta\text{K}\beta$  splitting and the  $\text{K}\beta'$  intensity of  $\text{MMOH}_Q$  are excellent indicators of local high-spin  $S=2$  iron sites that antiferromagnetically couple to form the  $S_{\text{tot}}=0$  cluster.

The  $\text{K}\beta$  mainline of  $\text{MMOH}_Q$  has distinct features that also resemble other assigned  $S=2$   $\text{Fe}^{\text{IV}}$  centers [58, 74]. In a recent study of the Fe  $\text{K}\beta$  signatures of various compounds, the  $\text{K}\beta$  emission spectra of the iron perovskites  $\text{LaFe}^{\text{III}}\text{O}_3$  and  $\text{SrFe}^{\text{IV}}\text{O}_3$  were reported [58]. In  $\text{LaFe}^{\text{III}}\text{O}_3$ , a typical  $S=5/2$  ferric mainline is observed, whereas the high-spin  $S=2$   $\text{Fe}^{\text{IV}}$  mainline of  $\text{SrFe}^{\text{IV}}\text{O}_3$  exhibits a similar  $\text{K}\beta_{1,3}$  mainline maxima, but a slightly decreased  $\text{K}\beta'$  intensity, similar to that observed for  $\text{MMOH}_Q$ . However, we do note that this perovskite sample does not have the same molecular and/or electronic properties as the non-heme carboxylate-bridged di-iron center of sMMO, and so only qualitative comparisons are made.

Perhaps the most closely related structure to  $\text{MMOH}_Q$  is an RNR intermediate. The more widely studied di-iron RNR forms a high-valent  $\text{Fe}^{\text{III}}\text{-Fe}^{\text{IV}}$  intermediate termed ‘X,’ that has a  $S_{\text{tot}}=1/2$  cluster spin state as described earlier. Similarly, the hetero-metallic Mn–Fe class I-c of RNRs also forms a high-valent intermediate  $\text{Mn}^{\text{IV}}\text{-Fe}^{\text{IV}}$  intermediate [75]. For the  $d^4$  Fe center of the  $\text{Mn}^{\text{IV}}\text{-Fe}^{\text{IV}}$  RNR, the Fe  $\text{K}\beta$  mainline does not exhibit any clear energy shifts of the  $\text{K}\beta_{1,3}$  for the local  $\text{Fe}^{\text{II}}$ ,  $\text{Fe}^{\text{III}}$  and  $\text{Fe}^{\text{IV}}$  centers, and the  $\text{Fe}^{\text{IV}}$   $\text{K}\beta'$  exhibits a decreased intensity relative to the lower Fe oxidation-state intermediates [74]. The Fe  $\text{K}\beta$  XES of the  $S=2$   $\text{Fe}^{\text{IV}}$  center from Mn–Fe RNR has similar intensity to what we have observed for  $\text{MMOH}_Q$  (Figs. S3 and S4), lending additional support to the idea that measured  $\text{K}\beta$  XES of  $\text{MMOH}_Q$  reflects two  $S=2$   $\text{Fe}^{\text{IV}}$  ions and not a beam damaged product yielding high-spin  $S=5/2$  ferric sites, which would further increase  $\text{K}\beta'$  intensity and  $\Delta\text{K}\beta$  splitting.[25, 50] Furthermore, the  $\Delta\text{K}\beta$  splitting observed in RNR is similar to that seen here in  $\text{MMOH}_Q$ , Fig. S4. It is important again to note the clear presence of a  $\text{K}\beta'$  feature in the  $S=2$   $\text{Fe}^{\text{IV}}$  RNR compared to the lack of a well-resolved  $\text{K}\beta'$  in the  $S=1$   $\text{Fe}^{\text{IV}}=\text{O}$  model reported here (Fig. S4), offering further support that the  $\text{K}\beta$  mainline observed for  $\text{MMOH}_Q$  is diagnostic of an  $S=2$  local spin states at the iron atoms.

## Conclusion and outlook

In summary, the high-spin Fe<sup>II</sup>, Fe<sup>III</sup> and Fe<sup>IV</sup> sites of sMMO do not exhibit oxidation-state-dependent energy shifts by Fe K $\beta$  XES. Modest differences are observed in the K $\beta$ ' due to spin state. Notably, the  $\Delta K\beta(K\beta_{1,3}-K\beta')$  splitting of sMMO remains fairly constant, which is very consistent with high-spin states for MMOH<sub>ox</sub>, MMOH<sub>red</sub>, and MMOH<sub>Q</sub>. Direct comparison of the MMOH<sub>Q</sub> mainline to a known  $S=1$  Fe<sup>IV</sup> complex has shown dramatic spectral differences due to changes in the apparent spin state. This approach, in line with Fe K $\beta$  XES spin state fingerprinting for ferrous and ferric iron, clearly distinguishes between the intermediate  $S=1$  spin of the model complex studied here and the apparent  $S_{loc}=2$  of MMOH<sub>Q</sub>. The K $\beta$  XES experiment offers a clear experimental approach to determine local spin state in ambiguous systems, where magnetic spin coupling may make such determinations challenging for other experimental methods.

In the realm of X-ray spectroscopic studies, XES has become a viable tool to monitor spin state and/or oxidation states in both static samples and in situ or operando studies. Some of the clearest utility of Fe K $\beta$  XES has been to discriminate between low- versus high-spin iron species [56–60]. The lack of significant shifts in K $\beta_{1,3}$  mainline energies for the different oxidation states of MMOH intermediates precludes one from attempting more advanced X-ray spectroscopic techniques, such as “site selective” XAS or EXAFS for further enhancement of the MMOH<sub>Q</sub> signal in the mixed sample through the means of energy selective detection [76, 77]. The mainline spectra reported here will be valuable in future studies of sMMO. With the previous success of using Mn K $\beta$  emission to fingerprint oxidation state during in situ measurements [70, 78], one hopes to be able to utilize the Fe K $\beta$  XES to fingerprint, monitor and validate electronic changes during in situ studies. Such XES approaches have been highlighted in X-ray free electron laser (XFEL) protein diffraction studies to monitor a catalytic site's oxidation-state [79, 80]. The present results on the Fe K $\beta$  mainlines of sMMO, however, demonstrate the potential difficulty of employing such methods for monitoring oxidation-state during in situ studies, where the K $\beta_{1,3}$  energy does not offer great sensitivity but the K $\beta$ ' itself offers spin state (correlated to the oxidation-state) information through its clear intensity changes. Given the inherently lower intensity of the K $\beta$ ' feature, using this feature to follow spin state will require data collection with high signal-to-noise to ensure accurate assignments. We do note that the K $\alpha$  ( $2p \rightarrow 1s$ ) emission spectrum is an order of magnitude more intense than the K $\beta$  emission, allowing for easier measurements. K $\alpha$  XES does not offer the same degree of electronic sensitivity,

and extraction of oxidation or spin-state information is challenging [54, 58]. The three states of sMMO do not exhibit significant energy differences in their K $\alpha$  spectra (Figure S6 in ref [24].) and more recent K $\alpha$  XES of MMOH<sub>red</sub> and MMOH<sub>ox</sub> collected during XFEL crystallography experiments exhibits only subtle changes in the linewidths and shapes [81]. While more challenging to collect, K $\beta$  XES appears to offer the most sensitivity to spin state and the best potential fingerprint for formation of the di-iron(IV) ( $S_{loc}=2$ ) cluster.

Fe K $\beta$  XES of sMMO has proven to be a valuable technique to probe the local spin states of the di-iron site in the catalytic cluster. Understanding and verifying that MMOH<sub>Q</sub> is, in fact,  $S_{loc}=2$  is important to both understanding and proposing mechanisms for reactivity. Further, the ability to clearly distinguish  $S=1$  from  $S=2$  iron sites via K $\beta$  XES has potential for experimentally assessing the role of two-state reactivity in a wide range of high-valent Fe<sup>IV</sup>-oxo complexes [48, 82]. Hence, the present results deepen our understanding of the electronic structure of MMOH<sub>Q</sub>, while providing experimental fingerprints to enable the study of a wide range of high-valent iron species in both molecular models and enzymes.

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1007/s00775-022-01953-4>.

**Acknowledgements** Various members of the Inorganic Spectroscopy Department at the MPI CEC are thanked for helpful assistance with X-ray data collection. Bernd Meinert (MPI KoFo) is thanked for assistance in Mössbauer data collection. Financial support was provided by the Max Planck Society and the International Max Planck Research School (IMPRS) Recharge. J.D.L. acknowledge the National Institutes of Health for funding (GM118030). The XES experiments were performed on beamline ID-26 at the European Synchrotron Radiation Facility (ESRF), Grenoble, France. We are grateful to Dr. Blanka Detlefs, Dr. Lucia Amidani and Dr. Pieter Glatzel at the ESRF for providing assistance in using beamline ID-26.

**Funding** Open Access funding enabled and organized by Projekt DEAL.

**Data availability** The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

## Declarations

**Conflict of interest** The authors have no competing interests to declare that are relevant to the content of this article.

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## References

- Hanson RS, Hanson TE (1996) *Microbiol Rev* 60:439–471. <https://doi.org/10.1128/mr.60.2.439-471.1996>
- Ross MO, Rosenzweig AC (2017) *J Biol Inorg Chem* 22:307–319. <https://doi.org/10.1007/s00775-016-1419-y>
- Sirajuddin S, Rosenzweig AC (2015) *Biochemistry* 54:2283–2294. <https://doi.org/10.1021/acs.biochem.5b00198>
- Culpepper MA, Rosenzweig AC (2012) *Crit Rev Biochem Mol* 47:483–492. <https://doi.org/10.3109/10409238.2012.697865>
- Cutsail GE III, Ross MO, Rosenzweig AC, DeBeer S (2021) *Chem Sci* 12:6194–6209. <https://doi.org/10.1039/d1sc00676b>
- Jodts RJ, Ross MO, Koo CW, Doan PE, Rosenzweig AC, Hoffman BM (2021) *J Am Chem Soc* 143:15358–15368. <https://doi.org/10.1021/jacs.1c07018>
- Peng W, Qu XY, Shaik S, Wang BJ (2021) *Nat Catal* 4:266–273. <https://doi.org/10.1038/s41929-021-00591-4>
- Ross MO, MacMillan F, Wang J, Nisthal A, Lawton TJ, Olafson BD, Mayo SL, Rosenzweig AC, Hoffman BM (2019) *Science* 364:566–570. <https://doi.org/10.1126/science.aav2572>
- Ro SY, Schachner LF, Koo CW, Purohit R, Remis JP, Kenney GE, Liauw BW, Thomas PM, Patrie SM, Kelleher NL, Rosenzweig AC (2019) *Nat Commun* 10:2675. <https://doi.org/10.1038/s41467-019-10590-6>
- Chang WH, Lin HH, Tsai IK, Huang SH, Chung SC, Tu IP, Yu SS, Chan SI (2021) *J Am Chem Soc* 143:9922–9932. <https://doi.org/10.1021/jacs.1c04082>
- Christopher WK, Frank JT, He Y, Amy CR (2022) *Science* 375:1287–1291. <https://doi.org/10.1126/science.abm3282>
- Stanley SH, Prior SD, Leak DJ, Dalton H (1983) *Biotechnol Lett* 5:487–492. <https://doi.org/10.1007/Bf00132233>
- Fox BG, Hendrich MP, Surerus KK, Andersson KK, Froland WA, Lipscomb JD, Münck E (1993) *J Am Chem Soc* 115:3688–3701. <https://doi.org/10.1021/ja00062a039>
- Wallar BJ, Lipscomb JD (1996) *Chem Rev* 96:2625–2658. <https://doi.org/10.1021/cr9500489>
- Liu KE, Wang DL, Huynh BH, Edmondson DE, Salifoglou A, Lippard SJ (1994) *J Am Chem Soc* 116:7465–7466. <https://doi.org/10.1021/ja00095a083>
- Liu KE, Valentine AM, Wang DL, Huynh BH, Edmondson DE, Salifoglou A, Lippard SJ (1995) *J Am Chem Soc* 117:10174–10185. <https://doi.org/10.1021/ja00146a002>
- Tinberg CE, Lippard SJ (2011) *Acc Chem Res* 44:280–288. <https://doi.org/10.1021/ar1001473>
- Shu L, Nesheim JC, Kauffmann K, Münck E, Lipscomb JD, Que L Jr (1997) *Science* 275:515–518. <https://doi.org/10.1126/science.275.5299.515>
- Lee SK, Nesheim JC, Lipscomb JD (1993) *J Biol Chem* 268:21569–21577. [https://doi.org/10.1016/S0021-9258\(20\)80579-1](https://doi.org/10.1016/S0021-9258(20)80579-1)
- Lee SK, Fox BG, Froland WA, Lipscomb JD, Münck E (1993) *J Am Chem Soc* 115:6450–6451. <https://doi.org/10.1021/ja00067a086>
- Banerjee R, Proshlyakov Y, Lipscomb JD, Proshlyakov DA (2015) *Nature* 518:431–434. <https://doi.org/10.1038/nature14160>
- Banerjee R, Meier KK, Münck E, Lipscomb JD (2013) *Biochemistry* 52:4331–4342. <https://doi.org/10.1021/bi400182y>
- Castillo RG, Banerjee R, Allpress CJ, Rohde GT, Bill E, Que L Jr, Lipscomb JD, DeBeer S (2017) *J Am Chem Soc* 139:18024–18033. <https://doi.org/10.1021/jacs.7b09560>
- Cutsail GE III, Banerjee R, Zhou A, Que L Jr, Lipscomb JD, DeBeer S (2018) *J Am Chem Soc* 140:16807–16820. <https://doi.org/10.1021/jacs.8b10313>
- Jacobs AB, Banerjee R, Dewese DE, Braun A, Babicz JT, Gee LB, Sutherlin KD, Bottger LH, Yoda Y, Saito M, Kitao S, Kobayashi Y, Seto M, Tamasaku K, Lipscomb JD, Park K, Solomon EI (2021) *J Am Chem Soc* 143:16007–16029. <https://doi.org/10.1021/jacs.1c05436>
- Banerjee R, Jones JC, Lipscomb JD (2019) *Ann Rev Biochem* 88:409–431. <https://doi.org/10.1146/annurev-biochem-013118-111529>
- Rosenzweig AC (2015) *Nature* 518:309–310. <https://doi.org/10.1038/nature14199>
- Whittington DA, Lippard SJ (2001) *J Am Chem Soc* 123:827–838. <https://doi.org/10.1021/ja003240n>
- Schulz CE, Castillo RG, Pantazis DA, DeBeer S, Neese F (2021) *J Am Chem Soc* 143:6560–6577. <https://doi.org/10.1021/jacs.1c01180>
- Que L Jr (2007) *Acc Chem Res* 40:493–500. <https://doi.org/10.1021/ar700024g>
- Krebs C, Fujimori DG, Walsh CT, Bollinger JM Jr (2007) *Acc Chem Res* 40:484–492. <https://doi.org/10.1021/ar700066p>
- Sinnecker S, Svendsen N, Barr EW, Ye S, Bollinger JM Jr, Neese F, Krebs C (2007) *J Am Chem Soc* 129:6168–6179. <https://doi.org/10.1021/ja067899q>
- Jasniewski AJ, Que L Jr (2018) *Chem Rev* 118:2554–2592. <https://doi.org/10.1021/acs.chemrev.7b00457>
- Que L Jr, Tolman WB (2008) *Nature* 455:333–340. <https://doi.org/10.1038/nature07371>
- Sturgeon BE, Burdi D, Chen SX, Huynh BH, Edmondson DE, Stubbe J, Hoffman BM (1996) *J Am Chem Soc* 118:7551–7557. <https://doi.org/10.1021/ja960399k>
- Xue G, Wang D, De Hont R, Fiedler AT, Shan X, Münck E, Que L Jr (2007) *Proc Natl Acad Sci USA* 104:20713–20718. <https://doi.org/10.1073/pnas.0708516105>
- Xue GQ, Fiedler AT, Martinho M, Münck E, Que L Jr (2008) *Proc Natl Acad Sci USA* 105:20615–20620. <https://doi.org/10.1073/pnas.0808512105>
- Puri M, Que L Jr (2015) *Acc Chem Res* 48:2443–2452. <https://doi.org/10.1021/acs.accounts.5b00244>
- Stoian SA, Xue G, Bominaar EL, Que L Jr, Münck E (2014) *J Am Chem Soc* 136:1545–1558. <https://doi.org/10.1021/ja411376u>
- Riggs-Gelasco PJ, Price JC, Guyer RB, Brehm JH, Barr EW, Bollinger JM Jr, Krebs C (2004) *J Am Chem Soc* 126:8108–8109. <https://doi.org/10.1021/ja048255q>
- Martinho M, Xue G, Fiedler AT, Que L Jr, Bominaar EL, Münck E (2009) *J Am Chem Soc* 131:5823–5830. <https://doi.org/10.1021/ja8098917>
- Rohde JU, In JH, Lim MH, Brennessel WW, Bukowski MR, Stubna A, Münck E, Nam W, Que L Jr (2003) *Science* 299:1037–1039. <https://doi.org/10.1126/science.299.5609.1037>
- Klinker EJ, Kaizer J, Brennessel WW, Woodrum NL, Cramer CJ, Que L Jr (2005) *Angew Chem Int Ed* 44:3690–3694. <https://doi.org/10.1002/anie.200500485>
- Rasheed W, Draksharapu A, Banerjee S, Young VG, Fan R, Guo Y, Ozerov M, Nehr Korn J, Krzystek J, Telsler J, Que L Jr (2018) *Angew Chem Int Ed* 57:9387–9391. <https://doi.org/10.1002/anie.201804836>



45. England J, Martinho M, Farquhar ER, Frisch JR, Bominaar EL, Münck E, Que L Jr (2009) *Angew Chem Int Ed* 48:3622–3626. <https://doi.org/10.1002/anie.200900863>
46. Warm K, Paskin A, Kuhlmann U, Bill E, Swart M, Haumann M, Dau H, Hildebrandt P, Ray K (2021) *Angew Chem Int Ed* 60:6752–6756. <https://doi.org/10.1002/anie.202015896>
47. Pestovsky O, Stoian S, Bominaar EL, Shan XP, Münck E, Que L Jr, Bakac A (2005) *Angew Chem Int Ed* 44:6871–6874. <https://doi.org/10.1002/anie.200502686>
48. Swart M, Costas M (2016) *Spin states in biochemistry and inorganic chemistry*. John Wiley and Sons Ltd., West Sussex
49. Xue G, De Hont R, Münck E, Que L Jr (2010) *Nat Chem* 2:400–405. <https://doi.org/10.1038/nchem.586>
50. Valentine AM, Tavares P, Pereira AS, Davydov R, Krebs C, Hoffman BM, Edmondson DE, Huynh BH, Lippard SJ (1998) *J Am Chem Soc* 120:2190–2191. <https://doi.org/10.1021/ja974169x>
51. Gherman BF, Dunitz BD, Whittington DA, Lippard SJ, Friesner RA (2001) *J Am Chem Soc* 123:3836–3837. <https://doi.org/10.1021/ja0055108>
52. Siegbahn PEM (1999) *Inorg Chem* 38:2880–2889. <https://doi.org/10.1021/ic981332w>
53. Han W-G, Noodleman L (2008) *Inorg Chim Acta* 361:973–986. <https://doi.org/10.1016/j.ica.2007.06.007>
54. Glatzel P, Bergmann U (2005) *Coord Chem Rev* 249:65–95. <https://doi.org/10.1016/j.ccr.2004.04.011>
55. Castillo RG, Hahn AW, Van Kuiken BE, Henthorn JT, McGale J, DeBeer S (2021) *Angew Chem Int Ed* 60:10112–10121. <https://doi.org/10.1002/anie.202015669>
56. Pollock CJ, Delgado-Jaime MU, Atanasov M, Neese F, DeBeer S (2014) *J Am Chem Soc* 136:9453–9463. <https://doi.org/10.1021/ja504182n>
57. Lee N, Petrenko T, Bergmann U, Neese F, DeBeer S (2010) *J Am Chem Soc* 132:9715–9727. <https://doi.org/10.1021/ja101281e>
58. Lafuerza S, Carluantuo A, Retegan M, Glatzel P (2020) *Inorg Chem* 59:12518–12535. <https://doi.org/10.1021/acs.inorgchem.0c01620>
59. Mara MW, Hadt RG, Reinhard ME, Kroll T, Lim H, Hartsock RW, Alonso-Mori R, Chollet M, Glowina JM, Nelson S, Sokaras D, Kunnus K, Hodgson KO, Hedman B, Bergmann U, Gaffney KJ, Solomon EI (2017) *Science* 356:1276–1280. <https://doi.org/10.1126/science.aam6203>
60. Schuth N, Mebs S, Huwald D, Wrzolek P, Schwalbe M, Hemschemeier A, Haumann M (2017) *Proc Natl Acad Sci USA* 114:8556–8561. <https://doi.org/10.1073/pnas.1706527114>
61. Kowalska JK, Lima FA, Pollock CJ, Rees JA, DeBeer S (2016) *Isr J Chem* 56:803–815. <https://doi.org/10.1002/ijch.201600037>
62. Peng G, Degroot FMF, Hamalainen K, Moore JA, Wang X, Grush MM, Hastings JB, Siddons DP, Armstrong WH, Mullins OC, Cramer SP (1994) *J Am Chem Soc* 116:2914–2920. <https://doi.org/10.1021/ja00086a024>
63. Kowalska JK, Hahn AW, Albers A, Schiewer CE, Bjornsson R, Lima FA, Meyer F, DeBeer S (2016) *Inorg Chem* 55:4485–4497. <https://doi.org/10.1021/acs.inorgchem.6b00295>
64. Lassalle-Kaiser B, Boron TT, Krewald V, Kern J, Beckwith MA, Delgado-Jaime MU, Schroeder H, Alonso-Mori R, Nordlund D, Weng T-C, Sokaras D, Neese F, Bergmann U, Yachandra VK, DeBeer S, Pecoraro VL, Yano J (2013) *Inorg Chem* 52:12915–12922. <https://doi.org/10.1021/ic400821g>
65. Rees JA, Bjornsson R, Kowalska JK, Lima FA, Schlesier J, Sippel D, Weyhermüller T, Einsle O, Kovacs JA, DeBeer S (2017) *Dalton Trans* 46:2445–2455. <https://doi.org/10.1039/C7DT00128B>
66. Lambert C, Chernev P, Klingan K, Leidel N, Sigfridsson KGV, Happe T, Haumann M (2014) *Chem Sci* 5:1187–1203. <https://doi.org/10.1039/C3SC52703D>
67. Banerjee R, Komor AJ, Lipscomb JD (2017) *Methods Enzymol* 596:239–290. <https://doi.org/10.1016/bs.mie.2017.07.016>
68. Massie AA, Denler MC, Cardoso LT, Walker AN, Hossain MK, Day VW, Nordlander E, Jackson TA (2017) *Angew Chem Int Ed* 56:4178–4182. <https://doi.org/10.1002/anie.201612309>
69. Cutsail GE III, Blaesi EJ, Pollock CJ, Bollinger JM Jr, Krebs C, DeBeer S (2020). *J Inorg Biochem*. <https://doi.org/10.1016/j.jinorgbio.2019.110877>
70. Fransson T, Chatterjee R, Fuller FD, Gul S, Weninger C, Sokaras D, Kroll T, Alonso-Mori R, Bergmann U, Kern J, Yachandra VK, Yano J (2018) *Biochemistry* 57:4629–4637. <https://doi.org/10.1021/acs.biochem.8b00325>
71. Eeckhout SG, Safonova OV, Smolentsev G, Biasioli M, Safonov VA, Vykhodtseva LN, Sikora M, Glatzel P (2009) *J Anal Atom Spectr* 24:215–223. <https://doi.org/10.1039/B808345M>
72. Jensen SC, Davis KM, Sullivan B, Hartzler DA, Seidler GT, Casa DM, Kasman E, Colmer HE, Massie AA, Jackson TA, Pushkar Y (2017) *J Phys Chem Lett* 8:2584–2589. <https://doi.org/10.1021/acs.jpcclett.7b01209>
73. Hendrich MP, Münck E, Fox BG, Lipscomb JD (1990) *J Am Chem Soc* 112:5861–5865. <https://doi.org/10.1021/ja00171a029>
74. Martinie RJ, Blaesi EJ, Krebs C, Bollinger JM Jr, Silakov A, Pollock CJ (2017) *J Am Chem Soc* 139:1950–1957. <https://doi.org/10.1021/jacs.6b11563>
75. Jiang W, Hoffart LM, Krebs C, Bollinger JM Jr (2007) *Biochemistry* 46:8709–8716. <https://doi.org/10.1021/bi700906g>
76. Glatzel P, Jacquamet L, Bergmann U, de Groot FM, Cramer SP (2002) *Inorg Chem* 41:3121–3127. <https://doi.org/10.1021/ic010709m>
77. de Groot FMF (2000) *Top Catal* 10:179–186. <https://doi.org/10.1023/a:1019140823425>
78. Ibrahim M, Fransson T, Chatterjee R, Cheah MH, Hussein R, Lassalle L, Sutherlin KD, Young ID, Fuller FD, Gul S, Kim IS, Simon PS, de Lichtenberg C, Chernev P, Bogacz I, Pham CC, Orville AM, Saichek N, Northen T, Batyuk A, Carbajo S, Alonso-Mori R, Tono K, Owada S, Bhowmick A, Bolotovskiy R, Mendez D, Moriarty NW, Holton JM, Dobbek H, Brewster AS, Adams PD, Sauter NK, Bergmann U, Zouni A, Messinger J, Kern J, Yachandra VK, Yano J (2020) *Proc Natl Acad Sci USA* 117:12624–12635. <https://doi.org/10.1073/pnas.2000529117>
79. Bergmann U, Kern J, Schoenlein RW, Wernet P, Yachandra VK, Yano J (2021) *Nat Phys Rev* 3:264–282. <https://doi.org/10.1038/s42254-021-00289-3>
80. Kern J, Alonso-Mori R, Tran R, Hattné J, Gildea RJ, Echols N, Glöckner C, Hellmich J, Laksmono H, Sierra RG, Lassalle-Kaiser B, Koroidov S, Lampe A, Han G, Gul S, DiFiore D, Milathianaki D, Fry AR, Miahnahri A, Schafer DW, Messerschmidt M, Seibert MM, Koglin JE, Sokaras D, Weng T-C, Sellberg J, Latimer MJ, Grosse-Kunstleve RW, Zwart PH, White WE, Glatzel P, Adams PD, Bogan MJ, Williams GJ, Boutet S, Messinger J, Zouni A, Sauter NK, Yachandra VK, Bergmann U, Yano J (2013) *Science* 340:491–495. <https://doi.org/10.1126/science.1234273>
81. Srinivas V, Banerjee R, Lebrétte H, Jones JC, Aurelius O, Kim I-S, Pham CC, Gul S, Sutherlin KD, Bhowmick A, John J, Bozkurt E, Fransson T, Aller P, Butryn A, Bogacz I, Simon P, Keable S, Britz A, Tono K, Kim KS, Park S-Y, Lee SJ, Park J, Alonso-Mori R, Fuller FD, Batyuk A, Brewster AS, Bergmann U, Sauter NK, Orville AM, Yachandra VK, Yano J, Lipscomb JD, Kern J, Högbom M (2020) *J Am Chem Soc* 142:14249–14266. <https://doi.org/10.1021/jacs.0c05613>
82. Shaik S (2020) *Isr J Chem* 60:938–956. <https://doi.org/10.1002/ijch.202000002>

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