Spectrophotometric Determination of Pipazethate HCl and Dextromethorphan HBr using Potassium Permanganate

Ayman Abou El-Fetouh Gouda¹, Ragaa El-Sheikh¹, Zeineb. El Shafey², Nagda. Hossny², Rham El-Azzazy²

¹Chemistry Department, Faculty of Science, Zagazig University, Zagazig, Egypt; ²Chemistry Department, Faculty of Science (Girl's), Al-Azhar University, Cairo, Egypt

ABSTRACT

Rapid, simple and sensitive validated spectrophotometric methods have been described for the assay of pipazethate HCl (PiCl) and dextromethorphan HBr (DEX) either in pure form or in pharmaceutical formulations. The proposed methods were based on the oxidation of the studied drugs by a known excess of potassium permanganate in acidic medium and estimating the unreacted permanganate with amaranth dye (method A), acid orange II (method B), indigocarmine (method C) and methylene blue (method D), in the same acid medium at a suitable λ_{max} =521, 485, 610 and 664 nm, respectively. Beer's law is obeyed in the concentration range of 2.0-16 and 2.0-15 µg mL⁻¹ for PiCl and DEX, respectively with correlation coefficient (*n*=6) ≥ 0.9993. The apparent molar absorptivity and sandell sensitivity values are in the range 1.062-1.484 × 10⁴, 3.35-4.51 × 10⁴ L mol⁻¹ cm⁻¹ and 29.36-41.03, 8.21-11.06 ng cm⁻² for PiCl and DEX, respectively. Different variables affecting the reaction were studied and optimized. The proposed methods were applied successfully to the determination of the examined drugs either in a pure or pharmaceutical dosage forms with good accuracy and precision. No interferences were observed from excipients and the results obtained were in good agreement with those obtained using the official methods. (*Int J Biomed Sci* 2008; 4 (4): 294-302)

Keywords: spectrophotometry; pipazethate HCl; dextromethorphan HBr; potassium permanganate; oxidation reactions; pharmaceutical formulations

INTRODUCTION

Pipazethate hydrochloride (PiCl), 10*H*-pyrido[3,2*b*][1,4]benzothiadiazine-10-carboxylic acid 2-(2-piperidinoethoxy)ethyl ester (1) is a bronchodilator that suppresses irritative and spasmodic cough by inhibiting the excitability of the cough center and the peripheral neural receptors in the respiratory passage. The response to the drug takes about 10–20 min and lasts for 4–6 h (Scheme I). Pipazethate has been determined using a limited number of techniques including; spectrophotometry (2-6), TLC (7), HPLC (8), conductimetry (9) and ISE (10). PiCl was used in determination of Mo (VI) in alloy steels and soil samples (11).

Dextromethorphan hydrobromide (DEX), [(+)-3-Methoxy-17-methyl-9 α , 13 α , 14 α -morphinan hydrobromide monohydrate] is a cough suppressant, used for the relief of non-productive cough; it has a central action on the cough centre in the medulla (12) (Fig. 1). Different methods reported for the determination of DEX in bulk drug, in dosage forms with other drugs in cough-cold products and in biological samples. HPLC have been reported (13,

Corresponding author: Ayman Abou El-Fetouh Gouda, Chemistry Department, Faculty of Science, Zagazig University, Zagazig, Egypt. Tel: +20101892624; Fax: +20552308213; E-mail: aymanchimca@yahoo.com. **Received** October 5, 2008; **Accepted** October 31, 2008

Copyright: © 2008 Ayman Abou El-Fetouh Gouda et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.5/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.



Figure 1. The chemical structure of the studied drugs.

14), spectrophotometry (4, 5), first and second-derivative technique UV-spectrophotometry (15-18), capillary electrophoresis (19, 20), GC (21) and LC (22, 23).

The purpose of the present work is the development of a simple spectrophotometric method for the determination of anti-tussive drugs; pipazethate hydrochloride (PiCl) and dextromethorphan hydrobromide (DEX) in bulk and in their pharmaceutical formulations, based on the discoloring redox reaction with an excess of KMnO₄, used as self indicator and the determination of unreacted oxidant by the decrease in absorbance of the dyes; amaranth dye (AM) (method A), acid orange II (AO) (method B), indigocarmine (indigo) (method C) and methylene blue (MB) (method D).

EXPERIMENTAL

Materials

Pure pipazethate HCl (PiCl) and dextromethorphan HBr (DEX) were obtained from the Egyptian International Pharmaceutical Industries Company (EIPICO). All pharmaceutical preparations were obtained from commercial sources in the local market.

Reagent

All chemicals used were of analytical grade and all solvents were for spectroscopic grade.

An a stock aqueous solution of 1.0×10^{-3} M of amaranth (AM) (E-Merck, Germany), acid orange II (AO) (E-Merck, Germany), indigocarmine (Aldrich), and methylene blue (MB) (E-Merck, Germany) were prepared by dissolving an accurate weight of dye in least amount of water and completed to the mark in a 100 mL calibrated flask with water. A 5.0×10^{-4} M is prepared solution. The stock solutions of dyes were allowed to stand at room temperature for a few weeks without any significant decay.

A stock solution of 5.0×10^{-3} M KMnO₄ (Aldrich) was prepared by dissolving 0.079 g of KMnO₄ (Aldrich) in 10 mL of warm bidistilled water then completed to the mark in a 100 mL calibrated flask. Standardized using sodium oxalate (24) and kept in a dark bottle. A 5.0×10^{-4} M solution of KMnO₄ was prepared by diluting the previously stock solution with water and 2.0 M H₂SO₄ was prepared.

Apparatus

All the absorption spectral measurements were made using Kontron 930 (UV-Visible) spectrophotometer (German) with scanning speed 200 nm/min, and band width 1.0 nm equipped with 10 mm matched quartz cells.

Standard Solutions

Stock solutions (100 μ g mL⁻¹) of the studied drugs were freshly prepared daily by dissolving 10 mg of the drug in distilled water and then, completed to the mark in a 100 mL calibrated flask with distilled water. Working standard solutions were prepared by suitable dilution of the stock.

METHOD

Recommended procedures

Pipette a 1.0 mL aliquot of the examined drugs solution (100 µg mL⁻¹) into a series of 10 mL calibrated flasks, followed by acidification by adding 0.5 mL of 2.0 M H₂SO₄. Two mL and 1.5 mL of 5.0×10^{-4} M KMnO₄ were added and heated in a water bath at $60 \pm 2^{\circ}$ C for 5.0 min and 10 min with PiCl and DEX, respectively. The mixture was cooled to laboratory temperature, then 1.0, 1.5, 1.2 and 2.0 mL of (5.0×10^{-4} M) of AM, AO Indigo and MB, respectively with PiCl and DEX. The volume was completed to 10 mL with water. The decrease in color intensities were measured spectrophotometrically at their corresponding maximum wavelengths λ_{max} values, 521, 485, 610 and 664 nm for AM, AO, Indigo and MB, respectively. The concentration of each drug was determined from a calibration graph constructed under the same conditions.

Applications for pharmaceutical formulations

Procedure for tablets. The contents of twenty tablets of (Selgon tablets, 20 mg PiCl per tab. or Tussilar tablets, 10 mg DEX per tab.), were crushed powdered, weighed out and the average weight of one tablet was determined. An accurate weight equivalent to 10 mg of pure drug was dissolved in 20 mL distilled water and then filtered. The filtrate was diluted to 100 mL with distilled water in a 100 mL calibrated flask. This solution was further diluted stepwise to the request concentration with water and then analyzed by the recommended procedure.

Procedure for drops. The contents of five bottles (Selgon drops, 40 mg PiCl per mL or Tussilar drops, 1.0 g DEX per 15 mL) were mixed and the average volume for one bottle was determined. An aliquot of the solution equivalent to 10 mg PiCl and DEX was quantitatively transferred to 100 mL calibrated flask and made up to the mark with bidistilled water. The above stated procedures described were applied to determine drug concentrations.

RESULTS AND DISCUSSION

The optimum conditions for color development for each method were established by varying the parameters one at a time, keeping the others fixed and observing the effect produced on the absorbance of the colored species.

Absorption spectra

The spectrophotometric method for the determination of PiCl and DEX is based on their oxidation with a known excess of KMnO₄ in acidic medium and subsequent determination of residual oxidant by reacting it with fixed amount of amaranth dye (AM), acid orange II (AO), indigocarmine (indigo) and methylene blue (MB) show characteristics λ_{max} values at 521, 485, 610 and 664 nm for AM, AO, indigo and MB methods, respectively (Fig. 2).

Effect of heating time

In order to obtain the highest and most stable absorbance, the effect of heating time on the oxidation reaction of PiCl and DEX was catalyzed by heating in a water bath at $60 \pm 2^{\circ}$ C for the periods ranging for 2.5-20 min. the time required to complete the reaction and maximum absorbance was obtained after 5.0 min for PiCl and 10 min for DEX. After oxidation process, the solution must be cooled at least for 3.0 min before addition of dye (Fig. 3).



Figure 2. Absorption spectra of the oxidation product between 8.0 μ g mL⁻¹ PiCl, KMnO₄ and , (a) AM, (b) AO, (c) Indigo and (d) MB by heating at 60 ± 2°C for 5.0 min.



Figure 3. Effect of heating time on the oxidation of 8.0 μ g mL⁻¹ PiCl-dye at the optimum wavelengths λ_{max} of each dye.

Effect of oxidant concentration

When a study on the effect of KMnO_4 on color development was performed, it was observed that in both cases the absorbance increased with increase in the volume of KMnO_4 (5.0 × 10⁻⁴ M). It reached maximum when 2.0 ml and 1.5 ml of KMnO_4 solution was added to a total volume of 10 ml for PiCl and DEX, respectively. The color intensity decreased above the upper limits. Therefore, 2.0 ml and 1.5 ml of KMnO_4 were used for all measurements (Fig. 4).

Effect of acid concentration

To study the effect of acid concentration, different types of acids were examined (H_2SO_4 , H_3PO_4 and CH_3COOH) to achieve maximum yield of redox reaction. The results in-

dicated that the sulphuric acid was the preferable acid with KMnO_4 as oxidant. The reaction was performed in a series of 10 mL volumetric flask containing 8.0 µg mL⁻¹ of the cited drugs, different volumes (0.1–2.5 mL) of 2.0 M H₂SO₄ and 2.0 and 1.5 mL of KMnO₄ (5.0 × 10⁻⁴ M) with PiCl and DEX, respectively were added. After 5.0 min for PiCl and 10 min for DEX heating time at 60 ± 2°C in a water bath, the solution was cooled for about 3.0 min; the dyes (1.0, 1.5, 1.2 and 2.0 mL of AM, AO, indigo and MB, respectively) were added, then complete to 10 mL total volume with water. It was found that the maximum absorbance was obtained at 0.5 mL of 2.0 M H₂SO₄. Above this volume, the absorbance decreased for PiCl, where as for DEX the absorbance remained constant. Therefore, a volume of 0.5 ml of 2.0 M H₂SO₄, was used for all measurements (Fig. 5).

Effect of dye concentration

In order to ascertain the linear relationship between the volume of added KMnO_4 and the decrease in absorbance of AM, AO, Indigo and MB, experiments were performed using 0.5 mL of 2.0 M H₂SO₄ with varying volumes of KMnO_4 . The decrease in absorbance was found to be linear up to 2.0 and 1.5 mL of 5.0×10^{-4} M KMnO₄ with optimum volumes (1.0, 1.5, 1.2 and 2.0 mL of AM, AO, indigo and MB, respectively) for 8.0 µg mL⁻¹ of PiCl and DEX (Fig. 6). The color was found to be stable up to 24 h.

Stoichiometric ratio

Job's method of continuous variation and the molar ratio method described by Yoe and Jones (25, 26), was em-



Figure 4. Effect of volume of 5.0×10^{-4} M KMnO₄ on the development of the reaction product: 8.0 µg mL⁻¹ DEX with MB and 10 µg mL⁻¹ PiCl with AO.

ployed to determined the stoichiometry of drug, oxidant and dyes. Keeping the sum of the molar concentration of both fixed, the ratio of the concentrations of each two in the mixture was varied and the absorbances of the mixture were recorded at the suitable wavelength against reagent blank. The maximum absorbance corresponds to the stoichiometric ratio. Stoichiometric ratio was found to be 1:1 for drug to oxidant; drug to dyes and oxidant to dyes as shown in (Fig. 7) (Table 1).

Validation of the proposed methods

Linearity. At described experimental conditions for PiCl and DEX determination, standard calibration curves



Figure 5. Effect of mL added of Sulfuric acid (2.0 M) on absorbance of PiCl with (5.0 \times 10⁻⁴ M) KMnO4 and dyes (5.0 \times 10⁻⁴ M).



Figure 6. Effect of added dyes $(5.0 \times 10^{-4} \text{ M})$ on absorbance of 10 µg mL⁻¹ of PiCl with KMnO₄ $(5.0 \times 10^{-4} \text{ M})$.

for PiCl and DEX with $KMnO_4$ and dyes, were constructed by plotting absorbance's vs. concentrations. The statistical parameters were given in the regression equation calculated from the calibration graphs, along with the standard



Figure 7. Continuous variations graph for the reaction between 5.0×10^{-4} M PiCl and 5.0×10^{-4} M KMnO₄ with dyes (5.0×10^{-4} M).

deviations of the slope (S_b) and the intercept (S_a) on the ordinate and the standard deviation residuals (S_{yy}) .

The linearity of calibration graphs was proved by the high values of the correlation coefficient (r) and the small values of the y-intercepts of the regression equations. The apparent molar absorptivities of the resulting colored ion-pair complexes and relative standard deviation of response factors for each proposed spectrophotometric method were also calculated and recorded in Table 1. The molar absorptivity of D>C>B>A method for PiCl, while for DEX the molar absorptivity of D>A>C> B method.

Sensitivity. The detection limits (LOD) for the proposed methods were calculated using the following equation (27): LOD = 3s / k

where *s* is the standard deviation of replicate determination values under the same conditions as for the sample analysis in the absence of the analyte and *k* is the sensitivity, namely the slope of the calibration graph. In accordance with the formula, the detection limits were found to be 0.30, 0.16, 0.21 and 0.18 μ g mL⁻¹ for A, B, C and D methods, respectively. Whereas; for DEX the detection limits were found to be 0.09, 0.26, 0.37 and 0.21 μ g mL⁻¹ for A, B, C and D methods, respectively.

The limits of quantitation, LOQ, defined as (27); LOQ = 10 s / k

Devemeters	PiCl				DEX			
rarameters —		AO	Indigo	MB	AM	AO	Indigo	MB
λ_{max} , (nm)	522	610	664	485	520	610	664	485
Beer's law linits (µg mL ⁻¹)	2.0-16	2.0-14	2.0-14	2.0-10	2.0-10	4.0-12	4.0-15	4.0-10
Ringbom Limits (µg mL-1)	2.5-15	3.0-13	3.0-12.5	2.5-9.5	3.0-9.0	5.0-10.5	5.0-14	5.5-9.5
Molar absorptiivity $\times 10^4$ (L mol ⁻¹ cm ⁻¹)	1.062	1.193	1.37	1.484	4.2	3.35	3.72	4.51
Sandell sensitivity (ng cm ⁻²)	41.03	36.52	31.80	29.36	8.82	11.06	9.96	8.21
Regression equation ^a								
Slope (b)	0.0497	0.0571	0.0678	0.0661	0.1812	0.21	0.168	0.118
Intercept (a)	0.0051	0.0003	-0.0137	0.014	-0.446	-0.742	-0.521	-0.736
Correlation coefficient (<i>r</i>)	0.9999	0.9999	0.9997	0.9998	0.9990	0.9991	0.9994	0.9995
$S_{v/x}$	0.3118	0.2737	0.3384	0.2687	0.331	0.380	0.308	0.361
SD of slope (S_b)	0.2647	0.234	0.32	0.2317	0.105	0.120	0.097	0.114
SD of intercept (S_a)	0.0249	0.0261	0.0339	0.0469	1.64	2.15	1.96	2.05
Detection limits (µg mL ⁻¹)	0.30	0.16	0.21	0.18	0.09	0.26	0.37	0.21
Quantification limit (µg mL-1)	0.99	0.53	0.69	0.60	0.30	0.87	1.23	0.70
RSD ^b %	0.79	0.83	0.68	0.54	0.92	1.04	0.58	0.79
t ^c	0.122	0.18	0.118	0.206	0.091	0.791	0.618	0.301
F °	1.895	1.756	2.394	1.28	1.058	2.40	1.435	1.03

Table 1. Analytical parameters and optical characteristics of the proposed methods with PiCl and DEX

 $^{a}A = a + b C$, where C is the concentration in μ g mL⁻¹; $^{b}Average of six determinations; <math>^{c}Calculated t$ - and F-value; tabulated t and F-value for five degrees of freedom; p=0.05 are 2.57 and 5.05.

According to this equation, the limit of quantitation were found to be 0.99, 0.53, 0.69 and 0.60 μ g mL⁻¹ for A, B, C and D methods, respectively. Whereas; for DEX the detection limits were found to be 0.30, 0.87, 1.23 and 0.70 μ g mL⁻¹ for A, B, C and D methods, respectively.

Specificity, Precision, and Accuracy. Specificity of Oxidation-reduction reaction and selective determination of PiCl and DEX with KMnO₄ and dyes could be possible. Percentage relative standard deviation (RSD %) as precision and percentage relative error (Er %) as accuracy of the suggested method were calculated. Precision was carried out by six determinations at four different concentrations in these spectrophotometric methods. The percentage relative error calculated using the following equation:

 $\text{Er }\% = [(\text{founded} - \text{added}) / \text{added}] \times 100$

The inter-day precision and accuracy results are shown in (Table 2). These results of accuracy and precision show that the proposed methods have good repeatability and reproducibility.

Robustness and Ruggedness. For the evaluation of method robustness, some parameters were interchanged; $KMnO_4$ concentration, dye concentration, wavelength range, and heating time. The capacity remained unaffected by small deliberate variations. Method ruggedness

was expressed as RSD % of the same procedure applied by two analysts and in two different instruments on different days. The results showed no statistical differences between different analysts and instruments suggesting that the developed methods were robust and rugged.

Interferences

In pharmaceutical analysis, it is important to test the selectivity towards excipients and additives added to the pharmaceutical preparations of PiCl and DEX. It is clear from the results obtained for the pharmaceutical preparations that the commonly encountered excipients such as starch, talc, glucose, alginate and stearate did not interfere indicating a high selectivity for determining the studied PiCl and DEX in its dosage forms.

Pharmaceutical applications

The proposed methods were successfully applied to determine the drugs studied PiCl and DEX in tablets and drops. Six replicate determinations were made. Moreover, to check the validity of the proposed methods, dosage forms were tested for possible interference with standard addition method. There was no significant difference between slopes of calibration curves and standard addition methods at four methods. Therefore it is

			PiCl		DEX			
Method	Added (μg mL ⁻¹)	Recovery %	Precision RSD % ^a	Accuracy Er %	Taken (µg mL ⁻¹)	Recovery %	Precision RSD % ^a	Accuracy Er %
А	4.0	100.1	0.84	0.10	2.0	100.09	0.68	0.09
	8.0	99.57	0.92	-0.425	4.0	99.92	0.90	-0.08
	12	99.95	0.77	-0.05	6.0	99.70	0.88	-0.30
	16	100.2	1.08	0.20	8.0	100.15	1.17	0.15
В	4.0	99.72	0.99	-0.275	4.0	99.20	0.76	-0.80
	6.0	99.15	0.85	-0.85	6.0	100.20	0.57	0.20
	8.0	98.94	1.12	-1.06	8.0	99.90	0.66	-0.10
	10	99.45	0.88	-0.55	10	100.31	0.39	0.31
С	3.0	100.12	0.76	0.133	4.0	99.70	1.08	-0.30
	6.0	99.85	0.79	-0.15	6.0	98.92	0.54	-1.08
	9.0	99.92	0.81	-0.078	9.0	99.30	0.45	-0.70
	12	100.15	1.23	0.15	12	100.10	0.61	0.10
D	2.0	100.07	1.06	0.07	4.0	100.67	0.57	0.67
	4.0	99.25	1.42	-0.75	5.0	100.50	0.72	0.50
	6.0	101.17	0.96	1.17	6.0	99.78	0.69	-0.22
	8.0	99.83	0.88	-0.17	8.0	100.08	0.94	0.083

Table 2. Evaluation of accuracy and precision data for PiCl and DEX obtained by the proposed methods

^aMean of six determination. RSD%, percentage relative standard deviation; Er%, percentage relative error.

concluded that the excipients in pharmaceutical dosage forms of PiCl and DEX such as starch, lactose, glucose, sugar, talc, sodium chloride, titanium dioxide, and magnesium stearate were not found any interference in the analysis of PiCl and DEX. The results were compared statistically by student's t- test (for accuracy) and variance ratio F- test (for precision) with official methods at 95% confidence level with five degrees of freedom (Table 3, 4). The results showed that the t and F- values were less than the critical value (27) indicating that there were no significant differences between the proposed and official methods. Because the proposed methods

		Selg	on tablets (20 mg/	tab.)	Selgon drops (40 mg/mL)			
Method	Taken (µg mL ⁻¹)	Added (ug mL ⁻¹)	Recovery ^a	Reference method	Added (ug mL ⁻¹)	Recovery ^a	Reference method	
А	4		99.80			97.85		
		2.0	100.50		2.0	99.50		
		4.0	100.35		4.0	101.00		
		6.0	100.20		6.0	100.25		
		8.0	99.65		8.0	100.60		
		10	100.80		10	99.25		
	Mean ^a \pm SD (p=0.05)		100.22 ± 0.832	99.70 ± 1.16		100.08 ± 0.67	100.50 ± 0.63	
	t ^b		0.364			0.457		
	F ^b		1.944			1.13		
В	2		99.51			98.95		
		2.0	99.96		2.0	99.50		
		4.0	100.01		4.0	100.40		
		6.0	99.10		6.0	100.55		
		8.0	99.85		8.0	99.50		
		10	100.30		10	99.90		
	Mean ^a \pm SD (p=0.05)		99.97 ± 0.94	99.70 ± 1.16		99.80 ± 0.61	100.50 ± 0.63	
	t ^b		0.181			0.798		
	F ^b		1.52			1.07		
С	2		99.60			99.35		
		2.0	100.45		2.0	99.70		
		4.0	100.15		4.0	100.40		
		6.0	100.30		6.0	99.20		
		8.0	99.85		8.0	100.50		
		10	99.60		10	99.75		
	Mean ^a \pm SD (p=0.05)		99.99 ± 0.86	99.70 ± 1.16		99.82 ± 0.53	100.50 ± 0.63	
	t ^b		1.444			0.826		
	F^{b}		1.82			1.413		
D	1.0		99.89			99.25		
		1.0	99.95		1.0	100.40		
		2.0	100.40		2.0	99.50		
		4.0	100.05		4.0	100.70		
		6.0	100.20		6.0	100.25		
		8.0	99.75		8.0	100.60		
	Mean ^a \pm SD (p=0.05)		100.04 ± 1.232	99.70 ± 1.16		100.12 ± 0.601	100.50 ± 0.63	
	t ^b		0.201			0.436		
	F^{b}		1.128			1.10		

 Table 3. Application of the standard addition technique for the determination of PiCl in dosage forms using the proposed methods

^aAverage of six determinations; ^bCalculated *t*- and *F*-value; tabulated *t* and *F*-value for five degrees of freedom; *p*=0.05 are 2.57 and 5.05.

		Tus	silar tablets (10 m	g/tab.)	Tussilar drops (1.0 g/15 mL)		
Method	Taken (μg mL ⁻¹)	Added (μg mL ⁻¹)	Recovery ^a %	Reference method	Added (µg mL-1)	Recovery ^a %	Reference method
А	4		99.80			97.85	
		2.0	100.50		2.0	99.50	
		4.0	100.35		4.0	101.00	
		6.0	100.20		6.0	100.25	
		8.0	99.65		8.0	100.60	
		10	100.80		10	99.25	
	Mean ^a \pm SD (p=0.05)		100.22 ± 0.432	99.92 ± 0.85		100.08 ± 0.67	100.18 ± 0.81
	t ^b		0.026				
	F^{b}		1.106				
В	2		99.51			98.95	
		2.0	99.96		2.0	99.50	
		4.0	100.01		4.0	100.40	
		6.0	99.10		6.0	100.55	
		8.0	99.85		8.0	99.50	
		10	100.30		10	99.90	
	Mean ^a \pm SD (p=0.05)		99.97 ± 0.423	99.92 ± 0.85		99.80 ± 0.61	100.18 ± 0.81
	t ^b		0.296			0.798	
	F^{b}		1.27			1.07	
С	2		99.60			99.35	
		2.0	100.45		2.0	99.70	
		4.0	100.15		4.0	100.40	
		6.0	100.30		6.0	99.20	
		8.0	99.85		8.0	100.50	
		10	99.60		10	99.75	
	Mean ^a \pm SD (p=0.05)		99.99 ± 0.36	99.92 ± 0.85		99.82 ± 0.53	100.18 ± 0.81
	t ^b		0.09			0.826	
	$F^{\mathfrak{b}}$		1.43			1.413	
D	1.0		99.63			100.04	
		1.0	99.91		1.0	99.50	
		2.0	100.16		2.0	100.40	
		4.0	99.10		4.0	100.55	
		6.0	99.85		6.0	99.95	
		8.0	100.20		8.0	99.90	
	Mean ^a \pm SD (p=0.05)		99.81 ± 0.41	99.92 ± 0.85		100.06 ± 0.38	100.18 ± 0.81
	t ^b		0.302			0.436	
	F^{b}		1.067			1.10	

Table 4 Application of the stondard addition to shall one for the	determination of DEV in decade former using the	a
Table 4. Application of the standard addition technique for the	determination of DEA in dosage forms using th	e proposed methods
		- p p

^aAverage of six determinations; ^bCalculated *t*- and *F*-value; tabulated *t and F*-value for five degrees of freedom; *p*=0.05 are 2.57 and 5.05.

Step 1:

 $\begin{array}{l} Durgs + KMnO_4 \rightarrow oxidation \ products \ of \ drugs + unreacted \ KMnO_4 \\ [excess] \end{array}$

Step 2:

unreacted $\text{KMnO}_4 + \text{Dyes} \rightarrow \text{oxidation products of dyes} + \text{unreacted dyes}$ (colored) measured spectrophotometrically

Figure 8. The possible sequences of Oxidation-reduction reaction

were more reproducible with high recoveries they can be recommended for routine analysis in majority of drug quality control laboratories.

Chemistry of colored species

The proposed methods are based on the oxidation of the cited drugs by excess of KMnO_4 to form oxidation products besides unreacted KMnO_4 (step 1), and followed by the determination of unreacted KMnO_4 by measuring the decrease in the absorbance of AM, AO, Indigo and MB dyes at their λ_{max} (step 2). The possible sequences of reactions are presented in Fig. 8.

CONCLUSION

The order of λ_{max} values among the proposed methods for the determination of the cited drugs is D>C>A>B. The higher λ_{max} of the visible spectrophotometric methods over reported UV and visible spectrophotometric methods is decisive and advantageous since interference from the excipients should be far less at higher wavelengths. The proposed methods are accurate and precise as indicated by good recoveries of the drugs and low RSD values. The proposed methods can be applied for routine analysis and in quality control laboratories for quantitative determination of the cited drugs both in the pure and dosage forms.

REFERENCES

- Martindale, The Extra Pharmacopoeia, ed. Reynolds JEF. London: Pharmaceutical Press. 30th edn. 1999; 8140.
- 2. Zarapker SS, Rele RV, Shah VM. Indian Drugs. 1987; 24: 445-449.
- 3. Zarapker SS, Rele RV, Doshi VJ. Indian Drugs. 1987; 24: 560-564.
- El-Shiekh R, Zahran F, Gouda AAEF. Spectrochimica Acta. A. 2007; 66: 1279-1287.
- Amin AS, El-Sheikh R, Zahran F, Gouda AAEF. Spectrochimica Acta. A. 2007; 67: 1088-1093.
- 6 El-Shiekh R, Amin AS, Zahran F, Gouda AAEF. J. AOAC Int. 2007; 90: 686-692.
- 7. Revanasiddappa HD, Ramappa PG. Indian Drugs. 1995; 32: 73-77.
- 8. Revanasiddappa HD, Ramappa PG. Indian Drugs. 1995; 32: 534-536.
- 9. Abdel-Ghani NT, Shoukry AF, El-Nashar RM. Analyst. 2001; 126: 79-85.
- Issa TM, Shoukry AF, El-Nashar RM. J. Pharm. Biomed. Anal. 2001; 26: 379-386.
- Melwanki MB, Seetharamappa J, Masti SP. Anal. Sci. 2001; 17: 1121-1124.
- United States Pharmacopoeia, 25th Review, The National Formulary, 19th Review, The United States Pharmacopoeia Convention, Rockville, MD. 2002; 975.
- 13. Shervington LA. Anal. Lett. 1999; 30: 927-944.
- Bendriss EK, Markoglou N, Wainer IW. J. Chromatogr. B: Biomed. Sci. Appl. 2001; 754: 209-215.
- Tantishaiyakul V, Poeaknapo C, Sribun P, Sirisuppanon K. J. Pharm. Biomed. Anal. 1998; 17: 237-243.
- 16. Ajay S, Piyush T. Indian Drugs. 1999; 36: 444-448.
- 17. Gangwal S, Trivedi P. Indian Drugs. 1999; 36: 568-571.
- Bratio MS, Kaskhedikar SG, Chaturvedi SC. Indian Drugs. 1999; 36: 702-705.
- 19. Suntornsuk L. Electrophoresis. 2001; 22: 139-143.
- Gomez MR, Olsina RA, Martinez LD, Silva MF. J. Pharm. Biomed. Anal. 2002; 30: 791-799.
- 21. Argekar AP, Raj SV, Kapadia SU. Indian Drugs. 1998; 35: 24-28.
- Jones DR, Gorski JC, Hamman MA, Hall SD. J. Chromatogr. B: Biomed. Sci. Appl. 1996; 678: 105-111.
- Bolden RD, Hoke SH, Eichhold TH, McCauley-Myers DL, et al. J. Chromatogr. B: Analyt. Technol. Biomed. Life Sci. 2002; 772: 1-10.
- Basset Denny, Jeffery Mendham JRCGH. Vogel Text Book of Quantitative Inorganic analysis. *London: Imperical Collage*. 4th Edn. 1986; 350.
- 25. Yoe jh, Jones al. Ind. End. Chem. Anal. 1944; 16: 111.
- 26. Job P. Ann. Chim. 1928; 9: 113.
- 27. Miller JN, Miller JC. Statistics and Chemometrics for Analytical Chemistry. 5th ed. *England: Prentice Hall.* 2005.