## CONCISE COMMUNICATIONS

# Ineffectiveness of Recombinant Interferon- $\beta_{serine}$ Nasal Drops for Prophylaxis of Natural Colds

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Two randomized, double-blind, placebo-controlled trials during early autumn of 1986 and 1987 evaluated the efficacy and tolerance of recombinant interferon- $\beta_{serine}$  (rIFN- $\beta_{ser}$ ) nasal drops for prevention of natural rhinovirus colds. In 1986,  $9 \times 10^6$  units of rIFN- $\beta_{ser}$  (139 subjects) or placebo (157) were adminstered once daily except Sundays for 4 w. Rhinovirus colds occurred in 2.8% of rIFN- $\beta_{ser}$  recipients and 6.0% of placebo recipients during the treatment period (52% reduction, P = .3). In 1987,  $24 \times 10^6$  units of rIFN- $\beta_{ser}$  (186) or placebo (197) were given daily for 25 consecutive days. Rhinovirus colds developed in 6.3% of rIFN- $\beta_{ser}$  recipients and 5.3% of placebo recipients. In each study, illness frequency and number of days with subjective colds did not differ between the groups. Recipients of nasal drops of rIFN- $\beta_{ser}$  at either dosage did not differ in tolerance from placebo recipients. The lack of both prophylactic efficacy and nasal toxicity are in contrast to prior observations with nasal sprays of rIFN- $\alpha_{ab}$ .

Seasonal prophylaxis studies have demonstrated that nasal sprays of recombinant interferon- $\alpha_{2b}$  (rIFN- $\alpha_{2b}$ ) are effective in preventing natural rhinovirus infections but are associated with excess rates of nasal irritation [1–4]. Recombinant interferon- $\beta_{serine}$ (rIFN- $\beta_{ser}$ ) has comparable antiviral activity against rhinovirus and coronavirus under in vitro conditions [5, 6]. A tolerance study [7] using 12 × 10<sup>6</sup> units/d for 25 d as a nasal spray found that rIFN- $\beta_{ser}$  appears to be better tolerated than previously studied alpha interferons. Two recent studies [7, 8] have shown rIFN- $\beta_{ser}$ , 6–10 × 10<sup>6</sup> units/d, to be effective in the prophylaxis of experimental rhinovirus colds. Consequently, we evaluated the efficacy of

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Written informed consent was obtained from all participants, and the guidelines for human experimentation of the U.S. Department of Health and Human Services and those of the University of Virginia Human Investigation Committee were followed in the conduct of these studies.

This work was supported by grants from Richardson-Vicks Research Center, Shelton, Connecticut; Triton Biosciences, Inc., Alameda, California; and by grant T32AI-07046 from the National Institute of Allergy and Infectious Diseases. rIFN- $\beta_{ser}$  nasal drops for preventing natural colds in two randomized, double-blind, placebo-controlled seasonal prophylaxis trials. In the first study, conducted during the fall of 1986, subjects receiving  $9 \times 10^6$  units/d (except Sundays) for 4 w tended to have fewer rhinovirus infections than placebo recipients. A subsequent 25-d, dose-rising tolerance trial revealed no increase in nasal examination abnormalities with daily doses as high as  $24 \times 10^6$  units (S. Sperber, F. Hayden, unpublished data). The second efficacy study was conducted in 1987 at this higher dosage.

#### **Materials and Methods**

Subjects. Healthy adult volunteers were recruited in Charlottesville from late August to early Septem-

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ber of 1986 and 1987 according to previously published criteria [7].

Interferon administration. Lyophilized rIFN- $\beta_{ser}$  (Triton Biosciences, Alameda, Calif) and a placebo with identical appearance, protein content, and pH were reconstituted daily. These solutions were transferred to sterile screw-cap vials, refrigerated or held on ice, and dispensed within 5–6 h. Neither the interferon nor the placebo contained preservative. Each vial served as the source of drug or placebo for 10 subjects. To ensure that active drug was administered, three vials of lyophilized rIFN- $\beta_{ser}$  from the same batch as used in the 1987 study that had been retained by the supplier were reconstituted in the same manner as above and found by Triton Biosciences to have the anticipated interferon activity within the limits of the assay.

Subjects reported each morning (except Sundays in the 1986 study) to one of two study sites for drug administration and symptom recording. Nasal drops were delivered by a study nurse with a calibrated pipette. The subjects were supine with the nasal passages in a vertical plane, and performed three 10-s head-turning maneuvers to help distribute the drops within the nasal passages. In 1986, rIFN- $\beta_{ser}$  9  $\times$ 10<sup>6</sup> units or placebo was administered daily except Sundays for 4 w as nasal drops (150 µl/nostril). In 1987, rIFN- $\beta_{ser}$  24  $\times$  10<sup>6</sup> units or placebo was administered as nasal drops (200 µl/nostril) for 25 consecutive d. Subjects were removed from the studies if they missed more than two doses or failed to appear on a Saturday and the following Monday in the 1986 study.

Monitoring of illness and infection. The presence and severity of colds and specific symptoms were determined using previously described methods [9]. A respiratory illness episode was defined as the presence of at least one respiratory symptom (excluding sneezing) on two consecutive days or two or more respiratory symptoms on one day. Nasal washings and throat swabs were processed for viral isolation [7, 9] if the subject reported a cold or met the criteria for a respiratory illness episode. Daily recording of symptoms was continued in 1987 by each subject at home for 2 w after the completion of drug administration; symptomatic participants were asked to call the project staff and virus cultures were obtained.

Monitoring of toxicity. Symptoms of nasal intolerance (table 1) and their severity were recorded daily. Rhinoscopic examinations were performed before and at the conclusion of the studies in all subjects and after 2 w of treatment in the 1987 study. Blood and urine samples were collected before initiating drug administration and during the last week of treatment for routine hematology, blood chemistries, and urinalysis studies.

Serum samples were obtained before and 2 w after the completion of drug administration for determination of IgG and IgM against rIFN- $\beta_{ser}$  by ELISA (performed in the laboratory of Dr. Anne Teitelbaum, Triton Biosciences). Samples were obtained from all subjects during the 1986 study and from a subset of 51 rIFN- $\beta_{ser}$  and 15 placebo recipients in 1987. Samples that tested positive were then tested for interferon neutralizing activity in a standard bioassay.

Peripheral blood 2'-5'oligoadenylate synthetase activity. During the 1987 study, heparinized blood samples from nine interferon and nine placebo recipients were collected for determination of 2'-

Year, group (n)	Symptoms*					
	Blood-tinged mucus	Burning	Dryness	Rhinoscopic signs <sup>†</sup>		
				Bleeding	Erosion	Ulcer
1986						
rIFN-β <sub>ser</sub> (138)	1	4	4	13	1	1
Placebo (149)	1	5	7	7	5	1
1987						
rIFN-β <sub>ser</sub> (186)	5	3	9	7	2	3
Placebo (197)	3	4	9	6	3	2

**Table 1.** Symptoms and rhinoscopic signs of nasal intolerance during 4-week prophylaxis with rIFN- $\beta_{ser}$  nasal drops by percentage of subjects.

\* Cumulative frequency during 4-w period.

<sup>†</sup> Cumulative frequency based on nasal examination findings at 2 and 4 w in 1987; 4 w only in 1986. 5'oligoadenylate synthetase activity [10], a marker of interferon activity, before initiation of drug administration and 2 w into treatment,  $\sim$ 24 h after a preceding dose. The assays were conducted by Amina Woods in the laboratory of Dr. Paul Lietman, Johns Hopkins University, Baltimore, using methods previously described [10].

Data analysis. Outcome parameters (symptom occurrence, illness episodes, and viral isolates) were considered evaluable for efficacy analysis if they occurred during the period beginning 2 d after initiation of medication through day two after completion of prophylaxis (figure 1, diagonal bar). Treatment groups were compared with analysis of variance. Proportions were compared with Fisher's exact test.

### Results

Subjects. In the 1986 study, 296 subjects were enrolled. Of these, 287 (138 rIFN- $\beta_{ser}$ , 149 placebo) completed at least 22 d of drug administration. Of the nine who dropped out (1 rIFN- $\beta_{ser}$ , 8 placebo), none was a result of adverse drug effects. The 1986 study was originally started with the preservative thimerosal added to the diluent. During symptom assessment on the second day of drug administration, many subjects complained of a foul taste or odor or nausea that lasted up to several hours after receiving the nasal drops. Further administration of the drug was halted after about half the subjects had received the second dose. Evaluation revealed that

14 13 12 = Placebo 11 10 = Period of efficacy nalysi 8 Number of Rhinovirus Colds 6 5 3 2 °7 16 18 20 22 24 26 28 6 10 12 14 2 8 ż Treatment Day Post-Treatment Day 15 14 13-В 12 11 10 9 Number of Rhinovirus 8 = Placebo Colds 7 ··· = rIFN - 8se Period of efficacy analysis 5 4 3 0 10 12 14 16 18 20 22 24 ź 4 6' 8 10 12 14 á n 2 à Treatment Day Post - Treatment Day

Figure 1. Cumulative incidence of rhinovirus colds during seasonal prophylaxis with rIFN- $\beta_{ser}$  and placebo. A, Fall 1986. B, Fall 1987, including 2-w follow-up period after prophylaxis. Diagonal bar = period for analysis of efficacy from 2 d after initiation of medication through 2 d after completion of prophylaxis.

the thimerosal content was 10-fold higher than the 10 ppm planned. All subjects were asymptomatic the next morning and the study was restarted without preservative after a drug-free period of 1-2 d. This first day of preservative-free drug was considered to be day 1 of drug administration, and drug was given for the complete 4-w period. No subjects withdrew from the study as a result of this problem.

In the 1987 study, 383 volunteers were enrolled. Of these, 363 (174 rIFN- $\beta_{ser}$ , 189 placebo) completed at least 23 d of drug administration and were considered evaluable for determination of efficacy. Seven subjects (4 rIFN- $\beta_{ser}$ , 3 placebo recipients) were removed from study at 2 w because of abnormalities on rhinoscopic examination. Each study group was comparable with respect to mean age (30–31 y), gender (male:female ratio, 0.4), and current smokers (22%–23%).

**Respiratory illness.** No differences in the frequency of respiratory illness episodes in recipients of rIFN- $\beta_{ser}$  compared with placebo were found in 1986 (32% vs. 30%) or in 1987 (32% vs. 31%). Similarly, when illness episodes were defined by the volunteer's subjective impression that he or she had a cold, the treatment groups did not differ in 1986 (15% vs. 17%) or in 1987 (20% vs. 21%). The distribution of respiratory illnesses over the treatment course was also similar for the two groups. During the 2-w period after completion of drug administration in 1987, the frequency of subjective colds (rIFN- $\beta_{ser}$ 3%, placebo 6%) and respiratory illness episodes (rIFN- $\beta_{ser}$  10%, placebo 8%) did not differ between groups.

For all subjects in both studies, the total symptom burden in recipients of rIFN- $\beta_{ser}$  was not significantly different compared with placebo recipients, although the total symptom burden tended to be about 30% lower among rIFN- $\beta_{ser}$  recipients in 1986 (mean score per day  $\pm$  SD, 0.2  $\pm$  0.3 vs. 0.3  $\pm$  0.6; P = .10). In neither study were there significant differences in any individual symptom between treatment groups. For subjects who had a respiratory illness episode in 1986, the symptom burden per episode tended to be reduced nearly one-third in ill interferon recipients compared with placebo (mean score per episode  $\pm$  SD, 15  $\pm$  16 vs. 22  $\pm$  23; .05 < P < .1), and the duration of illness tended to be shorter (mean duration  $\pm$  SD, 5  $\pm$  4 d vs. 7  $\pm$  5 d; .05 < P < .1). In 1987 there was no difference in illness severity or duration between interferon and placebo recipients.

Viral infections. Rhinovirus, the most common viral isolate in both studies, was recovered during 13% and 18% of respiratory illness episodes during 1986 and 1987, respectively, and during 28% of subjective colds each year. During the 4-w observation period in 1986, 6.0% (9/149) of all placebo recipients versus 2.9% (4/138) of all rIFN- $\beta_{ser}$  recipients had a rhinovirus-documented cold, representing a 52% reduction in colds in rIFN- $\beta_{ser}$  recipients (P = .3). Most of the rhinovirus colds occurred during the first 2 w of the study and none were identified during week 4 (figure 1A), suggesting that the study may have begun late in that year's rhinovirus season. The next most common isolate was enterovirus, which was identified in six rIFN- $\beta_{ser}$  and two placebo recipients. Six of these isolates were typed (by Dr. Mark Pallanch, Centers for Disease Control, Atlanta) as coxsackie virus A21 (four rIFN-B<sub>ser</sub> and two placebo recipients).

In 1987, rhinovirus-documented colds occurred in 6.3% (11/174) of rIFN- $\beta_{ser}$  recipients, compared with 5.3% (10/189) of placebo recipients during the 4-w period of drug administration (figure 1B). Coxsackievirus A9 was isolated during one additional placebo cold. During the 2 w after completion of drug administration in 1987, rhinovirus colds occurred in four rIFN- $\beta_{ser}$  recipients and one placebo recipient. Parainfluenza was isolated from two ill placebo recipients.

Tolerance. The nasal drops were well-tolerated in each study and no subject withdrew as a result of adverse nasal symptoms. In 1987 four interferon and three placebo recipients were withdrawn from the study after 2 w because of abnormalities on nasal examination (one erosion in each group and ulcers in three interferon and two placebo recipients), none of which was associated with significant clinical symptoms. The frequency of irritative complaints or of nasal examination abnormalities did not differ between groups over the entire period of drug administration in either study (table 1). Leukopenia did not develop in any rIFN- $\beta_{ser}$  recipients, and no significant abnormalities or change from baseline in any hematologic or other laboratory parameter occurred over the course of the study except for elevations in serum aminotransferase levels in one interferon recipient from each study (317 and 355 units/l). After the study, one of these subjects admitted to a history of extensive alcohol abuse. In both cases, the physical examination and other laboratory studies were unrevealing and the values returned to

normal after several weeks. None of the serum specimens obtained 2 w after completion of drug administration had detectable interferon-neutralizing activity.

Peripheral blood 2'-5'oligoadenylate synthetase activity. The mean  $\pm$  SD, 2'-5'oligoadenylate synthetase activity in peripheral blood obtained ~24 h after the preceding dose for the rIFN- $\beta_{ser}$  recipients was 83  $\pm$  33 pmol/10<sup>s</sup> cells per hour (n = 9) compared with 68  $\pm$  47 (n = 9) in the placebo recipients. The 95% confidence limits for the assay in healthy adults are 19-352 pmol/10<sup>s</sup> cells per hour [9].

*Efficacy of blinding.* The results of a questionnaire administered to subjects at one of the study sites at the end of the 1987 treatment period indicated that subjects were well blinded to the treatment they received. Of recipients in each group, 21% believed they were receiving an active drug, 18% believed they were receiving an inactive preparation, and 61% were unable to decide.

#### Discussion

These studies showed that administration of rIFN- $\beta_{ser}$  as nasal drops for 4 w is well-tolerated but ineffective in preventing rhinovirus colds. In contrast to the efficacy of comparable dosages of rIFN- $\alpha_2$ [9, 11], rIFN- $\beta_{ser}$  was disappointingly ineffective in preventing natural rhinovirus colds. The first study, in which there was a tendency toward reduction in rhinovirus colds, prompted us to conduct a second trial at a higher dosage. Despite the higher dosage and daily administration of drug, the second trial failed to show an effect on the frequency of rhinovirus colds or any clinical benefits. The lower efficacy of rIFN- $\beta_{ser}$  was not predicted by in vitro testing. Studies in cell culture have found comparable inhibitory effects of rIFN-a<sub>2b</sub> and rIFN-β<sub>ser</sub> against representative rhinovirus strains [5, 6]. Also, the rhinovirus isolates from rIFN- $\beta_{ser}$  recipients in the 1986 study had comparable sensitivities to rIFN-Bser as did isolates from placebo recipients [5], suggesting that resistance to the antiviral activity of this interferon did not develop in rIFN- $\beta_{ser}$  recipients. Recombinant rIFN- $\beta_{ser}$  administered by nasal drops also had no prophylactic effect against coxsackie A21 virus colds. In vitro testing of these isolates has shown that they are inhibited by rIFN- $\beta_{ser}$ , although they appear to be less susceptible than the rhinoviruses [5].

Possible explanations for the lower efficacy of

rIFN- $\beta_{ser}$  include properties of the interferon itself and the method of drug delivery. It is noteworthy that in the prior field studies rIFN- $\alpha_2$  was administered as nasal spray whereas our rIFN-Bser trials were conducted with nasal drops. Nasal drops of rIFN- $\beta_{ser}$ , however, were effective in preventing experimental rhinovirus colds [7], and one study [12] using rIFN- $\alpha_2$  as treatment for experimental rhinovirus colds found greater efficacy with drops than with spray. It is possible that the efficacy of nasal drops in the challenge models relates to the fact that the viral inoculum is also given by nasal drops. Another possible explanation for the discrepant rIFN- $\alpha_{2b}$ and rIFN- $\beta_{ser}$  results is that of differing pharmacokinetics, particularly that of increased nonspecific tissue binding or natural inhibitors of interferon-β in nasal secretions [7, 13].

Studies with parenterally administered interferons have documented increases in peripheral blood leukocyte 2'-5'oligoadenylate synthetase activity [10, 14] as a marker of interferon activity. In this study, no such increases were seen in recipients of intranasal rIFN- $\beta_{ser}$  compared with placebo, which probably indicates the lack of a systemic interferon effect. Whether this may correspond to the lack of a local antiviral effect in the nasal mucosa is uncertain, in part because comparable studies have not yet been done with intranasal rIFN- $\alpha_2$ .

Symptoms and signs of nasal irritation were not significantly increased in rIFN- $\beta_{ser}$  recipients compared with placebo, even at daily doses of  $24 \times 10^6$  units ( $600 \times 10^6$  units total). A prior tolerance trial [7] with rIFN- $\beta_{ser}$  administered as a nasal spray,  $12 \times 10^6$  units daily, found a significant increase (38%) in bleeding sites on rhinoscopy compared with that in placebo sprayers (12.5%). The decreased incidence of irritation in the current studies despite higher dosages may relate to mechanical trauma induced by the sprayer tip or repeated blast of spray in the earlier tolerance study or more efficient delivery of interferon to the mucosa by nasal sprays than by drops.

The nasal tolerance of rIFN- $\beta_{ser}$  in each of these studies was also more favorable than that observed in prior seasonal prophylaxis studies with lower dosages of nasal sprays of recombinant alpha interferons [2, 3]. Because the prior tolerance trial with rIFN- $\beta_{ser}$  as nasal spray [7] found qualitatively similar, but less frequent, histologic changes as seen with recombinant alpha interferons, it is possible that the better tolerance profile in the current studies is a combined result of a less biologically active interferon and a less traumatic or efficient delivery system. Definitive answers to these quesitons would require parallel studies using rIFN- $\beta_{ser}$  and rIFN- $\alpha_{2b}$ as nasal spray and drops to determine both efficacy and tolerance.

In summary, we found that despite its in vitro efficacy, high doses of rIFN- $\beta_{ser}$  nasal drops were ineffective in preventing naturally occurring respiratory illnesses and rhinovirus infections. In contrast to earlier studies with recombinant alpha interferons, rIFN- $\beta_{ser}$  administered over 25 d was remarkably well tolerated. Further studies of these interferons and the methods of delivery are necessary to explain the lack of prophylactic efficacy of rIFN- $\beta_{ser}$  against natural rhinovirus infections despite its activity in cell culture.

#### References

- Hayden FG. Intranasal interferons for control of respiratory viral infections. In: Revel M, ed. Clinical aspects of interferons. Boston: Kluwer Academic, 1988:3-16
- Douglas RM, Albrecht JK, Milles HB, Moore BW, Read R, Worswick DA, Woodward AJ. Intranasal interferon-α<sub>2</sub> prophylaxis of natural respiratory virus infection. J Infect Dis 1985;151:731-736
- Monto AS, Shope TC, Schwartz SA, Albrecht JK. Intranasal interferon-α2b for seasonal prophylaxis of respiratory infection. J Infect Dis 1986;154:128-133
- Monto AS, Albrecht JK, Schwartz SA. Demonstration of dose-response relationship in seasonal prophylaxis of re-

spiratory infections with alpha-2b interferon. Antimicrob Agents Chemother 1988;32:47-50

- Sperber SJ, Hayden FG. Comparative susceptibility of respiratory viruses to recombinant interferons α2b and β. J Interferon Res 1989;9:285-293
- Ahmad ALM, Tyrrell DAJ. Synergism between anti-rhinovirus antivirals: various human interferons and a number of synthetic compounds. Antiviral Res 1986;6:241-252
- 7. Sperber SJ, Levine PA, Innes DJ, Mills SE, Hayden FG. Tolerance and efficacy of intranasal administration of recombinant  $\beta_{serine}$  interferon in healthy adults. J Infect Dis **1988**;158:166-175
- Higgins PG, Al-Nakib W, Willman J, Tyrrell DAJ. Interferonβser as prophylaxis against experimental rhinovirus infection in volunteers. J Interferon Res 1986;6:153–159
- Farr BM, Gwaltney JM Jr, Adams KF, Hayden FG. Intranasal interferon-α2 for prevention of natural rhinovirus colds. Antimicrob Agents Chemother 1984;26:31-34
- Barouki FM, Witter FR, Griffin DE, Nadler PI, Woods A, Wood DL, Lietman PS. Time course of interferon levels, antiviral state, 2',5'-oligoadenylate synthetase and side effects in healthy men. J Interferon Res 1987;7:29-39
- Betts RF, Erb S, Roth F, Reichman RC, Treanor J, Beutner K, Dolin R. A field trial of intranasal interferon [abstract]. Proceedings of the 13th International Congress of Chemotherapy, Vienna, 1983
- Hayden FG, Gwaltney JM Jr. Intranasal interferon-α<sub>2</sub> treatment of experimental rhinoviral colds. J Infect Dis 1984; 150:174-180
- Harmon MW, Greeenberg SB, Couch RB. Effect of human nasal secretions on the antiviral activity of human fibroblast and leukocyte interferon. Proc Soc Exp Biol Med 1976;152:598-602
- Borden EC, Hawkins MJ, Sielaff KM, Storer BM, Schiesel JD, Smalley RV. Clinical and biological effects of recombinant interferon-β administered intravenously daily in phase I trial. J Interferon Res 1988;8:357-366