

# Acquisition of Antibiotic-Resistant Gram-negative Bacteria in the Benefits of Universal Glove and Gown (BUGG) Cluster Randomized Trial

Anthony D. Harris,<sup>1</sup> Daniel J. Morgan,<sup>2,3</sup> Lisa Pineles,<sup>2</sup> Larry Magder,<sup>2</sup> Lyndsay M. O'Hara,<sup>2</sup> and J. Kristie Johnson<sup>4</sup>

<sup>1</sup>University of Maryland School of Medicine, Baltimore, Maryland, USA, <sup>2</sup>Department of Epidemiology and Public Health, University of Maryland School of Medicine, Baltimore, Maryland, USA, <sup>3</sup>Veterans Affairs Maryland Health Care System, Baltimore, Maryland, USA, and <sup>4</sup>Department of Pathology, University of Maryland School of Medicine, Baltimore, Maryland, USA

**Background.** The Benefits of Universal Glove and Gown (BUGG) cluster randomized trial found varying effects on methicillin-resistant *Staphylococcus aureus* and vancomycin-resistant *Enterococcus* and no increase in adverse events. The aim of this study was to assess whether the intervention decreases the acquisition of antibiotic-resistant gram-negative bacteria.

**Methods.** This was a secondary analysis of a randomized trial in 20 hospital intensive care units. The intervention consisted of healthcare workers wearing gloves and gowns when entering any patient room compared to standard care. The primary composite outcome was acquisition of any antibiotic-resistant gram-negative bacteria based on surveillance cultures.

**Results.** A total of 40 492 admission and discharge perianal swabs from 20 246 individual patient admissions were included in the primary outcome. For the primary outcome of acquisition of any antibiotic-resistant gram-negative bacteria, the intervention had a rate ratio (RR) of 0.90 (95% confidence interval [CI], .71–1.12;  $P = .34$ ). Effects on the secondary outcomes of individual bacteria acquisition were as follows: carbapenem-resistant Enterobacteriaceae (RR, 0.86 [95% CI, .60–1.24;  $P = .43$ ), carbapenem-resistant *Acinetobacter* (RR, 0.81 [95% CI, .52–1.27;  $P = .36$ ), carbapenem-resistant *Pseudomonas* (RR, 0.88 [95% CI, .55–1.42];  $P = .62$ ), and extended-spectrum  $\beta$ -lactamase-producing bacteria (RR, 0.94 [95% CI, .71–1.24];  $P = .67$ ).

**Conclusions.** Universal glove and gown use in the intensive care unit was associated with a non-statistically significant decrease in acquisition of antibiotic-resistant gram-negative bacteria. Individual hospitals should consider the intervention based on the importance of these organisms at their hospital, effect sizes, CIs, and cost of instituting the intervention.

**Clinical Trials Registration.** NCT01318213.

**Keywords.** antibiotic resistance; barrier precautions; contact precautions.

Antibiotic resistance is associated with considerable morbidity, mortality, and costs [1, 2]. The estimated cost of antibiotic resistance in the United States is more than \$4 billion per year [2]. Healthcare-associated infections are the most common complication of hospital care, affecting an estimated 1 in every 20 inpatients [3]. Antibiotic-resistant gram-negative bacteria continue to emerge and rank highly on the list of pathogens causing national healthcare-associated infections [4, 5].

Tremendous controversy exists about the relative advantages and disadvantages of contact precautions [6, 7]. Previously, we published a cluster randomized trial titled the Benefits of Universal Glove and Gown (BUGG) that showed

no statistically significant effect on the composite primary outcome of methicillin-resistant *Staphylococcus aureus* (MRSA) or vancomycin-resistant *Enterococcus* (VRE) acquisition [8]. However, this composite outcome result was driven by differing effects on MRSA and VRE; there was a large and statistically significant decrease of individual patient MRSA acquisition and no effect on VRE acquisition. Importantly, the study also showed no increase in adverse events and improved hand hygiene compliance on room exit with the intervention.

Antibiotic resistant gram-negative bacteria are among the most important threats to human health, being categorized by the Centers for Disease Control and Prevention (CDC) as “urgent” and “serious” threats [9]. Interventions recommended by national organizations include the use of contact precautions to prevent the spread of these bacteria to other patients [10, 11]. However, no randomized trials have assessed the impact of contact precautions on antibiotic-resistant gram-negative bacteria.

In the current study, we used previously collected and stored perianal samples from the BUGG cluster randomized trial to assess if wearing gloves and gowns for all patient contact in the intensive care unit (ICU) reduces acquisition rates of antibiotic-resistant gram-negative bacteria, including carbapenem-resistant *Pseudomonas aeruginosa*, carbapenem-resistant

Received 17 October 2019; editorial decision 16 January 2020; accepted 21 January 2020; published online January 23, 2020.

Correspondence: A. D. Harris, University of Maryland School of Medicine, 10 S Pine St, MSTF 330, Baltimore, MD 21201 (aharris@som.umaryland.edu).

Clinical Infectious Diseases® 2021;72(3):431–7

© The Author(s) 2020. Published by Oxford University Press for the Infectious Diseases Society of America. This is an Open Access article distributed under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs licence (<http://creativecommons.org/licenses/by-nc-nd/4.0/>), which permits non-commercial reproduction and distribution of the work, in any medium, provided the original work is not altered or transformed in any way, and that the work is properly cited. For commercial re-use, please contact journals.permissions@oup.com  
DOI: 10.1093/cid/ciaa071

*Acinetobacter baumannii*, extended-spectrum  $\beta$ -lactamase (ESBL)-producing Enterobacteriaceae, and carbapenemase-producing Enterobacteriaceae (CPE).

## METHODS

### Study Design

This study is a secondary analysis of specimens collected in the BUGG study, a 20-hospital cluster randomized trial of universal glove and gown compared to standard practice. The study was conducted in medical, surgical, and medical-surgical ICUs varying in size from 9 to 36 beds and located across the United States in rural, urban, academic, and nonacademic settings. The primary outcome of the original trial was acquisition of MRSA or VRE. Details of the original study design have been previously published [8]. ICUs were randomized to either the intervention or control arm. The study had a baseline period from 1 September 2011 to 30 November 2011. The study period was from 4 January 2012 to 4 October 2012. The trial was conducted in accordance with the Consolidated Standards of Reporting Trials (CONSORT) guidelines [12].

### Intervention and Control Arms

The intervention occurred at the cluster level of the ICU. During the baseline period, all ICUs followed their usual standard of care, which consisted of healthcare workers following CDC contact precautions guidelines (gloves and gowns) for patients known to have antibiotic-resistant bacteria such as VRE and MRSA [11]. After the baseline period, ICUs were randomized, and during the study period, all healthcare workers (nurses, physicians, respiratory therapists, etc) in the 10 ICUs assigned to the intervention arm were required to wear gloves and gowns for all patient contact and when entering any patient room [11, 13]. The 10 control ICUs followed their usual standard of care during the study period. Compliance with glove and gown use was measured by 30-minute direct observation periods on a random sample of rooms. No hospitals performed active surveillance for antibiotic-resistant gram-negative bacteria. All hospitals isolated patients with carbapenem-resistant gram-negative bacteria. Twelve hospitals performed chlorhexidine bathing (5 in the control arm and 7 in the intervention arm) [14].

### Outcomes

All patients had ICU admission and ICU discharge perianal cultures. The primary outcome was acquisition of either carbapenem-resistant *Pseudomonas aeruginosa*, carbapenem-resistant *A. baumannii*, ESBL-producing Enterobacteriaceae, or CPE as a composite. Secondary outcomes were each of these antibiotic-resistant gram-negative bacteria analyzed individually. For each patient, acquisition was defined as having an initial ICU surveillance specimen that was negative for an antibiotic-resistant pathogen with a subsequent discharge surveillance specimen within the same ICU admission that was positive for

the same antibiotic-resistant pathogen. ICUs did not receive results of the surveillance cultures. Specimens were shipped to and processed at the University of Maryland using a method that did not affect bacterial yield or organism identification [15]. The same laboratory procedures were followed in the baseline and intervention. The same laboratory technicians handled all the specimens. The laboratory technicians were blinded to which study arms the specimens were from. For Enterobacteriaceae, we focused on acquired resistance to carbapenem and ESBLs. Since both of these resistance mechanisms occur by  $\beta$ -lactamases, we performed polymerase chain reaction for the detection of CTX-M, *Klebsiella pneumoniae* carbapenemase (KPC), New Delhi metallo- $\beta$ -lactamase (NDM), IMP, verona integron-encoded metallo- $\beta$ -lactamase (VIM), and oxacillinase (OXA) [16, 17]. For *A. baumannii* and *P. aeruginosa* carbapenem resistance, we cultured and performed susceptibility testing. Specimens were first enriched into brain-heart infusion broth with 2  $\mu$ g/mL of meropenem. These were incubated overnight at 37°C and plated to Ceftrimide (Becton Dickinson, Sparks, Maryland) for *P. aeruginosa* and *Acinetobacter* RambaCHROM agar (Molecular Toxicology, Boone, North Carolina) for the isolation of *A. baumannii*. Agar was used following manufacturer instructions. Susceptibility testing using the Vitek 2 assay (bioMérieux, Durham, North Carolina) was performed on all isolates identified as *P. aeruginosa* or *A. baumannii* following Clinical and Laboratory Standards Institute guidelines [18].

### Statistical Analysis

The statistical analysis plan was written and sealed prior to the analysis. The analysis was based on the outcome (acquisition yes/no) for each person seen in the study ICUs at either the baseline period (when standard contact precautions were used in all ICUs) or the study period (when half of the ICUs employed universal contact precautions). The probability that each person was classified as having acquired an infection during their ICU stay is a function of the acquisition rate in that ICU at that period, and the number of days between admission specimen collection and discharge specimen collection, which was approximately equal to the patient ICU length of stay. The rate of acquisition in an ICU at a given period was modeled as a multiplicative function of parameters related to period (baseline or study), contact precautions (whether that ICU was using universal or selective precautions during that period), and ICU (treated as a random effect). This corresponds to using a generalized linear mixed model for a binary outcome with a complementary log-log link, random effects for ICUs, and the log of the number of days between swabs as an offset term. The model was fit by maximum likelihood estimation using SAS Proc GLIMMIX. The model resulted in estimates of the mean rate of acquisition during the baseline period, the mean rate during the study period in ICUs that performed selective precautions, the mean rate during the study period in ICUs that performed

universal precautions, and the rate ratio (RR) due to the intervention. Rate ratios adjusted for the admission prevalence at each hospital and in each time period were also calculated. For ease of interpretation, we also present a rate difference, which is the difference in acquisition rates due to the intervention based on the model evaluated at the average ICU. Confidence intervals (CIs) for the rate differences were calculated using the delta method based on the parameter estimates and standard errors from the multiplicative model that we fit.

## RESULTS

Twenty ICUs participated in the study and none withdrew. Of the 26 180 patients enrolled in the original study, 20 246 patients had both admission and discharge swabs, including 4243 patients during the baseline period and 16 003 patients during the study period. A total of 40 492 perianal swabs were worked up, including 8486 swabs during the baseline period and 32 006 swabs during the study period. During the study period, compliance with obtaining perianal cultures at admission was 94.92%. Compliance with obtaining perianal cultures at discharge was 85.07%. Compliance with wearing gloves in the intervention ICUs was 86.18% (2787 of 3234), and compliance with gowns was 85.14% (2750 of 3230). In the control group, 10.52% of patients were on contact precautions. In the control ICUs, for patients on contact precautions, compliance with wearing gloves was 84.11% (556 of 661) and compliance with gowns was 81.21% (536 of 660).

Table 1 demonstrates the acquisition rates in the baseline and study periods for both the control group and the intervention group. Baseline rates for the primary outcome were similar in the intervention and control group prior to the randomization. Of note, acquisition rates increased in the study period for both intervention and control groups.

The effects of the intervention on the primary outcome and the secondary outcomes are shown in Table 2 as RRs and in Figure 1 as rate differences. For the primary outcome of acquisition of any antibiotic-resistant gram-negative bacteria, the intervention showed a decrease in acquisition: RR, 0.90 [95% CI, .71–1.12];  $P = .34$ ) and a rate difference of  $-2.1$  (95% CI,  $-5.9$

to 1.7;  $P = .34$ ). For each individual outcome, the RR was  $< 1$ , suggesting a decrease in the rate of acquisition of the antibiotic-resistant bacteria in the intervention group. None of these associations were statistically significant.

Figure 2 demonstrates the prevalence of positive ICU admission cultures by month in both the baseline and study periods. The results demonstrate an increase in positive patients on admission in the study period compared to the baseline period in both the intervention group and control group.

## DISCUSSION

Healthcare workers' use of gloves and gowns for all ICU patient contact was associated with a non-statistically significant decrease in acquisition rate of antibiotic-resistant gram-negative bacteria compared to ICUs using contact precautions only for patients known to be colonized with antibiotic-resistant bacteria.

The primary outcome rate ratio was 0.90, indicating a 10% decrease in acquisition rate in intervention units compared to control units. The CIs around this rate ratio ranged from 0.71 to 1.12, indicating that our findings are consistent with the possibility that the intervention resulted in a 29% reduction in acquisition rates, but also that our study is consistent with the possibility of no effect of the intervention. The estimated RRs were all  $< 1$  for the individual gram-negative bacteria, but all were  $> 0.80$ . The data indicate the largest potential benefit for carbapenem-resistant *A. baumannii* and the smallest potential benefit for ESBL-producing bacteria, which is consistent with the literature [19–21]. *Acinetobacter baumannii* has data supporting its persistence in the hospital environment and a strong association with risk of transmission to subsequent patients in the same room, which may explain our finding [21–23].

How to interpret these results and place these results in the context of our previous results for MRSA and VRE is challenging. The previous study showed a large statistically significant effect on MRSA acquisition and no effect on VRE. It is noteworthy that of the 6 outcomes/individual antibiotic-resistant bacteria analyzed in this study and the original study, for 5 we observed a reduction in the acquisition of antibiotic-resistant bacteria

**Table 1. Rate of Acquisition**

Organism	No. of Acquisitions/Total Days at Risk (Rate per 1000 Days)			
	Baseline Control Group (n = 2297)	Baseline Intervention Group (n = 1946)	Study Period Control Group (n = 7916)	Study Period Intervention Group (n = 8087)
<i>Pseudomonas</i>	13/10 041 (1.29)	16/8598 (1.86)	124/32 269 (3.84)	130/32 875 (3.95)
Carbapenemase-producing Enterobacteriaceae	31/9951 (3.12)	28/8478 (3.30)	103/32 327 (3.19)	92/33 189 (2.77)
<i>Acinetobacter</i>	16/9948 (1.61)	20/8616 (2.32)	147/32 042 (4.59)	142/32 579 (4.36)
ESBL-producing bacteria	47/9747 (4.82)	37/8294 (4.46)	253/31 323 (8.08)	248/31 901 (7.77)
Any	102/10 062 (10.14)	90/8667 (10.38)	585/32 830 (17.82)	566/33 576 (16.86)

Abbreviations: *Acinetobacter*, carbapenem-resistant *Acinetobacter baumannii*; CPE, carbapenemase-producing Enterobacteriaceae; ESBL, extended-spectrum  $\beta$ -lactamase; *Pseudomonas*, carbapenem-resistant *Pseudomonas aeruginosa*.

**Table 2. Effect of the Intervention of Universal Glove and Gown**

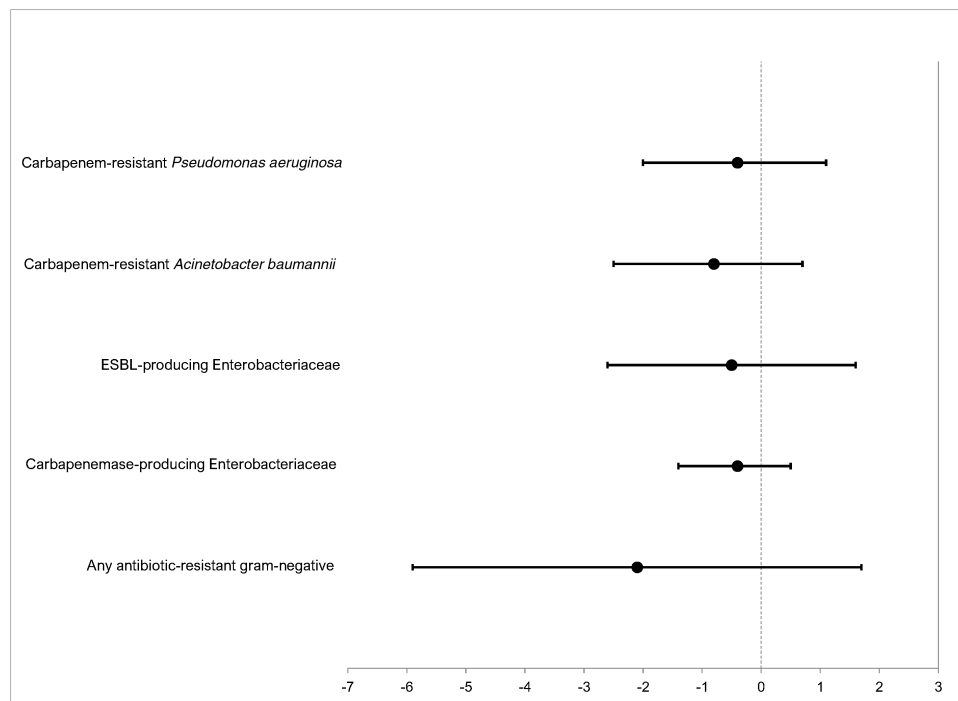
Organism	RR for Impact of the Intervention (95% CI)	PValue	RR for Impact of the Intervention Adjusted for Site-specific Admission Prevalence (95% CI)	PValue
<i>Pseudomonas</i> , carbapenem-resistant <i>Pseudomonas aeruginosa</i>	0.88 (.55–1.42)	.62	0.78 (.51–1.19)	.25
Carbapenemase-resistant Enterobacteriaceae	0.86 (.60–1.24)	.43	0.88 (.62–1.23)	.45
<i>Acinetobacter</i>	0.81 (.52–1.27)	.36	0.75 (.50–1.13)	.17
ESBL-producing bacteria	0.94 (.71–1.24)	.67	0.95 (.74–1.21)	.67
Any	0.90 (.71–1.12)	.34	0.90 (.73–1.10)	.31

Abbreviations: *Acinetobacter*, carbapenem-resistant *Acinetobacter baumannii*; CI, confidence interval; CRE, carbapenemase-resistant carbapenemase-producing Enterobacteriaceae; ESBL, extended-spectrum β-lactamase; RR, rate ratio.

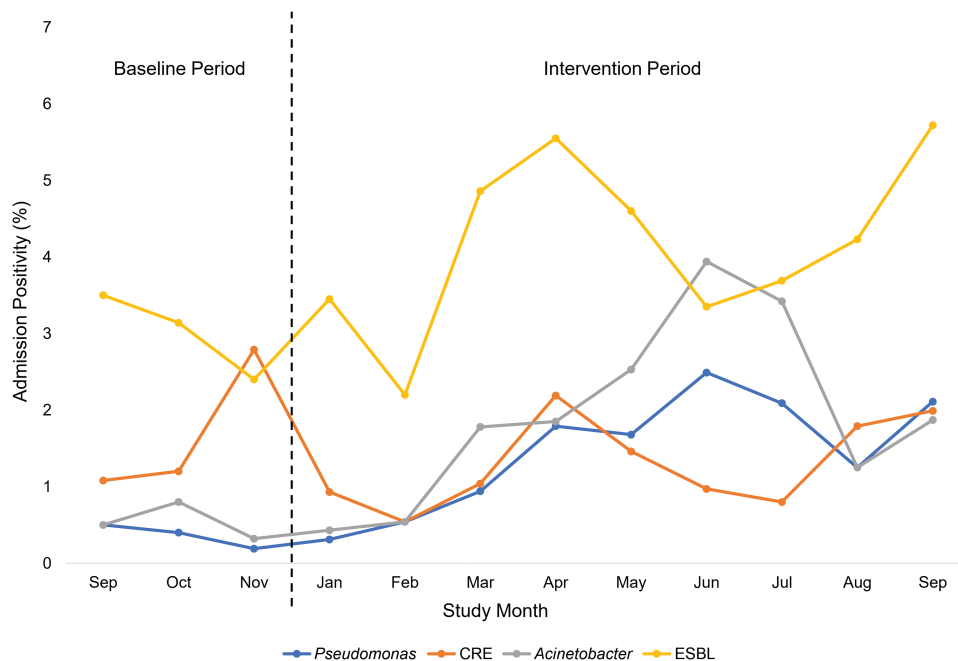
in the universal glove and gown arm. However, all effect sizes were small, all CIs except that for MRSA overlapped 1, and all *P* values except that for MRSA were >.05. Some would argue that these results are therefore inconclusive and that the original study was underpowered to detect an effect size of 10%–20%, which is what was seen with most of the outcomes. These points and the terminology we have used throughout this manuscript to interpret the results of our study are especially relevant, as many statistical experts and leading journals are moving away from a focus on *P* values and more of a focus on effect size and CIs [24–27].

The question as to what an individual hospital should do with these results is also challenging. One key data point for hospitals to consider is to address the question about how many patients

would not acquire antibiotic-resistant gram-negative bacteria in a typical ICU with a universal glove and gown policy. An average ICU has 16 beds and thus has a total of 5840 patient-days. Figure 1 demonstrates that the rate difference of –2.1 per 1000 patient-days means that, on average, the intervention would lead to 2.1 fewer patients acquiring antibiotic-resistant bacteria per 1000 patient-days. Thus, in an average ICU, over a 1-year period, 12 patients would be prevented from acquiring antibiotic-resistant gram-negative bacteria. However, there is uncertainty in this estimate, and our data are consistent with as many as 34 patients prevented, or 10 additional patients caused by universal precautions. The literature suggests that between 10% and 40% of these patients would develop subsequent infections within the same hospitalization with the acquired bacteria [10, 28–34].



**Figure 1.** Rate differences (per 1000 person-days) and 95% confidence intervals for the impact of universal glove and gown use, by organism. The rate difference provides a measure of the public health impact of the intervention and describes the number of cases that could be prevented. Abbreviations: ESBL, extended-spectrum β-lactamase.



**Figure 2.** Admission positivity rate per organism, by month. Abbreviations: CRE, carbapenemase-resistant Enterobacteriaceae; ESBL, extended-spectrum  $\beta$ -lactamase.

Antibiotic-resistant gram-negative bacteria have been estimated to increase hospital length of stay by 24% and current admission hospital costs by 29% [35]. Having an antibiotic-resistant bacterial infection has also been shown to increase subsequent readmissions, prescriptions, and inpatient days [36].

One of the few studies to assess the impact of different infection control interventions, including active surveillance and contact precautions on antibiotic-resistant gram-negative bacteria, is the Clinical Trial to Reduce Antibiotic Resistance in European Intensive Cares (MOSAR-ICU) trial [37]. This study was a cluster randomized stepped-wedge design with a sequential set of interventions, with the first being chlorhexidine bathing, the second being hand hygiene initiative, and the last being active surveillance and contact precautions. The outcomes were MRSA, VRE, and highly resistant Enterobacteriaceae. The study's main findings support the use of chlorhexidine bathing and improved hand hygiene for MRSA. Active surveillance and contact precautions for highly resistant bacteria after the chlorhexidine bathing and the hand hygiene intervention were implemented showed no effect on highly resistant Enterobacteriaceae rates. However, it is not clear that the study was powered to detect this difference.

Our study has several limitations: As indicated above, the study was underpowered to detect an effect size that was found. In addition, from the baseline period to the study period, acquisition rates of the gram-negative bacteria increased in both the control and intervention periods. We do not know entirely why this occurred. We think a major reason for this increase was the increase in colonization pressure/admission prevalence of these bacteria during the study period in both the control

and intervention sites and may be in part due to seasonal effects [38–40]. Colonization pressure has been well described to be a major driver of acquisition of antibiotic-resistant bacteria [41–43].

Our study has several strengths. The cluster randomized trial design provides stronger evidence than most studies currently used to support or negate infection control interventions, and the primary outcome measurement of acquisition was more objective than clinical culture positivity as used in other studies [44]. In addition, all ICUs enrolled completed the study, which is rare in a study of this size, and compliance with the intervention was high, which demonstrates the feasibility of implementing and sustaining the intervention. Moreover, our results are generalizable to a broad set of hospitals because the study was conducted in medical, surgical, and medical-surgical ICUs varying in size from 9 to 36 beds and located across the United States in rural, urban, academic, and nonacademic settings.

In conclusion, the association of universal glove and gown use in the ICU with acquisition of antibiotic-resistant gram-negative bacteria was inconclusive. The observed rate ratios for all 5 outcomes suggest that the intervention was protective; however, none were statistically significant. Individual hospitals should consider the cost-effectiveness of the intervention based on the effect sizes, CIs, and cost of instituting the intervention when they decide upon whether or not to adopt the intervention.

#### Notes

**Acknowledgments.** The authors thank Stephanie Hitchcock and Corey Sparkes for their outstanding work in the laboratory. The authors also



thank the Benefits of Universal Glove and Gown (BUGG) Trial primary investigators for their outstanding contributions: Syed K. Shahryar, MD, St Luke's Medical Center; Tamara Rasner, RN, BS, CIC, Top Echelon; Connie S. Price, MD, Denver Medical Center; Joseph J. Gadbaw, MD, Lawrence and Memorial Hospital; Marci L. Drees, MD, MS, Christiana Hospital; Daniel H. Kett, MD, Jackson Memorial Hospital; L. Silvia Muñoz-Price, Medical College of Wisconsin; Jesse T. Jacob, MD, Emory University Hospital Midtown; Loreen A. Herwaldt, MD, University of Iowa Hospitals and Clinics; Carol A. Sulis, MD, Boston Medical Center; Deborah S. Yokoe, MD, MPH, Brigham and Women's Hospital; Lisa Maragakis, MD, MPH, Johns Hopkins Hospital; Matthew E. Lissauer, MD, Robert Wood Johnson University Hospital; Marcus J. Zervos, MD, Henry Ford Hospital; David K. Warren, MD, Barnes Jewish Hospital; Robin L. Carver, RN, BSN, CIC, Premier, Inc; Deverick J. Anderson, MD, MPH, Duke University; Dave P. Calfee, MD, MS, Weill Cornell Medical College; Jason E. Bowling, MD, University Hospital; Nasia Safdar, MD, PhD, University of Wisconsin Hospitals and Clinics.

**Disclaimer.** The authors of this manuscript are responsible for its content. Statements in the manuscript do not necessarily represent the official views of or imply endorsement by the Agency for Healthcare Research and Quality (AHRQ) or the US Department of Health and Human Services (HHS). The sponsor had no role in study design; in the collection, analysis, and interpretation of data; in the writing of the report; or in the decision to submit the manuscript for publication.

**Financial support.** This work was supported by the AHRQ, HHS (grant number R18 HS024045).

**Potential conflicts of interest.** A. D. H. reports personal fees from UpToDate, outside the submitted work. D. J. M. reports grants from NIH, CDC, VA HSR&D, and AHRQ, outside the submitted work. All other authors: No reported conflicts of interest. All authors have submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest.

## References

- Zimlichman E, Henderson D, Tamir O, et al. Health care-associated infections: a meta-analysis of costs and financial impact on the US health care system. *JAMA Intern Med* **2013**; 173:2039–46.
- Talbot GH, Bradley J, Edwards JE Jr, Gilbert D, Scheld M, Bartlett JG; Antimicrobial Availability Task Force of the Infectious Diseases Society of America. Bad bugs need drugs: an update on the development pipeline from the Antimicrobial Availability Task Force of the Infectious Diseases Society of America. *Clin Infect Dis* **2006**; 42:657–68.
- Klevens RM, Edwards JR, Richards CL Jr, et al. Estimating health care-associated infections and deaths in U.S. hospitals, 2002. *Public Health Rep* **2007**; 122:160–6.
- Weiner LM, Webb AK, Limbago B, et al. Antimicrobial-resistant pathogens associated with healthcare-associated infections: summary of data reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2011–2014. *Infect Control Hosp Epidemiol* **2016**; 37:1288–301.
- World Health Organization. WHO publishes list of bacteria for which new antibiotics are urgently needed. Available at: <https://www.who.int/news-room/detail/27-02-2017-who-publishes-list-of-bacteria-for-which-new-antibiotics-are-urgently-needed>. Accessed 19 March 2019.
- Morgan DJ, Murthy R, Munoz-Price LS, et al. Reconsidering contact precautions for endemic methicillin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-resistant *Enterococcus* (VRE). *Infect Control Hosp Epidemiol* **2016**; 36:1163–72.
- Rubin MA, Samore MH, Harris AD. The importance of contact precautions for endemic methicillin-resistant *Staphylococcus aureus* and vancomycin-resistant enterococci. *JAMA* **2018**; 319:863–4.
- Harris AD, Pineles L, Belton B, et al; Benefits of Universal Glove and Gown (BUGG) Investigators. Universal glove and gown use and acquisition of antibiotic-resistant bacteria in the ICU: a randomized trial. *JAMA* **2013**; 310:1571–80.
- Centers for Disease Control and Prevention. The biggest antibiotic-resistant threats in the U.S. 2019. Available at: <https://www.cdc.gov/drugresistance/biggest-threats.html>. Accessed 7 August 2019.
- Yokoe DS, Anderson DJ, Berenholtz SM, et al; Society for Healthcare Epidemiology of America (SHEA). A compendium of strategies to prevent healthcare-associated infections in acute care hospitals: 2014 updates. *Infect Control Hosp Epidemiol* **2014**; 35:967–77.

- Siegel JD, Rhinehart E, Jackson M, Chiarello L; Health Care Infection Control Practices Advisory Committee. 2007 guideline for isolation precautions: preventing transmission of infectious agents in health care settings. *Am J Infect Control* **2007**; 35:S65–164.
- Campbell MK, Piaggio G, Elbourne DR, Altman DG, CONSORT Group. Consort 2010 statement: extension to cluster randomised trials. *BMJ* **2012**; 345:e5661.
- Siegel JD, Rhinehart E, Jackson M, Chiarello L; Healthcare Infection Control Practices Advisory Committee. Management of multidrug-resistant organisms in health care settings, 2006. *Am J Infect Control* **2007**; 35:S165–93.
- Morgan DJ, Pineles L, Shardell M, et al; BUGG Study Primary Investigators. Effect of chlorhexidine bathing and other infection control practices on the Benefits of Universal Glove and Gown (BUGG) trial: a subgroup analysis. *Infect Control Hosp Epidemiol* **2015**; 36:734–7.
- Green HP, Johnson JA, Furuno JP, et al. Impact of freezing on the future utility of archived surveillance culture specimens. *Infect Control Hosp Epidemiol* **2007**; 28:886–8.
- Poirel L, Walsh TR, Cuvillier V, Nordmann P. Multiplex PCR for detection of acquired carbapenemase genes. *Diagn Microbiol Infect Dis* **2011**; 70:119–23.
- Centers for Disease Control and Prevention. Multiplex real-time PCR detection of *Klebsiella pneumoniae* carbapenemase (KPC) and New Delhi metallo- $\beta$ -lactamase (NDM-1) genes. Available at: <https://www.cdc.gov/HAI/settings/lab/kpc-ndm1-lab-protocol.html#pp>. Accessed 15 March 2019.
- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing; 26th informational supplement. CLSI document M100-S26. Wayne, PA: CLSI, **2016**.
- R-GNOSIS. Main findings and results. Available at: <http://www.r-gnosis.eu/page/main-findings-results.php>. Accessed 7 August 2019.
- Harris AD, Johnson JK, Pineles L, O'Hara LM, Bonomo RA, Thom KA. Patient-to-patient transmission of *Acinetobacter baumannii* gastrointestinal colonization in the intensive care unit. *Antimicrob Agents Chemother* **2019**; 63. doi:10.1128/AAC.00392-19.
- Munoz-Price LS, Namias N, Cleary T, et al. *Acinetobacter baumannii*: association between environmental contamination of patient rooms and occupant status. *Infect Control Hosp Epidemiol* **2013**; 34:517–20.
- Thom KA, Johnson JK, Lee MS, Harris AD. Environmental contamination because of multidrug-resistant *Acinetobacter baumannii* surrounding colonized or infected patients. *Am J Infect Control* **2011**; 39:711–5.
- Rosa R, Arheart KL, Depascale D, et al. Environmental exposure to carbapenem-resistant *Acinetobacter baumannii* as a risk factor for patient acquisition of *A. baumannii*. *Infect Control Hosp Epidemiol* **2014**; 35:430–3.
- Wasserstein RL, Lazar NA. The ASA statement on P-values: context, process, and purpose. *Am Stat* **2016**; 70:129–33. Available at: <https://amstat.tandfonline.com/doi/full/10.1080/00031305.2016.1154108>. Accessed 7 August 2019.
- Wasserstein RL, Schirm AL, Lazar NA. Moving to a world beyond “p < 0.05.” *Am Stat* **2019**. Available at: <https://www.tandfonline.com/doi/full/10.1080/00031305.2019.1583913>. Accessed 7 August 2019.
- Harrington D, D'Agostino RB Sr, Gatsonis C, et al. New guidelines for statistical reporting in the journal. *N Engl J Med* **2019**; 381:285–6.
- Goodman SN, Berlin JA. The use of predicted confidence intervals when planning experiments and the misuse of power when interpreting results. *Ann Intern Med* **1994**; 121:200–6.
- Harris AD, Nemyo L, Johnson JA, et al. Co-carriage rates of vancomycin-resistant *Enterococcus* and extended-spectrum beta-lactamase-producing bacteria among a cohort of intensive care unit patients: implications for an active surveillance program. *Infect Control Hosp Epidemiol* **2004**; 25:105–8.
- Harris AD, Furuno JP, Roghmann MC, et al. Targeted surveillance of methicillin-resistant *Staphylococcus aureus* and its potential use to guide empiric antibiotic therapy. *Antimicrob Agents Chemother* **2010**; 54:3143–8.
- Harris AD, Jackson SS, Robinson G, et al. *Pseudomonas aeruginosa* colonization in the intensive care unit: prevalence, risk factors, and clinical outcomes. *Infect Control Hosp Epidemiol* **2016**; 37:544–8.
- Blanco N, Harris AD, Rock C, et al. Risk factors and outcomes associated with multidrug-resistant *Acinetobacter baumannii* upon intensive care unit admission. *Antimicrob Agents Chemother* **2017**; 62. doi:10.1128/AAC.01631-17.
- Borer A, Saidel-Odes L, Eskira S, et al. Risk factors for developing clinical infection with carbapenem-resistant *Klebsiella pneumoniae* in hospital patients initially only colonized with carbapenem-resistant *K. pneumoniae*. *Am J Infect Control* **2012**; 40:421–5.
- Schechner V, Kotlovsky T, Kazma M, et al. Asymptomatic rectal carriage of blaKPC producing carbapenem-resistant Enterobacteriaceae: who is prone to become clinically infected? *Clin Microbiol Infect* **2013**; 19:451–6.
- Emmanuel Martinez A, Widmer A, Frei R, et al. ESBL-colonization at ICU admission: impact on subsequent infection, carbapenem-consumption, and outcome. *Infect Control Hosp Epidemiol* **2019**; 40:408–13.

35. Mauldin PD, Salgado CD, Hansen IS, Durup DT, Bosso JA. Attributable hospital cost and length of stay associated with health care-associated infections caused by antibiotic-resistant gram-negative bacteria. *Antimicrob Agents Chemother* **2010**; 54:109–15.
36. Nelson RE, Jones M, Liu CF, et al. The impact of healthcare-associated methicillin-resistant *Staphylococcus aureus* infections on post-discharge healthcare costs and utilization. *Infect Control Hosp Epidemiol* **2015**; 36:534–42.
37. Derde LPG, Cooper BS, Goossens H, et al; MOSAR WP3 Study Team. Interventions to reduce colonisation and transmission of antimicrobial-resistant bacteria in intensive care units: an interrupted time series study and cluster randomised trial. *Lancet Infect Dis* **2014**; 14:31–9.
38. MacFadden DR, McGough SF, Fisman D, Santillana M, Brownstein JS. Antibiotic resistance increases with local temperature. *Nat Clim Chang* **2018**; 8:510–4.
39. Fisman DN. Seasonality of infectious diseases. *Annu Rev Public Health* **2007**; 28:127–43.
40. Perencevich EN, McGregor JC, Shardell M, et al. Summer peaks in the incidences of gram-negative bacterial infection among hospitalized patients. *Infect Control Hosp Epidemiol* **2008**; 29:1124–31.
41. Bonten MJM. Colonization pressure: a critical parameter in the epidemiology of antibiotic-resistant bacteria. *Crit Care* **2012**; 16:142.
42. Arvaniti K, Lathyris D, Ruimy R, et al. The importance of colonization pressure in multiresistant *Acinetobacter baumannii* acquisition in a Greek intensive care unit. *Crit Care* **2012**; 16:R102.
43. Ajao AO, Harris AD, Roghmann MC, et al. Systematic review of measurement and adjustment for colonization pressure in studies of methicillin-resistant *Staphylococcus aureus*, vancomycin-resistant enterococci, and clostridium difficile acquisition. *Infect Control Hosp Epidemiol* **2011**; 32:481–9.
44. Feng PJ, Kallen AJ, Ellingson K, Muder R, Jain R, Jernigan JA. Clinical incidence of methicillin-resistant *Staphylococcus aureus* (MRSA) colonization or infection as a proxy measure for MRSA transmission in acute care hospitals. *Infect Control Hosp Epidemiol* **2011**; 32:20–5.