



Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.



ELSEVIER

Vaccinology at the beginning of the 21st century

Andreas Wack and Rino Rappuoli

Today, the main challenges for vaccinologists include improving vaccines against as yet undefeated pathogens, rapid identification and response to emerging diseases and successful intervention in chronic diseases in which ongoing immune responses are insufficient. Reverse genetics and reverse vaccinology are now used to generate rapidly new vaccine strains and to mine whole genomes in the search for promising antigens. The rational design of adjuvants has become possible as a result of the discovery of the receptors that recognize microbial patterns and lead to dendritic cell activation. Antigen-loaded dendritic cells, DNA in naked, formulated or viral form, and other delivery systems are used to maximize immune responses. Although work on the 'easy' vaccines has already been completed, it is hoped that a combination of conceptual and technical innovation will enable the development of more complex and sophisticated vaccines in the future.

Addresses

Chiron Vaccines, Via Fiorentina 1, 53100 Siena, Italy

Corresponding author: Rappuoli, Rino (rino_rappuoli@chiron.com)

Current Opinion in Immunology 2005, 17:411–418

This review comes from a themed issue on
Host–pathogen interactions
Edited by Bali Pulendran and Robert A Seder

Available online 13th June 2005

0952-7915/\$ – see front matter

© 2005 Elsevier Ltd. All rights reserved.

DOI 10.1016/j.coi.2005.05.005

Introduction

Although the eradication of smallpox in the 1970s and of poliomyelitis hopefully in the coming years mark two of the most important milestones in medical history, we now face an unprecedented succession of new pathogens which jump species barriers to infect humans, and the frustration deriving from the inability to control devastating diseases such as HIV, malaria and tuberculosis. This review will cover the recent progress in vaccinology (Figure 1), largely resulting from dramatic technical innovation that is now reaching the clinic, as well as the huge challenges posed by old and new pathogens that are facing us.

The diseases to fight

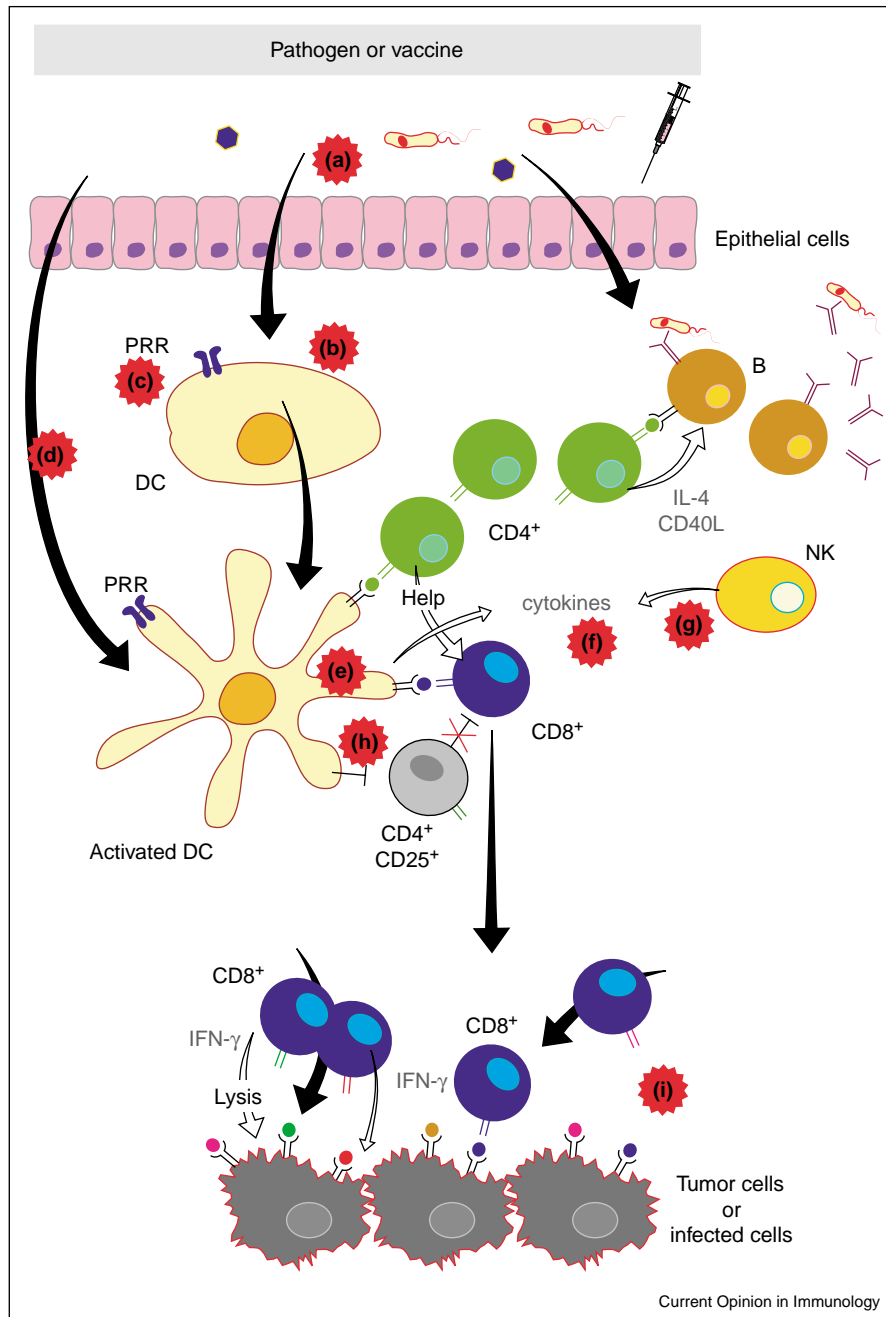
The three 'big killers', the pathogens that most heavily afflict global health, are HIV, mycobacterium and plas-

modium. Whereas the latter two represent long-term companions of the human species, HIV is a virus that spread in the human population only about 25 years ago and has a nonhuman primate origin [1]. In fact, most, if not all, of the recently emerging diseases go back to animal reservoirs, from which they infect humans through close contact during hunting and farming, in live animal markets or through food processing, preparation or consumption. An example is Ebola virus, which, after three outbreaks between 1976 and 1979, has appeared in the human population nine times since 1994, with each outbreak resulting from the handling of dead gorillas, chimpanzees or duikers, which in turn are thought to have been infected by an unknown natural host [2*]. The variant Creutzfeldt–Jacob disease reached the population through the food chain, from 'rendered' sheep and cow carcasses fed to cows which were subsequently consumed by humans. The severe acute respiratory syndrome (SARS)–coronavirus sequences from the earliest identified cases are identical to those found in palm civets and raccoon dogs in animal markets and farms [3*], strongly suggesting an animal origin for this disease.

Rapidly changing ecosystems and human behavior, an ever-increasing density of human and (farmed) animal populations and their close vicinity, poverty, a high degree of mobility and many other factors contribute to the more frequent occurrence and often rapid dissemination of new diseases. Another example of this is the arrival and persistence of West Nile virus in the USA, which is probably a result of increased mobility, an exceptionally broad host range and climatic changes [4]. Among the re-emerging diseases of the past few years, diphtheria and cholera should be mentioned [5], as well as the frequent appearance of multidrug-resistant bacteria and the cases of anthrax infection as a result of deliberate release in 2001.

In the case of influenza, concern for the advent of a new pandemic has been fuelled by reports of 89 human infections by the avian H5N1 virus strain (where H stands for hemagglutinin and N for neuraminidase, the two major surface glycoproteins of the virus) in 2004, leading to 52 deaths (http://www.who.int/csr/disease/avian_influenza/en/). This was the third time in the space of a few years (after the previous outbreaks of infection in 1997 and 2003) that H5N1 avian flu viruses caused disease and death in humans. An H9N2 avian strain caused infection in humans in 1999 and 2003. Because the world's population has not been exposed to these strains, it is immunologically naïve, and these or any other avian flu strains with new surface glycoproteins would meet no

Figure 1



Possible intervention points for vaccine improvement. Schematic view of a pathogen- or vaccine-induced immune response. Vaccines or pathogens cross epithelial barriers and are taken up by antigen-presenting cells such as DCs. Interaction between pattern recognition receptors (PRRs) and their agonists activate DCs, resulting in increased antigen presentation, cytokine production and co-stimulation. $CD4^+$ and $CD8^+$ T cells recognize antigen presented by DCs and are activated. Cognate interaction between primed $CD4^+$ T cells and B cells activates the B cells, resulting in clonal expansion and antibody production. DC activation also leads to inhibition of the regulatory effect of $CD4^+CD25^+$ Treg cells. Fully activated $CD8^+$ cells can target tumor cells and pathogen-infected cells. The letters (a–i) indicate processes where improved vaccines can lead to more efficient immune responses: (a) The site of administration influences the type of immune response and enables the usage of lower vaccine doses [19,20]. (b) Particulate antigen is taken up more easily by macrophages and DCs than soluble antigen [47]. (c) TLR agonists and other immunostimulants binding to PRRs increase the activation of DCs [32–35,36*,37*,38,40]. (d) DCs matured and loaded with antigen *in vitro* are efficient vaccines [41**,42,43]. (e) DNA vaccination leads to efficient antigen presentation on MHC class I [48,49,50*,51**,52,53]. Crosspresentation of antigen on MHC class I of host DCs is facilitated after vaccination with antigen-loaded DCs undergoing delayed apoptosis [44*]. (f) Cytokines can be added in protein or DNA form as natural adjuvants [32,33]. (g) Recruited NK cells can be an early source of Th1-driving cytokines [39*]. (h) Vaccines can break tolerance when the suppressive effect of $CD4^+CD25^+$ Treg cells is overcome [54*]. (i) Pre-existing T cells specific for tumor antigens not contained in the vaccine expand after vaccination and predominate in the antitumor response [55,56*].

resistance provided by existing immunity and could rapidly expand. Once an avian strain has developed a more efficient means of human-to-human transmission, devastating pandemics, such as those arising in 1918, 1957 and 1968, could ensue. It should be noted, however, that older studies found seroprevalence rates for avian flu strains among Chinese rural populations to be between 2% and 7% for H5 viruses and between 15% and 38% for other avian strains [6,7]. Thus, it might in fact be the quality of surveillance, rather than the frequency of outbreaks, that has increased over the past few years [8]. However, the contact between humans and pathogens in animal reservoirs is likely to intensify, and, consequently, the emergence or re-emergence of diseases will occupy vaccinologists frequently in the future.

How to react to new pathogens

One of the most important future challenges will be to respond promptly to emerging diseases such as those mentioned above. A striking example for rapid reaction was in the case of the SARS outbreak, where the genome sequence was publicly available in less than a month after the virus was identified [9]. This enabled the speedy development of diagnostic tools, as well as the identification and recombinant expression of targets for vaccines and therapeutic agents [10–12].

For the influenza virus, in addition to the annual definition of the relevant strains to be included in the vaccine for the following season, the World Health Organization closely monitors cases of avian flu (for further information, see the World Health Organization website indicated above), and prototype vaccines for these strains are being developed. Apart from almost complete lack of protection in the population, an additional threat of the avian flu H5N1 isolate from 2003 is that it kills embryonated eggs, the traditional virus growth substrate used for influenza vaccine production. Such problems can now be solved, owing to the discovery some years ago that influenza virus can be generated entirely from transfected DNA (reverse genetics [13,14]). In this particular case, Webby *et al.* [15•] have used polymerase chain reaction-based mutagenesis to replace the hemagglutinin cleavage site (which was shown to be the cause of high pathogenicity) of the H5N1 strain with the sequence from a nonpathogenic strain. Vero cells were then transfected with plasmids encoding the neuraminidase and mutated hemagglutinin from the circulating strain, together with the plasmids encoding the remaining proteins from the laboratory-optimized PR8 strain. The resulting vaccine strain was successfully grown in eggs and shown to be nonpathogenic and stable. Thus, the use of reverse genetics enables rapid production of a reference vaccine virus in response to the emergence of a new influenza variant [15•,16]. In addition, reverse genetics can be used for more far-reaching vaccination strategies, such as the construction of ‘consensus’ strains expressing conserved amino acid

sequences or more than one version of the surface glycoproteins, or additional immunoenhancing molecules, such as cytokines [8]. Much research effort is also being invested into the development of improved cell culture systems that can replace completely the use of embryonated eggs in vaccine production and would render the production process more flexible and controllable.

Several reports have addressed the question of how to stretch the available supply of vaccine doses in cases of shortage or in the face of a pandemic. Two studies indicated that intradermal, rather than intramuscular, application of 40% [17•] or 20% [18•] of the usual vaccine dose leads to equal or better immune responses. Both theoretical models [19] and trials [20] have shown that immunizing a high proportion of children, known to have a high rate of infection and an important role in transmission, also decreases the incidence of influenza in older age groups, a phenomenon known as herd immunity and described in previous trials in Michigan and Japan [21,22].

Reverse vaccinology

The genomic revolution has opened up a completely new approach to vaccine discovery. For pathogens that do not grow *in vitro*, the availability of the genome sequence has enabled the development of recombinant vaccines, as has been carried out for hepatitis B virus (HBV) and is underway for hepatitis C. In regard to bacteria, group B meningococcus posed insurmountable obstacles to conventional vaccinology approaches; these were eventually overcome by mining the information from the sequenced genome [23]. A total of 600 potential vaccine candidates were predicted by computer analysis, 350 of which were expressed and tested for immunogenicity [24]. Some of these candidates are now in clinical trials. This genome-based approach, called reverse vaccinology, is now used routinely in vaccine development, and is a major tool in the quest for vaccines against pneumococcus, group B streptococcus and chlamydia (see also Update).

Recently, genome sequencing of both *Plasmodium falciparum* [25] and its main vector, *Anopheles gambiae* [26], has sparked off new hopes for an efficient vaccine against malaria. For the rodent models of this disease, subtractive cDNA techniques were used to identify genes that are only expressed in pre-erythrocytic stages of the parasite [27,28]. *Plasmodium* mutants deficient in one of these genes, *uis3*, are blocked in their early liver-stage development and all subsequent stages, and therefore do not lead to disease. When *uis3*-deficient sporozoites are used as genetically attenuated vaccines in mice, they confer long-lasting, stage-specific protection [29••]. This is a promising example of how molecular approaches are employed for the rational design of new vaccines.

Another example of encouraging progress towards a malaria vaccine was reported by Alonso *et al.* [30•]. They

describe Phase IIb trials of a subunit vaccine consisting of a recombinant protein (expressed in yeast) composed of the carboxy-terminal of the *P. falciparum* circumsporozoite protein and the HBV surface antigen. This fusion protein, together with unfused HBV surface antigen proteins, forms particles. The final vaccine formulation includes the adjuvant AS02A, an oil in water emulsion containing the immunostimulants monophosphoryl lipid A (MPL) and *Quillaja saponaria* fraction 21. It had previously been shown that, indeed, a CD4⁺ T cell response to an epitope contained in the vaccine correlates with protection from infection and disease [31^{*}]. In this trial in children, the vaccine showed an efficacy of 30% and 58% in preventing clinical episodes and severe episodes, respectively [30^{*}].

Adjuvants

The past ten years have changed our vision of the immune response to pathogens. It has become clear that the degree and type of antigen-specific, clonal B and T cell responses (acquired immunity) depend crucially on the prior action of a more ancient system of pathogen detection (innate immunity). This system relies on the activation of antigen-presenting cells such as dendritic cells (DCs) upon recognition of patterns common to viruses and bacteria and largely absent in mammals. With the discovery of the involved pattern recognition receptors, among which the Toll-like receptors (TLRs) represent an important subgroup, immune-enhancing molecules or adjuvants can no longer be considered as the alchemistic 'immunologist's dirty secret' but have become amenable to rational design, providing a huge

potential for manipulating the immune response. As different TLR agonists elicit different types of immune responses (reviewed in [59]), future adjuvants might be able to tailor the immune response so that optimal protection to a given pathogen is induced. In fact, the number of clinical trials involving TLR agonists as new adjuvants is ever increasing [32,33] (Table 1).

The adjuvant function of nonmethylated cytidine-phosphate-guanosine (CpG) sequences, which are frequent in microbes but under-represented in humans and are agonists of TLR9, has been extensively demonstrated in animal models [34] and is currently being tested in several clinical trials. When CpGs are coadministered with licensed HBV or flu vaccines, the combination leads to increased antibody titers or increased (interferon- γ) IFN- γ production, as compared with the response to the vaccine alone [35,36^{*}]. An additional effect of CpG oligonucleotides appears to be the promotion of affinity maturation and, as a result, a higher overall affinity of the vaccine-specific antibody pool [37^{*}]. Similarly, the TLR4 agonist MPL has been shown in the past to enhance the immune response to HBV vaccination in humans [38]. In addition, both the malaria subunit vaccine mentioned above [30^{*}] and a licensed melanoma vaccine contain MPL. Another experimental vaccine that includes a TLR2 agonist is described below, in the section on synthetic vaccines.

Activation of DCs increases their ability to process and present antigen and to attract and activate T cells through cytokine secretion; consequently, several cytokines are

Table 1

Human TLR agonists used as adjuvants in vaccine formulations in clinical trials or licensed vaccines^a.

Receptor	Known natural agonist	Form used in vaccines	Vaccine type
TLR1 (with TLR2)	Lipopeptides		
TLR2	Lipopeptides Lipoteichoic acid Porins Zymosan		
TLR3	Double-stranded RNA		
TLR4	Lipopolysaccharide Heat shock proteins Fibrinogen Fibronectin	Monophosphoryl lipid A AS02 (MPL + saponin QS-21) AS04 (MPL + alum) RC-529 (MPL derivative)	Melanoma Malaria [30 [*]], HBV, HPV, HIV-1, cancer, tuberculosis HBV [38] HBV
TLR5	Flagellin		
TLR6 (with TLR2)	Lipoproteins		
TLR7	Unknown		
TLR8	Single stranded RNA		
TLR9	Bacterial DNA	CpG oligonucleotides	HBV, flu [35,36 [*] ,37 [*]] ISS (CpG linked to antigen DNA)
TLR10	Unknown		
TLR11	Components of uropathogenic bacteria		

^a Data from [32,33] unless otherwise indicated.

currently being tested for their adjuvant function. As mentioned above, the way the innate immune system is activated influences the type of the ensuing acquired immune response. Martin-Fontecha *et al.* [39[•]] showed that the ability of adjuvants to elicit a Th1 type response depends crucially on the recruitment of, and IFN- γ production by, natural killer (NK) cells, which indicates a possible mechanism of the way in which adjuvants direct the type of adaptive immune response induced downstream. Another vaccine approach is the use of heat shock proteins, which bind specifically and activate dendritic cells and, because they are loaded with endogenous peptides, can be purified from tumor cells and function as a combined antigen delivery system and adjuvant [40].

Antigen-loaded DCs as vaccines

Because the main targets of adjuvants are DCs, it is a logical step to evaluate their direct use as a vaccine. Despite the labor-intensive necessity of individual cell culture for each patient, this approach can be attractive where other approaches have failed, for instance as a therapeutic vaccine in HIV or cancer patients. When HIV patients were treated with autologous DCs loaded with autologous, inactivated HIV, both virus-specific CD4⁺ Th1 and CD8⁺ responses were induced and the plasma viral loads were reduced [41^{••}]. These results closely reflect previous findings from a similar vaccination of rhesus macaques [42], except for the lack of induction of neutralizing antibodies in the human study. DC vaccination is also being tested in a variety of cancer treatments [43].

In a study comparing the immunogenicity of DCs transfected with cytopathic or noncytopathic viral RNA, the former regimen was shown to be more efficient at inducing protective immune responses [44[•]]. This suggests that reprocessing of dying DCs by endogenous antigen-presenting cells enhances immunogenicity, either through additional danger signals triggered by the cell damage or by the increased level of crosspresentation by endogenous DCs. The same mechanism might be at work in mycobacterium infection of macrophages, where apoptosis was shown to enhance crosspresentation by bystander DCs [45]. Interestingly, following fractionation of the cytoplasm of dying cells, uric acid was identified as a highly efficient endogenous danger signal that enhances immunogenicity [46], and might explain the above results.

DNA vaccines

The high expectations associated with DNA vaccination, as a result of promising data obtained in mice, were somewhat tempered by disappointing early results when DNA was tested as a vaccine in humans. Therefore, the latest generation of DNA vaccines rely on improved delivery either through use of microparticles [47] or through viral vectors. A particularly promising approach is a heterologous prime-boost strategy, where adminis-

tration of plasmid DNA is followed by recombinant virus (modified vaccinia virus Ankara [MVA] or adenovirus) expressing the same antigen. This regimen induced strong T cell responses against *P. falciparum* in naïve adults [48] and enhanced the response in Gambian men who are constantly exposed to *P. falciparum* [49]. Although partial protection against challenge with a different *P. falciparum* strain was observed, no significant differences in the infection rate was found in the Gambian trial [50[•]]. In spite of these setbacks, a very similar regimen has been shown also to be highly immunogenic against HBV, tuberculosis and HIV, and vaccination only with MVA expressing the mycobacterium A85 antigen elicited strong T cell responses [51^{••}].

Because the induction of T cells is considered to be crucial in anti-tumor immune responses, a huge number of trials are presently being conducted to test DNA vaccination regimens for anticancer treatment [52,53]. When viral vehicles (vaccinia or adenovirus) were compared with loaded DCs in terms of their ability to overcome established tolerance and induce immune responses in a transgenic mouse model, the viral formulations were able to do so, whereas DCs required repeated administration of TLR agonists or irrelevant virus, or removal of suppressive CD4⁺CD25⁺ Treg cells [54[•]]. Such models of established tolerance might prove useful for testing the success of vaccines in the face of long-term antigen exposure, as in the case of cancer or chronic diseases.

Two studies analyzed in detail the T cell response after vaccination with a recombinant canarypox virus expressing melanoma-specific T cell epitopes [55[•],56[•]]. Focusing on the blood and metastases from a patient with complete regression, these studies reconfirmed earlier observations that the frequency of anti-tumor T cells can be relatively high. Although vaccination leads to a slight increase in vaccine-specific T cells, these remain only a small fraction of total anti-tumor T cells. By contrast, T cells directed against epitopes not contained in the vaccine represent the vast majority of anti-tumor cells, and their frequency increases both in the blood and in metastases. Thus, it appears that with the appropriate vaccine regimen, the inefficiency of pre-existing specific T cells to combat the tumor was reversed in an indirect manner, presumably by activating another subset of T cells. It remains to be clarified, however, how much of this reactivation is due to the action of vaccine-induced T cells and how much is a general, antigen-independent immune-enhancing effect. In any case, for the development of therapeutic vaccines, it will be vital to understand how the balance can be tipped back from a state of tolerance to a successful immune response.

Synthetic vaccines

The development of vaccines aimed at the polysaccharide (PS) capsule of bacteria is one of the great

achievements in vaccinology. So far, the PS used in large-scale vaccine production has been purified from the pathogen itself, grown in large quantities – an approach that is costly and difficult to control. Through great simplification of the carbohydrate chemistry involved, Verez-Bencomo *et al.* [57^{*}] have now demonstrated the first large-scale production of an anti-*Haemophilus influenzae* type B vaccine, consisting of synthetic PS conjugated to tetanus toxoid protein carrier. This vaccine has been shown to be as efficient as commercially available vaccines in inducing protective levels of antibody titers in infants.

An entirely synthetic vaccine with a branched structure containing a TLR2 ligand, a CD4⁺ T cell epitope and either a CD8⁺ T cell or a B cell epitope has been shown to elicit strong CD8⁺ T cell and B cell responses, respectively [58^{**}]. Here, the minimal requirements for an efficient vaccine are met in a single molecule: targeting to and activation of DCs, T cell help and activation of antigen-specific CD8⁺ T cells or B cells.

Conclusions

The world of vaccines is undergoing dramatic changes. Never before have such sophisticated techniques and an in-depth knowledge of immunological processes been at hand to exploit fully the potential of protecting from, as well as curing, diseases through vaccination. A formidable task in the future will be the development of effective therapeutic vaccines, in situations where chronic antigen exposure by itself does not elicit a sufficiently strong immune response, as is the case in cancer and chronic infectious diseases. Compared with vaccines against self-limiting infections, where the aim is to be as good as the real pathogen (but less harmful), this requires the development of vaccines that are better than the natural antigens in inducing immunity. All of our knowledge will be necessary to succeed in this challenge.

Update

A recent article describes the use of multigenome analysis and screening against a large panel of strains to identify a universal group B streptococcus vaccine. Although none of the single antigens contained in the vaccine elicits protection against all strains, the combination of four proteins is able to cover a wide range of strains [60^{*}].

Acknowledgements

We are grateful to Giorgio Corsi for artwork.

References and recommended reading

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
 - of outstanding interest
1. Gao F, Bailes E, Robertson DL, Chen Y, Rodenburg CM, Michael SF, Cummins LB, Arthur LO, Peeters M, Shaw GM *et al.*: **Origin of HIV-1 in the chimpanzee Pan troglodytes troglodytes.** *Nature* 1999, **397**:436-441.
 2. Leroy EM, Rouquet P, Formenty P, Souquiere S, Kilbourne A, Froment JM, Bermejo M, Smit S, Karesh W, Swanepoel R *et al.*: **Multiple Ebola virus transmission events and rapid decline of central African wildlife.** *Science* 2004, **303**:387-390.
 - Human Ebola virus outbreaks consist of parallel epidemics caused by several different viral strains which appear to be transmitted to humans through handling of gorilla, chimpanzee or duiker carcasses.
 3. Chinese SMEC: **Molecular evolution of the SARS coronavirus during the course of the SARS epidemic in China.** *Science* 2004, **303**:1666-1669.
 - The Chinese SARS consortium demonstrates that virus sequences from the earliest patients are identical to those found in palm civets, and seven out of 11 cases had documented contact with wild animals. In addition, the isolated case in December 2003 was caused by a virus with highest sequence homology to palm civets, again suggesting the animal origin of the SARS coronavirus.
 4. Granwehr BP, Lillibridge KM, Higgs S, Mason PW, Aronson JF, Campbell GA, Barrett AD: **West Nile virus: where are we now?** *Lancet Infect Dis* 2004, **4**:547-556.
 5. Rappuoli R: **From Pasteur to genomics: progress and challenges in infectious diseases.** *Nat Med* 2004, **10**:1177-1185.
 6. Shortridge KF: **Pandemic influenza: a zoonosis?** *Semin Respir Infect* 1992, **7**:11-25.
 7. Profeta ML, Palladino G: **Serological evidence of human infections with avian influenza viruses. Brief report.** *Arch Virol* 1986, **90**:355-360.
 8. Palese P: **Influenza: old and new threats.** *Nat Med* 2004, **10**:S82-S87.
 9. Stadler K, Massignani V, Eickmann M, Becker S, Abrignani S, Klenk HD, Rappuoli R: **SARS – beginning to understand a new virus.** *Nat Rev Microbiol* 2003, **1**:209-218.
 10. Yang ZY, Kong WP, Huang Y, Roberts A, Murphy BR, Subbarao K, Nabel GJ: **A DNA vaccine induces SARS coronavirus neutralization and protective immunity in mice.** *Nature* 2004, **428**:561-564.
 11. Bisht H, Roberts A, Vogel L, Bukreyev A, Collins PL, Murphy BR, Subbarao K, Moss B: **Severe acute respiratory syndrome coronavirus spike protein expressed by attenuated vaccinia virus protectively immunizes mice.** *Proc Natl Acad Sci USA* 2004, **101**:6641-6646.
 12. Traggiai E, Becker S, Subbarao K, Kolesnikova L, Uematsu Y, Gismondo MR, Murphy BR, Rappuoli R, Lanzavecchia A: **An efficient method to make human monoclonal antibodies from memory B cells: potent neutralization of SARS coronavirus.** *Nat Med* 2004, **10**:871-875.
 13. Fodor E, Devenish L, Engelhardt OG, Palese P, Brownlee GG, Garcia-Sastre A: **Rescue of influenza A virus from recombinant DNA.** *J Virol* 1999, **73**:9679-9682.
 14. Neumann G, Watanabe T, Ito H, Watanabe S, Goto H, Gao P, Hughes M, Perez DR, Donis R, Hoffmann E *et al.*: **Generation of influenza A viruses entirely from cloned cDNAs.** *Proc Natl Acad Sci USA* 1999, **96**:9345-9350.
 15. Webby RJ, Perez DR, Coleman JS, Guan Y, Knight JH, Govorkova EA, McClain-Moss LR, Peiris JS, Rehg JE, Tuomanen EI *et al.*: **Responsiveness to a pandemic alert: use of reverse genetics for rapid development of influenza vaccines.** *Lancet* 2004, **363**:1099-1103.
 - This paper demonstrates the rapid generation of a genetically attenuated influenza H5N1 reference vaccine strain by use of reverse genetics.
 16. Subbarao K, Chen H, Swayne D, Mingay L, Fodor E, Brownlee G, Xu X, Lu X, Katz J, Cox N *et al.*: **Evaluation of a genetically modified reassortant H5N1 influenza A virus vaccine candidate generated by plasmid-based reverse genetics.** *Virology* 2003, **305**:192-200.
 17. Belshe RB, Newman FK, Cannon J, Duane C, Treanor J, Van Hoecke C, Howe BJ, Dubin G: **Serum antibody responses after intradermal vaccination against influenza.** *N Engl J Med* 2004, **351**:2286-2294.
 - See annotation to [18^{*}].

18. Kenney RT, Frech SA, Muenz LR, Villar CP, Glenn GM: **Dose sparing with intradermal injection of influenza vaccine.** *N Engl J Med* 2004, **351**:2295-2301.
- These two studies show that intradermal, instead of intramuscular, vaccination enables the use of considerably less antigen to induce a similar degree of protection.
19. Weycker D, Edelsberg J, Elizabeth Halloran M, Longini IM Jr, Nizam A, Ciuryla V, Oster G: **Population-wide benefits of routine vaccination of children against influenza.** *Vaccine* 2005, **23**:1284-1293.
20. Piedra PA, Gaglani MJ, Kozinetz CA, Herschler G, Riggs M, Griffith M, Fewlass C, Watts M, Hessel C, Cordova J *et al.*: **Herd immunity in adults against influenza-related illnesses with use of the trivalent-live attenuated influenza vaccine (CAIV-T) in children.** *Vaccine* 2005, **23**:1540-1548.
21. Reichert TA, Sugaya N, Fedson DS, Glezen WP, Simonsen L, Tashiro M: **The Japanese experience with vaccinating schoolchildren against influenza.** *N Engl J Med* 2001, **344**:889-896.
22. Monto AS, Davenport FM, Napier JA, Francis T Jr: **Modification of an outbreak of influenza in Tecumseh, Michigan by vaccination of schoolchildren.** *J Infect Dis* 1970, **122**:16-25.
23. Mora M, Veggi D, Santini L, Pizza M, Rappuoli R: **Reverse vaccinology.** *Drug Discov Today* 2003, **8**:459-464.
24. Pizza M, Scarlato V, Masignani V, Giuliani MM, Arico B, Comanducci M, Jennings GT, Baldi L, Bartolini E, Capecchi B *et al.*: **Identification of vaccine candidates against serogroup B meningococcus by whole-genome sequencing.** *Science* 2000, **287**:1816-1820.
25. Gardner MJ, Hall N, Fung E, White O, Berriman M, Hyman RW, Carlton JM, Pain A, Nelson KE, Bowman S *et al.*: **Genome sequence of the human malaria parasite *Plasmodium falciparum*.** *Nature* 2002, **419**:498-511.
26. Holt RA, Subramanian GM, Halpern A, Sutton GG, Charlab R, Nussskern DR, Wincker P, Clark AG, Ribeiro JM, Wides R *et al.*: **The genome sequence of the malaria mosquito *Anopheles gambiae*.** *Science* 2002, **298**:129-149.
27. Matuschewski K, Ross J, Brown SM, Kaiser K, Nussenzweig V, Kappe SH: **Infectivity-associated changes in the transcriptional repertoire of the malaria parasite sporozoite stage.** *J Biol Chem* 2002, **277**:41948-41953.
28. Kaiser K, Matuschewski K, Camargo N, Ross J, Kappe SH: **Differential transcriptome profiling identifies *Plasmodium* genes encoding pre-erythrocytic stage-specific proteins.** *Mol Microbiol* 2004, **51**:1221-1232.
29. Mueller AK, Labaied M, Kappe SH, Matuschewski K: **Genetically modified *Plasmodium* parasites as a protective experimental malaria vaccine.** *Nature* 2005, **433**:164-167.
- In this study, reverse genetics was used to generate a *Plasmodium* knockout strain with a developmental block at the early liver stage. If used as a live attenuated vaccine in a malaria mouse model, this strain confers protection against infectious sporozoite challenge.
30. Alonso PL, Sacarlal J, Aponte JJ, Leach A, Macete E, Millman J, Mandomando I, Spiessens B, Guinovart C, Espasa M *et al.*: **Efficacy of the RTS, S/AS02A vaccine against *Plasmodium falciparum* infection and disease in young African children: randomised controlled trial.** *Lancet* 2004, **364**:1411-1420.
- This vaccine contains a yeast-expressed hybrid molecule composed of a part of the circumsporozoite protein and the HBV surface antigen, formulated in a complex adjuvant. Significant efficacy, in terms of prevention of clinical episodes and of severe malaria, was observed.
31. Reece WH, Pinder M, Gothard PK, Milligan P, Bojang K, Doherty T, Plebanski M, Akinwunmi P, Everaere S, Watkins KR *et al.*: **A CD4(+) T-cell immune response to a conserved epitope in the circumsporozoite protein correlates with protection from natural *Plasmodium falciparum* infection and disease.** *Nat Med* 2004, **10**:406-410.
- Using an enzyme-linked immunospot assay on T cells previously expanded *in vitro*, the authors identified a conserved CD4⁺ T cell epitope in the circumsporozoite protein with predictive value: an IFN- γ response of CD4⁺ T cells to this epitope positively correlated with protection from malaria.
32. Engers H, Kieny MP, Malhotra P, Pink JR: **Third meeting on Novel Adjuvants Currently in or Close to Clinical Testing World Health Organization—Organisation Mondiale de la Sante, Fondation Merieux, Annecy, France, 7-9 January 2002.** *Vaccine* 2003, **21**:3503-3524.
33. Pink JR, Kieny MP: **4th meeting on Novel Adjuvants Currently in/close to Human Clinical Testing World Health Organization—organisation Mondiale de la Sante Fondation Merieux, Annecy, France, 23-25, June 2003.** *Vaccine* 2004, **22**:2097-2102.
34. Klinman DM, Currie D, Gursel I, Verthelyi D: **Use of CpG oligodeoxynucleotides as immune adjuvants.** *Immunol Rev* 2004, **199**:201-216.
35. Halperin SA, Van Nest G, Smith B, Abtahi S, Whiley H, Eiden JJ: **A phase I study of the safety and immunogenicity of recombinant hepatitis B surface antigen co-administered with an immunostimulatory phosphorothioate oligonucleotide adjuvant.** *Vaccine* 2003, **21**:2461-2467.
36. Cooper CL, Davis HL, Morris ML, Efler SM, Adhami MA, Krieg AM, Cameron DW, Heathcote J: **CPG 7909, an immunostimulatory TLR9 agonist oligodeoxynucleotide, as adjuvant to engerix-B(R) HBV vaccine in healthy adults: a double-blind phase I/II study.** *J Clin Immunol* 2004, **24**:693-701.
- See annotation to [37*].
37. Siegrist CA, Pihlgren M, Tougne C, Efler SM, Morris ML, AlAdhami MJ, Cameron DW, Cooper CL, Heathcote J, Davis HL *et al.*: **Co-administration of CpG oligonucleotides enhances the late affinity maturation process of human anti-hepatitis B vaccine response.** *Vaccine* 2004, **23**:615-622.
- In these two clinical trials, CpG was shown to be an efficient adjuvant when used in combination with licensed vaccines.
38. Jacques P, Moens G, Desombere I, Dewijngaert J, Leroux-Roels G, Wettendorff M, Thoelen S: **The immunogenicity and reactogenicity profile of a candidate hepatitis B vaccine in an adult vaccine non-responder population.** *Vaccine* 2002, **20**:3644-3649.
39. Martin-Fontecha A, Thomsen LL, Brett S, Gerard C, Lipp M, Lanzavecchia A, Sallusto F: **Induced recruitment of NK cells to lymph nodes provides IFN-gamma for T(H)1 priming.** *Nat Immunol* 2004, **5**:1260-1265.
- The authors showed that the ability of some adjuvants to induce Th1 priming correlates with the degree of recruitment of NK cells, which represent an early source of IFN- γ . This might provide a rationale for the search for efficient Th1-inducing adjuvants.
40. Lewis JJ: **Therapeutic cancer vaccines: using unique antigens.** *Proc Natl Acad Sci USA* 2004, **101**(Suppl. 2):14653-14656.
41. Lu W, Arraes LC, Ferreira WT, Andrieu JM: **Therapeutic dendritic-cell vaccine for chronic HIV-1 infection.** *Nat Med* 2004, **10**:1359-1365.
- This study followed a previous one in macaques [42] and showed that injection of autologous DCs loaded with autologous inactivated HIV induces HIV-specific CD4⁺ and CD8⁺ T cell responses and leads to a prolonged tenfold reduction in virus load in about half of the patients.
42. Lu W, Wu X, Lu Y, Guo W, Andrieu JM: **Therapeutic dendritic-cell vaccine for simian AIDS.** *Nat Med* 2003, **9**:27-32.
43. Cerundolo V, Hermans IF, Salio M: **Dendritic cells: a journey from laboratory to clinic.** *Nat Immunol* 2004, **5**:7-10.
44. Racanelli V, Behrens SE, Aliberti J, Rehmann B: **Dendritic cells transfected with cytopathic self-replicating RNA induce crosspriming of CD8+ T cells and antiviral immunity.** *Immunity* 2004, **20**:47-58.
- The surprising observation in this study is that induction of immunity and crosspresentation of antigen is greatly facilitated by the death of the injected antigen-carrying DCs. This suggests that danger signals from dying cells might have an adjuvant effect in vaccinations, as well as in immune responses.
45. Schaible UE, Winau F, Sieling PA, Fischer K, Collins HL, Hagens K, Modlin RL, Brinkmann V, Kaufmann SH: **Apoptosis facilitates antigen presentation to T lymphocytes through MHC-I and CD1 in tuberculosis.** *Nat Med* 2003, **9**:1039-1046.
46. Shi Y, Evans JE, Rock KL: **Molecular identification of a danger signal that alerts the immune system to dying cells.** *Nature* 2003, **425**:516-521.

47. O'Hagan DT, Singh M, Ulmer JB: **Microparticles for the delivery of DNA vaccines.** *Immunol Rev* 2004, **199**:191-200.
48. McConkey SJ, Reece WH, Moorthy VS, Webster D, Dunachie S, Butcher G, Vuola JM, Blanchard TJ, Gothard P, Watkins K *et al.*: **Enhanced T-cell immunogenicity of plasmid DNA vaccines boosted by recombinant modified vaccinia virus Ankara in humans.** *Nat Med* 2003, **9**:729-735.
49. Moorthy VS, Pinder M, Reece WH, Watkins K, Atabani S, Hannan C, Bojang K, McAdam KP, Schneider J, Gilbert S *et al.*: **Safety and immunogenicity of DNA/modified vaccinia virus ankara malaria vaccination in African adults.** *J Infect Dis* 2003, **188**:1239-1244.
50. Moorthy VS, Imoukhuede EB, Milligan P, Bojang K, Keating S, Kaye P, Pinder M, Gilbert SC, Walraven G, Greenwood BM *et al.*: **A randomised, double-blind, controlled vaccine efficacy trial of DNA/MVA ME-TRAP against malaria infection in Gambian adults.** *PLoS Med* 2004, **1**:e33.
- In spite of the induction of strong T cell responses by this DNA-MVA prime-boost vaccine, it was not effective at reducing the natural malaria infection rate in this trial.
51. McShane H, Pathan AA, Sander CR, Keating SM, Gilbert SC, Huygen K, Fletcher HA, Hill AV: **Recombinant modified vaccinia virus Ankara expressing antigen 85A boosts BCG-primed and naturally acquired antimycobacterial immunity in humans.** *Nat Med* 2004, **10**:1240-1244.
- The authors observed antigen-specific IFN- γ responses of short duration induced by vaccination with MVA expressing the conserved mycobacterial antigen 85A. This T cell response is substantially higher and sustained in vaccinees who received bacillus Calmette-Guérin followed by MVA three weeks later, indicating that this prime-boost regimen is a promising tuberculosis vaccine strategy.
52. Liu M, Acres B, Balloul JM, Bizouarne N, Paul S, Slos P, Squiban P: **Gene-based vaccines and immunotherapeutics.** *Proc Natl Acad Sci USA* 2004, **101**(Suppl. 2):14567-14571.
53. Stevenson FK, Ottensmeier CH, Johnson P, Zhu D, Buchan SL, McCann KJ, Roddick JS, King AT, McNicholl F, Savelyeva N *et al.*: **DNA vaccines to attack cancer.** *Proc Natl Acad Sci USA* 2004, **101**(Suppl. 2):14646-14652.
54. Yang Y, Huang CT, Huang X, Pardoll DM: **Persistent Toll-like receptor signals are required for reversal of regulatory T cell-mediated CD8 tolerance.** *Nat Immunol* 2004, **5**:508-515.
- This transgenic adoptive transfer model of tolerance enables the comparison of different vaccine regimens in their ability to break tolerance. Viral vaccines are most efficient in overcoming established tolerance. DC-based vaccine strategies that are equivalent in priming naïve T cells were able to break tolerance only when TLR ligands were added or after removal of regulatory T cells.
55. Lurquin C, Lethe B, De Plaen E, Corbiere V, Theate I, van Baren N, Coulie PG, Boon T: **Contrasting frequencies of antitumor and anti-vaccine T cells in metastases of a melanoma patient vaccinated with a MAGE tumor antigen.** *J Exp Med* 2005, **201**:249-257.
- See annotation to [56*].
56. Germeau C, Ma W, Schiavetti F, Lurquin C, Henry E, Vigneron N, Brasseur F, Lethe B, De Plaen E, Velu T *et al.*: **High frequency of antitumor T cells in the blood of melanoma patients before and after vaccination with tumor antigens.** *J Exp Med* 2005, **201**:241-248.
- These two studies showed that in the blood and metastases of vaccinated cancer patients, T cells specific for the vaccine antigen might represent only a small fraction of total tumor-specific T cells, both before and after vaccination. This suggests that a prevaccine, spontaneous antitumor T cell response exists, becomes ineffective but can be reactivated indirectly by the vaccine.
57. Verez-Bencomo V, Fernandez-Santana V, Hardy E, Toledo ME, Rodriguez MC, Heynngnezz L, Rodriguez A, Baly A, Herrera L, Izquierdo M *et al.*: **A synthetic conjugate polysaccharide vaccine against Haemophilus influenzae type b.** *Science* 2004, **305**:522-525.
- The large-scale production of a conjugate vaccine containing synthetic polysaccharides has been achieved by simplification of the polysaccharide chemistry involved.
58. Jackson DC, Lau YF, Le T, Suhrbier A, Deliyannis G, Cheers C, Smith C, Zeng W, Brown LE: **A totally synthetic vaccine of generic structure that targets Toll-like receptor 2 on dendritic cells and promotes antibody or cytotoxic T cell responses.** *Proc Natl Acad Sci USA* 2004, **101**:15440-15445.
- This elegant study showed how all of the components required for an efficient vaccine can be included in one synthetic molecule: a T helper epitope, a TLR agonist as an adjuvant and either a CD8⁺ T cell or a B cell epitope. The resultant vaccines confer protection and are able to break tolerance in mice.
59. Pulendran B: **Variation of the immune response with dendritic cells and pathogen recognition receptors.** *J Immunol* 2005, **174**:2457-2465.
60. Maione D, Margalit Y, Ros I, Rinaudo D, Massignani V, Mora M, Scarselli M, Tettelin H, Brettoni C, Iacobini ET *et al.*: **Identification of a universal group B streptococcus vaccine by multiple genome screen.** *Science*, in press.
- The use of different group B streptococcus strains for genome analysis and screening enables the development of a universal vaccine. The single vaccine components are not present in all strains, and do not protect against all strains, but the combination of them confers wide range protection.