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Expression of PEG10 Is Associated with Poor Survival and Tumor Recurrence in Hepatocellular Carcinoma

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Purpose

Paternally expressed gene 10 (*PEG10*), first identified as an imprinted gene, is paternally expressed and maternally silenced. In hepatocellular carcinoma (HCC), PEG10 has been identified as a potential target gene located within the amplified 7q21 locus. The purpose of this study was to investigate the expression of PEG10 protein in HCC and evaluate its prognostic significance.

Materials and Methods

PEG10 protein expression was examined by immunohistochemistry in tumor tissues from 218 HCC patients undergoing curative resection. Furthermore, the relationships between PEG10 expression and clinicopathologic features or postoperative survival of HCC patients were evaluated. The median follow-up period was 119.8 months for survivors.

Results

PEG10 expression was observed in 148 of the 218 HCCs (67.9%) and was significantly correlated with younger age, female, higher Edmondson grade, microvascular invasion, intrahepatic metastasis, higher American Joint Committee on Cancer T-stage, and higher α -fetoprotein level. PEG10 expression was an independent predictor of early recurrence (p=0.013), and it showed an unfavorable influence on recurrence-free survival (p < 0.001). A subgroup analysis showed that among patients with α -fetoprotein \leq 20 ng/mL (80 patients), the PEG10-positive group also showed an unfavorable influence on recurrence-free survival (p=0.002). Moreover, a multivariate survival analysis identified PEG10 as an independent predictor of shorter recurrence-free survival (p=0.005). PEG10 expression showed an unfavorable influence on overall survival (p=0.007) but was not an independent predictor of shorter overall survival (p=0.128).

Conclusion

PEG10 protein could be a potential biomarker predicting early recurrence and recurrencefree survival in HCC patients after curative resection, even in those with normal serum α -fetoprotein levels.

> Key words PEG10, Hepatocellular carcinoma, Recurrence

Introduction

encompasses a wide variety of cell types, effective therapeutic approaches require a better understanding of molecular mechanisms that drive oncogenesis and clarify the responsiveness of different cancer phenotypes to potential therapeutic agents [3].

Paternally expressed gene 10 (PEG10) was first described by Ono et al. [4] as an imprinted gene with an active paternal allele but silent maternal allele and is generally not expressed in the normal adult liver. It has been suggested that PEG10

resection, and resistance to chemotherapy [1,2]. It is a necessity to identify patients with poor prognosis for timely intervention. Since cancer is a heterogeneous disease that in the

The overall prognosis of hepatocellular carcinoma (HCC)

patients remains poor due to many risk factors, including late

diagnosis, high incidence of tumor recurrence after curative

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Fig. 1. Immunostaining of paternally expressed gene 10 in normal liver shows no immunoreactivity in normal hepatocytes (horseradish peroxidase staining, ×200).

is derived from a retrotransposon that was previously integrated into the mammalian genome [4]. PEG10 was overexpressed in a variety of human cancers, including leukemia, breast cancer, prostate cancer, pancreas cancer, gallbladder cancer, and HCC [5-7]. Furthermore, PEG10 was a poorprognosis related biomarker for gallbladder adenocarcinoma [6]. In HCC, *PEG10* has been identified as a potential target gene located within the amplified 7q21 locus [8-10]. A functional role for PEG10 in the growth-promoting activities has been demonstrated in the HCC cells [7]. Ip et al. [8] reported a significant progressive trend of increasing PEG10 expression from the putative pre-malignant adjacent livers to early resectable HCCs, and to late inoperable HCCs. However, the prognostic significance of PEG10 protein expression in HCC has not yet been elucidated. In this study, we investigated PEG10 protein expression by immunohistochemistry in order to identify a marker associated with tumor recurrence and to evaluate the prognostic significance of PEG10 in 218 HCC patients with longterm follow-up.

Materials and Methods

1. Tissue samples

Liver tissue samples from 218 surgically resected HCC were collected between July 2000 and May 2006 from Samsung Medical Center, Seoul, Korea. The cases were selected based on the following criteria: pathological diagnosis of HCC and curative resection of tumor without preoperative or postoperative adjuvant therapy. Curative resection was defined by a complete resection of all tumor nodules with clear microscopic resection margins and no residual tumors as indicated by a computed tomography scan one month after surgery. Tumor differentiation was determined based on the criteria proposed by Edmondson and Steiner [11]. A microvascular invasion was considered present when at least one or more endothelial cells or tunica media of the vessel surrounded the neoplastic cell group. Intrahepatic metastasis and multicentric occurrence were defined in accordance to the previously reported criteria [12]. All patients were staged using the American Joint Committee on Cancer (AJCC) staging system [13] and Barcelona Clinic Liver Cancer (BCLC) staging classification [14]. Early recurrence after curative resection of HCC was defined as an intrahepatic or extrahepatic recurrence within 2 years [15]. The Institutional Review Board of Samsung Medical Center



Fig. 2. Immunostaining of paternally expressed gene 10 in hepatocellular carcinomas showing positive cytoplasmic expression (A) or negative expression (B) (horseradish peroxidase staining, ×200).

Table 1. Correlation between PEG10 expression and theclinicopathologic features in 218 hepatocellular carcinomas

Age (yr)0.046 ≤ 55 13095 (73.1)>558853 (60.2)Gender0.037Female3932 (82.1)Male179116 (64.8)Tumor size (cm)0.772 ≤ 5.0 8458 (69.0)Edmondson grade0.021I188 (44.4)II151101 (66.9)III4939 (79.6)Microvascular invasion0.001(-)9452 (55.3)(+)12496 (77.4)Major portal vein invasion0.008(-)208142 (68.3)(+)162102 (63.0)(+)5646 (82.1)Multicentric occurrece> 0.999(-)209142 (67.9)(+)951 (56.0)28564 (75.3)33931 (79.5)432 (66.7)C117 (64.3)B9267 (72.8)C117 (65.6)<3.5191127 (66.5)<3.52721 (77.8)AIP level (ny /L)*< 0.014Non-viral2715 (55.6)HBV171122 (71.3)HCV2011 (55.0)Liver cirrhosis0.012(-)10767 (62.6)(-)11481 (73.0)	Variable	No.	PEG10 expression	p-value
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$\begin{array}{c c c c c c c } & 209 & 142 (67.9) \\ (+) & 9 & 6 (66.7) \\ \hline \\ AJCC T-stage & 0.011 \\ 1 & 91 & 51 (56.0) \\ 2 & 85 & 64 (75.3) \\ 3 & 39 & 31 (79.5) \\ 4 & 3 & 2 (66.7) \\ \hline \\ BCLC stage & 0.390 \\ \hline \\ 0-A & 115 & 74 (64.3) \\ B & 92 & 67 (72.8) \\ \hline \\ C & 11 & 7 (63.6) \\ \hline \\ Albumin level (g/dL) & 0.240 \\ > 3.5 & 191 & 127 (66.5) \\ < 3.5 & 27 & 21 (77.8) \\ \hline \\ AFP level (ng/mL)^{a)} & < 0.001 \\ \leq 200 & 126 & 70 (55.6) \\ > 200 & 84 & 73 (86.9) \\ \hline \\ Etiology & 0.114 \\ Non-viral & 27 & 15 (55.6) \\ HBV & 171 & 122 (71.3) \\ HCV & 20 & 11 (55.0) \\ \hline \\ Liver cirrhosis & 0.102 \\ (-) & 107 & 67 (62.6) \\ (+) & 111 & 81 (73.0) \\ \hline \end{array}$	Multicentric occurr	ence	× ,	> 0.999
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$\begin{array}{c ccccc} 2 & 85 & 64 & (75.3) \\ 3 & 39 & 31 & (79.5) \\ 4 & 3 & 2 & (66.7) \\ \hline BCLC stage & 0.390 \\ \hline 0-A & 115 & 74 & (64.3) \\ B & 92 & 67 & (72.8) \\ \hline C & 11 & 7 & (63.6) \\ \hline Albumin level (g/dL) & 0.240 \\ > 3.5 & 191 & 127 & (66.5) \\ < 3.5 & 27 & 21 & (77.8) \\ \hline AFP level (ng/mL)^{a)} & < 0.001 \\ < 200 & 126 & 70 & (55.6) \\ > 200 & 84 & 73 & (86.9) \\ \hline Etiology & 0.114 \\ \hline Non-viral & 27 & 15 & (55.6) \\ \hline HBV & 171 & 122 & (71.3) \\ \hline HCV & 20 & 11 & (55.0) \\ \hline Liver cirrhosis & 0.102 \\ (-) & 107 & 67 & (62.6) \\ (+) & 111 & 81 & (73.0) \\ \hline \end{array}$	1	91	51 (56.0)	
$\begin{array}{ccccccc} 3 & 39 & 31 (79.5) \\ 4 & 3 & 2 (66.7) \\ \hline BCLC stage & 0.390 \\ \hline 0-A & 115 & 74 (64.3) \\ B & 92 & 67 (72.8) \\ C & 11 & 7 (63.6) \\ \hline Albumin level (g/dL) & 0.240 \\ > 3.5 & 191 & 127 (66.5) \\ \leq 3.5 & 27 & 21 (77.8) \\ \hline AFP level (ng/mL)^{a)} & < 0.001 \\ \leq 200 & 126 & 70 (55.6) \\ > 200 & 84 & 73 (86.9) \\ \hline Etiology & 0.114 \\ Non-viral & 27 & 15 (55.6) \\ HBV & 171 & 122 (71.3) \\ HCV & 20 & 11 (55.0) \\ \hline Liver cirrhosis & 0.102 \\ (-) & 107 & 67 (62.6) \\ (+) & 111 & 81 (73.0) \\ \hline \end{array}$	2	85	64 (75.3)	
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$\begin{array}{c c c c c c c } 0-A & 115 & 74 (64.3) \\ B & 92 & 67 (72.8) \\ \hline C & 11 & 7 (63.6) \\ \hline Albumin level (g/dL) & 0.240 \\ > 3.5 & 191 & 127 (66.5) \\ \leq 3.5 & 27 & 21 (77.8) \\ \hline AFP level (ng/mL)^{a)} & <0.001 \\ \leq 200 & 126 & 70 (55.6) \\ > 200 & 84 & 73 (86.9) \\ \hline Etiology & 0.114 \\ \hline Non-viral & 27 & 15 (55.6) \\ \hline HBV & 171 & 122 (71.3) \\ \hline HCV & 20 & 11 (55.0) \\ \hline Liver cirrhosis & 0.102 \\ (-) & 107 & 67 (62.6) \\ (+) & 111 & 81 (73.0) \\ \hline \end{array}$	BCLC stage			0.390
B92 $67 (72.8)$ C117 (63.6)Albumin level (g/dL)0.240> 3.5191127 (66.5) ≤ 3.5 2721 (77.8)AFP level (ng/mL)a)< 0.001	0-A	115	74 (64.3)	
C117 (63.6)Albumin level (g/dL)0.240> 3.5191127 (66.5) ≤ 3.5 2721 (77.8)AFP level (ng/mL) ^{a)} < 0.001	В	92	67 (72.8)	
Albumin level (g/dL) 0.240 > 3.5 191 127 (66.5) \leq 3.5 27 21 (77.8) AFP level (ng/mL) ^{a)} < 0.001 \leq 200 126 70 (55.6) > 200 84 73 (86.9) Etiology 0.114 Non-viral 27 15 (55.6) HBV 171 122 (71.3) HCV 20 11 (55.0) Liver cirrhosis 0.102 (-) 107 67 (62.6) (+) 111 81 (73.0)	С	11	7 (63.6)	
> 3.5 191 127 (66.5) \leq 3.5 27 21 (77.8) AFP level (ng/mL) ^a) < 0.001	Albumin level (g/d	łL)		0.240
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	> 3.5	191	127 (66.5)	
$\begin{array}{c c c c c c c } AFP \mbox{ level } (ng/mL)^{a)} & < 0.001 \\ \leq 200 & 126 & 70 (55.6) \\ > 200 & 84 & 73 (86.9) \\ \hline Etiology & 0.114 \\ Non-viral & 27 & 15 (55.6) \\ HBV & 171 & 122 (71.3) \\ HCV & 20 & 11 (55.0) \\ \hline Liver cirrhosis & 0.102 \\ (-) & 107 & 67 (62.6) \\ (+) & 111 & 81 (73.0) \\ \hline \end{array}$	≤ 3.5	27	21 (77.8)	
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	AFP level (ng/mL)	a)		< 0.001
> 200 84 73 (86.9) Etiology 0.114 Non-viral 27 15 (55.6) HBV 171 122 (71.3) HCV 20 11 (55.0) Liver cirrhosis 0.102 (-) 107 67 (62.6) (+) 111 81 (73.0)	≤ 200	126	70 (55.6)	
Etiology 0.114 Non-viral 27 15 (55.6) HBV 171 122 (71.3) HCV 20 11 (55.0) Liver cirrhosis 0.102 (-) 107 67 (62.6) (+) 111 81 (73.0)	> 200	84	73 (86.9)	
Non-viral 27 15 (55.6) HBV 171 122 (71.3) HCV 20 11 (55.0) Liver cirrhosis 0.102 (-) 107 67 (62.6) (+) 111 81 (73.0)	Etiology			0.114
HBV 171 122 (71.3) HCV 20 11 (55.0) Liver cirrhosis 0.102 (-) 107 67 (62.6) (+) 111 81 (73.0)	Non-viral	27	15 (55.6)	
HCV 20 11 (55.0) Liver cirrhosis 0.102 (-) 107 67 (62.6) (+) 111 81 (73.0)	HBV	171	122 (71.3)	
Liver cirrhosis 0.102 (-) 107 67 (62.6) (+) 111 81 (73.0)	HCV	20	11 (55.0)	
(-) 107 67 (62.6) (+) 111 81 (73.0)	Liver cirrhosis		× /	0.102
(+) 111 81 (73.0)	(-)	107	67 (62.6)	
	(+)	111	81 (73.0)	

Table 1. Continued

Variable	No.	PEG10 expression	p-value
Early recurrence (≤	2 yr)		< 0.001
(–) ^{b)}	61	32 (52.5)	
(+)	122	97 (79.5)	
Late recurrence (> 2	2 yr)		0.863
(-) ^{b)}	61	32 (52.5)	
(+)	35	19 (54.3)	

Values are presented as number (%). PEG10, paternally expressed gene 10; AJCC, American Joint Committee on Cancer; BCLC, Barcelona Clinic Liver Cancer; AFP, α -fetoprotein; HBV, hepatitis B virus; HCV, hepatitis C virus. ^aPartial data were not available, ^bNo early or late recurrence.

granted approval for this study.

Patients were followed every 3 months after surgery. Tumor recurrence was detected by three phase dynamic computed tomography scans or a magnetic resonance imaging. The median follow-up period was 119.8 months (range, 14.0 to 151.4 months) for survivors and the follow-up period for recurrence was at least 24 months. Recurrence-free survival (RFS) was defined as from the date of the operation until the detection of tumor recurrence.

Two pathologists reviewed the slides to identify the representative tumor areas and marked the formalin-fixed paraffin-embedded blocks. Two tissue cores with a diameter of 2.0 mm were punched from the marked areas of the donor tissue block and re-embedded into the recipient paraffin blocks at defined positions. The two cores of the normal liver tissue from 12 patients with metastatic colon carcinoma of the liver were included in each array block, as controls. Each tissue microarray block contained up to 60 tissue cores.

2. Immunohistochemistry

The tissue microarray block was cut into $4-\mu$ m sections and immunohistochemical staining was performed as previously described [16]. The sections were subjected to antigen retrieval in 0.01 mol/L citrate buffer at pH 6.0 in a pressure cooker for 30 minutes. The sections were then incubated with a mouse monoclonal antibody to PEG10 (1:400, clone 4C10A7, MAB10294, Abnova Co., Taipei, Taiwan) for 30 minutes at room temperature. A negative control was obtained by replacing the primary antibody with isotype-matched irrelevant antibody. In order to validate the concordance between tissue microarrays and whole tumor sections, we further detected PEG10 expression for 40 corresponding

Variable	Univariate model			Multivariate model		
Variable	Coefficient	OR (95% CI)	p-value	Coefficien	t OR (95% CI)	p-value
Age (≤ 55 yr vs. > 55 yr)	-0.134	0.874 (0.470-1.627)	0.672	-	-	-
Gender (female vs. male)	-0.059	0.943 (0.413-2.152)	0.889	-	-	-
Tumor size (≤ 5.0 cm vs. > 5.0 cm)	0.805	2.237 (1.162-4.306)	0.016	0.206	1.229 (0.532-2.840)	0.630
Edmondson grade (I+II vs. III)	0.803	2.232 (0.992-5.022)	0.052	-	-	-
Microvascular invasion (no vs. yes)	1.494	4.456 (2.317-8.569)	< 0.001	0.619	1.858 (0.820-4.211)	0.138
Major portal vein invasion (no vs. yes)	1.564	4.779 (0.591-38.619)	0.142	-	-	-
Intrahepatic metastasis (no vs. yes)	2.665	14.362 (4.263-48.389)	< 0.001	1.929	6.880 (1.815-26.085)	0.005
Multicentric occurrence (no vs. yes)	1.133	3.103 (0.365-26.373)	0.300	-	-	-
AJCC T-stage (1 vs. 2+3+4)	1.623	5.067 (2.614-9.820)	< 0.001	-	-	-
BCLC stage (0+A vs. B+C)	1.185	3.271 (1.713-6.248)	< 0.001	-	-	-
Albumin level (> 3.5 g/dL vs. \leq 3.5 g/dL)	1.333	3.791 (1.080-13.305)	0.038	1.325	3.762 (0.839-16.867)	0.083
AFP level ($\leq 200 \text{ ng/mL vs.} > 200 \text{ ng/mL})^{a}$	0.685	1.984 (1.016-3.875)	0.045	-	-	-
Etiology (non-viral vs. viral)	1.765	5.841 (2.253-15.145)	< 0.001	1.584	4.876 (1.657-14.347)	0.004
Liver cirrhosis (no vs. yes)	0.529	1.697 (0.911-3.162)	0.096	-	-	-
PEG10 expression (- vs. +)	1.257	3.516 (1.804-6.856)	< 0.001	0.987	2.682 (1.233-5.833)	0.013

Table 2. Univariate and multivariate logistic regression models for predicting early tumor recurrence in 183 hepatocellular carcinomas

OR, odds ratio; CI, confidence interval; AJCC, American Joint Committee on Cancer; BCLC, Barcelona Clinic Liver Cancer; AFP, α-fetoprotein; PEG10, paternally expressed gene 10. ^{a)}Partial data were not available.

whole tumor sections randomly chosen from the 218 cases.

PEG10 expression was assessed by two independent pathologists (C.-K.P. and H.B.), who were blinded to the clinicopathologic data, and any discrepancies were resolved by a consensus. A nearly homogeneous cytoplasmic immunostaining, with moderate staining intensity, was observed. For determining PEG10 expression, the proportion of stained tumor cells was semi-quantitatively assessed and each sample was scored on a scale of 0 to 4 (0, < 1%; 1, 1%-25%; 2, 26%-50%; 3, 51%-75%; 4, > 75%). Duplicate tissue cores for each tumor showed high levels of homogeneity for the proportion of stained cells. When there were differences between the duplicate tissue cores, the higher score was taken.

3. Statistical analysis

All statistical analyses were performed using SPSS ver. 18 statistical software (SPSS Inc., Chicago, IL). The association between PEG10 expression and clinicopathologic variables was analyzed using the chi-square test or Fisher exact test. The logistic regression analysis was used to predict tumor recurrence. Survival analysis was performed using the Kaplan-Meier method, and the difference in survival rates was assessed by the log-rank test. The Cox proportional hazards regression model was used to identify factors that were independently associated with survival. Significant prognostic factors identified by univariate analysis were entered into a multivariate analysis. A p-value of < 0.05 was considered to be statistically significant.

Results

1. Clinicopathologic features of patients

The mean patient age was 51.9 years (range, 17 to 76 years), 179 patients were men and 39 women. One hundred and seventy-one patients (78.4%) were infected with hepatitis B virus and 20 (9.2%) with hepatitis C virus. No viral marker was recognized in 27 patients (12.4%). One hundred and fifty-seven patients (72.0%) had suffered from tumor recurrence, 122 patients (56.0%) from early recurrence, and 35 patients (16.1%) from late recurrence. Eighty-nine patients (40.8%) died of HCC. Sixteen of the 105 deaths in this study were due to non-HCC causes. Ten of the 16 deaths were due to hepatic failure; five due to non-hepatic causes, and one due to unknown cause.

2. PEG10 protein expression in HCC

Immunohistochemistry revealed that immunoreactivity for PEG10 was localized only in the cytoplasm of the tumor



Fig. 3. Kaplan-Meier survival curves showing recurrence-free survival among all patients (A), patients with α -fetoprotein (AFP) $\leq 20 \text{ ng/mL}$ (B), patients with tumor size $\leq 5.0 \text{ cm}$ (C), and patients at American Joint Committee on Cancer (AJCC) T-stage 1 (D) according to the paternally expressed gene 10 (PEG10) expression.

cells with moderate staining intensity. No immunoreactivity was found in normal hepatocytes (Fig. 1). We regarded PEG10 as positive when $\geq 1\%$ of the tumor cells showed cytoplasmic immunoreactivity (Fig. 2). PEG10 protein expression was observed in 148 of the 218 HCCs (67.9%). PEG10 expression was significantly correlated with younger age (p=0.046), female (p=0.037), higher Edmondson grade (p=0.021), microvascular invasion (p=0.001), intrahepatic metastasis (p=0.008), higher AJCC T-stage (p=0.011), and

higher α -fetoprotein level (p < 0.001). PEG10 expression was associated with early recurrence (p < 0.001), but not with late recurrence (p=0.863) (Table 1).

3. Prediction of early recurrence in HCC

Univariate analyses revealed that early recurrence was significantly correlated with larger tumor size (p=0.016), microvascular invasion (p < 0.001), intrahepatic metastasis

Variable	Recurrence-fre	e survival	Overall surv	Overall survival	
	HR (95% CI)	p-value	HR (95% CI)	p-value	
Age (≤ 55 yr vs. > 55 yr)	0.908 (0.659-1.253)	0.558	1.104 (0.751-1.623)	0.614	
Gender (female vs. male)	0.999 (0.667-1.495)	0.996	1.307 (0.777-2.198)	0.312	
Tumor size (≤ 5.0 cm vs. > 5.0 cm)	1.721 (1.252-2.366)	0.001	2.487 (1.694-3.653)	< 0.001	
Edmondson grade (I+II vs. III)	1.621 (1.130-2.327)	0.009	1.709 (1.113-2.623)	0.014	
Microvascular invasion (no vs. yes)	2.396 (1.723-3.332)	< 0.001	3.608 (1.996-4.716)	< 0.001	
Major portal vein invasion (no vs. yes)	3.045 (1.545-5.999)	0.001	5.249 (2.620-10.517)	< 0.001	
Intrahepatic metastasis (no vs. yes)	4.154 (2.935-5.879)	< 0.001	5.870 (3.946-8.732)	< 0.001	
Multicentric occurrence (no vs. yes)	1.382 (0.677-2.821)	0.374	0.611 (0.194-1.925)	0.400	
AJCC T-stage (1 vs. 2+3+4)	2.534 (1.814-3.540)	< 0.001	3.266 (2.103-5.072)	< 0.001	
BCLC stage (0+A vs. B+C)	2.072 (1.511-2.841)	< 0.001	3.084 (2.067-4.600)	< 0.001	
Albumin level (> 3.5 g/dL vs. $\leq 3.5 \text{ g/dL}$)	1.845 (1.193-2.855)	0.006	2.463 (1.508-4.022)	< 0.001	
AFP level ($\leq 200 \text{ ng/mL vs.} > 200 \text{ ng/mL})^{a}$	1.502 (1.089-2.072)	0.013	1.325 (0.895-1.962)	0.159	
Etiology (non-viral vs. viral)	2.674 (1.447-4.941)	0.002	1.463 (0.783-2.733)	0.233	
Liver cirrhosis (no vs. yes)	1.307 (0.954-1.790)	0.095	1.099 (0.750-1.613)	0.628	
PEG10 expression (- vs. +)	2.041 (1.425-2.921)	< 0.001	1.859 (1.186-2.914)	0.007	

Table 3. Univariate analyses of recurrence-free survival and overall survival in 218 hepatocellular carcinomas

HR, hazard ratio; CI, confidence interval; AJCC, American Joint Committee on Cancer; BCLC, Barcelona Clinic Liver Cancer; AFP, α -fetoprotein; PEG10, paternally expressed gene 10. ^aPartial data were not available.

(p < 0.001), higher AJCC T-stage (p < 0.001), higher BCLC stage (p < 0.001), lower albumin level (p=0.038), higher α -fetoprotein level (p=0.045), viral etiology (p < 0.001), and PEG10 expression (p < 0.001). Since AJCC T-stage and BCLC stage were associated with vascular invasion, we did not perform multiple analyses with these variables to avoid potential bias. An evaluation of the significant weight of the serum α -fetoprotein level was not performed due to missing data (n=178). On multivariate analyses, intrahepatic metastasis (p=0.005), viral etiology (p=0.004), and PEG10 expression (p=0.013) were independent predictors of early recurrence (Table 2).

4. Correlation between PEG10 expression and clinical outcome

The 3-, 5-, 7-, and 9-year RFS and the overall survival (OS) rates for 218 HCC patients were 38.9%, 34.6%, 28.3%, 27.6% and 74.6%, 65.2%, 56.4%, 50.2%, respectively. Univariate analyses revealed that larger tumor size, Edmondson grade III, microvascular invasion, major portal vein invasion, intrahepatic metastasis, higher AJCC T-stage, higher BCLC stage, and lower albumin level showed unfavorable influences on both RFS and OS. Higher α -fetoprotein level and viral etiology showed unfavorable influences on RFS. PEG10 expression showed an unfavorable influence on RFS (p < 0.001) (Table 3). The 5-year RFS rate of the PEG10-positive group was significantly lower than that of the PEG10-negative

group (25.4% vs. 53.7%) (Fig. 3A). The mean RFS of the PEG10-positive group and PEG10-negative group were 40.6 and 76.9 months, respectively. A subgroup analysis showed that among patients with α -fetoprotein ≤ 20 ng/mL (n=80), the PEG10-positive group (n=36) also showed an unfavorable influence on RFS (p=0.002) (Fig. 3B). Further analysis revealed that among patients with tumor size ≤ 5.0 cm (n=134) and patients at AJCC T-stage 1 (n=91), the PEG10-positive groups (n=90 and n=51, respectively) tended to show unfavorable influences on RFS (p=0.072 and p=0.079, respectively) (Fig. 3C and D).

PEG10 expression showed an unfavorable influence on OS (p=0.007) (Table 3). The 5-year OS rate of the PEG10-positive group was significantly lower than that of the PEG10-negative group (59.7% vs. 76.8%) (Fig. 4). Mean OS of the PEG10-positive group and PEG10-negative group were 88.1 and 112.7 months, respectively.

On multivariate analyses, intrahepatic metastasis and lower albumin level were found to be independent predictors of both shorter RFS and shorter OS. Microvascular invasion and viral etiology were found to be independent predictors of shorter RFS. PEG10 expression (p=0.005) was found to be an independent predictor of shorter RFS, but not of OS (p=0.128). PEG10-positive patients were more likely to suffer from recurrence than PEG10-negative patients (hazard ratio, 1.724) (Table 4).

Variable	Recurrence-free	e survival	Overall sur	Overall survival		
	HR (95% CI)	p-value	HR (95% CI)	p-value		
Tumor size (≤ 5.0 cm vs. > 5.0 cm)	1.252 (0.866-1.811)	0.231	1.383 (0.887-2.156)	0.152		
Edmondson grade (I+II vs. III)	0.897 (0.595-1.354)	0.605	0.951 (0.593-1.524)	0.834		
Microvascular invasion (no vs. yes)	1.568 (1.038-2.369)	0.033	1.561 (0.900-2.706)	0.113		
Major portal vein invasion (no vs. yes)	1.023 (0.468-2.236)	0.955	1.500 (0.692-3.251)	0.304		
Intrahepatic metastasis (no vs. yes)	2.466 (1.593-3.818)	< 0.001	3.647 (2.187-6.082)	< 0.001		
Albumin level (> 3.5 g/dL vs. $\leq 3.5 \text{ g/dL}$)	1.606 (1.016-2.541)	0.043	2.511 (1.509-4.178)	< 0.001		
Etiology (non-viral vs. viral)	2.192 (1.173-4.097)	0.014	-	-		
PEG10 expression (- vs. +)	1.724 (1.183-2.514)	0.005	1.441 (0.900-2.306)	0.128		

Table 4. Multivariate analyses of recurrence-free survival and overall survival in 218 hepatocellular carcinomas

HR, hazard ratio; CI, confidence interval; PEG10, paternally expressed gene 10.



Fig. 4. Kaplan-Meier survival curves showing overall survival for paternally expressed gene 10 (PEG10) expression in 218 hepatocellular carcinomas.

Discussion

PEG10 might be involved in oncogenesis by increasing the apoptotic resistance of the tumor cells. In HCC, PEG10 decreased cell death through interaction with seven in absentia homolog 1, which is a mediator of apoptosis [17]. Moreover, PEG10 knockdown inhibited the proliferation of the HepG2 HCC cells [5], while targeted disruption of the mouse *PEG10* gene caused early embryonic lethality due to defects in the placenta [18]. Recent reports demonstrated that PEG10 expression can be regulated by proto-oncogene

c-MYC [5], by E2F transcription factors that modulate the cell cycle [19], and by the sex hormone androgen [20]. The exact mechanism by which PEG10 signals is unclear, but it is known to interact with tumor growth factor β type I receptor ALK1 [21].

Although PEG10 overexpression in HCC was previously reported [7-10], the prognostic significance of PEG10 protein expression in HCC has not yet been reported. In this study, we found that PEG10 expression was significantly associated with higher Edmondson grade, microvascular invasion, intrahepatic metastasis, higher AJCC T-stage, and higher α fetoprotein level. Moreover, PEG10 expression was an independent predictor of early recurrence and shorter RFS. PEG10 expression could be used as a tool for early detection of HCC recurrence.

Measurement of the serum α -fetoprotein level is widely used for routine surveillance and noninvasive diagnosis of HCC, as well as to evaluate the prognosis and monitor recurrence following treatment [22]. As a limitation, elevated serum α -fetoprotein is observed in only 60% to 70% of HCC patients and, to a lesser extent (33%-65%), in patients with smaller HCCs [23]. Cucchetti et al. [24] asserted that preoperative high serum α -fetoprotein level (> 60 ng/mL) could be a risk factor for recurrence after resection in patients with liver cirrhosis. However, there is no effective marker to monitor recurrence and guide treatment for α -fetoprotein– negative ($\leq 20 \text{ ng/mL}$) patients after hepatectomy. In this study, tumor recurrence was detected in 51 of 80 α -fetoprotein-negative patients (63.8%). We found that within α -fetoprotein–negative patients, the PEG10-positive group still showed an unfavorable influence on RFS (p=0.002). PEG10 might be a useful biomarker to monitor recurrence for α -fetoprotein–negative HCC patients.

HCC patients with tumor size ≤ 5.0 cm and patients at AJCC T-stage 1 are generally considered to have relatively low risk for recurrence [25]; however, in this study, 91 of 134

(67.9%) with \leq 5.0 cm and 53 of 91 (58.2%) AJCC T-stage 1 showed recurrence. Within these populations, we found that the PEG10-positive groups tended to show unfavorable influences on RFS (p=0.072 and p=0.079, respectively).

Conclusion

Our findings indicate, for the first time, that PEG10 protein could be a potential biomarker predicting early recurrence and RFS in HCC patients after curative resection, even in those with normal serum α -fetoprotein levels. These findings also indicate possible new lines of research for the development of therapeutic approaches targeting PEG10. Further work is needed to explore functional roles of the *PEG10* gene, which might play in the development and progression of HCC.

Conflicts of Interest

Conflict of interest relevant to this article was not reported.

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