



Challenges of Antimicrobial Resistance and Stewardship in Solid Organ Transplant Patients

Miranda So^{1,2} · Laura Walti¹

Accepted: 15 March 2022 / Published online: 30 April 2022

© The Author(s), under exclusive licence to Springer Science+Business Media, LLC, part of Springer Nature 2022

Abstract

Purpose of Review Without effective antimicrobials, patients cannot undergo transplant surgery safely or sustain immunosuppressive therapy. This review examines the burden of antimicrobial resistance in solid organ transplant recipients and identifies opportunities for antimicrobial stewardship.

Recent Findings Antimicrobial resistance has been identified to be the leading cause of death globally. Multidrug-resistant pathogens are associated with significant morbidity and mortality in transplant recipients. Methicillin-resistant *S. aureus* affects liver and lung recipients, causing bacteremia, pneumonia, and surgical site infections. Vancomycin-resistant enterococci is a nosocomial pathogen primarily causing bacteremia in liver recipients. Multidrug-resistant Gram-negative pathogens present urgent and serious threats to transplant recipients. Extended-spectrum beta-lactamase-producing *E. coli* and *K. pneumoniae* commonly cause bacteremia and intra-abdominal infections in liver and kidney recipients. Carbapenemase-producing Enterobacterales, mainly *K. pneumoniae*, are responsible for infections early-post transplant in liver, lung, kidney, and heart recipients. *P. aeruginosa* and *A. baumannii* continue to be critical threats. While there are new antimicrobial agents targeting resistant pathogens, judicious prescribing is crucial to minimize emerging resistance. The full implications of the COVID-19 global pandemic on antimicrobial resistance in transplant recipients remain to be understood. Currently, there are no established standards on the implementation of antimicrobial stewardship interventions, but strategies that leverage existing antimicrobial stewardship program structure while tailoring to the needs of transplant recipients may help to optimize antimicrobial use.

Summary Clinicians caring for transplant recipients face unique challenges tackling emerging antimicrobial resistance. Coordinated antimicrobial stewardship interventions in collaboration with appropriate expertise in transplant and infectious diseases may mitigate against such threats.

Introduction

Antimicrobials are life-saving medications that are essential to modern medicine. Antimicrobial resistance (AMR) is a global health threat that requires urgent multisectoral,

multinational, and interdisciplinary action to address—the so-called One Health approach [1, 2]. AMR has been described as the “silent pandemic,” further exacerbated by antibiotics used to treat bacterial coinfections and secondary infections associated with the COVID-19 global pandemic [3–6]. AMR disproportionately affects those who are most vulnerable, including immunocompromised patients and solid organ transplant (SOT) recipients [7]. Without effective antimicrobials, transplant surgery cannot be conducted safely, and antirejection immunosuppressants cannot be implemented, as untreatable infections will negate the life-saving purpose of organ transplantation [7]. This review briefly describes the epidemiology of AMR (among bacterial pathogens) from a global perspective, examines the burden of AMR in SOT recipients, and discusses the challenges in the wider context of the COVID-19 pandemic. It

This article is part of the Topical Collection on *Antimicrobial Development and Drug Resistance*

✉ Miranda So
miranda.so@uhn.ca

¹ Toronto General Hospital, University Health Network, 9th Floor Munk Building, Room 800, 585 University Avenue, Toronto, ON M5G 2N2, Canada

² Leslie Dan Faculty of Pharmacy, University of Toronto, Toronto, ON, Canada

also highlights opportunities for antimicrobial stewardship initiatives in SOT patients to mitigate against the threats of AMR.

Epidemiology of Antimicrobial Resistance: A Global Perspective

This review follows the international consensus published in 2012 for definitions of acquired resistance [8]. Multidrug resistance (MDR) is defined as nonsusceptibility to at least one agent in three or more antibiotic classes [8]. Extensively drug resistance (XDR) is defined as nonsusceptibility to at least one agent in all but two or fewer antibiotic classes (i.e., bacterial isolates remain susceptible to only one or two classes) [8]. Pandrug resistance (PDR) is defined as nonsusceptibility to all licensed, routinely available antibiotics [8]. Although these definitions have some limitations, they have been applied to the SOT population [9].

Globally, AMR has been identified as a leading cause of death, with the highest burden in low-resource countries [10•]. There are notable regional differences in AMR rates and epidemiology, potentially associated with antibiotic use [7, 10•, 11, 12]. With concerted efforts, AMR rates for some pathogens such as methicillin-resistant *Staphylococcus aureus* (MRSA) and vancomycin-resistant enterococci (VRE) have stabilized, though primarily in high-income countries only, and not in low-resource settings [10•, 13, 14]. Globally, MRSA have shifted from healthcare settings to the community, whereas VRE remain primarily a nosocomial pathogen [15–18, 19••]. Furthermore, VRE with daptomycin- and linezolid-non-susceptibility have been described [20]. Vulnerable patients, such as those with frequent healthcare exposure due to immunocompromised state, are most at risk for contracting infections caused by MDR organisms [19••, 21, 22, 23••]. In contrast to MRSA and VRE, prevalence of MDR Gram-negative bacilli continues to rise and causes significant morbidity and mortality [24••]. Enterobacterales, mainly *Escherichia coli* and *Klebsiella pneumoniae*, are producers of betalactamases, such as cephalosporinases and carbapenemases, whereas non-fermenters such as *Pseudomonas aeruginosa* carry multiple mechanisms of resistance, and *Acinetobacter baumannii* resistance is conferred through carbapenemase (CRAB) [24••]. New antibiotics targeting MDR Gram-negative pathogens, such as ceftazidime-avibactam, ceftolozane-tazobactam, cefiderocol, meropenem-vaborbactam, among others, have become available in recent years [24••, 25••]. However, none of these agents provides universal and predictable activity, and treatment-emergent resistance has been observed [24••, 25••]. Antimicrobials with suboptimal pharmacokinetics, potential efficacy concerns, and toxicity issues such as polymyxins, aminoglycosides, and tigecycline are still being used to treat infections by MDR pathogens

based on their expected spectrum of activity, despite their undesirable profiles [24••, 25••].

Epidemiology and Risk Factors of Antimicrobial Resistance in Solid Organ Transplant Patients

Gram-Positive Bacteria

Despite a decrease in MRSA infections between 2005 and 2016, they remain concerning for SOT patients [7, 26•]. MRSA infections are most common in liver and lung transplant recipients [26•, 27•, 28], and the most common syndromes are bloodstream infections, pneumonia, surgical site infections, and intra-abdominal infections [29•]. In liver recipients with *S. aureus* bacteremia, the prevalence of MRSA ranged from 26.3 to 100% [28]. In a meta-analysis of 23 studies that evaluated infections of MRSA relative to colonization status, 17 (74%) pertained to liver recipients [30]. Pre-transplant, the pooled prevalence of MRSA colonization was 8.5% (95% CI 3.2–15.8%), compared to 9.4% (95% CI 3.0–18.5%) post-transplant [30]. MRSA colonization status was associated with increased risk of infections. The pooled risk ratio of infection was 5.5 (95% CI 2.36–12.90) from pre-transplant colonization and 10.56 (5.58–19.95) from post-transplant colonization [30]. In a recent single-center cohort study between 2008 and 2018 of 351 liver candidates, 5.4% (19/351) of the entire cohort were MRSA-positive, among whom two experienced infections [31]. In contrast, a single-center cohort of 369 pediatric SOT patients admitted between 2009 and 2014 found liver candidates to have the lowest risk of being colonized with MRSA (odds ratio 0.22, 95% CI 0.06–0.81), whereas lung candidates had the highest (odds ratio 18.7, 95% CI 1.9–182.3) [32]. Other risk factors for MRSA infections are summarized in Table 1 [26•, 33–37].

Vancomycin-resistant enterococcus (VRE) infections have been decreasing between 2012 and 2017 [7, 29•, 38]. A meta-analysis of VRE colonization status relative to infection rate in SOT patients estimated the pooled prevalence as 11.9% (95% CI 6.8–18.2) [30]. VRE colonization status was associated with infections, and liver recipients were the most commonly affected group [30]. Pre-transplant colonization was associated with VRE infection (risk ratio 6.65, 95% CI 2.54–17.41), as was post-transplant colonization (risk ratio 7.93, 95% CI 2.36–26.67) in a meta-analysis [30]. A single-center study of liver recipients identified 35% (123/351) of the entire cohort as VRE-positive, among whom 46% (57/123) experienced VRE infections [31]. A single-center cohort between 2008 and 2017 of 536 liver recipients reported 58 episodes of enterococcal bacteremia among 42 patients (7.8%), and VRE was the causative pathogen in 26

Table 1 Summary of risk factors for infections due to MDR pathogens in SOT patients

MDR pathogen	Risk factors	Most commonly affected SOT recipients
Methicillin-resistant <i>S. aureus</i>	Colonization status, alcoholic cirrhosis, decreased prothrombin ratio, recent surgical intervention, prolonged operating time, CMV seronegative status, primary CMV infection, prior antibiotic exposure, length of hospital and ICU stay, donor derived infection	Liver, lung, heart
Vancomycin-resistant enterococci (VRE)	Colonization status, post-transplant dialysis, length of hospital stay, donor-derived infection	Liver, heart
Extended spectrum beta-lactamase producing Enterobacterales (<i>E. coli</i>, <i>K. pneumoniae</i>)	Colonization status, history of infection due to ESBL-producing organism, post-transplant treatment with corticosteroid or treatment for acute rejection, exposure to antibiotics, including 3 rd generation cephalosporin, renal replacement therapy post-transplant, donor-derived infection	Liver, kidney, heart
Carbapenemase-producing Enterobacterales, mainly <i>K. pneumoniae</i> (KPC)	Colonization status, renal replacement therapy post-transplant, high model for end-stage liver disease (MELD) score at transplant, ureteral stent placement, re-transplantation, donor-derived infection	Liver, lung, kidney, kidney-pancreas
Multidrug-resistant or extremely drug resistant <i>P. aeruginosa</i>	Colonization status, cystic fibrosis, prior transplant, intensive care admission, septic shock, donor-derived infection	Lung, liver
Carbapenem-resistant <i>A. baumannii</i> (CRAB)	High pre-transplant blood urea nitrogen, hypoalbuminemia, prolonged operating time, mechanical ventilation, intensive care admission, donor-derived infection	Abdominal organs, lung

episodes (45%) [39]. Post-transplant dialysis and length of post-transplant hospitalization were associated with VRE bacteremia, with odds ratio 3.95 (95% CI 1.51–10.34) and 1.03 (95% CI 1.01–1.04), respectively [39]. A recent analysis of surgical site infections in liver transplant patients reported to the National Health Care Safety Network between 2015 and 2018 found prevalence of VRE to be 69.4% (84/121) among *E. faecium* isolates [40]. As VRE remains mainly a nosocomial pathogen, unit-level outbreaks affecting SOT patients have been described, with biofilm formation as a contributing factor [41]. VRE colonization status was not a risk factor for infections in non-liver recipients in a large single-center cohort study [42]. See Table 1 for a summary of risk factors for VRE infections.

Gram-Negative Bacteria

Infections due to MDR Gram-negative bacilli (GNB), particularly extended-spectrum beta-lactamase (ESBL) and carbapenemase-producing Gram-negative pathogens, have been on the rise in the last decade and affecting SOT patients [7, 23••, 27•, 43••, 44••, 45]. Prevalence of MDR GNB pathogens varies by organ group [9, 27•, 46••]. A meta-analysis of 1089 SOT patients from publications in 1994–2003 reported the pooled prevalence of ESBL

Enterobacterales colonization to be 18% (95% CI 5–36%) [47•]. The pooled prevalence from 3 studies in Europe was 15% (95% CI 3–34%), and from one North American study, it was 31.4% [47•]. Liver recipients had a pooled prevalence of 17% (95% CI 3–39%), and in kidney recipients, it was 23.5% [47•]. A single-center study from China assessing prevalence of ESBL phenotype in SOT recipients between 2007 and 2010 identified 80 MDR Gram-negative isolates among 350 patients [48]. Phenotypic expression of ESBL was found in 52.5% of the isolates (42/80), and 42.3% (33/80) of patients had ESBL Gram-negative bacterial infections [48]. A 10-year cohort study from France between 2001 and 2010 of 710 recipients reported pre-transplant colonization rate to be 4.1% (29/710), and 5.5% (39/710) developed infections caused by ESBL Enterobacterales within 4 months post-transplant, with intra-abdominal infections being the most common syndrome [49]. In kidney recipients, urinary tract infection was the most common presentation (70.9% [129/182]), and 11.4% (19/166) of the causative Enterobacterales were ESBL-producing [50]. A multicenter cohort study of SOT patients with ESBL Enterobacterales bacteremia between 2005 and 2015 assessed 988 episodes of bacteremia due to Enterobacterales [44••]. Among them, 40% (395) were caused by ESBL-producing organisms [44••]. Risk factors for ESBL

bacteremia were history of ESBL-positive cultures (adjusted odds ratio [aOR] 12.57, 95% CI 3.23–50.33), post-transplant corticosteroid (aOR 1.30, 95% CI 1.03–1.65), acute rejection treated with corticosteroid (aOR 1.18, 95% CI 1.16–1.19) [44••]. Exposure to antimicrobials associated with ESBL bacteremia include 3rd generation cephalosporin (aOR 1.95, 95% CI 1.48–2.57), echinocandins (aOR 1.61, 95% CI 1.16–1.19), and trimethoprim-sulfamethoxazole (aOR 1.35, 95% CI 1.10–1.64) [44••]. A clinical prediction tool for ESBL Enterobacterales bacteremia was developed based on a multicenter cohort of 897 SOT patients admitted between 2005 and 2018 [45]. Predictors selected in the final model were prior colonization or infection with Enterobacterales, recent antimicrobial exposure, severity of illness, and immunosuppressive regimen, but the model had not been externally validated [45]. See Table 1 for a summary of risk factors for ESBL-Enterobacterales infections.

Carbapenemase-producing Enterobacterales (CPE) is a critical threat associated with healthcare-acquired infections affecting SOT patients early post-transplant [7, 23••, 46••]. CPE refers to the mechanism of resistance (genotype), while carbapenem-resistant Enterobacterales (CRE) refers to the phenotypic definition based on susceptibility pattern. *Klebsiella pneumoniae* is the most common CPE pathogen, with carbapenemase (KPC) being the most common mechanism of resistance [46••]. Colonization is reported in 2–18% of SOT patients, whereas acquisition after transplant was reported in 5–27% of patients [23••]. In a five-center serial point-prevalence survey conducted in the USA, 154 patients were screened for CPE between 2019 and 2020, among whom 92 (60%) were SOT patients, and 7 (8%) recipients were colonized [51]. The average rate of infection is estimated to be 10% in liver, 5–10% in lung, and around 5% in kidney recipients, although sources of data are limited to small, single-center studies [23••, 46••]. A retrospective study from Italy evaluated 828 SOT patients admitted in 2011–2014, among them KPC colonization was identified in 5.4% (45/828) of patients and 4.5% (35/828) had infections due to KPC [52]. Post-transplant colonization was reported in 88.9% (40/45) of patients [52]. In a multicenter international cohort study of 60 SOT patients with CRE colonization and/or infection pre-transplant (30 liver and 17 heart recipients), KPC was the most commonly detected CRE organism [53]. Post-transplant infection occurred in 40% (24/60) of the patients, with 62% having surgical site infection and 46% having bacteremia [53]. In a single-center cohort of kidney recipients between 2017 and 2019, prevalence of early (90-day post-transplant) KPC infection was 10.4% (43/419), which included pneumonia, surgical site infections, urinary tract infections, and bacteremia [54]. Site of infection are generally related to the type of graft, potentially with secondary bacteremia [9, 23••]. For liver recipients, intra-abdominal infections such as abscesses, infected

bilomas, hematomas, or biliary complications (cholangitis, biliary leakage) are common presentations [9]. Risk factors for CPE infections are summarized in Table 1 [9, 23••].

The so-called difficult-to-treat resistant *Pseudomonas aeruginosa* remains a threat to SOT patients [7, 9, 46••, 55••]. Its mechanisms of resistance include efflux pump, plasmid-encoded extended-spectrum beta-lactamases, inducible chromosomal cephalosporinase AmpC, carbapenemases (metallo-beta-lactamases, and oxacillinase), and inactivation of OprD porin [9, 56]. Lung recipients are at high risk of *P. aeruginosa* colonization, with prevalence estimated to be 28% pre-transplant and 38% post-transplant [57]. Patients with cystic fibrosis (CF) have pre-transplant and post-transplant colonization risks as high as 70%, and re-colonization by the same strain post-transplant was 53% in one study [58–61]. For non-CF patients, colonization prevalence was reported to be 2.2% and 26% for pre- and post-transplant, respectively [59]. Pneumonia caused by MDR *P. aeruginosa* was reported in 33% of lung recipients early post-transplant in a retrospective study [62], and complications such as fatal empyema have been described [63•, 64]. SOT is a risk factor for antibiotic-resistant *P. aeruginosa* bloodstream infection [65], which accounted for 37–63% of *P. aeruginosa* bacteremia in SOT patients [66, 67]. The incidence of *P. aeruginosa* bacteremia in liver recipients ranged from 0.5 to 14.4%, and carbapenem- and quinolone-resistant *P. aeruginosa* was reported in 12.7% of patients, whereas the prevalence of XDR *P. aeruginosa* isolates was 9.3% [67–69]. Another study of abdominal transplant recipients with *P. aeruginosa* infections reported the prevalence of MDR isolates to be 46.3% (25/54) [70]. Risk factors for MDR or XDR *P. aeruginosa* infection are summarized in Table 1 [9].

Carbapenem-resistant *Acinetobacter baumannii* (CRAB) is a critical threat in healthcare-associated infections [7]. Data describing the overall prevalence of CRAB colonization and infections in SOT patients are limited, and while geographical variation has been described, the prevalence in the USA appears to be low [9, 46••]. In a cohort of abdominal transplant recipients in China between 2013 and 2020, carbapenem-resistant Gram-negative pathogens were reported 10.5% (153/1452) of patients, and CRAB accounted for 31% (47/153) of the isolates [71]. A single-center study in Turkey of 41 SOT recipients with *A. baumannii* infections between 2011 and 2017 reported 58.5% and 41.5% of the isolates to be MDR and XDR, respectively [72]. In a cohort of lung recipients in South Korea between 2012 and 2016, 57% (55/96) had *A. baumannii* infections, and among those isolates, 93% (51/55) were MDR [73]. Risk factors for antibiotic-resistant *A. baumannii* infections are summarized in Table 1 [73, 74].

In patients with CF, *Burkholderia cepacia* complex (BCC), particularly *B. cenocepacia*, are associated with accelerated pulmonary function decline [75, 76]. Pre-transplant

colonization and infection with BCC is associated with post-transplant infection, chronic allograft dysfunction (CLAD), and poor survival outcomes compared to BCC-negative status [75, 76]. BCC tend to localize within macrophages, which may contribute to discordance between treatment response and in vitro susceptibility [75]. Resistance to aminoglycosides, fluoroquinolones, sulfamethoxazole-trimethoprim (SMX-TMP), ceftazidime, and meropenem necessitates combination therapy, including ceftazidime-avibactam [46••, 77]. Local epidemiology varies, with incidence of BCC in Canadian centers being higher than in the USA (7.2% vs. 4.5%) [75]. Facility outbreaks have been described, highlighting the risk of nosocomial transmission [75]. Similar to BCC, *Stenotrophomonas maltophilia* is a therapeutic challenge, with options for susceptible strains limited to SMX-TMP and levofloxacin, while potential alternatives for emerging resistance may include minocycline, levofloxacin, ceftazidime, or ceftazidime-avibactam [25••, 46••]. MDR *S. maltophilia* is associated with poor outcomes in patients who received lung transplant for CF; however, the true epidemiology of *S. maltophilia* and its impact is less clear due to limited data [46••, 60, 63•].

Impact of Donor-Derived MDR Pathogens on SOT Patients

Donor-derived infections are defined as any infection present in the donor that is transmitted to one or more recipient [78••]. There are several possible ways donors colonized or infected with MDR organisms may impact transplant recipients: acceptance for organ utilization, modification of peritransplant surgical antibiotic prophylaxis regimens targeting MDR organisms, and selection of empirical antibiotic regimens for donor-derived infections [9, 78••, 79•, 80•, 81•, 82•, 83, 84, 85•]. In a retrospective cohort of 4 US transplant centers between 2015 and 2016, 15% (64/440) of deceased donors grew an MDR organism on culture [80•]. Risk factors for donors' acquisition of an MDR organism included hepatitis C viremia, need for dialysis, prior hematopoietic stem cell transplant, and exposure to antibiotics with a narrow Gram-negative spectrum [80•]. In a related study, among 440 donors, 7% (29/440) had MDR organism on hospital culture, with 2% (7/440) being MDR Gram-negative organisms, which was associated with a significant reduction in the number of organs transplanted per donor [82•]. Furthermore, organs were accepted significantly further down the match list, potentially reducing donor pool over time [82•]. The authors found that 14% (61/440) of donors received potentially redundant antibiotics, prompting suggestion for need for antimicrobial stewardship among donors during their terminal hospitalization [81•]. A multicenter retrospective cohort study evaluated 658 SOT patients identified 14%

(93/658) to have had a donor with MDR organisms [86•]. Donor MDR status was associated with a significantly increased hazard of infection compared to those with negative donor cultures (adjusted hazard ratio [aHR] 1.63, 95% CI 1.01–2.62) but were not associated with graft failure or death (aHR 0.45, 95% CI 0.15–1.36) [86•]. A single-center study of 28 abdominal transplant recipients in China between 2015 and 2020 reported a significantly lower survival rate if they had MDR Gram-negative (KPC, CRAB) donor-derived infections, compared with MDR Gram-positive (VRE) infections [87]. Isolation of resistant multidrug-resistant Gram-negative organisms from donor respiratory culture does not impact non-lung transplant recipient [88].

Mortality Associated with Antibiotic Resistance by Pathogen and Organ Type

Colonization and infections due to MDR organisms are associated with major impact on the outcomes of SOT recipients. However, in the absence of network-based, prospective registry data, the quality of available reports was limited by lack of standardized definitions to attribute morbidity and mortality to MDR organisms. Many were subject to bias due to retrospective, single-center design over varying study periods. For CPE infections, the reported mortality post-transplant rates varied. Driven by local epidemiology, depending on organ type, and infectious syndromes, the 1-year mortality ranged from less than 10% to over 80%, which was significantly higher than patients without CPE infections [23••, 43••, 52]. For ESBL-producing Gram-negative infections in liver and kidney transplant patients, bacteremia-related mortality was 26% [89]. Among patients with MRSA bacteremia, transplant status was not associated with higher 90-day mortality (adjusted odds ratio 0.74, 95% CI 0.44–1.25), but was associated with higher risk of septic shock and acute respiratory distress syndrome in a retrospective single-center study [90]. Others have reported higher long-term (1-year) mortality rate in lung recipients with MRSA infections compared with those who had methicillin-susceptible *S. aureus* infections [91]. Among heart transplant recipients, relative to those without infections, MDR pathogens and XDR pathogens infections were associated with 11-fold and 26-fold higher hazards of death, respectively [92•]. In a single-center study from 2011 to 2016, the most common MDR pathogens were ESBL-*E. coli* and *K. pneumoniae*, while *P. aeruginosa* and carbapenem-resistant *K. pneumoniae* were the most common XDR pathogens [92•]. In liver recipients, colonization and infection with MDR organisms were associated with increased risk of death (hazard ratio 2.57, $p < 0.001$) [31]. In kidney recipients, bacteremia due to *P. aeruginosa*, but not other MDR pathogens, was a significant risk factor for unfavorable outcome (defined as death, graft

nephrectomy, or return to dialysis), adjusted OR 46.11 (95% CI 3.9–552.2) [50]. Others reported carbapenem-resistant *K. pneumoniae* infection to be an independent risk factor for 1-year mortality, OR 6.75 (95% CI 1.05–43.4) [54].

Impact of the COVID-19 Global Pandemic on Antimicrobial Resistance and SOT Patients

SOT candidates and recipients have been affected by the direct risks of COVID-19 infection, as well as the secondary effects from the pandemic. Overall decrease in resources available at hospitals has led to reduction in non-urgent surgery and outpatient care, potentially adding to the negative impact on the mental status of SOT patients, contributing to decreased adherence to medical appointments [93–95]. Another potential impact has been a decrease in living and deceased organ donation worldwide, and its deleterious effect on transplant candidates' waitlist on morbidity and mortality [96–98]. Effects of the pandemic on AMR in general and subpopulations like SOT are difficult to predict. Increased antimicrobial use in COVID-19 patients could attribute to emergence of resistance [99, 100]. The overall prevalence of bacterial coinfection was estimated to be approximately 7%, and higher at 8.1% in critically ill patients, yet 70% of patients received antibiotics [5, 6, 101]. As a result of efforts to tackle the pandemic, resources for infectious diseases, antimicrobial stewardship (AMS), and infection prevention and control were stretched to their limits. In spite of that, recent European and UK surveillance data reported a decrease in AMU over the last 2 years [102, 103].

Severe COVID-19 infection with respiratory failure requiring prolonged mechanical ventilation or use of extracorporeal membrane oxygenation is associated with an increased risk of secondary infections and emergence of MDR pathogens [101, 104, 105]. Most studies addressing severely ill COVID-19 patients reported high rates of MDR pathogens [106, 107]. Furthermore, COVID-19 patient unit-level outbreaks due to MDR pathogens have been reported [108–110]. In contrast, decline in MDR rates for the entire hospital was reported in an Italian hospital [111]. Lung transplantation has emerged as a long-term solution for COVID-19 patients with non-reversible lung fibrosis [112–114]. Since these patients have had heavy exposure to the healthcare environment and high pretransplant infection rates, MDR rates in this population is potentially higher than non-COVID lung recipients [112]. Although short-term outcome seems similar in the limited literature when compared to lung transplant for other indications, long-term data are missing and most studies did not specifically report on infections in the posttransplant period [112, 114]. The

overall effect of the pandemic on AMR remains to be determined. Changes in the local epidemiology in healthcare settings could potentially lead to higher risks of acquisition of MDR pathogens among exposed individuals such as SOT recipients.

Challenges in Antimicrobial Stewardship in SOT Patients

Antimicrobial stewardship (AMS) is defined as a coordinated set of interventions to improve and measure the appropriate use of antimicrobials by promoting the selection of the optimal antimicrobial regimen including dosing, duration of therapy, and route of administration [115]. While new antimicrobials targeting MDR pathogens have become available in the last few years, clinicians are urged to be judicious in prescribing these agents to preserve their effectiveness [24, 25, 116]. Furthermore, global shortage of antimicrobials remains an ongoing challenge in healthcare [117]. Facing MDR and antimicrobial shortage, clinicians have had to resort to less desirable antimicrobials with worse adverse effects profile, such as polymyxins or tigecycline [24, 25]. Clinicians may also need to mitigate complex interactions between immunosuppressants and antimicrobials, carefully balancing the need to treat an infection with the goal of minimizing negative impact on the allograft. For it would be extremely unfortunate for a patient, after receiving a life-saving/altering organ transplant, only to lose the allograft or worse, succumb to an infection caused by an antibiotic-resistant pathogen. To ensure sustainability of safe and effective antimicrobials for current and future patients, coherent actions to promote responsible use of antimicrobials under the auspices of quality improvement are crucial [118].

Quality assurance programs from the US Center for Disease Prevention and Control (CDC), the Centers for Medicare and Medicaid, The Joint Commission, UK's National Institute for Health and Care Excellence, and Accreditation Canada, among others, have established quality standards for AMS programs across a wide spectrum of patient settings, including institutional and outpatient care [119, 120, 121, 122]. For example, the CDC's Core Elements of Antibiotic Stewardship encompass accountability structure, leadership, team membership, interventions, antimicrobial use monitoring and reporting for AMS programs in hospitals (Table 2) [119]. However, there are no established standards on how to implement AMS programs specific to SOT patients, and clinical data evaluating the safety and effectiveness of AMS interventions in this population remain scarce [19]. Diagnostic uncertainty and atypical presentation of infection in immunosuppressed patients are often the rationale for initiating antimicrobials [19]. Guidelines for

Table 2 Summary of AMS interventions tailored to SOT patients

US CDC core elements of hospital-based AMS programs	Leveraging existing AMS framework	Suggested additional components to address the needs of SOT patients
Hospital leadership commitment	Dedicate necessary human, financial and information technology resources AMS program leadership report to hospital leadership	Resources to facilitate AMS interventions in SOT patients Engage SOT program leadership and key stakeholders Including clinical expertise in SOT and transplant infectious diseases to be a part of the interdisciplinary AMS team, with sufficient resources and support
Accountability	Appoint a leader or co-leaders, such as a physician and pharmacist, responsible for program management and outcomes Appoint a pharmacist, ideally as the co-leader of the stewardship program, to lead implementation efforts to improve antibiotic use	Reporting of AMS activities and key performance indicators that reflect interventions implemented in SOT patients Determine key performance indicators that are feasible, valid, and meaningful for local key stakeholders in SOT Ensure key performance indicators reflect interventions implemented in SOT patients
Pharmacy expertise		
AMS interventions	Implement interventions, such as prospective audit and feedback or preauthorization, to improve antibiotic use Priority interventions include prospective audit and feedback, preauthorization, and facility-specific treatment recommendations. Facility-specific treatment guidelines can be important in enhancing the effectiveness of prospective audit and feedback and preauthorization Other priority interventions are infection- based, provider-based, pharmacy-based, microbiology-based, and nursing- based interventions	Establish process for audit and feedback with prescribers and clinicians in SOT patients Antibiogram for SOT patients Develop local guidelines specific for SOT patients using local epidemiology data. Start with a focused topic with a defined scope, and implement it and scale up as per local context. Refine the process before expanding to another topic or syndrome Engage patient, caregiver, SOT prescribers, nursing and pharmacy to identify interventions that best meet local needs Examples of suggestions tailored to SOT patients' needs: <ul style="list-style-type: none"> • Antifungal stewardship • Asymptomatic bacteruria in kidney transplant recipients • Syndrome-based interventions on empirical and targeted therapy • Adding rapid diagnostics to part of a bundle of interventions to guide tailoring of antimicrobial therapy
Tracking antimicrobial prescribing, resistance and <i>Clostridioides difficile</i> infection rate	Monitor antibiotic prescribing, impact of interventions, and other important outcomes like <i>C. difficile</i> infection and resistance patterns	Track data from SOT unit. Adapt, validate and utilize existing data extraction process to meet the needs of SOT patients
Reporting	Regularly report information on antibiotic use and resistance to prescribers, pharmacists, nurses, and hospital leadership Consider provider-level reporting (acknowledging that this has not been well studied for hospital antibiotic use)	Report data from SOT units, consider organ team level and/or provider level data reporting if feasible. Engage SOT clinicians in the process
Education	Educate prescribers, pharmacists, and nurses about adverse reactions from antibiotics, antibiotic resistance and optimal prescribing Case-based education, or “handshake stewardship”	Engage SOT clinicians using case-based format to discuss approach to infection management, including when to consult expertise from transplant infectious diseases team

Adapted from: CDC. Core Elements of Hospital Antibiotic Stewardship Programs. Atlanta, GA: US Department of Health and Human Services, CDC; 2019 and So M, Hand J, Forrest G, Pouch SM, Te H, Ardura MI, et al. White paper on antimicrobial stewardship in solid organ transplant recipients. *Am J Transplant.* 2022;22(1):96–112

management of certain infectious syndromes, such as those from the American Society of Transplantation, are available, but the heterogeneity of SOT patients makes defining appropriateness of antimicrobial regimen and identifying suitable outcome measures or key performance indicators difficult [19••, 123]. Clinical trials evaluating optimal duration of therapy tend to exclude SOT patients, limiting applicability of the evidence [19••]. Lastly, technical challenges in source control attainment, donor-derived infections, and the so-called net state of immunosuppression present additional barriers to designing and implementing AMS programs addressing the specific needs of SOT patients [19••, 124]. A 2016 survey of US transplant centers found that only 74% of institutional ASPs included coverage for adult SOT recipients [125•]. The extent to which CDC-recommended AMS interventions were implemented varied, and despite high needs for antimicrobials, strategies to assess antimicrobial prescribing quality in this population were limited [125•, 126]. A cross-sectional study of hospitals from 10 states in the USA evaluated the appropriateness of antimicrobial use in 1566 patients; immunocompromised patients (including SOT and stem cell transplant recipients) accounted for < 5% of the study cohort [126].

Potential Strategies to Optimize Antimicrobial Use and Mitigate the Threats of Antimicrobial Resistance

In 2020, 91% of hospitals in the USA have met all core elements of AMS programs recommended by the CDC [127••]. Therefore, leveraging existing framework and building on AMS programs already in place may be an efficient use of resources to address the unique AMS needs of SOT patients. Potential “add-on” components may include adding expertise in SOT and transplant infectious diseases to the interdisciplinary AMS team, monitoring and reporting antimicrobial use in SOT care units, and investing in resources to engage SOT leadership and clinicians to become stakeholders in SOT-specific AMS interventions [19••]. Locally developed guidelines constitute a key AMS strategy; therefore, focusing on organ-specific guidelines tailored to the SOT program’s epidemiology could be a reasonable starting point [19••, 115•]. Adjudication of adherence to surgical prophylaxis guidelines in transplant procedures is another reasonable intervention, focusing on organ-specific guidance [128, 129•]. Specific to the USA, the current National Healthcare Safety Network Antimicrobial Use Option Report does not include data from SOT care units for benchmarking, although this may change in the future. Potential interventions tailored to SOT patients within an existing hospital-based AMS program are summarized in Table 2 [19••, 119••]. Beyond the in-patient setting, antimicrobial

prescribed in the ambulatory clinics, where SOT patients receive often life-long care and follow-up monitoring, presents a crucial opportunity to explore the next horizon of AMS in SOT patients.

Conclusion

Rising antimicrobial resistance, particularly among difficult-to-treat Gram-negative pathogens, brings unique challenges to clinicians caring for SOT patients. While new antimicrobial agents targeting these pathogens have become available in recent years, without a coherent approach to ensure optimal and judicious use, they are at risk of losing their effectiveness due to emerging resistance. Antimicrobial stewardship programs tailored to the specific needs of SOT patients are important strategies to mitigate such threats to patient care.

Compliance with Ethical Standards

Conflict of Interest Miranda So and Laura Walti do not have any conflict of interests to disclose.

Human and Animal Rights and Informed Consent This article does not contain any studies with human or animal subjects performed by any of the authors.

References

Papers of particular interest, published recently, have been highlighted as:

- Of importance
 - Of major importance
1. Interagency Coordination Group on Antimicrobial Resistance. No time to wait: securing-the future from drug-resistant infections. 2019.
 2. Guardabassi L, Butaye P, Dockrell DH, Fitzgerald JR, Kuijper EJ. Escmid Study Group for Veterinary Microbiology. One Health: a multifaceted concept combining diverse approaches to prevent and control antimicrobial resistance. *Clin Microbiol Infect.* 2020;26(12):1604–5.
 3. Imperial College London. Antimicrobial resistance: a silent epidemic United Kingdom: Imperial College London. 2021. Available from: <https://www.imperial.ac.uk/stories/antimicrobial-resistance/>. Accessed 27 Dec 2021.
 4. Langford BJ, So M, Leung V, Raybardhan S, Lo J, Kan T, et al. Predictors and microbiology of respiratory and bloodstream bacterial infection in patients with COVID-19: living rapid review update and meta-regression. *Clin Microbiol Infect.* 2021.
 5. Langford BJ, So M, Raybardhan S, Leung V, Soucy JR, Westwood D, et al. Antibiotic prescribing in patients with COVID-19: rapid review and meta-analysis. *Clin Microbiol Infect.* 2021;27(4):520–31.

6. Langford BJ, So M, Raybardhan S, Leung V, Westwood D, MacFadden DR, et al. Bacterial co-infection and secondary infection in patients with COVID-19: a living rapid review and meta-analysis. *Clin Microbiol Infect.* 2020;26(12):1622–9.
7. Center for Disease Control and Prevention. Antibiotic resistance threats in the United States 2019. Atlanta, GA: National Center for Emerging and Zoonotic Infectious Diseases, Centers for Disease Control and Prevention. 2019. Available from: <http://www.cdc.gov/drugresistance/Biggest-Threats.html>. Accessed 27 Dec 2021.
8. Magiorakos AP, Srinivasan A, Carey RB, Carmeli Y, Falagas ME, Giske CG, et al. Multidrug-resistant, extensively drug-resistant and pandrug-resistant bacteria: an international expert proposal for interim standard definitions for acquired resistance. *Clin Microbiol Infect.* 2012;18(3):268–81.
9. Aguado JM, Silva JT, Fernandez-Ruiz M, Cordero E, Fortun J, Gudiol C, et al. Management of multidrug resistant Gram-negative bacilli infections in solid organ transplant recipients: SET/GESITRA-SEIMC/REIPI recommendations. *Transplant Rev. (Orlando).* 2018;32(1):36–57.
10. Murray CJL, Ikuta KS, Sharara F, Swetschinski L, Robles Aguilar G, Gray A, et al. Global burden of bacterial antimicrobial resistance in 2019: a systematic analysis. *The Lancet.* 2022;399(10325):629–55. **This review highlights the global burden of AMR remains high and disproportionately affects population in low-resource settings.**
11. Van Duin D, Doi Y. The global epidemiology of carbapenemase-producing Enterobacteriaceae. *Virulence.* 2017;8(4):460–9.
12. Goossens H, Ferech M, Vander Stichele R, Elseviers M. Outpatient antibiotic use in Europe and association with resistance: a cross-national database study. *The Lancet.* 2005;365(9459):579–87.
13. Jernigan JA, Hatfield KM, Wolford H, Nelson RE, Olubajo B, Reddy SC, et al. Multidrug-resistant bacterial infections in U.S. hospitalized patients, 2012–2017. *New England J Med.* 2020;382(14):1309–19.
14. ECDC. European Centre for Disease Prevention and Control. Antimicrobial resistance in the EU, EEA (EARS-Net) - annual epidemiological report. 2019 Stockholm European Centre for Disease Prevention and Control Sweden 2020.
15. Hassoun A, Linden PK, Friedman B. Incidence, prevalence, and management of MRSA bacteremia across patient populations—a review of recent developments in MRSA management and treatment. *Crit Care.* 2017;21(1):211.
16. Mitevska E, Wong B, Surewaard BGJ, Jenne CN. The prevalence, risk, and management of methicillin-resistant *Staphylococcus aureus* infection in diverse populations across Canada: a systematic review. *Pathogens.* 2021;10(4).
17. Petersen A, Larssen KW, Gran FW, Enger H, Haeggman S, Makitalo B, et al. Increasing incidences and clonal diversity of methicillin-resistant *Staphylococcus aureus* in the Nordic countries - results from the Nordic MRSA surveillance. *Front Microbiol.* 2021;12:668900.
18. Johnstone J, Chen C, Rosella L, Adomako K, Policarpio ME, Lam F, et al. Patient- and hospital-level predictors of vancomycin-resistant Enterococcus (VRE) bacteremia in Ontario. *Canada Am J Infect Control.* 2018;46(11):1266–71.
19. So M, Hand J, Forrest G, Pouch SM, Te H, Ardura MI, et al. White paper on antimicrobial stewardship in solid organ transplant recipients. *Am J Transplant.* 2022;22(1):96–112. **A white paper outlining the current landscape of antimicrobial stewardship from the literature and identifies gaps in knowledge for future research in SOT patients.**
20. Greene MH, Harris BD, Nesbitt WJ, Watson ML, Wright PW, Talbot TR, et al. Risk factors and outcomes associated with acquisition of daptomycin and linezolid-nonsusceptible vancomycin-resistant Enterococcus. *Open Forum Infect Dis.* 2018;5(10):ofy185.
21. Cassini A, Högberg LD, Plachouras D, Quattrocchi A, Hoxha A, Simonsen GS, et al. Attributable deaths and disability-adjusted life-years caused by infections with antibiotic-resistant bacteria in the EU and the European Economic Area in 2015: a population-level modelling analysis. *Lancet Infect Dis.* 2019;19(1):56–66.
22. Righi E, Peri AM, Harris PN, Wailan AM, Liborio M, Lane SW, et al. Global prevalence of carbapenem resistance in neutropenic patients and association with mortality and carbapenem use: systematic review and meta-analysis. *J Antimicrob Chemother.* 2017;72(3):668–77.
23. Giannella M, Bartoletti M, Conti M, Righi E. Carbapenemase-producing Enterobacteriaceae in transplant patients. *J Antimicrob Chemother.* 2021;76(Suppl 1):i27–i39. **A comprehensive review of the impact of AMR on SOT patients focusing on Enterobacteriaceae.**
24. Paul M, Carrara E, Retamar P, Tangden T, Bitterman R, Bonomo RA, et al. European Society of clinical microbiology and infectious diseases (ESCMID) guidelines for the treatment of infections caused by Multidrug-resistant Gram-negative bacilli (endorsed by ESICM -European Society of intensive care Medicine). *Clin Microbiol Infect.* 2021. **Comprehensive guidance on antimicrobial options to treat MDR Gram-negative organisms.**
25. Tamma PD, Aitken SL, Bonomo RA, Mathers AJ, van Duin D, Clancy CJ. Version 2.0 Infectious Diseases Society of America Guidance on the treatment of AmpC β -lactamase-producing Enterobacteriales, Carbapenem-resistant *Acinetobacter baumannii*, and *Stenotrophomonas maltophilia* infections. 2021. <https://www.idsociety.org/practice-guideline/amr-guidance-2.0/>. Accessed 2 Jan 2022. **Comprehensive guidance on antimicrobial options to treat MDR Gram-negative organisms.**
26. Pereira MR, Rana MM, for the A.S.T. ID Community of Practice. Methicillin-resistant *Staphylococcus aureus* in solid organ transplantation - Guidelines from the American Society of Transplantation Infectious Disease Community of Practice. *Clin Transplant.* 2019;33(9):e13611. **Guidance on the risk factors and management of MRSA infections in SOT patients.**
27. Bartoletti M, Giannella M, Tedeschi S, Viale P. Multidrug-resistant bacterial infections in solid organ transplant candidates and recipients. *Infect Dis Clin North Am.* 2018;32(3):551–80. **Brief review on the risk and management of infections from MDR pathogens in SOT patients.**
28. Liu T, Zhang Y, Wan Q. Methicillin-resistant *Staphylococcus aureus* bacteremia among liver transplant recipients: epidemiology and associated risk factors for morbidity and mortality. *Infect Drug Resist.* 2018;11:647–58.
29. Garzoni C, Vergidis P. A. S. T. Infectious diseases community of practice. Methicillin-resistant, vancomycin-intermediate and vancomycin-resistant *Staphylococcus aureus* infections in solid organ transplantation. *Am J Transplant.* 2013;13 Suppl 4:50–8. **An informative guideline on resistant *S. aureus* infections in SOT patients.**
30. Ziakas PD, Pliakos EE, Zervou FN, Knoll BM, Rice LB, Mylonakis E. MRSA and VRE colonization in solid organ transplantation: a meta-analysis of published studies. *Am J Transplant.* 2014;14(8):1887–94.
31. Ferstl PG, Filmann N, Heilgental EM, Schnitzbauer AA, Bechstein WO, Kempf VAJ, et al. Colonization with multidrug-resistant organisms is associated with increased mortality in liver transplant candidates. *PLoS One.* 2021;16(1):e0245091.
32. Paulsen G, Blum S, Danziger-Isakov L. Epidemiology and outcomes of pretransplant methicillin-resistant *Staphylococcus*

- aureus screening in pediatric solid organ transplant candidates. *Pediatr Transplant*. 2018:e13246.
33. Singh N, Paterson DL, Chang FY, Gayowski T, Squier C, Wagener MM, et al. Methicillin-resistant *Staphylococcus aureus*: the other emerging resistant Gram-positive coccus among liver transplant recipients. *Clin Infect Dis*. 2000;30(2):322–7.
 34. Florescu DF, McCartney AM, Qiu F, Langnas AN, Botha J, Mercer DF, et al. *Staphylococcus aureus* infections after liver transplantation. *Infection*. 2012;40(3):263–9.
 35. Schneider CR, Buell JF, Gearhart M, Thomas M, Hanaway MJ, Rudich SM, et al. Methicillin-resistant *Staphylococcus aureus* infection in liver transplantation: a matched controlled study. *Transpl Proc*. 2005;37(2):1243–4.
 36. Bert F, Bellier C, Lassel L, Lefranc V, Durand F, Belghiti J, et al. Risk factors for *Staphylococcus aureus* infection in liver transplant recipients. *Liver Transpl*. 2005;11(9):1093–9.
 37. Hashimoto M, Sugawara Y, Tamura S, Kaneko J, Matsui Y, Moriya K, et al. Impact of new methicillin-resistant *Staphylococcus aureus* carriage postoperatively after living donor liver transplantation. *Transpl Proc*. 2007;39(10):3271–5.
 38. Nellore A, Huprikar S. Practice AICo. Vancomycin-resistant *Enterococcus* in solid organ transplant recipients: guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. *Clin Transplant*. 2019:e13549.
 39. Kim YJ, Jun YH, Choi HJ, You YK, Kim DG, Choi JY, et al. Impact of Enterococcal Bacteremia in liver transplant recipients. *Transplant Proc*. 2019;51(8):2766–70.
 40. Chea N, Sapiano MRP, Zhou L, Epstein L, Guh A, Edwards JR, et al. Rates and causative pathogens of surgical site infections attributed to liver transplant procedures and other hepatic, biliary, or pancreatic procedures, 2015–2018. *Transpl Infect Dis*. 2021;23(4):e13589.
 41. Kreidl P, Mayr A, Hinterberger G, Berktold M, Knabl L, Fuchs S, et al. Outbreak report: a nosocomial outbreak of vancomycin resistant enterococci in a solid organ transplant unit. *Antimicrob Resist Infect Control*. 2018;7:86.
 42. McFarlane AC, Kabbani D, Bakal JA, Smith SW. Clinical impact of vancomycin-resistant enterococci colonization in nonliver solid organ transplantation and its implications for infection control strategies: a single-center, 10-year retrospective study. *Transpl Infect Dis*. 2021;23(6):e13747.
 - 43.●● Giannella M, Bartoletti M, Campoli C, Rinaldi M, Coladonato S, Pascale R, et al. The impact of carbapenemase-producing Enterobacteriaceae colonization on infection risk after liver transplantation: a prospective observational cohort study. *Clin Microbiol Infect*. 2019;25(12):1525–31. **Informative cohort study on the impact of MDR Enterobacteriaceae on liver recipients.**
 - 44.●● Anesi JA, Lautenbach E, Tamma PD, Thom KA, Blumberg EA, Alby K, et al. Risk factors for extended-spectrum beta-lactamase-producing Enterobacteriales bloodstream infection among solid-organ transplant recipients. *Clin Infect Dis*. 2021;72(6):953–60. **Informative study on ESBL Enterobacteriales bacteremia in SOT patients.**
 45. Wang R, Han JH, Lautenbach E, Tamma PD, Thom KA, Alby K, et al. Clinical prediction tool for extended-spectrum beta-lactamase-producing enterobacteriales as the etiology of a bloodstream infection in solid organ transplant recipients. *Transpl Infect Dis*. 2021;23(4):e13599.
 - 46.●● Pouch SM, Patel G. A. S. T. Infectious diseases community of practice. Multidrug-resistant Gram-negative bacterial infections in solid organ transplant recipients—Guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. *Clin Transplant*. 2019;33(9):e13594. **Comprehensive review and guidance on the management of MDR Gram-negative organisms in SOT patients.**
 - 47.● Alevizakos M, Kallias A, Flokas ME, Mylonakis E. Colonization with extended-spectrum beta-lactamase-producing Enterobacteriaceae in solid organ transplantation: a meta-analysis and review. *Transpl Infect Dis*. 2017;19(4). **Informative review on the impact of colonization with ESBL Enterobacteriaceae in SOT patients.**
 48. Men TY, Wang JN, Li H, Gu Y, Xing TH, Peng ZH, et al. Prevalence of multidrug-resistant Gram-negative bacilli producing extended-spectrum beta-lactamases (ESBLs) and ESBL genes in solid organ transplant recipients. *Transpl Infect Dis*. 2013;15(1):14–21.
 49. Bert F, Larroque B, Paugam-Burtz C, Dondero F, Durand F, Marcon E, et al. Pretransplant fecal carriage of extended-spectrum beta-lactamase-producing Enterobacteriaceae and infection after liver transplant. *France Emerg Infect Dis*. 2012;18(6):908–16.
 50. Tsikala-Vafea M, Basoulis D, Pavlopoulou I, Darema M, Deliolanis J, Daikos GL, et al. Bloodstream infections by Gram-negative bacteria in kidney transplant patients: Incidence, risk factors, and outcome. *Transpl Infect Dis*. 2020;22(6):e13442.
 51. Chan JL, Nazarian E, Musser KA, Snively EA, Fung M, Doernberg SB, et al. Prevalence of carbapenemase-producing organisms among hospitalized solid organ transplant recipients, five US hospitals, 2019–2020. *Transpl Infect Dis*. 2022:e13785.
 52. Pagani N, Corcione S, Lupia T, Scabini S, Filippini C, Angilletta R, et al. Carbapenemase-producing *Klebsiella pneumoniae* colonization and infection in solid organ transplant recipients: a single-center, retrospective study. *Microorganisms*. 2021;9(11).
 53. Taimur S, Pouch SM, Zubizarreta N, Mazumdar M, Rana M, Patel G, et al. Impact of pre-transplant carbapenem-resistant Enterobacteriales colonization and/or infection on solid organ transplant outcomes. *Clin Transplant*. 2021;35(4):e14239.
 54. Zhang F, Zhong J, Ding H, Pan J, Yang J, Lan T, et al. Analysis of risk factors for carbapenem-resistant *Klebsiella pneumoniae* infection and its effect on the outcome of early infection after kidney transplantation. *Front Cell Infect Microbiol*. 2021;11:726282.
 - 55.●● Tamma PD, Aitken SL, Bonomo RA, Mathers AJ, van Duin D, Clancy CJ. IDSA Guidance on the treatment of antimicrobial-resistant Gram-negative infections: version 1.0: a focus on extended-spectrum β -lactamase producing Enterobacteriales, carbapenem-resistant Enterobacteriales, and *Pseudomonas aeruginosa* with difficult-to-treat resistance. *Clin Infect Dis*. 2020. Accessed 2 Jan 2022. <https://www.idsociety.org/practice-guideline/amr-guidance/>. **Comprehensive guidance on the antimicrobial management of MDR Gram-negative pathogens.**
 56. Mojica MF, Rossi M-A, Vila AJ, Bonomo RA. The urgent need for metallo- β -lactamase inhibitors: an unattended global threat. *Lancet Infect Dis*. 2022;22(1):e28–34.
 57. Vos R, Vanaudenaerde BM, Geudens N, Dupont LJ, Van Raemdonck DE, Verleden GM. Pseudomonal airway colonisation: risk factor for bronchiolitis obliterans syndrome after lung transplantation? *Eur Respir J*. 2008;31(5):1037–45.
 58. Elborn JS. Cystic fibrosis. *The Lancet*. 2016;388(10059):2519–31.
 59. Botha P, Archer L, Anderson RL, Lordan J, Dark JH, Corris PA, et al. *Pseudomonas aeruginosa* colonization of the allograft after lung transplantation and the risk of bronchiolitis obliterans syndrome. *Transplantation*. 2008;85(5):771–4.
 60. Holm AE, Schultz HHL, Johansen HK, Pressler T, Lund TK, Iversen M, et al. Bacterial re-colonization occurs early after lung transplantation in cystic fibrosis patients. *J Clin Med*. 2021;10(6).
 61. Shteinberg M, Haq IJ, Polineni D, Davies JC. Cystic fibrosis. *The Lancet*. 2021;397(10290):2195–2111.
 62. Campos S, Caramori M, Teixeira R, Afonso J, Carraro R, Strabelli T, et al. Bacterial and fungal pneumonias after lung transplantation. *Transpl Proc*. 2008;40(3):822–4.

- 63.● Vazirani J, Crowhurst T, Morrissey CO, Snell GI. Management of multidrug resistant infections in lung transplant recipients with cystic fibrosis. *Infect Drug Resist.* 2021;14:5293–301. **Informative review on MDR organisms in patients with cystic fibrosis and potential lung transplant candidates.**
64. Carugati M, Piazza A, Peri AM, Cariani L, Brilli M, Girelli D, et al. Fatal respiratory infection due to ST308 VIM-1-producing *Pseudomonas aeruginosa* in a lung transplant recipient: case report and review of the literature. *BMC Infect Dis.* 2020;20(1):635.
65. Herrera S, Bodro M, Soriano A. Predictors of multidrug resistant *Pseudomonas aeruginosa* involvement in bloodstream infections. *Curr Opin Infect Dis.* 2021;34(6):686–92.
66. Johnson LE, D'Agata EM, Paterson DL, Clarke L, Qureshi ZA, Potoski BA, et al. *Pseudomonas aeruginosa* bacteremia over a 10-year period: multidrug resistance and outcomes in transplant recipients. *Transpl Infect Dis.* 2009;11(3):227–34.
67. Bodro M, Sabe N, Tubau F, Llado L, Baliellas C, Gonzalez-Costello J, et al. Extensively drug-resistant *Pseudomonas aeruginosa* bacteremia in solid organ transplant recipients. *Transplantation.* 2015;99(3):616–22.
68. Liu T, Zhang Y, Wan Q. *Pseudomonas aeruginosa* bacteremia among liver transplant recipients. *Infect Drug Resist.* 2018;11:2345–56.
69. Bodro M, Sabe N, Tubau F, Llado L, Baliellas C, Roca J, et al. Risk factors and outcomes of bacteremia caused by drug-resistant ESKAPE pathogens in solid-organ transplant recipients. *Transplantation.* 2013;96(9):843–9.
70. Su H, Ye Q, Wan Q, Zhou J. Predictors of mortality in abdominal organ transplant recipients with *Pseudomonas aeruginosa* infections. *Ann Transplant.* 2016;21:86–93.
71. Wu D, Chen C, Liu T, Jia Y, Wan Q, Peng J. Epidemiology, susceptibility, and risk factors associated with mortality in carbapenem-resistant Gram-negative bacterial infections among abdominal solid organ transplant recipients: a retrospective cohort study. *Infect Dis Ther.* 2021;10(1):559–73.
72. Serifoglu I, Er Dedekarginoglu B, Savas Bozbas S, Akcay S, Haberal M. Clinical characteristics of *Acinetobacter baumannii* infection in solid-organ transplant recipients. *Exp Clin Transplant.* 2018;16 Suppl 1(Suppl 1):171–5.
73. Oh DH, Kim YC, Kim EJ, Jung IY, Jeong SJ, Kim SY, et al. Multidrug-resistant *Acinetobacter baumannii* infection in lung transplant recipients: risk factors and prognosis. *Infect Dis (Lond).* 2019;51(7):493–501.
74. Kitazono H, Rog D, Grim SA, Clark NM, Reid GE. *Acinetobacter baumannii* infection in solid organ transplant recipients. *Clin Transplant.* 2015;29(3):227–32.
75. Mitchell AB, Glanville AR. The impact of resistant bacterial pathogens including *Pseudomonas aeruginosa* and *Burkholderia* on lung transplant outcomes. *Semin Respir Crit Care Med.* 2021;42(3):436–48.
76. Yeung JC, Machuca TN, Chaparro C, Cypel M, Stephenson AL, Solomon M, et al. Lung transplantation for cystic fibrosis. *J Heart Lung Transplant.* 2020;39(6):553–60.
77. Chien YC, Liao CH, Sheng WH, Chien JY, Huang YT, Yu CJ, et al. Clinical characteristics of bacteraemia caused by *Burkholderia cepacia* complex species and antimicrobial susceptibility of the isolates in a medical centre in Taiwan. *Int J Antimicrob Agents.* 2018;51(3):357–64.
- 78.●● Wolfe CR, Ison MG. A. S. T. Infectious Diseases Community of Practice. Donor-derived infections: guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. *Clin Transplant.* 2019;33(9):e13547. **Comprehensive guideline on managing donor-derived infections in SOT patients.**
- 79.● Malinis M, Boucher HW. A. S. T. Infectious Diseases Community of Practice. Screening of donor and candidate prior to solid organ transplantation-guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. *Clin Transplant.* 2019;33(9):e13548. **Review on screening of donor and candidates regarding potential risks of donor-derived infections.**
- 80.● Anesi JA, Blumberg EA, Han JH, Lee DH, Clauss H, Climaco A, et al. Risk factors for multidrug-resistant organisms among deceased organ donors. *Am J Transplant.* 2019;19(9):2468–78. **Informative study on donor-derived infections due to MDR organisms.**
- 81.● Anesi JA, Lautenbach E, Han J, Lee DH, Clauss H, Climaco A, et al. Antibiotic utilization in deceased organ donors. *Clin Infect Dis.* 2021;73(7):1284–7. **Informative study on the impact of donor-derived MDR organisms on antibiotic consumption.**
- 82.● Anesi JA, Han JH, Lautenbach E, Lee DH, Clauss H, Climaco A, et al. Impact of deceased donor multidrug-resistant bacterial organisms on organ utilization. *Am J Transplant.* 2020;20(9):2559–66. **Informative study on the impact of MDR organisms on organ utilization.**
83. Lewis JD, Sifri CD. Multidrug-resistant bacterial donor-derived infections in solid organ transplantation. *Curr Infect Dis Rep.* 2016;18(6):18.
84. Groff LT, Reed EE, Coe KE, El Boghdadly Z, Keller BC, Whitson BA, et al. Effectiveness of short vs long-course perioperative antibiotics in lung transplant recipients with donor positive respiratory cultures. *Transpl Infect Dis.* 2021;23(3):e13518.
- 85.● Bunsow E, Los-Arcos I, Martin-Gomez MT, Bello I, Pont T, Berastegui C, et al. Donor-derived bacterial infections in lung transplant recipients in the era of multidrug resistance. *J Infect.* 2020;80(2):190–6. **Informative study on MDR organisms in donor-derived infections among lung recipients.**
- 86.● Anesi JA, Blumberg EA, Han JH, Lee DH, Clauss H, Hasz R, et al. Impact of donor multidrug-resistant organisms on solid organ transplant recipient outcomes. *Transpl Infect Dis.* 2021:e13783. **Informative study on MDR organisms donor-derived infections in outcomes of SOT recipients.**
87. Xiao J, Wu D, Jia Y, Wan Q, Peng J. Impact of donor-derived multi-drug-resistant organism infections on abdominal solid organ transplantation recipients in China. *Transplant Proc.* 2021;53(6):1853–7.
88. Benamu E, Pereira MR, Taimur S, Jacobs SE, Friedman AL, Jenkins SG, et al. Isolation of antibiotic-resistant Gram-negative organisms from donor respiratory culture does not impact non-lung solid organ recipient management. *Clin Transplant.* 2019;33(8):e13646.
89. Aguiar EB, Maciel LC, Halpern M, de Lemos AS, Ferreira ALP, Basto ST, et al. Outcome of bacteremia caused by extended-spectrum β -lactamase-producing Enterobacteriaceae after solid organ transplantation. *Transpl Proc.* 2014;46(6):1753–6.
90. Eichenberger EM, Ruffin F, Sharma-Kuinkel B, Dagher M, Park L, Kohler C, et al. Bacterial genotype and clinical outcomes in solid organ transplant recipients with *Staphylococcus aureus* bacteremia. *Transpl Infect Dis.* 2021;23(6):e13730.
91. Shields RK, Clancy CJ, Minces LR, Kwak EJ, Silveira FP, Abdel Massih RC, et al. *Staphylococcus aureus* infections in the early period after lung transplantation: epidemiology, risk factors, and outcomes. *J Heart Lung Transplant.* 2012;31(11):1199–206.
- 92.● Bhatt PJ, Ali M, Rana M, Patel G, Sullivan T, Murphy J, et al. Infections due to multidrug-resistant organisms following heart transplantation: epidemiology, microbiology, and outcomes. *Transpl Infect Dis.* 2020;22(1):e13215. **Informative study on infections due to MDR organisms in heart recipients.**
93. Lambooy S, Krishnasamy R, Pollock A, Hilder G, Gray NA. Telemedicine for outpatient care of kidney transplant and CKD patients. *Kidney International Reports.* 2021;6(5):1265–72.

94. Reuken PA, Rauchfuss F, Albers S, Settmacher U, Trautwein C, Bruns T, et al. Between fear and courage: attitudes, beliefs, and behavior of liver transplantation recipients and waiting list candidates during the COVID-19 pandemic. *Am J Transplant*. 2020;20(11):3042–50.
95. Kröncke S, Lund LK, Buchholz A, Lang M, Briem-Richter A, Grabhorn EF, et al. Psychosocial situation, adherence, and utilization of video consultation in young adult long-term pediatric liver transplant recipients during COVID-19 pandemic. *Pediatric Transplant*. 2021;25(8).
96. Millan DAC, Fajardo-Cediel W, Tobar-Roa V, Garcia-Perdomo HA, Autran-Gomez AM. Strategies to mitigate the impact of COVID 19 pandemic on organ donation and kidney transplantation in Latin America. *Curr Urol Rep*. 2021;22(12):59.
97. Salvalaggio PR, Ferreira GF, Caliskan Y, Vest LS, Schnitzler MA, de Sandes-Freitas TV, et al. An International survey on living kidney donation and transplant practices during the COVID-19 pandemic. *Transplant Infectious Disease*. 2021;23(2):e13526-e.
98. Russo FP, Izzy M, Rammohan A, Kirchner VA, Di Maira T, Belli LS, et al. Global impact of the first wave of COVID-19 on liver transplant centers: a multi-society survey (EASL-ESOT/ELITA-ILTS). *J Hepatol*. 2022;76(2):364–70.
99. Monnet DL, Harbarth S. Will coronavirus disease (COVID-19) have an impact on antimicrobial resistance? *Euro Surveill*. 2020;25(45).
- 100.● Clancy CJ, Buehrle DJ, Nguyen MH. PRO: The COVID-19 pandemic will result in increased antimicrobial resistance rates. *JAC Antimicrob Resist*. 2020;2(3):dlaa049. **Part of a Pro/Con debate on the potential impact of COVID-19 pandemic on emergence of MDR organisms.**
- 101.● Lansbury L, Lim B, Baskaran V, Lim WS. Co-infections in people with COVID-19: a systematic review and meta-analysis. *J Infect*. 2020;81(2):266–75. **Important systematic review/meta-analysis on bacterial co-infections in patients with COVID-19.**
102. ECDC. European Centre for Disease Prevention and Control. Antimicrobial consumption in the EU/EEA (ESAC-Net)-Annual Epidemiological Report 2020. Stockholm: ECDC; 2021. 2020.
103. Rezel-Potts E, L'Esperance V, Gulliford MC. Antimicrobial stewardship in the UK during the COVID-19 pandemic: a population-based cohort study and interrupted time-series analysis. *Br J Gen Pract*. 2021;71(706):e331–8.
104. Pasero D, Cossu AP, Terragni P. Multi-drug resistance bacterial infections in critically ill patients admitted with COVID-19. *Microorganisms*. 2021;9(8).
- 105.● O'Toole RF. The interface between COVID-19 and bacterial healthcare-associated infections. *Clin Microbiol Infect*. 2021;27(12):1772–6. **Informative review of COVID-19 and emergence of nosocomial MDR infections.**
- 106.● Aurilio C, Sansone P, Paladini A, Barbarisi M, Coppolino F, Pota V, et al. Multidrug resistance prevalence in COVID Area. *Life (Basel)*. 2021;11(7). **Informative review on MDR and COVID-19.**
107. Polly M, de Almeida BL, Lennon RP, Cortês MF, Costa SF, Guimarães T. Impact of the COVID-19 pandemic on the incidence of multidrug-resistant bacterial infections in an acute care hospital in Brazil. *Am J Infect Control*. 2022;50(1):32–8.
108. Porretta AD, Baggiani A, Arzilli G, Casigliani V, Mariotti T, Mariottini F, et al. Increased risk of acquisition of New Delhi metallo-beta-lactamase-producing carbapenem-resistant Enterobacterales (NDM-CRE) among a cohort of COVID-19 patients in a teaching hospital in Tuscany, Italy. *Pathogens*. 2020;9(8).
109. Nori P, Szymczak W, Puius Y, Sharma A, Cowman K, Gialanella P, et al. Emerging co-pathogens: New Delhi metallo-beta-lactamase producing Enterobacterales infections in New York City COVID-19 patients. *Int J Antimicrob Agents*. 2020;56(6):106179.
110. Gottesman T, Fedorowsky R, Yerushalmi R, Lellouche J, Nutman A. An outbreak of carbapenem-resistant *Acinetobacter baumannii* in a COVID-19 dedicated hospital. *Infect Prev Pract*. 2021;3(1):100113.
111. Bentivegna E, Luciani M, Arcari L, Santino I, Simmaco M, Martelletti P. Reduction of multidrug-resistant (MDR) bacterial infections during the COVID-19 Pandemic: a retrospective study. *Int J Environ Res Public Health*. 2021;18(3).
112. Bharat A, Machuca TN, Querrey M, Kurihara C, Garza-Castillon R, Kim S, et al. Early outcomes after lung transplantation for severe COVID-19: a series of the first consecutive cases from four countries. *Lancet Respir Med*. 2021;9(5):487–97.
113. King CS, Mannem H, Kukreja J, Aryal S, Tang D, Singer JP, et al. Lung transplantation for patients with COVID-19. *Chest*. 2022;161(1):169–78.
114. Yeung JC, Cypel M, Chaparro C, Keshavjee S. Lung transplantation for acute COVID-19: the Toronto Lung Transplant Program experience. *CMAJ*. 2021;193(38):E1494–7.
- 115.● Barlam TF, Cosgrove SE, Abbo LM, MacDougall C, Schuetz AN, Septimus EJ, et al. Implementing an antibiotic stewardship program: guidelines by the Infectious Diseases Society of America and the Society for Healthcare Epidemiology of America. *Clin Infect Dis*. 2016;62(10):e51–77. **Comprehensive guideline on the implementation antimicrobial stewardship in hospital setting.**
116. Sfeir MM. The GAIN Act legislation to combat antimicrobial resistance: where do we stand? *Infect Control Hosp Epidemiol*. 2018;39(12):1499–500.
117. Shafiq N, Pandey AK, Malhotra S, Holmes A, Mendelson M, Malpani R, et al. Shortage of essential antimicrobials: a major challenge to global health security. *BMJ Glob Health*. 2021;6(11).
118. Dyar OJ, Huttner B, Schouten J, Pulcini C, Esgap. What is antimicrobial stewardship? *Clin Microbiol Infect*. 2017;23(11):793–8.
- 119.●● Centers for Disease Control and Prevention. Core elements of antibiotic stewardship. Atlanta, GA, USA: U.S. Department of Health & Human Services. 2022. [Available from: <https://www.cdc.gov/antibiotic-use/core-elements/index.html>]. Accessed 3 Jan 2022. **Practical guidance on the implementation of antimicrobial stewardship programs in hospital setting.**
- 120.● Centers for Medicare & Medicaid Services. Omnibus burden reduction (conditions of participation) final rule CMS-3346-F. 2019. [Available from: <https://www.cms.gov/newsroom/fact-sheets/omnibus-burden-reduction-conditions-participation-final-rule-cms-3346-f>]. Accessed 3 Jan 2022. **Requirements for antimicrobial stewardship programs per US Centers for Medicare and Medicaid.**
- 121.● The Joint Commission. New antimicrobial stewardship standard. R3 Report Requirement, Rationale, Reference [Internet]. 2016; (8). Available from: <https://www.jointcommission.org/standards/r3-report/r3-report-issue-8-new-antimicrobial-stewardship-standard/>. Accessed 3 Jan 2022. **The US Joint Commission requirements for antimicrobial stewardship programs.**
- 122.● The Joint Commission. Antimicrobial Stewardship in Ambulatory Health Care. R3 report requirement, rationale, reference [Internet]. 2019 May 5, 2020;(23). Available from: <https://www.jointcommission.org/en/standards/r3-report/r3-report-issue-23-antimicrobial-stewardship-in-ambulatory-health-care/>. Accessed 3 Jan 2022. **The US Joint Commission requirements for antimicrobial stewardship programs in ambulatory setting.**
123. Green M, Blumberg EA, Danziger-Isakov L, Huprikar S, Kotton CN, Kumar D. Foreword: 4th edition of the American Society of Transplantation Infectious Diseases Guidelines. *Clin Transplant*. 2019;33(9):e13642.
124. Fishman JA. Infection in organ transplantation. *Am J Transplant*. 2017;17(4):856–79.
- 125.● Seo SK, Lo K, Abbo LM. Current state of antimicrobial stewardship at solid organ and hematopoietic cell transplant centers in the United States. *Infect Control Hosp Epidemiol*.

- 2016;37(10):1195–200. **Survey of antimicrobial stewardship interventions in transplant centers in the US.**
126. Magill SS, O'Leary E, Ray SM, Kainer MA, Evans C, Bamberg WM, et al. Assessment of the appropriateness of antimicrobial use in US hospitals. *JAMA Netw Open.* 2021;4(3):e212007.
- 127.●● Centers for Disease Control and Prevention. Antibiotic Use in the United States, 2021 Update: Progress and Opportunities. 2021. **Updated CDC antibiotic threats report on prevalence and impact of MDR organisms in the US.**
128. Abbo LM, Grossi PA, The AST ID Community of Practice. Surgical site infections: guidelines from the American Society of Transplantation Infectious Diseases Community of Practice. *Clinical Transplantation.* 2019;33(9):e13589.
- 129.● Yau J, Dann J, Geyston J, Hall HC, Pelletier S, Sifri CD. Eliminated routine postorthotopic liver transplant antibiotics in uncomplicated patients leads to equivalent safety outcomes. *Antimicrobial Stewardship Healthcare Epidemiol.* 2022;2(1). **A study on reducing antimicrobial exposure in liver recipients.**

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.