Check for updates

OPEN ACCESS

EDITED BY Saeed Ahmad Khan, Kohat University of Science and Technology, Pakistan

REVIEWED BY Riya Biswas, Auckland University of Technology, New Zealand Abimanyu Sugumaran, SRM Institute of Science and Technology, India Shibing Wang, Zhejiang Provincial People's Hospital, China

*CORRESPONDENCE Aws Alshamsan, aalshamsan@ksu.edu.sa,

⁺These authors have contributed equally to this work

SPECIALTY SECTION

This article was submitted to Experimental Pharmacology and Drug Discovery, a section of the journal Frontiers in Pharmacology

RECEIVED 30 April 2022 ACCEPTED 29 July 2022 PUBLISHED 24 August 2022

CITATION

Alshememry AK, Alsaleh NB, Alkhudair N, Alzhrani R and Alshamsan A (2022), Recent nanotechnology advancements to treat multidrugresistance pancreatic cancer: Preclinical and clinical overview. *Front. Pharmacol.* 13:933457. doi: 10.3389/fphar.2022.933457

COPYRIGHT

© 2022 Alshememry, Alsaleh, Alkhudair, Alzhrani and Alshamsan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Recent nanotechnology advancements to treat multidrug-resistance pancreatic cancer: Pre-clinical and clinical overview

Abdullah K. Alshememry^{1,2†}, Nasser B. Alsaleh^{2,3†}, Nora Alkhudair⁴, Rami Alzhrani⁵ and Aws Alshamsan^{1,2}*

¹Department of Pharmaceutics, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia, ²Nanobiotechnology Unit, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia, ³Department of Pharmacology and Toxicology, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia, ⁴Department of Clinical Pharmacy, College of Pharmacy, King Saud University, Riyadh, Saudi Arabia, ⁵Department of Pharmaceutics and Pharmaceutical Technology, College of Pharmacy, Taif University, Taif, Saudi Arabia

Pancreatic cancer (PC) remains one of the most lethal and incurable forms of cancer and has a poor prognosis. One of the significant therapeutic challenges in PC is multidrug resistance (MDR), a phenomenon in which cancer cells develop resistance toward administered therapy. Development of novel therapeutic platforms that could overcome MDR in PC is crucial for improving therapeutic outcomes. Nanotechnology is emerging as a promising tool to enhance drug efficacy and minimize off-target responses via passive and/or active targeting mechanisms. Over the past decade, tremendous efforts have been made to utilize nanocarriers capable of targeting PC cells while minimizing off-target effects. In this review article, we first give an overview of PC and the major molecular mechanisms of MDR, and then we discuss recent advancements in the development of nanocarriers used to overcome PC drug resistance. In doing so, we explore the developmental stages of this research in both pre-clinical and clinical settings. Lastly, we discuss current challenges and gaps in the literature as well as potential future directions in the field.

KEYWORDS

multidrug resistance, drug delivery, paclitaxel, tumor microenvironment, pancreatic cancer, non-clinical studies, clinical studies

Introduction

Pancreatic cancer (PC) accounts for 2.5% of all cancer cases worldwide, making it the fourth leading cause of cancer mortality (Siegel et al., 2019). An estimated 495,773 patients were diagnosed with PC in 2021 (Cancer.Net, 2022), with approximately 50% of the cases were diagnosed with metastatic disease (Bray et al., 2018). The location of the tumor

TABLE 1 Pancreatic cancer pathological types. Pancreatic cancer pathologically originates from exocrine or endocrine cells. The prevalence and common mutations are different depending on the tumor type and cell origin. Invasive ductal adenocarcinoma is a common type of PC in cells with an exocrine origin.

Туре	Category	Prevalence	Mutations
Invasive ductal adenocarcinoma	Exocrine	95%	KRAS, P16/CDKN2A, TP53, SMAD4
Acinar cell carcinoma	Exocrine	1-2%	APC/β-catenin
Serous Cystadenocarcinoma	Exocrine	3%	VHL
Neuroendocrine (PENT)	Endocrine	5%	MEN1

KRAS: Kirsten rat sarcoma viral oncogene homolog; CDKN2A: Cyclin-dependent kinase inhibitor 2A; TP53: Tumor protein p53; SMAD: an acronym from the fusion of *Caenorhabditis elegans Sma* genes and the *Drosophila Mad*; APC: Adenomatous polyposis coli; VHL: Von Hippel-Lindau; MEN1: Multiple endocrine neoplasia type 1.

within the pancreas can influence the symptoms and clinical presentation (Ryan et al., 2014; Ducreux et al., 2015; Tempero et al., 2019). Almost 65% of PC tumors develop in the head and the neck of the pancreas, and patients commonly present with jaundice and abdominal pain due to bile obstruction (Ducreux et al., 2015; Tempero et al., 2019). In some cases, PC can either be located in the pancreas body (15%), and the tail (10%), or can be a multifocal tumor (2%). Furthermore, patients can present with other non-specific symptoms, including late-onset type II diabetes, abdominal pain, weight loss, and steatorrhea (Gheorghe et al., 2020).

Pathologically, PC originates from two types of cells: exocrine or endocrine cells (Bardeesy and DePinho, 2002; Ghaneh et al., 2007; Corbo et al., 2012; Aguirre and Collisson, 2017). Over 95% of all PC types are of exocrine origin, where the majority of these cases are pancreatic adenocarcinomas (PDAC) (Cowgill and Muscarella, 2003; Li and Jiao, 2003). Less common cells are those of acinar and cystadenocarcinoma origin (Table 1). The endocrine tumors include neuroendocrine tumors (PNET), which account for 5% of all PC cases (Hackeng et al., 2016). PNETs grow slowly and are less aggressive than the PDAC (Bardeesy and DePinho, 2002; Ghaneh et al., 2007). PDAC is a complex disease with poorly differentiated histological features. The high intertumoral heterogeneity, genomic instability, and stromal desmoplasia formation cause significant challenges in the early diagnosis and treatment of PDAC (Ghaneh et al., 2007; Corbo et al., 2012; Aguirre and Collisson, 2017). Pancreatic intraepithelial neoplasia (PanIN) is the most common early non-invasive precursor lesion of PDAC, in addition to mucinous cystic neoplasm and intraductal papillary mucinous neoplasm. The precursor's progression from low-grade to highgrade is due to genetic and epigenetic alterations that lead to the formation of invasive PDAC. When PDAC lesions form, the genetic mutations will continue to progress beyond primary mutations (Ghaneh et al., 2007; Rishi et al., 2015; Hackeng et al., 2016; Dreyer et al., 2017).

Drug resistance, both intrinsic (innate) and acquired (in response to drug therapy), is a key contributing factor to the poor prognosis of PC (Binenbaum et al., 2015). The survival rate of PC has remained almost unchanged for several decades, and it

is considered among the lowest, with a 5-year survival rate for a maximum of 9% of cases for all stages combined (Rawla et al., 2019; Siegel et al., 2019). Such a low rate of survival is attributed to two major factors: 1) late diagnosis of typically advanced/ metastasized and unresectable PC due to a lack of early diagnostic biomarkers; and 2) lack of effective therapeutic intervention (Siegel et al., 2019). There are fundamental reasons why pancreatic tumors are difficult to treat in PC. Firstly, pancreatic tumors, such as PDAC, are solid tumors comprised of a dense stromal environment of cancerous cells, non-cancerous cells such as fibroblasts, and a dense extracellular matrix. The density of these arrangements impedes drug permeation. Additionally, if a drug can permeate the cancerous stroma, the drug molecules are typically unable to differentiate between cancerous and non-cancerous cells due to the dense tumor stroma (Heidemann and Kirschner, 1978; Sofuni et al., 2005). An important determinant of peritoneal metastasis is the anatomical position of the primary tumor (Baretti et al., 2019). In some cases, cells directly attach to and invade organs and tissues in the peritoneal cavity (Avula et al., 2020) or result in intraperitoneal metastases via blood vessels or lymphatic absorption through the hematogenous route (Ge et al., 2017). In most cases of PC, metastasis has already occurred by the time of diagnosis. Furthermore, even when surgical intervention is applicable, following chemoradiotherapy, rapid relapse is often seen due to the presence of pancreatic satellite cells that promote carcinogenesis (Hosoki, 1983; Sofuni et al., 2005; Zhi et al., 2014; Binenbaum et al., 2015).

The stromal desmoplastic reactions induced by the pancreatic stellate cells (PSC), when activated by growth factors, lead to the secretion of collagen, hyaluronic acid, and other components of the extracellular matrix. Thus, PSCs induce stromal fibrosis, reduce cellular vascularity, and induce hypoxia. The stromal barrier created by PSC elevates the interstitial fluid pressure and compresses the blood vessels, preventing passive transportation of chemotherapeutic agents and eventually leading to treatment failure (Erkan et al., 2012; Hackeng et al., 2016). Efficacy of Trans-arterial chemoembolization (TACE) with TACE + radiofrequency ablation (RFA) and/or ¹²⁵I radioactive seed implantation for unresectable pancreatic

10.3389/fphar.2022.933457

cancer are other approaches that have been studied retrospectively and TACE combined with either ¹²⁵I seed implantation and/or RFA, was shown to have improved treatment response and overall survival rate compared with TACE alone (Das et al., 2019). However, in the same study, the overall survival rates were better in the case of RFA. To summarize, based on these previous data, the sequence of use and the best combination therapy in the case of PC remains to be investigated.

While there are numerous drugs available for the treatment of PC, two nanoparticle formulations, Abraxane® and Onivyde® (irinotecan liposome injection) have been approved as of July 2022 by the US FDA for the treatment of metastasized cancers, in combination with gemcitabine. Abraxane® is a Paclitaxel Albumin-stabilized Nanoparticle Formulation, approved for the treatment of metastasized PC in combination with gemcitabine hydrochloride as a first-line treatment. More recently, the use of nanotechnology for delivering drugs to targeted sites in the body is an inexpensive and effective system for treating diseases and other conditions. Indeed, the use of nanotechnology can revolutionize the treatment of cancer, by enabling diagnostic tools for early detection of the disease (Hu et al., 2021) as well as improving drug delivery. In PC, the delivery and distribution of drugs to the tumor are compromised due to intrinsic physical and biochemical barriers, which result in increased interstitial fluid pressure, vascular compression, and hypoxia. Moreover, therapies based on targeting immune responses including therapeutic vaccines, immune checkpoint inhibition, and CAR-T cell therapy often do not show expected responses due to a highly immunosuppressive tumor microenvironment. These two factors present as a major challenge for developing effective therapies against PC. Nanoparticles have been extensively studied as delivery platforms and adjuvants for cancer and other disease therapies. Knowledge gained through using nanocarrier-based systems in other cancer types, combined with the ability to modulate nanocarriers toward targeting multiple MDR mechanisms simultaneously provides an opportunity to enable improvement in drug delivery and enhancing therapeutic outcomes for PC.

Mechanisms of resistance in pancreatic cancer

Numerous alterations at the genetic, epigenetic, and protein levels are implicated in PC drug resistance (Binenbaum et al., 2015). Although different therapy regimens exist, the current standard of care regimen therapy for PC remains largely dependent on gemcitabine (GEM), which is considered a gold standard in chemotherapy. GEM is a nucleoside analog, which provides only a modest clinical benefit (Burris et al., 1997). Multiple combination regimens of chemotherapeutics such as GEM with 5-fluorouracil (5-FU), cisplatin or paclitaxel (PTX), or FOLFIRINOX (i.e., fluorouracil, leucovorin, irinotecan, and oxaliplatin alone or alongside targeted therapies [e.g., cetuximab and bevacizumab]) have failed to demonstrate significant clinical benefits (Berlin et al., 2002; Cascinu et al., 2006; Heinemann et al., 2006). Despite the initial response to chemotherapy in the different forms of PC, the rapid development of drug resistance remains a major challenge in the treatment of PC (Binenbaum et al., 2015).

It is now well established that a variety of cancers mediate their aggressiveness and resistance to chemoradiotherapy via modulating key cellular regulatory pathways that control cell proliferation and differentiation, inflammation, and programmed cell death pathways, including apoptosis and autophagy (Xia et al., 2014). PC shows a significant upregulation of ATP binding cassette (ABC) transporters ABCB4, ABCB11, ABCC1, ABCC3, ABCC5, ABCC10, and ABCG2 at the RNA level in tumors relative to the normal pancreas (Mohelnikova-Duchonova et al., 2013). Additionally, drug efflux pump MDR1/P-gp is highly expressed in PC cells (O'Driscoll et al., 2007), which may play a critical role in the development of resistance to chemotherapeutic agents. Therefore, it is equally important to understand the underlying molecular mechanisms of PC drug resistance for unraveling novel therapeutic interventions with improved efficacy. In addition, somatic mutations in key genes such as many proto-oncogenes (e.g., Ras, Myc, Cdk4) are critical in the initiation and progression of malignant tumors. However, cancer treatment is even more challenging because tumor exposure to therapy, including chemoradiotherapy and targeted therapy, is often associated with further mutations and the development of compensatory mechanisms that render cancer refractoriness to therapy and increased aggressiveness and metastasis (Binenbaum et al., 2015). Therefore, understanding the molecular mechanisms of drug resistance is crucial in order to intervene and eventually win the battle against cancer.

The main mutations of PDAC include KRAS, CDKN2A, TP53, and SMAD4 (Hackeng et al., 2016). Over 90% of PC is associated with KRAS mutations, most commonly KRAS^{G12D}, during both the initial stage (i.e., precursor lesions that develop into invasive pancreatic ductal adenocarcinoma, also known as pancreatic intraepithelial neoplasia) and progression stage (di Magliano and Logsdon, 2013). There have been efforts to discover new PDAC targeting agents. A phase 1 clinical trial (NCT04117087) used a long peptide vaccine combined with Nivolumab and Ipilimumab for resected MMR-p colorectal and pancreatic cancer patients. Interestingly, Govindan et al. discovered that AMG 510 is a novel small molecule that can bind specifically and irreversibly in KRAS^{G12C} (Govindan et al., 2019; Alzhrani et al., 2021). KRAS^{G12C} is a mutation that is predominantly found in non-small lung cancer. However, the use of AMG 510 is limited in PDAC because KRAS^{G12C} mutation only accounts for 2%. These findings will motivate the scientific



FIGURE 1

Major molecular contexts underlying drug resistance in pancreatic cancer. This figure illustrates the three major molecular contexts underlying drug resistance in pancreatic cancer. These include: 1) the process of epithelial-mesenchymal transition (EMT) in which cancer cells lose their epithelial phenotype (such as cell-to-cell contact) to gain more aggressive and metastatic mesenchymal phenotypes; 2) expansion of subpopulation of pancreatic cancer stem cells (PCSCs); and 3) dynamic tumor microenvironment (TME). The tumor stroma surrounding cancer cells and PCSCs consist non-cellular components including extracellular matrix (ECM) and multiple cell types including CAF, MDSC, Th-cells, Treg, TAM and DC. Cytokines released from the different cell types help maintaining the TME in dynamic state that supports tumor growth and metastasis. Th-cells exist in TME but are suppressed. DCs, which are key in the processing and presentation of cancer neoantigens, exist in TME but they are suppressed as well. EMT: epithelial-mesenchymal transition; PCSC: pancreatic cancer stem cell: TME: tumor microenvironment: CAF: Cancer-associated fibroblast: MDSC: myeloid-derived suppressor cell; Th-cells: Helper T cells; Treg: Regulatory T-cell; TAM: tumor-associated macrophage; DC: Dendritic cell.

community to develop new drugs that can target inactive KRAS^{G12D} and KRAS^{G12V}, which account for 80% of PDAC (Govindan et al., 2019; Alzhrani et al., 2021).

It is worth noting that tremendous efforts over the past years have revealed numerous molecular components and intricate signaling networks that are deregulated in PC, contributing to chemoresistance. These include, but are not limited, drug transporters (e.g., hENT and hCNT), intracellular enzymes (e.g., deoxycytidine kinase), dCK, which is critical for GEM bioactivation, DNA repair mechanisms (e.g., excision repair cross-complementation 1), ERCC1, antioxidant response (e.g., Nrf and HSPs), signaling pathways that regulate cell-cycle and programmed cell death (e.g., Nuclear Factor κB [NFκB], MAPK, PI3K/Akt and p53), epigenetic components (e.g., histone deacetylase), HDAC and more recently, noncoding RNAs (ncRNAs) [e.g., microRNAs (miRNAs), long noncoding RNAs (lncRNAs) and circular RNAs (circRNAs)] (Binenbaum et al., 2015; Xie et al., 2020a; Lin et al., 2020; Pandya et al., 2020). More recently, differential upregulation and functional role of the transmembrane mucin MUC4 in PC is an attractive target for

immunotherapy and MUC4 β encapsulation in polyanhydride nanoparticles has been shown to provide long-term protection against rapid phagocytic and proteolytic clearance in circulation. Stable MUC4 β release from these nanoparticles and its immunogenic capacity has recently been demonstrated by Liu and colleagues in mice models (Liu et al., 2021). Discussing the specific molecular mechanism is beyond the scope of this review and is discussed in detail elsewhere, but here we give an overall overview and describe three key themes involved in PC drug resistance. These are epithelial-mesenchymal transition (EMT), expansion of pancreatic cancer stem cells (PCSCs), and dynamic state of the tumor microenvironment (TME), as illustrated in (Figure 1) (Binenbaum et al., 2015; Zeng et al., 2019).

Epithelial-mesenchymal transition (EMT) is a dynamic process that involves the transitioning of differentiated epithelial cells into the mesenchymal cell phenotype. During this transformation, cells lose key features of epithelial cells such as intercellular junctions while simultaneously undergoing cytoskeletal rearrangement of the mesenchymal cell phenotype (Nieto et al., 2016). EMT is particularly critical during cancer progression and the generation of stem-like cells with a high capacity for metastasis (Nieto et al., 2016). Notably, previous literature has demonstrated the key role of EMT in conferring drug resistance against chemoradiotherapy and targeted therapy in a variety of solid tumors, including PC (Shah et al., 2007; Wang et al., 2009; Singh and Settleman, 2010; Nieto et al., 2016). For instance, chemoresistance to GEM in PC cells has been associated with increased expression of mesenchymal markers such as vimentin and ZEB1, in contrast to GEM-sensitive cells with a high E-cadherin expression: a common epithelial phenotype maker (Li et al., 2009). Similar findings have been found with other chemotherapeutics, such as 5-FU and cisplatin (Arumugam et al., 2009). Although the underlying molecular mechanisms of EMT-induced chemoresistance are not fully understood, accumulating evidence suggests the involvement of multiple signaling pathways, including NFkB, TGFB, and Notch pathways (Ellenrieder et al., 2001; Min et al., 2008; Wang et al., 2009). Another key molecular regulator of EMT in PC cells is miRNAs: small non-coding RNAs that regulate gene expression (Yonemori et al., 2017). A multitude of miRNAs has been shown to be implicated in regulating major cellular signaling pathways involved in the development of PC (Yonemori et al., 2017). For instance, a low expression of miR-200 was found in the GEM-resistant but not in the GEM-sensitive PC cell lines (Li et al., 2009). Together, these findings indicate the critical role of EMT in the development of PC drug resistance in transformed cells.

Previous evidence has demonstrated that cancers are heterogenous in nature, with subpopulations of cells of varying phenotypes. Cancer stem cells (CSCs) represent a small subpopulation of tumor cells with a high capacity for self-renewing, differentiation, and tumor progression (Yu et al., 2012). Indeed, CSCs display a high capacity for

clinical success (Ho et al., 2020). For instance, within the stromal cells, cancer-associated fibroblasts (CAFs), which are

the major source of ECM, are associated with

tumorigenesis when transplanted into a host compared to other tumor subpopulations (Yu et al., 2012). Several cellular surface markers have been identified to be expressed by CSCs in various cancers, including CD44, CD24, CD133, aldehyde dehydrogenase1 (ALDH1), and epithelial-specific antigen (ESA) (Yu et al., 2012). In PC, previous work has demonstrated that CD44, CD24, and ESA-expressing PCSCs, represented only by a small percentage of the cancer cell population (~1%), were associated with high tumorigenic potential, and it is likely that PCSCs at least partially mediate chemoradiotherapy-induced drug resistance, as evident in PCSC enrichment in response to chemoradiotherapy treatment (Du et al., 2011). Similar findings were also found in another report, demonstrating that within a pancreatic tumor, a subset of undifferentiated, stem-like cells expressing the surface marker CD133, was associated with high tumorigenic potential and chemoresistance (Hermann et al., 2007). Although there is an overlap between the two subpopulations of PCSCs in the two reports (CD44/CD24/ESA-vs. CD133-expressing PCSCs), they were not identical (Hermann et al., 2007). Additionally, another subset of cells with exclusive migratory properties expressing CD133 and CXCR4 were identified at the invasive front of pancreatic tumors. It was found that without these CD133+/ CXCR4⁺ cells, metastasis was almost completely abrogated (Hermann et al., 2007). Together, these findings indicate the existence of different subsets of PCSCs within pancreatic tumors with a high capacity for tumorigenesis and drug resistance. Such findings demonstrate the heterogenic nature of cancers which has been long disregarded in cancer research. It is worth noting that EMT transformed cells can share cellular and molecular features of CSCs, which could explain an increased tumorigenic profile of EMT transformed cells as well as drug resistance. Nonetheless, these cells, unlike PCSCs, were sensitive to conventional chemotherapy. Furthermore, it is now widely accepted that subpopulations of CSCs exist in a dynamic state (quiescent vs. slow-vs. rapid-cycling CSCs), which is constantly influenced by signals and cues from their surroundings (the tumor microenvironment (TME). With their large plasticity and ability to switch to a quiescent state, CSCs can resist chemoradiotherapy (Batlle and Clevers, 2017).

The tumor microenvironment (TME) consists of cellular and non-cellular components (extracellular matrix (ECM)) and plays a crucial role in drug resistance. Indeed, one major obstacle in the treatment of PC is the dense fibrotic stroma, also known as desmoplastic stroma, surrounding tumor cells that act as a physical barrier. As such, they prevent drugs from infiltrating tumor core cells (Dauer et al., 2017). Thus, the dense fibrotic stroma is considered a histopathological hallmark of PC. It is worth noting that TME is composed of several types of cells, including fibroblasts, pancreatic stellate cells (myofibroblast-like cells), and a variety of immune cells and despite the considerable efforts made to target a multitude of cellular and noncellular components within TME, yet it was associated with limited immunosuppressive properties (Kraman et al., 2010). Depletion of CAFs was associated with enhanced efficacy of immune checkpoint inhibitors (ICIs) (Feig et al., 2013). In addition to CAFs, other cell types within the TME including regulatory T-cells (Treg), myeloid-derived suppressor cell (MDSC), and tumor-associated macrophage (TAM) also possess immunosuppressive properties (Ho et al., 2020). Cancer immunotherapy has revolutionized the field over the past couple of decades. Notable success has been made against multiple solid cancers, particularly those with high tumor mutational burden (TMB) within the tumor genome, such as melanoma and lung cancer (Goodman et al., 2017; Yousefi et al., 2017). This success has not been evident in PC (Schizas et al., 2020), which has largely been attributed to the nonimmunogenic nature of most types of PC. In other words, most PC types are not readily recognized by the immune cells due to low TMB and thus a low number of neoantigens, a tumorassociated antigen, and subsequent presences of TILs (Bailey et al., 2016; Danilova et al., 2019). Additionally, as mentioned earlier, PC is typically contained in a highly dense fibrotic stroma consisting of multiple cell types with immunosuppressive properties (Ho et al., 2020; Schizas et al., 2020). Unsurprisingly, together, these two factors make cancer immunotherapy less effective in PC. Utilizing the properties of TME for controlling local therapeutic delivery is an area of active research (Alshememry et al., 2017).

Nanotechnology for the treatment of pancreatic cancer

Nanomaterials and nanoparticles (NPs) are extremely small (1-100 nm) in size and hence can directly interact with biological molecules (Hosein et al., 2013). Engineered NPs are an excellent tool for drug delivery due to their unique structural properties which include a large surface-to-mass ratio, capacity to be modulated to bind different cellular targets, and ability to carry different cargo including proteins, nucleotides, and drugs. Advances in the field of nanotechnology have created tremendous prospects for improving therapeutic drug delivery (Schroeder et al., 2011; Melancon et al., 2012; Prabhu and Patravale, 2012). Currently, the use of nanotechnology in drug delivery typically involves a combination of nanomaterials and a drug of interest, and a significant number of nanoplatforms are being employed and are under testing in the different phases of clinical trials (Ogawa and Miura, 2014; Rebelo et al., 2017). These combinations utilize different types of nanomaterials such as polymeric NPs, liposomes, amphiphilic polymer NPs, small interfering RNA (siRNA), graft polymers, dendrimers, thermo-responsive polymers, mixed micelles, ultrasoundresponsive nano-emulsions, carbon nanotubes, quantum dots and inorganic NPs (magnetic-hybrid NPs, and gold NPs) (Manzur et al., 2017; Rebelo et al., 2017; Sielaff and Mousa, 2018).

Nanotechnology has been largely utilized in cancer research to improve the delivery of drugs to the tumor site exploiting the leaky vasculature of the tumor via passive or enhanced permeability and retention (EPR) effect. However, a major challenge in PC is its hypovascularization, and hence, other strategies must be utilized, such as using targeted (active) delivery (Xie et al., 2020b). As a result of this insufficient vascularization, ineffective distribution of drugs may account for much of the chemotherapy resistance seen in PC treatment. The use of nanotherapy may bypass the inefficient vascularization issues by delivering chemotherapeutic drugs directly to the pancreatic tissue, for example, by targeting stromal hedgehog receptors rather than relying on blood flow, a methodology that provides a tunable distribution of chemotherapeutic agents throughout pancreatic tumor tissue, thereby increasing drug solubility, half-life, and stability (Jiang et al., 2020). Moreover, NPs delivering chemotherapeutic drugs can bypass multidrug-resistant (MDR) efflux pumps present on the surface of most tumor cells (Lobato-Mendizabal and Ruiz-Arguelles, 1990; Blanco et al., 2011; McCarroll et al., 2014). This, among other reasons that will be discussed in later sections, supports the use of nanotechnology in PC treatment.

Nano-based intervention to overcome MDR-PC in pre-clinical and clinical settings: Key examples

The use of NPs has several advantages, including minimizing MDR and drug-related toxicities. Nanoparticles can target a wide range of physiological and metabolic characteristics of the targeted tissues, thereby increasing biodistribution and bioavailability of drugs and enhancing their plasma half-life and EPR (Parhi et al., 2012; Poon et al., 2015; Gad et al., 2016; Liu et al., 2022). Recently, the potency of the chemotherapeutic agents for the treatment of PC has been improved through the use of RNA interference (RNAi) technologies, including miRNA and siRNA, which selectively suppress the expression of target genes leading to increased drug efficacy and enhancing anti-cancer activity (Tang et al., 2021). Integrating RNAi with NPs can therefore be extremely effective at treating PC (Hiss et al., 2007; Gurunathan et al., 2018). Studies have recently shown that nano-sized exosomes are an efficient RNAi carrier, making them an attractive delivery cargo to cancerous cells (Farran and Nagaraju, 2020) Exosomes have intrinsic advantages over liposomes, as they are less toxic and can be dosed at higher concentrations in the blood to work as molecular cargos which could be used to inhibit oncogenes, activate tumor suppressor genes and modulate immune responses to control tumor cell growth (Oliveira et al., 2021) (Zhao et al., 2021b). As researchers continue to uncover the cellular and molecular basis of PC drug resistance, nanotechnology used for the delivery of drugs can be further engineered to provide effective solutions for mitigating MDR in the treatment of PC (Zhang et al., 2017). Table 2 summarizes key nano-based systems used for the delivery of antitumor drugs to mitigate MDR.

Albumin-based nanoparticles in PC

Albumin-based nanoparticles can be utilized as theranostics (i.e., to deliver therapeutic agents and simultaneously used for diagnosis). Albumin is the most abundant plasma protein and known ligand to be associated with a caveolae-mediated endocytosis mechanism. Albumin-based nanoparticles when internalized by the cell via caveolae-mediated endocytosis, can overcome the issue of MDR by bypassing and evading ATPbinding cassette (ABC) transporters, which are responsible for the efflux of anticancer drugs and subsequent MDR once released into the cytoplasm (Yuan et al., 2016). Nanoparticle albumin (Nab) is made by mixing human albumin in an aqueous medium under high pressure to form 100-200 nm albumin NPs. These NPs are mixed with chemotherapeutic alkaloids such as PTX (Macarulla et al., 2019). As shown in Table 3, Nab-based delivery systems are the most extensively studied nanocarrier system in the treatment of PC in human clinical trials (Von Hoff et al., 2011; Hosein et al., 2013; Von Hoff et al., 2013; Goldstein et al., 2015; Vogel et al., 2016; Macarulla et al., 2019). A common example of albumin-based nanoparticles is Nab-PTX, an albumin-binding PTX and a microtubule-stabilizing agent known to enhance microtubule polymerization during mitosis leading to cell cycle arrest in the G2/M phases. Based on these properties, Nab-PTX can stop rapid and uncontrollable cell division and help overcome MDR receptor-mediated endocytosis (Demidenko et al., 2008; Guo et al., 2018). Importantly, Nab-PTX has been shown to mitigate MDR in PC (Guo et al., 2018). In 2018, Guo et al. created GEM-resistant pancreatic cells by inducing lower rates of hENT1 expression. These cells were then exposed to free GEM or GEM delivered using human-serum albumin nanoparticles (HSA-NPs). Their results showed that GEM-HSA-NPs was more effective at slowing pancreatic cell proliferation and triggering apoptosis in comparison to free GEM alone, without any increase in toxicity, as shown in vivo studies. In phase III clinical trial (MPACT) on previously untreated patients with metastatic PC, a combination of Nab-PTX and GEM increased the median OS of patients receiving nab-PTX and gemcitabine to 8.7 vs. 6.6 months in patients treated with gencitabine alone (p <0.0001) (van Horssen et al., 2006; Libutti et al., 2010). Furthermore, the combination showed an increase in the cumulative delivery of gemcitabine by 2.8-fold compared to gemcitabine alone (Von Hoff et al., 2011; Von Hoff et al.,

TABLE 2 Selected examples of nano-based formulations used to overcome MDR in tumor cells.

Type of nanocarrier	Objective	Cargo	Cell/Animal model	Effects on MDR	Outcomes/results
M1Exo-GEM-DFX (Zhao et al., 2021a)	M1Exo was engineered as a drug carrier to co-delivery DFX and GEM to overcome the chemoresistance of GEM and improve its therapeutic potential	DFX and GEM	PANC-1 cells	Inhibit cell P-glycoprotein expression	M1Exo-GEM-DFX was able to overcome GEM resistance induced by P-glycoprotein expression <i>in vitro</i>
s (DGL) _n @Apt NPs(Chen et al., 2022)	Modulation of PDAC stromal structure and send chemotherapy drugs to the deep tumor vis the use of Aptamer-decorated hypoxia-responsive nanoparticle s (DGL) _n @Apt	GEM + STAT3 inhibitor (HJC0152)	Pan02 multicellular spheroids (MCSs) cells / Pan02 xenograft mice	Inhibition of the STAT3 pathway	Triggered by hypoxia, the ultra-small dual-loaded DGL NPs exhibited excellent deep-tumor penetration, promoted drugs endocytosis, and autophagy induction
PEG-Gem-cisPt-MSNs (Tarannum et al., 2022)	Development of two versions of mesoporous silica nanoparticles (MSNs), a dual loaded PEG-functionalized NPs, and MSNs containing Sonic Hedgehog (SHh) inhibitor for stroma modulation and improved delivery	GEM + CisPt	HPAF II and Miapaca-2 cells / HPAF II xenograft mice	Inhibition of sonic hedgehog (SHh) signaling pathway	The sequential combination of CyP- MSNs followed by PEG- Gem-cisPt- MSNs led to (i) effective stromal modulation (ii) increased access to secondary PEG- Gem-cisPt-MSNs at the tumor site (iii) enhanced therapeutic performance in HPAF II xenograft mice
TPMILs (Obaid et al., 2022)	Development of cetuximab (anti-EGFR mAb) targeted photoactivable multi- inhibitor liposomes (TPMILs) co-loaded with lapidated benzoporphyrin derivative (BPD-PC) photosensitizer and irinotecan to remediate desmoplasia, a major contributor to chemoresistance	(BPD-PC) + irinotecan	MIA PaCa-2 + PCAF tumor model	Reduction in stromal collagen density and collagen fiber alignment	Synchronized chemotherapeutic and a photodynamic insult to PDAC tissue was achieved with doubled overall survival
HSA NPs(Guo et al., 2018)	Enhancing the antitumor effect of GEM by the encapsulation into HSA- NPs to overcome GEM resistance in GEM-resistant PC induced by low hENT1 gene expression	GEM	BxPC-3 and SW1990 cells/ patient-derived xenograft BALB/c-nu/nu mice model	Inhibit cell proliferation, arrest cell cycle, and trigger apoptosis	GEM-loaded HSA-NPs was able to overcome GEM-resistance induced by low hENT1 expression <i>in vitro</i> and <i>in vivo</i>
HSA NPs(Han et al., 2017)	Development of tumor microenvironment targeting HSA-GEM/ IR780 complex with the redox-responsive release of GEM using GFLG cleavable peptide	GEM + IR780 (NIR dye)	BxPC-3 cells	Induction of apoptosis and Inhibition of cells proliferation	The developed theranostic nanoplatform showed high tissue accumulation and retention with: (i) targeted intracellular drug release, (ii) enhanced tumor inhibition activity (iii) insignificant side effects
Pheophorbide-a conjugated albumin NPs(Yu et al., 2017)	Inhibit PC with lymphatic metastases by the combination of chemotherapy with photodynamic therapy (PDT)	GEM	(BxPC-3-LN7) cells	Increase in drug accumulation in primary tumors as well as metastatic lymph nodes	Developed triple functional system efficiently controlled the release of GEM from the modified NPs and possessed imaging-guided theranostic properties
Nanovector- albumin- bound PTX (MSV/nAb-	Enhancing drug transport by increasing caveolin-1	PTX + GEM		Increase cellular uptake as a result of GEM-induced	GEM enhanced the transport of MSV/nAb-

(Continued on following page)

TABLE 2 (Continued) Selected examples of nano-based formulations used to overcome MDR in tumor cells.

Type of nanocarrier	Objective	Cargo	Cell/Animal model	Effects on MDR	Outcomes/results
PTX) (Borsoi et al., 2017)	expression (albumin transporter) via combination therapy of MSV/nAb-PTX with GEM		L3.6 pl human cells/ L3.6 pl—bearing nu/nu nude mice	high cav-1 expression, which leads to increased transport of nAb-PTX into tumor tissue	PTX in GEM-resistant pancreatic ductal adenocarcinoma
Chitosan coated solid- lipid NPs (c-SLN) (Thakkar et al., 2018)	To use nano-encapsulated c-SLNs combinations to determine the efficacy of the ACS therapeutic regimen	Aspirin (ASP)+ curcumin (CUR)+free sulforaphane (SFN); ACS	Panc-1 and MIA PaCa-2 cells/LSL-Kras ^{G12D/+} ; Pdx- 1 ^{Cre/+} transgenic mouse model	Increase in drug efficacy	Due to enhanced bioavailability of the combined ACS chemopreventive agents, the dosage for this therapeutic regimen can substantially be reduced, which by virtue reduces any potential serious side effects
SN38 (irinotecan active metabolite) polymeric prodrug-based NPs(Wang et al., 2017)	Development of a nano- based system for effective synergistic therapy to overcome fibroblast- induced drug resistance	GDC-0449 (hedgehog pathway inhibitor)	BxPC-3 cells and MIA PaCa-2 cells/PSCs and BxPC-3—bearing BALB/c nude mice	Increase in drug efficacy by modulating the fibroblast-enriched tumor microenvironment	size-tunable nanoparticles were obtained and controllable loading efficiency, which was directly correlated to the length of the hydrophobic SN38 block
(PLGA-ORM NPs) (Khan et al., 2015)	Providing effective endosomal release to the cytosol	Ormeloxifene	(HPAF-II, AsPC-1, BxPC- 3, Panc-1, and MiaPaca)/a BxPC-3 xenograft mice model	Increase in drug efficacy	PLGA-ORM NPs showed substantial antitumor efficacy and effective endosomal release resulted in PC tumor suppression
PLGA-PEG NPs(Elgogary et al., 2016)	Targeting the glutamine metabolism	BPTES	P8, A6L, A32, P198, E3, P215, P10, and JD13D human PC cells/Foxn1nu athymic tumor-bearing nude mice	Increase in drug accumulation	Combination therapy of BPTES-loaded NPs and metformin were shown to be effective in blocking the metabolism of glutamine and glucose
Redox-responsive Apt/ CPP-CPTD NPs(He et al., 2018)	Development of sequentially responsive NPs with redox-responsive on-demand drug release and ECM-responsive tumor penetration	Camptothecin prodrug, CPTD	MIA PaCa-2 cells/MIA PaCa-2 orthotopic human PC xenograft bearing nude mice	Enhance cytotoxicity and cellular accumulation	Formulated NPs showed selective accumulation at the tumor site with mild <i>in vitro</i> cytotoxicity and good <i>in vivo</i> antitumor efficacy
PLGA NPs(Lucero-Acuna et al., 2014)	Enhanced PH-427 delivery to the PC harboring K-ras mutation to overcome the protective stromal layer surrounding the pancreatic tumor	PH-427 (AKT/ PDK1 inhibitor)	MiaPaCa-2 harboring K-ras mutation/Orthotopic MiaPaCa-2—bearing mice	Increase in cellular uptake and drug efficacy	PH-427- loaded PLGA NPs resulted in the enhanced therapeutic effect of PH-427 <i>in vitro</i> and <i>in vivo</i>
PEGylated colloidal gold NPs(Libutti et al., 2018)	Targeting components of the tumor microenvironment responsible for creating high interstitial fluid pressure to improve the delivery of anticancer drugs	TNF and a PTX prodrug	Genetically engineered mice with pancreatic ductal adenocarcinoma	Increase in drug efficacy by tumor IFP reduction	The combination of TNF (targeting tumor vasculature) with PTX (either loaded on the NPs or administered separately) increased the efficacy of the cytotoxic agent
Superparamagnetic iron oxide nanoparticle (SPION) (Khan et al., 2019)	Development of (SPION) loaded with curcumin (SP- CUR), which is known for its anti-inflammatory and antitumor activity, to overcome GEM resistance and enhance its therapeutic potential <i>in vitro</i> and <i>in</i> <i>vivo</i>	Curcumin + GEM	Panc-1, HPAF, CPSC, and HPSC cells / HPAF-II human PSCs—bearing athymic Nu/Nu mice	Suppression of sonic hedgehog (SHH) signaling pathway and oncogenic CXCR4/CXCL12 signaling axis	Efficient delivery of curcumin was achieved, which also played a role in sensitizing cells to standard GEM therapy
		Cisplatin			

(Continued on following page)

Type of nanocarrier	Objective	Cargo	Cell/Animal model	Effects on MDR	Outcomes/results
Nanogels (NGs) (Soni et al., 2019)	Development of Cisplatin- loaded mAb-coated NGs for targeted delivery to PCs and the evaluation of antitumor activity in combination with GEM		T3M4/Luc cells/ T3M4/ Luc—bearing Nu-Nu nude mice	Increase in drug efficacy by targeted therapy using an anti-STn antibody (TKH2 mAb)	Enhanced drug delivery, as well as synergistic cytotoxic effect, was observed after sequential exposure of PC cells to GEM followed by CDDP
Fucose-bound liposomes (Yoshida et al., 2012)	Development of liposomal formulation functionalized with L-fucose to target the fucosylated antigens highly expressed on the surface of cancer cells to enhance cisplatin delivery	Cisplatin	BxPC-3, AsPC-1, PK59, and HuCCT1 cell lines/ Subcutaneous model:AsPC- 1-bearing mice; Liver metastasis and orthotopic models: BxPC-3-Luc- bearing mice	Increase in cellular uptake and cytotoxicity	Cisplatin-loaded Fucose- bound liposomes were effectively delivered to PC cells and resulted in effective inhibition of tumor growth as well as extending survival in the mouse xenograft models
Au-GO@ZC-DOX stealth nanovesicles (Thapa et al., 2018)	development of pH- triggered stealth nanovesicles for chemophototherapy	DOX	Panc-1 cells and Mia PaCa- 2 cells/PANC-1- bearing BALB/c nude mice	Increase in cellular uptake and cytotoxicity	The multi-componential nanovesicle showed effective Macrophage opsonization inhibition, resulting in anti-cancer and anti-migration effects
HA-SMA Micelles (Kesharwani et al., 2015)	Development of functionalized micelles with HA to target the PC overexpressed CD44 receptors to overcome MDR	3, 4- difluorobenzylidene curcumin (CDF)	MiaPaCa-2 and AsPC-1 cells	Inhibition of NF-κB in CD44 ⁺ cells	The developed nanosystem showed remarkable colloidal stability and sustained drug release and potent anticancer activity
Polymeric Micelles (Xu et al., 2015)	Development (TPGS-GEM) prodrug micelles to protect the drug from enzymatic metabolism	TPGS-GEM (prodrug)	BxPC-3 cells	Enhanced drug efficacy as the micellar formulation protected the drug from enzymatic metabolism	Long circulation half-life of GEM was obtained in addition to enhanced anticancer activity
Ultra-pH-sensitive micelles (UPSM) (Kong et al., 2019)	Development of UPSM improved pH buffer capacity for simultaneous inhibition of lysosomal acidification and enhancement of therapeutic delivery	Triptolide prodrug-	KRAS mutant PANC-1 and MIA PaCa-2/MIA PaCa-2- luc—bearing BALB/C nude mice	Disruption of lysosomal catabolism and growth inhibition of KRAS mutant	The newly developed nanosystem revealed more efficient lysosomal catabolism when compared with conventional lysosomotropic agents. In addition, pH-sensitive UPSM showed significant cytotoxicity when compared to non-pH- sensitive micelles

TABLE 2 (Continued) Selected examples of nano-based formulations used to overcome MDR in tumor cells.

2013; Goldstein et al., 2015). Based on the findings of the trial, GEM plus nab-PTX became the first therapy recommended by the National Institute for Health and Care Excellence in the United Kingdom for the management of previously untreated metastatic PC (Von Hoff et al., 2011).

Metal-based nanoparticles in PC

Metal-based NPs such as gold, silver, iron, platinum, and titanium are commonly used for imaging, as drug delivery carriers, and as radiosensitizers in radiation, proton, or photodynamic therapy (Klebowski et al., 2018). Due to their inherent physicochemical properties, metal NPs could overcome MDR via different mechanisms (Sharma et al., 2018), as shown in Figure 2. The endosomal-based cellular uptake mechanism can be considered one of the main advantages of metal-based NPs in overcoming MDR. This is achieved by selectively releasing intracellular drugs after evading the membrane-embedded multidrug efflux pumps (Ayers and Nasti, 2012). Khan *et al.* developed a nanoformulation of superparamagnetic iron oxide nanoparticles (SPION) loaded with curcumin (SP-CUR), an antiinflammatory and anti-tumorigenic compound. They tested the activity of this nanoformulation in combination with GEM on GEM-resistant pancreatic cells. Efficient delivery of curcumin was achieved using their formulation, and they found that SP-

TABLE 3 Selected examples of clinical trials applying nano-based formulations in PC treatment.

Study	Testing	Study type	Dosing regimen	Median survival	Main outcomes
Von Hoff et al., 2011 (Von Hoff et al., 2011)	GEM + Nab-PTX	Phase I/II	67 patients 100, 125, and 150 mg/m ² nab-paclitaxel plus 1,000 mg/m ² gemcitabine on days 1, 8, and 15 every 28 days	12.2 months	Response rate 48%, Overall Survival (OS) 12.2, and 1-year survival rate 48%
NCT00844649					
Hosein et al., 2013 (Hosein et al., 2013)	nab-PTX	Phase II	19 patients were treated with nab- paclitaxel 100 mg/m ² on days 1, 8, and 15 of a 28-day cycle	7.3 months	6 months OS 58%, median OS 7.3 months
Von Hoff et al., 2013 (Von Hoff et al., 2013)	GEM + Nab- PTX vs. GEM	Phase III	Group 1: 431 patients given nab-PTX (125 mg/m ²) followed by GEM (1,000 mg/m ²) on days 1, 8, and 15 every 4 weeks	Median survival was 8.5 months in GEM + Nab- PTX vs. 6.7 months in the GEM group	Group 1 vs. Group 2: Median survival 8.5 vs. 6.7 months; 1-year survival rate 35vs. 22%; 2-year survival rate 9 vs. 4%; higher risks of peripheral neuropathy and myelosuppression with group 1 compared to group 2
NCT00844649			Group 2: 430 patients given GEM monotherapy (1,000 mg/m ²) weekly for 7 of 8 weeks (cycle 1) and then on days 1, 8, and 15 every 4 weeks (cycle 2)		
Goldstein et al., 2015 (Goldstein et al., 2015)	GEM + Nab- PTX vs. GEM	Phase III	Patients (n = 861) randomly assigned to receive GEM + Nab-PTX vs. GEM	Median survival was 8.7 months in GEM + Nab- PTX vs. 6.6 months in the GEM group	OS and long-term survival (>3 years) were higher amongst GEM + Nab-PTX compared to the GEM monotherapy group
Update on OS of NCT00844649					
Vogel et al., 2016 (Vogel et al., 2016)	GEM + Nab- PTX vs. GEM	Phase III	Patients randomly assigned to receive GEM + Nab-PTX vs. GEM alone	Median survival was 9.8 months in GEM + Nab- PTX vs. 7.5 months in the GEM group	OS 8% with GEM + Nab-PTX vs. 4% from GEM alone; Overall Response Rate 27 vs. 9% with GEM + Nab-PTX vs. GEM alone respectively
Sub-analysis of NCT00844649					
Macarulla et al., 2019 (Macarulla et al., 2019)	GEM + Nab- PTX vs. GEM	Phase I/II	6 groups inducted in phase I and 2 groups in phase II both using GEM + Nab-PTX at different doses (100 g/ m ² or 125 mg/m ² Nab-PTX + 1,000 mg/m ² GEM)	NA	Improvement in overall survival irrespective of the dose of Nab-PTX used
NCT02382263					
Libutti et al., 2010 (Libutti et al., 2010)	CYT-6091 (colloidal gold)	Phase I	3 participants were given 50 mg/m ² to 600 mg/m ² of rhTNF via the CYT-6091 delivery system	NA	CYT-6091 delivery system led to great tumor tissue concentration of rhTNF compared to normal tissues
NCT00356980					
Stathopoulos et al., 2005 (Stathopoulos et al., 2005)	Lipoplatin	Phase I	Dose starting at 25 mg/m 2 and was increased by 25–125 mg/m 2	NA	No significant nephrotoxicity or systemic toxicity noted with this preparation
Greek trial					
Stathopoulos et al., 2005 (Stathopoulos et al., 2005)	Lipoplatin	Phase II	GEM dose 1,000 mg/m ² and the lipoplatin dose was escalated from 25 mg/m^2 to 125 mg/m^2	3 months	Partial response (>50% tumor reduction) was seen in 2 patients. Stable disease (<25-50% reduction in the tumor) was seen in 14 patients
Greek trial					
Syrigos et al., 2002 (Syrigos et al., 2002)	Docetaxel and liposomal doxorubicin	Phase II	21 patients given docetaxel (80 mg/ m²), and liposomal doxorubicin (30 mg/m²) was administered on day 1, every 3 weeks	10 months	Median survival 10, 1-year survival 33.3%
Greek trial					
Hamaguchi et al., 2007 (Hamaguchi et al., 2007)	NK105 (PTX- polymeric micelles)	Phase I	Initially given 10 mg/m ² and successively increased the dose	NA	The size of liver mets reduced by 90% in patients receiving a dose of 150 $ \rm mg/m^2$ or higher dose



CUR increased the effectiveness of GEM therapy through suppressing two signal transduction pathways that are implemented in MDR: 1) sonic hedgehog (SHH) and 2) oncogenic CXCR4/CXCL12. Following their endosomal escape, SPION particles showed increased cellular internalization where they were observed to be more closely associated with cytosol/mitochondria to prevent lysosomal degradation (Khan et al., 2019). In 2010, Libutti et al. used a novel drug delivery system, CYT-6091, surface-modified colloidal gold nanoparticles, to increase tumor levels of rhTNF and reduce its systemic metabolism and toxicity (Libutti et al., 2010). CYT-6091 is constructed by combining rhTNF and thiolyated glycol to the surface of 27 nm gold colloidal particles. CYT9061 was studied at doses between 50 mg/m2 to 600 mg/m2 in a phase I clinical trial conducted with three patients with pancreatic adenocarcinoma. Using the gold colloidal nanoparticles, researchers were able to administer extremely high concentrations of rhTNF compared to when given in isolation: the highest isolated tolerated concentration of pure rhTNF is 1 mg per cycle. After treatment, examination by electron microscope of normal and tumor tissues was carried out and found gold particles isolated within tumor tissues or at anticipated clearance sites (Ladd et al., 2017). These findings suggest that gold-based, colloidal nano-delivery systems can be used to deliver chemotherapy drugs to target tissues. However, subsequent clinical trials remain to be conducted to prove their efficacy and safety.

Polymeric micelles in PC

Polymeric micelles, including both hydrophobic and hydrophilic moieties (Manzur et al., 2017), are the largest class of nanomaterials being investigated for PC treatment in preclinical settings. These nanoparticles contain linear or highly branched and symmetrical polymers, with hydrophilic dendritic extensions and a hydrophobic core. The terminal groups at the dendritic extensions can be adjusted to allow for better solubility depending upon the carrier medium and allow for superior anchorage and permeability into target tissues (Chiba and Twyman, 2017; Ladd et al., 2017). Polymeric micelles are NPs formed by the self-assembly of amphiphilic block copolymers when present in certain solvents with a surfactant concentration above a critical micelle concentration (c.m.c). They are efficient drug delivery systems for cancer treatment with the ability to inhibit P-gp action, alter drug internalization, enable selective drug targeting, and enable subcellular localization. Polymeric micelles can circumvent MDR through a combination of mechanisms, including the EPR effect, endosomal-triggered active internalization, and drug escape (Kapse-Mistry et al., 2014). In our literature review, we found at least seven published studies using polymeric micelles for the delivery of anti-tumor drugs in clinical trials, including Genexol[®]-PM, NK105, NC-4016, NK012, NC-6004, NK911, and SP1049C (Gong et al., 2012). NK105 is a nanoparticle formulation that incorporates PTX into a "core-shelltype" polymeric micelle. In a phase I trial conducted by Hamaguchi et al. (2007), the pharmacokinetics of NK105, a micelle carrier for PTX, was studied. The study included 19 patients with advanced PC who were given an initial dose of 10 mg/m² of NK105; this dose increased successively. The size of metastatic liver tumors was reduced by 90% in patients given a 150 mg/m² dose of the drug (Hamaguchi et al., 2007).

Lipid-based nanoformulations in PC

Lipid-based nanoformulations have been extensively studied for the delivery of antitumor agents, including natural products (Kashyap et al., 2019). Liposomes are lipid bilayer systems that can cross the lipid bilayer to selectively target cancer cells based on biomarkers, overly expressed on PC cells (Urey et al., 2017). There are several strategies whereby liposomes can enhance drug bioavailability and efficacy in drug-resistant cancer. These include 1) liposomes modified for controlled and on-demand release; and 2) ligand-targeted liposomes such as immunoliposomes, which facilitate intracellular drug delivery into tumor cells. Liposomes can also directly inhibit P-gp through endocytosis and consequently enhance intracellular drug accumulation (Kapse-Mistry et al., 2014). In 2012, the use of liposomes in PC treatment was explored by Yoshida et al. They targeted fucosylated antigens that are highly expressed on the surface of PC cells. They engineered L-fucose-bound liposomes that encapsulated Cy5.5 or cisplatin. In vitro studies on CA19-9 expressing PC cells showed that L-fucose-bound liposomes encapsulating either Cy5.5 or cisplatin were effectively delivered and in mouse xenograft models, cisplatin-loaded liposomes were successfully delivered to PC cells and inhibited tumor growth (Yoshida et al., 2012). Furthermore, in the second-line setting of metastatic PDAC following administration of GEM-based regimens, nanoliposomes irinotecan (nal-IRI) was approved by the FDA in combination with 5-FU and leucovorin (5-FU/LV) in 2015. In phase III, the NAPOLI-1 trial, the median progressionfree survival (PFS) was 3.1 and 1.5 months (p < 0.001) in patients who received nal-IRI + 5-FU/LV and patient's 5-FU/LV alone, respectively (Wang-Gillam et al., 2016). In the final OS analysis of the NAPOLI-1 trial, the median OS was increased by 2 months (p =0.042) in the nal-IRI + 5-FU/LV group (Wang-Gillam et al., 2016).

Nanogels in PC

Nanogels can utilize the unique characteristics of tumor microenvironments such as pH and temperature, to release

drugs within the cell, resulting in efficient drug delivery (Damaghi et al., 2013). In a study by Damaghi et al., a nanogel-based platform for PC therapy was reported (Soni et al., 2019). They developed a cisplatin-loaded, mAb-coated nanogel for targeted delivery and used it in combination with GEM. In vitro results revealed an increase in drug efficacy. Additionally, enhanced drug delivery and synergistic cytotoxic effect were observed after sequential exposure of PC cells to GEM. Together, these studies have all demonstrated the advantage and improved therapeutic outcomes with the use of nanomaterials and nano-drug platforms, particularly and most importantly against MDR in experimental models and clinical trials (Kesharwani et al., 2015; Borsoi et al., 2017; Guo et al., 2018; Kong et al., 2019). Such advantage to the use of nanomaterials is mediated through a wide range of mechanisms, including enhanced cellular uptake, evading endosomal-lysosomal drug breakdown, inhibition of drug efflux, and increasing plasma halflife.

Key limitations on the clinical translation of nanomedicine in PC

There has been a tremendous effort to understand the structural and functional properties of nanoparticles directed against cancer but their translation to clinical practice has been largely limited. This can primarily be attributed to a poor understanding of the biological barriers and nanomaterial behavior inside the body and cells, as well as the overemphasis and relying on animal models during preclinical evaluation, which does not necessarily represent the same disease phenotype in humans (Gonzalez-Valdivieso et al., 2021).

There are several major challenges in the treatment of PC, which need to be overcome to make the use of nanotherapies a success against PCs. These include off-target toxicity, low bioavailability of chemotherapeutic drugs, and undesirable pharmacokinetics. One way to address these obstacles is through the use of nanotechnology as an effective vehicle for chemotherapeutic drugs. Currently, only 16 nano-based cancer drugs are approved by FDA and around 75 nanoformulations are being investigated in clinical trials (He et al., 2019). It is extremely important to narrow the gap between preclinical toxicity studies and toxicity studies in patients, as nanomedicines have been shown to exert additional unintended and often toxic effects on normal cellular function. Moreover, there has been a lack of convincing data on the process of excretion of nanomedicines from the human system, as most data is available from animal disease models. Nanomedicines can pose safety issues at different levels (apart from the intrinsic toxicity of the API itself). Furthermore, the biodistribution of nanoparticles changes unpredictably resulting in uptake and accumulation in certain organs, which may result in target off-target effects and local

overexposure. Indeed, some nanoparticles have a known tendency to accumulate in lymphoid organs and kidneys (for some polymer-bound drugs) (Metselaar and Lammers, 2020).

Furthermore, there remains a general lack of understanding on the cost-effectiveness, manufacturing, and scaling up, as well as regulation with regard to using nanomedicines for cancer (van der Meel et al., 2017). As the science behind the structuralfunctional relationship provides clarity on the interaction of nanomedicines *in vivo*, the regulatory challenges must be addressed simultaneously to bring these potentially gamechanging therapeutics to the frontline against fighting pancreatic cancer.

Conclusion and future directions

Numerous noncarriers have been developed and investigated for the treatment of PC to overcome the problem of MDR seen with chemotherapy and other therapeutic options, however, with limited success Even though nano-based carriers show great promise in treating various cancers, they have several limitations, including potential toxicity, difficult scalability, and low loading efficiency that could be responsible for their low success rate reaching clinical settings. Notably, the albumin-based nanocarrier was the most successful in clinical studies for PC. This is mainly because albumin nanoparticles were successful in encapsulating widely used chemotherapeutics that are less soluble/insoluble in water. albumin is highly biocompatibility Additionally, and biodegradability, making it an attractive material for drug delivery applications. The development of future therapies for cancer and nano-based therapeutics should not be limited by designing nanocarriers only for passive targeting of cancerous cells. Internalization of chemotherapeutic agents into tumor cells can be further improved via the utilization of an active targeting approach to enhance drug delivery efficacy. Additionally, research into novel biomarkers to enable active targeting will empower delivery strategies of nanocarriers to combat cancer resistance.

While we have certainly made huge progress in understanding the drug resistance mechanisms in PC and the signaling pathways responsible for PC cell metastases, investigations on the use of nanomedicine in this field lag behind. Currently, the majority of work in the field of nanomedicine is largely focused on increasing drug stability, accumulation, and targeting, which is yet of critical importance, particularly in nucleoside transporter (e.g., ENT1 and CNT1)-mediated drug resistance against GEM (Hung et al., 2015; Poon et al., 2015). Future studies utilizing nanotechnology against MDR pancreatic cancers should integrate multiple modalities and exploit the rapidly accumulating mechanistic knowledge in this cancer model (e.g., targeting PCSCs, dual delivery of potential drug modalities, etc.). In addition, utilizing the endogenous properties of the TME to trigger the release of cancer therapeutics from nanocarriers should be considered during the delivery system design. Such a design will add another dimension of controlled

release that can impact clinical efficacy, where adverse effects can be minimized while retaining therapeutic benefits.

The main goal of PC treatment is to enhance the efficiency of drug delivery and minimize drug resistance. Despite the tremendous effort in making novel nanocarriers in pre-clinical settings, the development of clinical translation to the bedside remains laborious. Extrapolation of scientific findings from animals to humans is extremely challenging, mainly due to differences in physiology and anatomy between species, making direct extrapolation unreliable. Furthermore, unlike the experimental settings in clinical studies, animals are designed with syngeneic backgrounds, and disease models are designed to produce as homogenous a population as possible. On the other hand, heterogeneity is the basis of ineffectiveness in clinical trials. Moreover, individual variability in lifestyle and disease progression plays key roles in the overall efficacy, unlike the well-controlled animal experiments.

The promise of nanomedicine will be realized by moving away from designing a targeting strategy against a single target to including targeting approaches that address multiple signaling mechanisms and molecular targets, considering the complexity of both the human physiology and the tumor microenvironment, including the development of MDR mechanisms.

Author contributions

Conception of the work, draft, and revision of the work: AKA, NBA, NA, RA, and AA.

Acknowledgments

The authors are thankful to the Research Center, College of Pharmacy, and the Deanship of Scientific Research, King Saud University for their support to conduct this work.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors, and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Aguirre, A. J., and Collisson, E. A. (2017). Advances in the genetics and biology of pancreatic cancer. *Cancer J.* 23 (6), 315–320. doi:10.1097/PPO. 00000000000286

Alshememry, A. K., El-Tokhy, S. S., and Unsworth, L. D. (2017). Using properties of tumor microenvironments for controlling local, on-demand delivery from biopolymer-based nanocarriers. *Curr. Pharm. Des.* 23 (35), 5358–5391. doi:10. 2174/1381612823666170522100545

Alzhrani, R., Alsaab, H. O., Vanamala, K., Bhise, K., Tatiparti, K., Barari, A., et al. (2021). Overcoming the tumor microenvironmental barriers of pancreatic ductal adenocarcinomas for achieving better treatment outcomes. *Adv. Ther.* 4 (6), 2000262. doi:10.1002/adtp.202000262

Arumugam, T., Ramachandran, V., Fournier, K. F., Wang, H., Marquis, L., Abbruzzese, J. L., et al. (2009). Epithelial to mesenchymal transition contributes to drug resistance in pancreatic cancer. *Cancer Res.* 69 (14), 5820–5828. doi:10.1158/0008-5472.CAN-08-2819

Avula, L. R., Hagerty, B., and Alewine, C. (2020). Molecular mediators of peritoneal metastasis in pancreatic cancer. *Cancer Metastasis Rev.* 39 (4), 1223–1243. doi:10.1007/s10555-020-09924-4

Ayers, D., and Nasti, A. (2012). Utilisation of nanoparticle technology in cancer chemoresistance. J. Drug Deliv. 2012, 265691. doi:10.1155/2012/265691

Bailey, P., Chang, D. K., Nones, K., Johns, A. L., Patch, A. M., Gingras, M. C., et al. (2016). Genomic analyses identify molecular subtypes of pancreatic cancer. *Nature* 531 (7592), 47–52. doi:10.1038/nature16965

Bardeesy, N., and DePinho, R. A. (2002). Pancreatic cancer biology and genetics. *Nat. Rev. Cancer* 2 (12), 897–909. doi:10.1038/nrc949

Baretti, M., Pulluri, B., Tsai, H. L., Blackford, A. L., Wolfgang, C. L., Laheru, D., et al. (2019). The significance of ascites in patients with pancreatic ductal adenocarcinoma: A case-control study. *Pancreas* 48 (4), 585–589. doi:10.1097/mpa.00000000001262

Batlle, E., and Clevers, H. (2017). Cancer stem cells revisited. Nat. Med. 23 (10), 1124-1134. doi:10.1038/nm.4409

Berlin, J. D., Catalano, P., Thomas, J. P., Kugler, J. W., Haller, D. G., and Benson, A. B., 3rd (2002). Phase III study of gemcitabine in combination with fluorouracil versus gemcitabine alone in patients with advanced pancreatic carcinoma: Eastern Cooperative Oncology Group Trial E2297. J. Clin. Oncol. 20 (15), 3270–3275. doi:10.1200/JCO.2002.11.149

Binenbaum, Y., Na'ara, S., and Gil, Z. (2015). Gemcitabine resistance in pancreatic ductal adenocarcinoma. *Drug resist. updat.* 23, 55–68. doi:10.1016/j. drup.2015.10.002

Blanco, E., Hsiao, A., Mann, A. P., Landry, M. G., Meric-Bernstam, F., and Ferrari, M. (2011). Nanomedicine in cancer therapy: Innovative trends and prospects. *Cancer Sci.* 102 (7), 1247–1252. doi:10.1111/j.1349-7006.2011.01941.x

Borsoi, C., Leonard, F., Lee, Y., Zaid, M., Elganainy, D., Alexander, J. F., et al. (2017). Gemcitabine enhances the transport of nanovector-albumin-bound paclitaxel in gemcitabine-resistant pancreatic ductal adenocarcinoma. *Cancer Lett.* 403, 296–304. doi:10.1016/j.canlet.2017.06.026

Bray, F., Ferlay, J., Soerjomataram, I., Siegel, R. L., Torre, L. A., and Jemal, A. (2018). Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *Ca. Cancer J. Clin.* 68 (6), 394–424. doi:10.3322/caac.21492

Burris, H. A., 3rd, Moore, M. J., Andersen, J., Green, M. R., Rothenberg, M. L., Modiano, M. R., et al. (1997). Improvements in survival and clinical benefit with gemcitabine as first-line therapy for patients with advanced pancreas cancer: A randomized trial. *J. Clin. Oncol.* 15 (6), 2403–2413. doi:10.1200/JCO.1997.15.6.2403

Cancer.Net (2022). Pancreatic Cancer: Statistics [Online]. Available: https://www.cancer.net/cancer-types/pancreatic-cancer/statistics (Accessed July 15, 2022).

Cascinu, S., Verdecchia, L., Valeri, N., Berardi, R., and Scartozzi, M. (2006). New target therapies in advanced pancreatic cancer. *Ann. Oncol.* 17 (5), v148–152. doi:10.1093/annonc/mdj971

Chen, H., Guo, Q., Chu, Y., Li, C., Zhang, Y., Liu, P., et al. (2022). Smart hypoxiaresponsive transformable and charge-reversible nanoparticles for the deep penetration and tumor microenvironment modulation of pancreatic cancer. *Biomaterials* 287, 121599. doi:10.1016/j.biomaterials.2022.121599

Chiba, F., and Twyman, L. J. (2017). Effect of terminal-group functionality on the ability of dendrimers to bind proteins. *Bioconjug. Chem.* 28 (8), 2046–2050. doi:10. 1021/acs.bioconjchem.7b00350

Corbo, V., Tortora, G., and Scarpa, A. (2012). Molecular pathology of pancreatic cancer: From bench-to-bedside translation. *Curr. Drug Targets* 13 (6), 744–752. doi:10.2174/138945012800564103

Cowgill, S. M., and Muscarella, P. (2003). The genetics of pancreatic cancer. Am. J. Surg. 186 (3), 279–286. doi:10.1016/s0002-9610(03)00226-5

Damaghi, M., Wojtkowiak, J. W., and Gillies, R. J. (2013). pH sensing and regulation in cancer. *Front. Physiol.* 4, 370. doi:10.3389/fphys.2013.00370

Danilova, L., Ho, W. J., Zhu, Q., Vithayathil, T., De Jesus-Acosta, A., Azad, N. S., et al. (2019). Programmed cell death ligand-1 (PD-L1) and CD8 expression profiling identify an immunologic subtype of pancreatic ductal adenocarcinomas with favorable survival. *Cancer Immunol. Res.* 7 (6), 886–895. doi:10.1158/2326-6066.CIR-18-0822

Das, S. K., Wang, J. L., Li, B., Zhang, C., and Yang, H. F. (2019). Clinical effectiveness of combined interventional therapy as a salvage modality for unresectable pancreatic carcinoma. *Oncol. Lett.* 18 (1), 375–385. doi:10.3892/ol. 2019.10323

Dauer, P., Nomura, A., Saluja, A., and Banerjee, S. (2017). Microenvironment in determining chemo-resistance in pancreatic cancer: Neighborhood matters. *Pancreatology* 17 (1), 7–12. doi:10.1016/j.pan.2016.12.010

Demidenko, Z. N., Kalurupalle, S., Hanko, C., Lim, C. U., Broude, E., and Blagosklonny, M. V. (2008). Mechanism of G1-like arrest by low concentrations of paclitaxel: Next cell cycle p53-dependent arrest with sub G1 DNA content mediated by prolonged mitosis. *Oncogene* 27 (32), 4402–4410. doi:10.1038/onc. 2008.82

di Magliano, M. P., and Logsdon, C. D. (2013). Roles for KRAS in pancreatic tumor development and progression. *Gastroenterology* 144 (6), 1220–1229. doi:10. 1053/j.gastro.2013.01.071

Dreyer, S. B., Chang, D. K., Bailey, P., and Biankin, A. V. (2017). Pancreatic cancer genomes: Implications for clinical management and therapeutic development. *Clin. Cancer Res.* 23 (7), 1638–1646. doi:10.1158/1078-0432.CCR-16-2411

Du, Z., Qin, R., Wei, C., Wang, M., Shi, C., Tian, R., et al. (2011). Pancreatic cancer cells resistant to chemoradiotherapy rich in "stem-cell-like" tumor cells. *Dig. Dis. Sci.* 56 (3), 741–750. doi:10.1007/s10620-010-1340-0

Ducreux, M., Cuhna, A. S., Caramella, C., Hollebecque, A., Burtin, P., Goere, D., et al. (2015). Cancer of the pancreas: ESMO clinical practice guidelines for diagnosis, treatment and follow-up. *Ann. Oncol.* 26 (5), v56–68. doi:10.1093/ annonc/mdv295

Elgogary, A., Xu, Q., Poore, B., Alt, J., Zimmermann, S. C., Zhao, L., et al. (2016). Combination therapy with BPTES nanoparticles and metformin targets the metabolic heterogeneity of pancreatic cancer. *Proc. Natl. Acad. Sci. U. S. A.* 113 (36), E5328–E5336. doi:10.1073/pnas.1611406113

Ellenrieder, V., Hendler, S. F., Boeck, W., Seufferlein, T., Menke, A., Ruhland, C., et al. (2001). Transforming growth factor beta1 treatment leads to an epithelialmesenchymal transdifferentiation of pancreatic cancer cells requiring extracellular signal-regulated kinase 2 activation. *Cancer Res.* 61 (10), 4222–4228.

Erkan, M., Hausmann, S., Michalski, C. W., Fingerle, A. A., Dobritz, M., Kleeff, J., et al. (2012). The role of stroma in pancreatic cancer: Diagnostic and therapeutic implications. *Nat. Rev. Gastroenterol. Hepatol.* 9 (8), 454–467. doi:10.1038/nrgastro. 2012.115

Farran, B., and Nagaraju, G. P. (2020). Exosomes as therapeutic solutions for pancreatic cancer. *Drug Discov. Today* 25 (12), 2245–2256. doi:10.1016/j.drudis. 2020.09.037

Feig, C., Jones, J. O., Kraman, M., Wells, R. J., Deonarine, A., Chan, D. S., et al. (2013). Targeting CXCL12 from FAP-expressing carcinoma-associated fibroblasts synergizes with anti-PD-L1 immunotherapy in pancreatic cancer. *Proc. Natl. Acad. Sci. U. S. A.* 110 (50), 20212–20217. doi:10.1073/pnas.1320318110

Gad, A., Kydd, J., Piel, B., and Rai, P. (2016). Targeting cancer using polymeric nanoparticle mediated combination chemotherapy. *Int. J. Nanomed. Nanosurg.* 2 (3), 10.16966/2470-3206.116. doi:10.16966/2470-3206.116

Ge, W., Chen, G., and Fan, X. S. (2017). Pathway of peritoneal carcinomatosis maybe hematogenous metastasis rather than peritoneal seeding. *Oncotarget* 8 (25), 41549–41554. doi:10.18632/oncotarget.14607

Ghaneh, P., Costello, E., and Neoptolemos, J. P. (2007). Biology and management of pancreatic cancer. *Postgrad. Med. J.* 56 (8), 478–497. doi:10.1136/gut.2006. 103333

Gheorghe, G., Bungau, S., Ilie, M., Behl, T., Vesa, C. M., Brisc, C., et al. (2020). Early diagnosis of pancreatic cancer: The key for survival. *Diagn. (Basel)* 10 (11), E869. doi:10.3390/diagnostics10110869

Goldstein, D., El-Maraghi, R. H., Hammel, P., Heinemann, V., Kunzmann, V., Sastre, J., et al. (2015). nab-Paclitaxel plus gemcitabine for metastatic pancreatic cancer: long-term survival from a phase III trial. *J. Natl. Cancer Inst.* 107 (2), dju413. doi:10.1093/jnci/dju413 Gong, J., Chen, M., Zheng, Y., Wang, S., and Wang, Y. (2012). Polymeric micelles drug delivery system in oncology. *J. Control. Release* 159 (3), 312–323. doi:10.1016/j. jconrel.2011.12.012

Gonzalez-Valdivieso, J., Girotti, A., Schneider, J., and Arias, F. J. (2021). Advanced nanomedicine and cancer: Challenges and opportunities in clinical translation. *Int. J. Pharm.* 599, 120438. doi:10.1016/j.ijpharm.2021.120438

Goodman, A. M., Kato, S., Bazhenova, L., Patel, S. P., Frampton, G. M., Miller, V., et al. (2017). Tumor mutational burden as an independent predictor of response to immunotherapy in diverse cancers. *Mol. Cancer Ther.* 16 (11), 2598–2608. doi:10. 1158/1535-7163.MCT-17-0386

Govindan, R., Fakih, M., Price, T., Falchook, G., Desai, J., Kuo, J., et al. (2019). OA02. 02 Phase 1 study of safety, tolerability, PK and efficacy of AMG 510, a novel KRASG12C inhibitor, evaluated in NSCLC. *J. Thorac. Oncol.* 14 (10), S208. doi:10. 1016/j.jtho.2019.08.412

Guo, Z., Wang, F., Di, Y., Yao, L., Yu, X., Fu, D., et al. (2018). Antitumor effect of gemcitabine-loaded albumin nanoparticle on gemcitabine-resistant pancreatic cancer induced by low hENT1 expression. *Int. J. Nanomedicine* 13, 4869–4880. doi:10.2147/IJN.S166769

Gurunathan, S., Kang, M. H., Qasim, M., and Kim, J. H. (2018). Nanoparticlemediated combination therapy: Two-in-One approach for cancer. *Int. J. Mol. Sci.* 19 (10), E3264. doi:10.3390/ijms19103264

Hackeng, W. M., Hruban, R. H., Offerhaus, G. J., and Brosens, L. A. (2016). Surgical and molecular pathology of pancreatic neoplasms. *Diagn. Pathol.* 11 (1), 47. doi:10.1186/s13000-016-0497-z

Hamaguchi, T., Kato, K., Yasui, H., Morizane, C., Ikeda, M., Ueno, H., et al. (2007). A phase I and pharmacokinetic study of NK105, a paclitaxel-incorporating micellar nanoparticle formulation. *Br. J. Cancer* 97 (2), 170–176. doi:10.1038/sj.bjc. 6603855

Han, H., Wang, J., Chen, T., Yin, L., Jin, Q., and Ji, J. (2017). Enzyme-sensitive gemcitabine conjugated albumin nanoparticles as a versatile theranostic nanoplatform for pancreatic cancer treatment. *J. Colloid Interface Sci.* 507, 217–224. doi:10.1016/j.jcis.2017.07.047

He, H., Liu, L., Morin, E. E., Liu, M., and Schwendeman, A. (2019). Survey of clinical translation of cancer nanomedicines-lessons learned from successes and failures. *Acc. Chem. Res.* 52 (9), 2445–2461. doi:10.1021/acs.accounts.9b00228

He, X., Chen, X., Liu, L., Zhang, Y., Lu, Y., Zhang, Y., et al. (2018). Sequentially triggered nanoparticles with tumor penetration and intelligent drug release for pancreatic cancer therapy. *Adv. Sci.* 5 (5), 1701070. doi:10.1002/advs.201701070

Heidemann, S. R., and Kirschner, M. W. (1978). Induced formation of asters and cleavage furrows in oocytes of *Xenopus laevis* during *in vitro* maturation. *J. Exp. Zool.* 204 (3), 431-444. doi:10.1002/jez.1402040314

Heinemann, V., Quietzsch, D., Gieseler, F., Gonnermann, M., Schonekas, H., Rost, A., et al. (2006). Randomized phase III trial of gemcitabine plus cisplatin compared with gemcitabine alone in advanced pancreatic cancer. *J. Clin. Oncol.* 24 (24), 3946–3952. doi:10.1200/JCO.2005.05.1490

Hermann, P. C., Huber, S. L., Herrler, T., Aicher, A., Ellwart, J. W., Guba, M., et al. (2007). Distinct populations of cancer stem cells determine tumor growth and metastatic activity in human pancreatic cancer. *Cell Stem Cell* 1 (3), 313–323. doi:10. 1016/j.stem.2007.06.002

Hiss, D. C., Gabriels, G. A., and Folb, P. I. (2007). Combination of tunicamycin with anticancer drugs synergistically enhances their toxicity in multidrug-resistant human ovarian cystadenocarcinoma cells. *Cancer Cell Int.* 7, 5. doi:10.1186/1475-2867-7-5

Ho, W. J., Jaffee, E. M., and Zheng, L. (2020). The tumour microenvironment in pancreatic cancer - clinical challenges and opportunities. *Nat. Rev. Clin. Oncol.* 17 (9), 527–540. doi:10.1038/s41571-020-0363-5

Hosein, P. J., de Lima Lopes, G., Jr., Pastorini, V. H., Gomez, C., Macintyre, J., Zayas, G., et al. (2013). A phase II trial of nab-Paclitaxel as second-line therapy in patients with advanced pancreatic cancer. *Am. J. Clin. Oncol.* 36 (2), 151–156. doi:10.1097/COC.0b013e3182436e8c

Hosoki, T. (1983). Dynamic CT of pancreatic tumors. *AJR. Am. J. Roentgenol.* 140 (5), 959–965. doi:10.2214/ajr.140.5.959

Hu, X., Xia, F., Lee, J., Li, F., Lu, X., Zhuo, X., et al. (2021). Tailor-made nanomaterials for diagnosis and therapy of pancreatic ductal adenocarcinoma. *Adv. Sci.* 8 (7), 2002545. doi:10.1002/advs.202002545

Hung, S. W., Marrache, S., Cummins, S., Bhutia, Y. D., Mody, H., Hooks, S. B., et al. (2015). Defective hCNT1 transport contributes to gemcitabine chemoresistance in ovarian cancer subtypes: Overcoming transport defects using a nanoparticle approach. *Cancer Lett.* 359 (2), 233–240. doi:10.1016/j.canlet.2015. 01.017

Jiang, B., Zhou, L., Lu, J., Wang, Y., Liu, C., You, L., et al. (2020). Stroma-targeting therapy in pancreatic cancer: One coin with two sides? *Front. Oncol.* 10, 576399. doi:10.3389/fonc.2020.576399

Kapse-Mistry, S., Govender, T., Srivastava, R., and Yergeri, M. (2014). Nanodrug delivery in reversing multidrug resistance in cancer cells. *Front. Pharmacol.* 5, 159. doi:10.3389/fphar.2014.00159

Kashyap, D., Tuli, H. S., Yerer, M. B., Sharma, A., Sak, K., Srivastava, S., et al. (2019). Natural product-based nanoformulations for cancer therapy: Opportunities and challenges. *Semin. Cancer Biol.* 69, 5–23. doi:10.1016/j.semcancer.2019.08.014

Kesharwani, P., Banerjee, S., Padhye, S., Sarkar, F. H., and Iyer, A. K. (2015). Hyaluronic acid engineered nanomicelles loaded with 3, 4-difluorobenzylidene curcumin for targeted killing of CD44+ stem-like pancreatic cancer cells. *Biomacromolecules* 16 (9), 3042–3053. doi:10.1021/acs.biomac.5b00941

Khan, S., Chauhan, N., Yallapu, M. M., Ebeling, M. C., Balakrishna, S., Ellis, R. T., et al. (2015). Nanoparticle formulation of ormeloxifene for pancreatic cancer. *Biomaterials* 53, 731–743. doi:10.1016/j.biomaterials.2015.02.082

Khan, S., Setua, S., Kumari, S., Dan, N., Massey, A., Hafeez, B. B., et al. (2019). Superparamagnetic iron oxide nanoparticles of curcumin enhance gemcitabine therapeutic response in pancreatic cancer. *Biomaterials* 208, 83–97. doi:10.1016/j. biomaterials.2019.04.005

Klebowski, B., Depciuch, J., Parlinska-Wojtan, M., and Baran, J. (2018). Applications of noble metal-based nanoparticles in medicine. *Int. J. Mol. Sci.* 19 (12), E4031. doi:10.3390/ijms19124031

Kong, C., Li, Y., Liu, Z., Ye, J., Wang, Z., Zhang, L., et al. (2019). Targeting the oncogene KRAS mutant pancreatic cancer by synergistic blocking of lysosomal acidification and rapid drug release. ACS Nano 13 (4), 4049–4063. doi:10.1021/ acsnano.8b08246

Kraman, M., Bambrough, P. J., Arnold, J. N., Roberts, E. W., Magiera, L., Jones, J. O., et al. (2010). Suppression of antitumor immunity by stromal cells expressing fibroblast activation protein-alpha. *Science* 330 (6005), 827–830. doi:10.1126/science.1195300

Ladd, E., Sheikhi, A., Li, N., van de Ven, T. G. M., and Kakkar, A. (2017). Design and synthesis of dendrimers with facile surface group functionalization, and an evaluation of their bactericidal efficacy. *Molecules* 22 (6), E868. doi:10.3390/ molecules22060868

Li, D., and Jiao, L. (2003). Molecular epidemiology of pancreatic cancer. Int. J. Gastrointest. Cancer 33 (1), 3-14. doi:10.1385/IJGC:33:1:3

Li, Y., VandenBoom, T. G., 2nd, Kong, D., Wang, Z., Ali, S., Philip, P. A., et al. (2009). Up-regulation of miR-200 and let-7 by natural agents leads to the reversal of epithelial-to-mesenchymal transition in gemcitabine-resistant pancreatic cancer cells. *Cancer Res.* 69 (16), 6704–6712. doi:10.1158/0008-5472.CAN-09-1298

Libutti, S. K., Paciotti, G. F., Byrnes, A. A., Alexander, H. R., Jr., Gannon, W. E., Walker, M., et al. (2010). Phase I and pharmacokinetic studies of CYT-6091, a novel PEGylated colloidal gold-rhTNF nanomedicine. *Clin. Cancer Res.* 16 (24), 6139–6149. doi:10.1158/1078-0432.CCR-10-0978

Libutti, S. K., Tamarkin, L., and Nilubol, N. (2018). Targeting the invincible barrier for drug delivery in solid cancers: Interstitial fluid pressure. *Oncotarget* 9 (87), 35723–35725. doi:10.18632/oncotarget.26267

Lin, Z., Lu, S., Xie, X., Yi, X., and Huang, H. (2020). Noncoding RNAs in drugresistant pancreatic cancer: A review. *Biomed. Pharmacother*. 131, 110768. doi:10. 1016/j.biopha.2020.110768

Liu, L., Kshirsagar, P., Christiansen, J., Gautam, S. K., Aithal, A., Gulati, M., et al. (2021). Polyanhydride nanoparticles stabilize pancreatic cancer antigen MUC4β. *J. Biomed. Mat. Res. A* 109 (6), 893–902. doi:10.1002/jbm.a.37080

Liu, L., Kshirsagar, P. G., Gautam, S. K., Gulati, M., Wafa, E. I., Christiansen, J. C., et al. (2022). Nanocarriers for pancreatic cancer imaging, treatments, and immunotherapies. *Theranostics* 12 (3), 1030–1060. doi:10.7150/thno.64805

Lobato-Mendizabal, E., and Ruiz-Arguelles, G. J. (1990). Leukemia and malnutrition. II. The magnitude of maintenance chemotherapy as a prognostic factor in the survival of patients with standard-risk acute lymphoblastic leukemia. *Rev. Invest. Clin.* 42 (2), 81–87.

Lucero-Acuna, A., Jeffery, J. J., Abril, E. R., Nagle, R. B., Guzman, R., Pagel, M. D., et al. (2014). Nanoparticle delivery of an AKT/PDK1 inhibitor improves the therapeutic effect in pancreatic cancer. *Int. J. Nanomedicine* 9, 5653–5665. doi:10.2147/IJN.S68511

Macarulla, T., Pazo-Cid, R., Guillen-Ponce, C., Lopez, R., Vera, R., Reboredo, M., et al. (2019). Phase I/II trial to evaluate the efficacy and safety of nanoparticle albumin-bound paclitaxel in combination with gemcitabine in patients with pancreatic cancer and an ECOG performance status of 2. J. Clin. Oncol. 37 (3), 230–238. doi:10.1200/JCO.18.00089

Manzur, A., Oluwasanmi, A., Moss, D., Curtis, A., and Hoskins, C. (2017). Nanotechnologies in pancreatic cancer therapy. *Pharmaceutics* 9 (4), E39. doi:10. 3390/pharmaceutics9040039

McCarroll, J., Teo, J., Boyer, C., Goldstein, D., Kavallaris, M., and Phillips, P. A. (2014). Potential applications of nanotechnology for the diagnosis and treatment of pancreatic cancer. *Front. Physiol.* 5, 2. doi:10.3389/fphys.2014.00002

Melancon, M. P., Stafford, R. J., and Li, C. (2012). Challenges to effective cancer nanotheranostics. J. Control. Release 164 (2), 177–182. doi:10.1016/j.jconrel.2012.07.045

Metselaar, J. M., and Lammers, T. (2020). Challenges in nanomedicine clinical translation. *Drug Deliv. Transl. Res.* 10 (3), 721-725. doi:10.1007/s13346-020-00740-5

Min, C., Eddy, S. F., Sherr, D. H., and Sonenshein, G. E. (2008). NF-kappaB and epithelial to mesenchymal transition of cancer. *J. Cell. Biochem.* 104 (3), 733–744. doi:10.1002/jcb.21695

Mohelnikova-Duchonova, B., Brynychova, V., Oliverius, M., Honsova, E., Kala, Z., Muckova, K., et al. (2013). Differences in transcript levels of ABC transporters between pancreatic adenocarcinoma and nonneoplastic tissues. *Pancreas* 42 (4), 707–716. doi:10.1097/MPA.0b013e318279b861

Nieto, M. A., Huang, R. Y., Jackson, R. A., and Thiery, J. P. (2016). Emt: 2016. Cell 166 (1), 21–45. doi:10.1016/j.cell.2016.06.028

O'Driscoll, L., Walsh, N., Larkin, A., Ballot, J., Ooi, W. S., Gullo, G., et al. (2007). MDR1/P-glycoprotein and MRP-1 drug efflux pumps in pancreatic carcinoma. *Anticancer Res.* 27 (4B), 2115–2120.

Obaid, G., Bano, S., Thomsen, H., Callaghan, S., Shah, N., Swain, J. W. R., et al. (2022). Remediating desmoplasia with EGFR-targeted photoactivable multiinhibitor liposomes doubles overall survival in pancreatic cancer. *Adv. Sci.* (*Weinh*)., 24, e2104594. doi:10.1002/advs.202104594

Ogawa, K., and Miura, T. (2014). Aphid polyphenisms: trans-generational developmental regulation through viviparity. *Front. Physiol.* 5, 1. doi:10.3389/fphys.2014.00001

Oliveira, C., Calmeiro, J., Carrascal, M. A., Falcão, A., Gomes, C., Miguel Neves, B., et al. (2021). Exosomes as new therapeutic vectors for pancreatic cancer treatment. *Eur. J. Pharm. Biopharm.* 161, 4–14. doi:10.1016/j.ejpb.2021.02.002

Pandya, G., Kirtonia, A., Sethi, G., Pandey, A. K., and Garg, M. (2020). The implication of long non-coding RNAs in the diagnosis, pathogenesis and drug resistance of pancreatic ductal adenocarcinoma and their possible therapeutic potential. *Biochim. Biophys. Acta. Rev. Cancer* 1874 (2), 188423. doi:10.1016/j. bbcan.2020.188423

Parhi, P., Mohanty, C., and Sahoo, S. K. (2012). Nanotechnology-based combinational drug delivery: An emerging approach for cancer therapy. *Drug Discov. Today* 17 (17-18), 1044–1052. doi:10.1016/j.drudis.2012.05.010

Poon, C., He, C., Liu, D., Lu, K., and Lin, W. (2015). Self-assembled nanoscale coordination polymers carrying oxaliplatin and gemcitabine for synergistic combination therapy of pancreatic cancer. *J. Control. Release* 201, 90–99. doi:10. 1016/j.jconrel.2015.01.026

Prabhu, P., and Patravale, V. (2012). The upcoming field of theranostic nanomedicine: An overview. J. Biomed. Nanotechnol. 8 (6), 859-882. doi:10. 1166/jbn.2012.1459

Rawla, P., Sunkara, T., and Gaduputi, V. (2019). Epidemiology of pancreatic cancer: Global trends, etiology and risk factors. *World J. Oncol.* 10 (1), 10–27. doi:10. 14740/wjon1166

Rebelo, A., Molpeceres, J., Rijo, P., and Reis, C. P. (2017). Pancreatic cancer therapy review: From classic therapeutic agents to modern nanotechnologies. *Curr. Drug Metab.* 18 (4), 346–359. doi:10.2174/1389200218666170201151135

Rishi, A., Goggins, M., Wood, L. D., and Hruban, R. H. (2015). Pathological and molecular evaluation of pancreatic neoplasms. *Semin. Oncol.* 42 (1), 28–39. doi:10. 1053/j.seminoncol.2014.12.004

Ryan, D. P., Hong, T. S., and Bardeesy, N. (2014). Pancreatic adenocarcinoma. N. Engl. J. Med. 371 (11), 1039–1049. doi:10.1056/NEJMra1404198

Schizas, D., Charalampakis, N., Kole, C., Economopoulou, P., Koustas, E., Gkotsis, E., et al. (2020). Immunotherapy for pancreatic cancer: A 2020 update. *Cancer Treat. Rev.* 86, 102016. doi:10.1016/j.ctrv.2020.102016

Schroeder, A., Heller, D. A., Winslow, M. M., Dahlman, J. E., Pratt, G. W., Langer, R., et al. (2011). Treating metastatic cancer with nanotechnology. *Nat. Rev. Cancer* 12 (1), 39–50. doi:10.1038/nrc3180

Shah, A. N., Summy, J. M., Zhang, J., Park, S. I., Parikh, N. U., and Gallick, G. E. (2007). Development and characterization of gemcitabine-resistant pancreatic tumor cells. *Ann. Surg. Oncol.* 14 (12), 3629–3637. doi:10.1245/s10434-007-9583-5

Sharma, A., Goyal, A. K., and Rath, G. (2018). Recent advances in metal nanoparticles in cancer therapy. J. Drug Target. 26 (8), 617–632. doi:10.1080/1061186X.2017.1400553

Siegel, R. L., Miller, K. D., and Jemal, A. (2019). Cancer statistics, 2019. Ca. Cancer J. Clin. 69 (1), 7–34. doi:10.3322/caac.21551

Sielaff, C. M., and Mousa, S. A. (2018). Status and future directions in the management of pancreatic cancer: Potential impact of nanotechnology. J. Cancer Res. Clin. Oncol. 144 (7), 1205–1217. doi:10.1007/s00432-018-2651-3

Singh, A., and Settleman, J. (2010). EMT, cancer stem cells and drug resistance: An emerging axis of evil in the war on cancer. *Oncogene* 29 (34), 4741–4751. doi:10. 1038/onc.2010.215

Sofuni, A., Iijima, H., Moriyasu, F., Nakayama, D., Shimizu, M., Nakamura, K., et al. (2005). Differential diagnosis of pancreatic tumors using ultrasound contrast imaging. J. Gastroenterol. 40 (5), 518–525. doi:10.1007/s00535-005-1578-z

Soni, K. S., Thomas, D., Caffrey, T., Mehla, K., Lei, F., O'Connell, K. A., et al. (2019). A polymeric nanogel-based treatment regimen for enhanced efficacy and sequential administration of synergistic drug combination in pancreatic cancer. *J. Pharmacol. Exp. Ther.* 370 (3), 894–901. doi:10.1124/jpet.118.255372

Stathopoulos, G. P., Boulikas, T., Vougiouka, M., Deliconstantinos, G., Rigatos, S., Darli, E., et al. (2005). Pharmacokinetics and adverse reactions of a new liposomal cisplatin (Lipoplatin): phase I study. *Oncol. Rep.* 13 (4), 589–595. doi:10.3892/or.13. 4.589

Syrigos, K. N., Michalaki, B., Alevyzaki, F., Machairas, A., Mandrekas, D., Kindilidis, K., et al. (2002). A phase-II study of liposomal doxorubicin and docetaxel in patients with advanced pancreatic cancer. *Anticancer Res.* 22 (6B), 3583–3588.

Tang, S., Hang, Y., Ding, L., Tang, W., Yu, A., Zhang, C., et al. (2021). Intraperitoneal siRNA nanoparticles for augmentation of gemcitabine efficacy in the treatment of pancreatic cancer. *Mol. Pharm.* 18 (12), 4448–4458. doi:10.1021/ acs.molpharmaceut.1c00653

Tarannum, M., Holtzman, K., Dreau, D., Mukherjee, P., and Vivero-Escoto, J. L. (2022). Nanoparticle combination for precise stroma modulation and improved delivery for pancreatic cancer. *J. Control. Release* 347, 425–434. doi:10.1016/j. jconrel.2022.05.019

Tempero, M. A., Malafa, M. P., Chiorean, E. G., Czito, B., Scaife, C., Narang, A. K., et al. (2019). Pancreatic adenocarcinoma, version 1.2019. *J. Natl. Compr. Canc. Netw.* 17 (3), 202–210. doi:10.6004/jnccn.2019.0014

Thakkar, A., Desai, P., Chenreddy, S., Modi, J., Thio, A., Khamas, W., et al. (2018). Novel nano-drug combination therapeutic regimen demonstrates significant efficacy in the transgenic mouse model of pancreatic ductal adenocarcinoma. *Am. J. Cancer Res.* 8 (10), 2005–2019.

Thapa, R. K., Ku, S. K., Choi, H. G., Yong, C. S., Byeon, J. H., and Kim, J. O. (2018). Vibrating droplet generation to assemble zwitterion-coated gold-graphene oxide stealth nanovesicles for effective pancreatic cancer chemo-phototherapy. *Nanoscale* 10 (4), 1742–1749. doi:10.1039/c7nr07603g

Urey, C., Hilmersson, K. S., Andersson, B., Ansari, D., and Andersson, R. (2017). Development and *in vitro* characterization of a gemcitabine-loaded MUC4-targeted immunoliposome against pancreatic ductal adenocarcinoma. *Anticancer Res.* 37 (11), 6031–6039. doi:10.21873/anticanres.12050

van der Meel, R., Lammers, T., and Hennink, W. E. (2017). Cancer nanomedicines: Oversold or underappreciated? *Expert Opin. Drug Deliv.* 14 (1), 1–5. doi:10.1080/17425247.2017.1262346

van Horssen, R., Ten Hagen, T. L., and Eggermont, A. M. (2006). TNF-Alpha in cancer treatment: Molecular insights, antitumor effects, and clinical utility. *Oncologist* 11 (4), 397–408. doi:10.1634/theoncologist.11-4-397

Vogel, A., Rommler-Zehrer, J., Li, J. S., McGovern, D., Romano, A., and Stahl, M. (2016). Efficacy and safety profile of nab-paclitaxel plus gemcitabine in patients with metastatic pancreatic cancer treated to disease progression: A subanalysis from a phase 3 trial (MPACT). *BMC Cancer* 16 (1), 817. doi:10. 1186/s12885-016-2798-8

Von Hoff, D. D., Ervin, T., Arena, F. P., Chiorean, E. G., Infante, J., Moore, M., et al. (2013). Increased survival in pancreatic cancer with nab-paclitaxel plus gemcitabine. *N. Engl. J. Med.* 369 (18), 1691–1703. doi:10.1056/NEJMoa1304369

Von Hoff, D. D., Ramanathan, R. K., Borad, M. J., Laheru, D. A., Smith, L. S., Wood, T. E., et al. (2011). Gemcitabine plus nab-paclitaxel is an active regimen in patients with advanced pancreatic cancer: A phase I/II trial. *J. Clin. Oncol.* 29 (34), 4548–4554. doi:10.1200/JCO.2011.36.5742

Wang, L., Liu, X., Zhou, Q., Sui, M., Lu, Z., Zhou, Z., et al. (2017). Terminating the criminal collaboration in pancreatic cancer: Nanoparticle-based synergistic therapy for overcoming fibroblast-induced drug resistance. *Biomaterials* 144, 105–118. doi:10.1016/j.biomaterials.2017.08.002

Wang, Z., Li, Y., Kong, D., Banerjee, S., Ahmad, A., Azmi, A. S., et al. (2009). Acquisition of epithelial-mesenchymal transition phenotype of gemcitabineresistant pancreatic cancer cells is linked with activation of the notch signaling pathway. *Cancer Res.* 69 (6), 2400–2407. doi:10.1158/0008-5472.CAN-08-4312 Wang-Gillam, A., Li, C. P., Bodoky, G., Dean, A., Shan, Y. S., Jameson, G., et al. (2016). Nanoliposomal irinotecan with fluorouracil and folinic acid in metastatic pancreatic cancer after previous gemcitabine-based therapy (NAPOLI-1): A global, randomised, open-label, phase 3 trial. *Lancet* 387 (10018), 545–557. doi:10.1016/S0140-6736(15)00986-1

Xia, Y., Shen, S., and Verma, I. M. (2014). NF-KB, an active player in human cancers. *Cancer Immunol. Res.* 2 (9), 823–830. doi:10.1158/2326-6066.CIR-14-0112

Xie, V. K., He, J., and Xie, K. (2020a). Protein arginine methylation promotes therapeutic resistance in human pancreatic cancer. *Cytokine Growth Factor Rev.* 55, 58–69. doi:10.1016/j.cytogfr.2020.07.011

Xie, Y., Hang, Y., Wang, Y., Sleightholm, R., Prajapati, D. R., Bader, J., et al. (2020b). Stromal modulation and treatment of metastatic pancreatic cancer with local intraperitoneal triple miRNA/siRNA nanotherapy. *ACS Nano* 14 (1), 255–271. doi:10.1021/acsnano.9b03978

Xu, Y., Meng, H., Du, F., Lu, W., Liu, S., Huang, J., et al. (2015). Preparation of intravenous injection nanoformulation of VESylated gemcitabine by co-assembly with TPGS and its anti-tumor activity in pancreatic tumor-bearing mice. *Int. J. Pharm.* 495 (2), 792–797. doi:10.1016/j.ijpharm.2015.09.030

Yonemori, K., Kurahara, H., Maemura, K., and Natsugoe, S. (2017). MicroRNA in pancreatic cancer. J. Hum. Genet. 62 (1), 33-40. doi:10.1038/jhg.2016.59

Yoshida, M., Takimoto, R., Murase, K., Sato, Y., Hirakawa, M., Tamura, F., et al. (2012). Targeting anticancer drug delivery to pancreatic cancer cells using a fucosebound nanoparticle approach. *PLoS One* 7 (7), e39545. doi:10.1371/journal.pone. 0039545

Yousefi, H., Yuan, J., Keshavarz-Fathi, M., Murphy, J. F., and Rezaei, N. (2017). Immunotherapy of cancers comes of age. *Expert Rev. Clin. Immunol.* 13 (10), 1001–1015. doi:10.1080/1744666X.2017.1366315 Yu, X., Zhu, W., Di, Y., Gu, J., Guo, Z., Li, H., et al. (2017). Triple-functional albumin-based nanoparticles for combined chemotherapy and photodynamic therapy of pancreatic cancer with lymphatic metastases. *Int. J. Nanomedicine* 12, 6771–6785. doi:10.2147/IJN.S131295

Yu, Z., Pestell, T. G., Lisanti, M. P., and Pestell, R. G. (2012). Cancer stem cells. *Int. J. Biochem. Cell Biol.* 44 (12), 2144–2151. doi:10.1016/j.biocel.2012.08.022

Yuan, Y., Cai, T., Xia, X., Zhang, R., Chiba, P., and Cai, Y. (2016). Nanoparticle delivery of anticancer drugs overcomes multidrug resistance in breast cancer. *Drug Deliv.* 23 (9), 3350–3357. doi:10.1080/10717544.2016.1178825

Zeng, S., Pottler, M., Lan, B., Grutzmann, R., Pilarsky, C., and Yang, H. (2019). Chemoresistance in pancreatic cancer. *Int. J. Mol. Sci.* 20 (18), E4504. doi:10.3390/ ijms20184504

Zhang, M., Liu, E., Cui, Y., and Huang, Y. (2017). Nanotechnology-based combination therapy for overcoming multidrug-resistant cancer. *Cancer Biol. Med.* 14 (3), 212–227. doi:10.20892/j.issn.2095-3941.2017.0054

Zhao, Y., Zheng, Y., Zhu, Y., Zhang, Y., Zhu, H., and Liu, T. (2021a). M1 macrophage-derived exosomes loaded with Gemcitabine and deferasirox against chemoresistant pancreatic cancer. *Pharmaceutics* 13 (9), 1493. doi:10. 3390/pharmaceutics13091493

Zhao, Y., Zheng, Y., Zhu, Y., Zhang, Y., Zhu, H., and Liu, T. (2021b). M1 macrophage-derived exosomes loaded with gemcitabine and deferasirox against chemoresistant pancreatic cancer. *Pharmaceutics* 13 (9), 1493. doi:10. 3390/pharmaceutics13091493

Zhi, X., Tao, J., Xie, K., Zhu, Y., Li, Z., Tang, J., et al. (2014). MUC4-induced nuclear translocation of beta-catenin: A novel mechanism for growth, metastasis and angiogenesis in pancreatic cancer. *Cancer Lett.* 346 (1), 104–113. doi:10.1016/j. canlet.2013.12.021