

## Effect of Sulfonylureas Administered Centrally on the Blood Glucose Level in Immobilization Stress Model

Naveen Sharma<sup>1</sup>, Yun-Beom Sim<sup>1</sup>, Soo-Hyun Park<sup>1</sup>, Su-Min Lim<sup>1</sup>, Sung-Su Kim<sup>1</sup>, Jun-Sub Jung<sup>1</sup>, Jae-Seung Hong<sup>2</sup>, and Hong-Won Suh<sup>1</sup>

<sup>1</sup>Department of Pharmacology, Institute of Natural Medicine, <sup>2</sup>Department of Physical Education, College of Natural Medicine, College of Medicine, Hallym University, Chuncheon 200-702, Korea

Sulfonylureas are widely used as an antidiabetic drug. In the present study, the effects of sulfonylurea administered supraspinally on immobilization stress-induced blood glucose level were studied in ICR mice. Mice were once enforced into immobilization stress for 30 min and returned to the cage. The blood glucose level was measured 30, 60, and 120 min after immobilization stress initiation. We found that intracerebroventricular (i.c.v.) injection with 30  $\mu$ g of glyburide, glipizide, glimepiride or tolazamide attenuated the increased blood glucose level induced by immobilization stress. Immobilization stress causes an elevation of the blood corticosterone and insulin levels. Sulfonylureas pretreated i.c.v. caused a further elevation of the blood corticosterone level when mice were forced into the stress. In addition, sulfonylureas pretreated i.c.v. alone caused an elevation of the plasma insulin level. Furthermore, immobilization stress-induced insulin level was reduced by i.c.v. pretreated sulfonylureas. Our results suggest that lowering effect of sulfonylureas administered supraspinally against immobilization stress-induced increase of the blood glucose level appears to be primarily mediated via elevation of the plasma insulin level.

**Key Words:** Blood glucose, Brain, Immobilization stress, Sulfonylurea

### INTRODUCTION

Stress can have various effects on the human biological systems. Stress is omnipresent in modern life and has great effect on cognitive and emotional functions [1]. Stress is defined as a state of disharmony or threatened homeostasis and results in various physiological and behavioral changes. The immobilization stress model is the most commonly employed models to mimic potent physical and psychological stress [2]. Moreover, the neuroanatomical regions mediating stress response, involving a variety of neurotransmitter systems, have been identified. Among all the brain regions, the PVN is a region that plays a central role in the regulation of the HPA axis during the stress [3,4]. It has been characterized that stress influences brain activity and promotes long-term changes in various neural systems [5]. For example, stress stimulates the release of vari-

ous hormones, these increased blood glucose levels [6]. Stress leads to the activation of sympathetic nervous system (SNS) and hypothalamic-pituitary-adrenal (HPA) responses [7]. The SNS activation produces catecholamines that are known as major stress hormones, especially epinephrine and norepinephrine. Increased activity of the SNS plays a role in the regulation of glucose metabolism. The alterations in glucose and lipid metabolism induced by stress may cause the development of diabetes mellitus [8]. The main regulator of the HPA axis is a corticotrophin-releasing hormone, finally causing a glucocorticoid secretion by the adrenal cortex [9]. Glucocorticoid usually increases the gluconeogenesis activity, leading to an increase of the blood glucose level [10].

Sulfonylurea is widely used oral drugs for the treatment of type II diabetes mellitus. Many studies have also reported that the major blood glucose-lowering activity of sulfonylurea appears to be primarily through enhance  $\beta$ -cell responsiveness. Also, increase intracellular cyclic AMP; exert insulinotropic effects by closing ATP-dependent potassium channels [11,12]. Its action of blood glucose is suppressed glucagon secretion, increase peripheral insulin sensitivity without affecting insulin binding [13]. Skillman and Feldman, reported that the sulfonylurea potentiate the biologic effect of the insulin, increasing the deficient numbers

Received April 7, 2014, Revised October 10, 2014,  
Accepted February 17, 2015

Corresponding to: Hong-Won Suh, Department of Pharmacology, Institute of Natural Medicine, College of Medicine, Hallym University, 39 Hallymdaehak-gil, Chuncheon 200-702, Korea. (Tel) 82-33-248-2614, (Fax) 82-33-248-2612, (E-mail) hwsuh@hallym.ac.kr



This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Copyright © Korean J Physiol Pharmacol & MEDrang Inc.

**ABBREVIATIONS:** i.c.v., intracerebroventricular; PVN, paraventricular nucleus; HPA, hypothalamic-pituitary-adrenal; SNS, sympathetic nervous system; AMP, Adenosine monophosphate; ATP, Adenosine triphosphate; STZ, Streptozotocin.

of insulin receptors and stimulating insulin secretion on skeletal muscle, fat and liver [14].

The peripheral anti-diabetic actions of sulfonylurea have been well studied and characterized. Chari et al. [15] demonstrated that the activation of central lactate metabolism after i.c.v administration of lactate, lowers the glucose production in uncontrolled diabetes and diet induced insulin resistance. Park et al. [16] have recently demonstrated that both morphine and  $\beta$ -endorphin administered i.c.v. acutely increase the blood glucose level. Furthermore, Sim et al. [17] reported that the anti-diabetic action of repaglinide appears to be, at least, mediated via the brain and the spinal cord as revealed in both D-glucose-fed and STZ-treated models. However, the roles of sulfonylureas directly administered into the brain in the regulation of stress-induced blood glucose level have not been well characterized yet. Thus, the present study was designed to examine the effect of sulfonylurea administered supraspinally on the blood glucose level induced by immobilization stress in mice.

**METHODS**

These experiments were approved by the Hallym University Animal Care and Use Committee (Registration Number: Hallym 2009-05-01). All procedures were conducted in accordance with the ‘Guide for Care and Use of Laboratory Animals’ published by the National Institutes of Health and

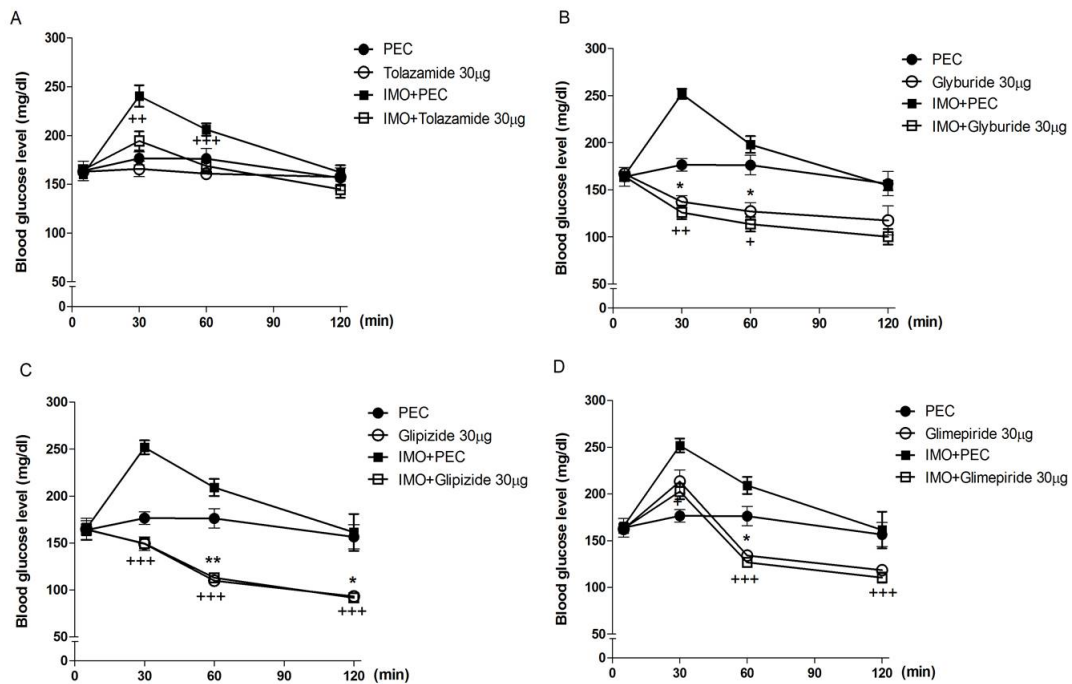
the ethical guidelines of the International Association for the Study of Pain.

**Experimental animals**

Male ICR mice (MJ Co., Seoul, Korea) weighing 20~25 g were used for all the experiments. Animals were housed 5 per cage in a room maintained at 22±0.5°C with an alternating 12 h light – dark cycle. Food and water were available *ad libitum*. The animals were allowed to adapt to the laboratory for at least 2 h before testing and were only used once. Experiments were performed during the light phase of the cycle (10:00~17:00). The animals were fasted for 16 hours. The behavioral scores were verified by repeat trials, and the observers of pain-induced behavior were blind to drug treatments.

**Intracerebroventricular injection**

The intracerebroventricular (i.c.v.) administration followed the method described by Haley [18]. Each mouse was grasped firmly without anesthesia by the loose skin behind the head. The skin was pulled taut. A 30-gauge needle attached to a 25  $\mu$ l syringe was inserted perpendicularly through the skull into the brain and solution was injected. The injection site was 2 mm from either side of the midline on a line drawn through the anterior base of the ears. The i.c.v. injection volumes were 5  $\mu$ l, and the injection sites



**Fig. 1.** Effect of tolazamide, glyburide, glipizide or glimepiride administered intracerebroventricularly on the blood glucose level in immobilization stress model. Mice were pretreated i.c.v. with 30  $\mu$ g of tolazamide, glyburide, glipizide or glimepiride for 10 min. Then mice were enforced into immobilization stress for 30 min and returned to the cage. The blood glucose level was measured at 30, 60 and 120 min after immobilization stress initiated, as shown in Fig. 1A, B, C and D respectively. The blood was collected from tail-vein. The vertical bars indicate the standard error of mean. (A: +++p<0.001, ++p<0.01; compared to PEC+IMO, B: +++p<0.001; B: \*p<0.05; compared to PEC, ++p<0.01 compared to PEC+IMO, +p<0.05; compared to PEC+IMO; C: \*\*p<0.01, \*p<0.05; compared to PEC, +++p<0.001 compared to PEC+IMO; D: A: \*p<0.05; compared to PEC, +++p< 0.001 compared to PEC+IMO). The number of animals used in the present study was 7~8 for each group.

were verified by injecting a similar volume of 1% methylene blue solution and determining the distribution of the injected dye in the ventricular space. The success rate for prior injections with this technique was over 95%.

### Immobilization stress procedure

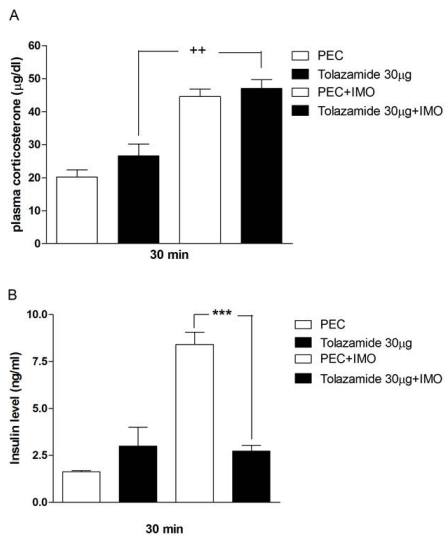
The mice were subjected to restraint stress as described in a previous study [19]. In brief, restraint was carried out by placing the mouse in a 50 ml corning tube, and adjusting it with an iron nail on the outside, which crossed in the caudal part of the animal. Adequate ventilation was provided by means of holes at the sides of the tubes. The mice were stressed by restraint for 30min. In the stress model there was a single exposure.

### Measurement of blood glucose level

The blood glucose level was measured at 30, 60 and 120 min after the immobilization stress initiated. The blood was collected shortly as much as possible with a minimum volume (1  $\mu$ l) from the tail-vein. The glucose level was measured using Accu-Chek Performa blood glucose monitoring system (glucometer) (Mannheim, Baden-Württemberg, Germany).

### Corticosterone assay and blood sampling

The plasma corticosterone level was determined by the fluorometric determination method [20]. Four hundred microliters of blood were collected by puncturing the retro-orbital venous plexus. Plasma was separated by centrifugation and stored at 80°C until assayed.



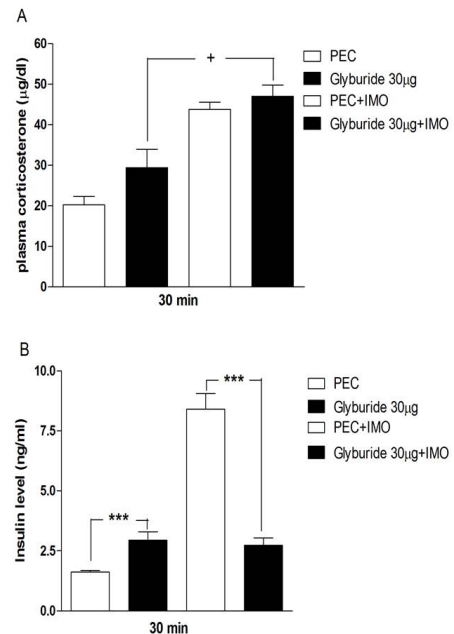
**Fig. 2.** Effect of tolazamide administered intracerebroventricularly plasma corticosterone and insulin levels in immobilization stress model. Mice were pretreated i.c.v. with 30  $\mu$ g of tolazamide for 10 min. Then mice were enforced into immobilization stress for 30 min and returned to the cage. The blood was collected from tail-vein. (A) Plasma corticosterone and (B) insulin levels were measured 30 min after immobilization stress initiated. The vertical bars indicate the standard error of mean (A: ++ $p < 0.01$  tolazamide compared to tolazamide+IMO, B: \*\*\* $p < 0.001$ ; compared to PEC+IMO). The number of animals used in the present study was 7~8 for each group.

### Insulin ELISA assay

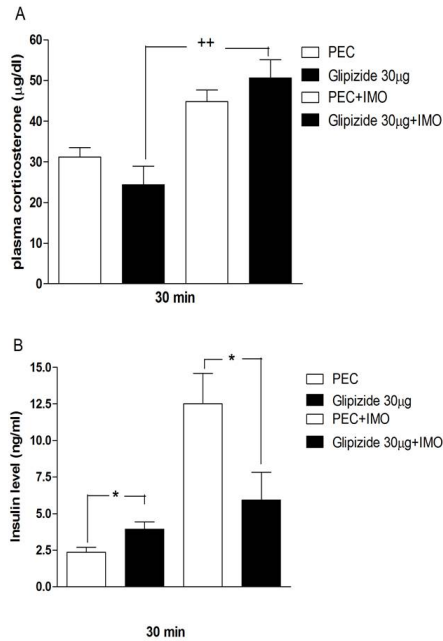
In Mouse Insulin ELISA, biotin conjugated anti insulin, and standard or sample are incubated in monoclonal anti-insulin-coated wells. After horse radish peroxidase (HRP) conjugated streptavidin remaining in the wells are reacted with a substrate chromogen reagent and the reaction is stopped by addition of an acidic solution, and absorbance is measured spectrophotometrically at 450 nm.

### Drugs

Tolazamide, glyburide, glipizide and glimepiride were purchased from Sigma Chemical Co. (St. Louis, MO, USA). Tolazamide, glyburide, glipizide and glimepiride were prepared following steps: (A) 1 g of pioglitazone and rosiglitazone was dissolved in 0.5 ml of ethanol plus 0.5 ml of polyethylene glycol 400. (B) Separately, 100 mg of sodium carboxymethylcellulose was dissolved in 9 ml of distilled water. (C) Finally, Solution (A) and Solution (B) were vigorously mixed. This solution (PEC) excluding tolazamide, glyburide, glipizide and glimepiride were used as vehicle control. All drugs were prepared just before use. Blood glucose meter, lancing device and strips were purchased from Roche Diagnostics (Accu-Chek Performa, Germany).



**Fig. 3.** Effect of glyburide administered intracerebroventricularly on the plasma corticosterone and insulin levels in immobilization stress model. Mice were pretreated i.c.v. with 30  $\mu$ g of glyburide for 10 min. Then mice were enforced into immobilization stress for 30 min and returned to the cage. The blood was collected from tail-vein. (A) Plasma corticosterone and (B) insulin levels were measured 30 min after immobilization stress initiated. The vertical bars indicate the standard error of mean (A: + $p < 0.05$ ; glyburide compared to glyburide+IMO, \* $p < 0.05$ ; compared to PEC+PBS, +++ $p < 0.001$ , ++ $p < 0.01$ , + $p < 0.05$ ; B: \*\*\* $p < 0.001$ ; PEC and PEC IMO compared to glyburide and glyburide+IMO, respectively). The number of animals used in the present study was 7~8 for each group.



**Fig. 4.** Effect of glipizide administered intracerebroventricularly on the plasma corticosterone and insulin levels in immobilization stress model. Mice were pretreated i.c.v. with 30 µg of glipizide for 10 min. Then mice were enforced into immobilization stress for 30 min and returned to the cage. The blood was collected from tail-vein. (A) Plasma corticosterone and (B) insulin levels were measured 30 min after immobilization stress initiated. The vertical bars indicate the standard error of mean (A:  $^{++}p < 0.01$  glyburide compared to glyburide+IMO, B:  $^{*}p < 0.05$ ; PEC and PEC+IMO compared to glyburide and glyburide+IMO, respectively). The number of animals used in the present study was 7~8 for each group.

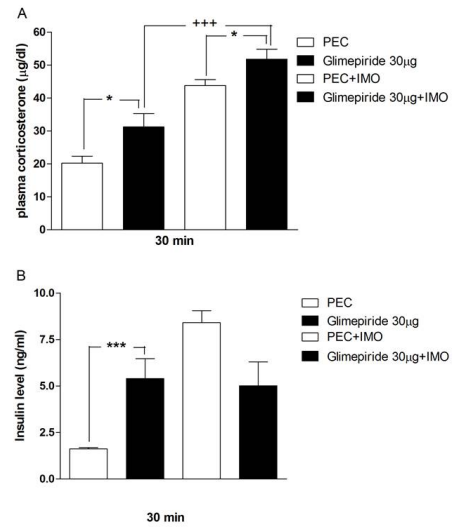
### Statistical analysis

Statistical analysis was carried out by student t test GraphPad Prism Version 4.0 for Windows (GraphPad Software, San Diego, CA, USA). p-values less than 0.05 were considered to indicate statistical significance. All values were expressed as the mean±S.E.M. In our study, we established the mean blood glucose value of the control group through many experiments under matching conditions. Selected mice of established blood glucose level were then used in replication experiments.

## RESULTS

### Effects of sulfonylureas administered intracerebroventricularly on the blood glucose level in immobilization stress model

Mice were pretreated i.c.v. with 30 µg of tolazamide, glyburide, glipizide or glimepiride for 10 min. Then mice were enforced into immobilization stress for 30 min and returned to the cage. The blood glucose level was measured at 30, 60 and 120 min after immobilization stress initiated. As shown in Fig. 1, tolazamide, glyburide, glipizide and glimepiride pretreated i.c.v. attenuated the elevation of the blood glucose level induced by immobilization stress, respectively.



**Fig. 5.** Effect of glimepiride administered intracerebroventricularly on the plasma corticosterone and insulin levels in immobilization stress model. Mice were pretreated i.c.v. 30 µg of glimepiride for 10 min. Then mice were enforced into immobilization stress for 30 min and returned to the cage. The blood was collected from tail-vein. (A) Plasma corticosterone and (B) insulin levels were measured 30 min after immobilization stress initiated. The vertical bars indicate the standard error of mean. (A:  $^{+++}p < 0.001$ ; glimepiride compared to glimepiride+IMO,  $^{*}p < 0.05$ ; PEC and PEC+IMO compared to glimepiride and glimepiride +IMO respectively, B:  $^{***}p < 0.001$ ; PEC compare to glimepiride). The number of animals used in the present study was 7~8 for each group.

### Effects of sulfonylureas pretreated i.c.v. on the blood corticosterone and insulin levels in immobilization stress model

Mice were pretreated i.c.v. with 30 µg of tolazamide, glyburide, glipizide or glimepiride for 10 min. Then mice were enforced into immobilization stress for 30 min and returned to the cage. The blood corticosterone and insulin levels were measured at 30 min after immobilization stress initiated. As shown in Figs. 2A, 3A, 4A and 5A, immobilization stress caused an elevation of plasma corticosterone level. In addition, tolazamide, glyburide, glipizide and glimepiride pretreated i.c.v. caused a further enhancement of the blood corticosterone level induced by immobilization stress. As shown in Figs. 2B, 3B, 4B and 5B, immobilization stress caused an elevation of plasma insulin level. In addition, tolazamide, glyburide, glipizide and glimepiride also increased plasma insulin level. However, tolazamide, glyburide, glipizide and glimepiride pretreated i.c.v. caused attenuated the blood insulin level induced by immobilization stress.

## DISCUSSION

In the present study, the possible inhibitory action of sulfonylureas administered supraspinally against the elevation of the blood glucose level induced by immobilization stress was studied in ICR mice. We found in the present study that all of the second-generation sulfonylureas administered supraspinally exerts an inhibitory action against the increased blood glucose level induced by immobili-

zation stress. However, first-generation sulfonylureas, except tolazamide, administered supraspinally does not alter the blood glucose level in immobilization stress model. These results suggest that some sulfonylureas exert their hypoglycemic effects at least by acting directly on the brain regions. Furthermore, from the findings of the present study, it can be speculated that, in addition to the direct action on pancreatic  $\beta$  cells, orally administered tolazamide, glyburide, glipizide and glimepiride may exert their anti-diabetic effects, at least, via acting on the brain. Although the exact reasons for differential actions of first-generation and second-generation sulfonylureas against immobilization stress-induced hyperglycemic effect are not currently known, earlier several studies have suggested that a second generation of sulfonylureas is far more potent than the first generation of sulfonylureas [12].

Several studies have demonstrated that the stress may cause increase in blood corticosterone level [21-23], this may cause increase in blood glucose level to meet energy demand during stress. Magariños et al. [24] demonstrated that, the stress-induced rise in plasma glucose may be attributed to corticosterone mediated enhanced metabolism to meet the increased demands of the body organs during stress. This elevation in corticosterone in stress condition may cause stress-induced hyperglycemia. In the present study, we found that the corticosterone level was significantly enhanced after 30 minutes of immobilization stress. In addition to this immobilization stress-induced corticosterone level is further increased by the i.c.v administration of sulfonylureas. These findings indicate that the corticosterone level increased by immobilization stress is just to supply the increased demand of energy to the organs during stress. Our results suggest that corticosterone appears not to be involved in the attenuation of blood glucose level by sulfonylureas. However corticosterone may help in blood glucose homeostasis against lowering effect of sulfonylureas.

Further, we examined whether insulin mediated pathway is involved in regulation of blood glucose level by i.c.v treatment with sulfonylureas. Ricart-Jané et al. [25] found that, a single immobilization stress application significantly increases the circulating level of corticosterone, glucose, insulin, glycerol, and ketone bodies, which is the typical response to acute stress. We found in the present study that the increase in blood glucose after acute immobilization stress is accompanied by an increase in plasma insulin level. The elevation of insulin level appears to maintain a homeostatic regulation of the blood glucose in immobilization stress. Sulfonylureas exert their anti-diabetic pharmacological effects in type 2 diabetes mellitus by improving glucose tolerance, stimulating insulin secretion peripheral tissues [26]. At in vitro, it is well known that sulfonylureas appear to block ATP-sensitive  $K^+$  channels at nanomolar concentrations in  $\beta$ -cells [27]. Sulfonylurea sensitive  $K^+$  channels inhibition lead to inhibition of gamma-aminobutyric acid (GABA) release, resulting in increase of insulin secretion [28,29]. We found in the present study that sulfonylureas administered supraspinally causes an elevation of plasma insulin level, suggesting that the activation of supraspinally located ATP-sensitive  $K^+$  channels may be responsible for an elevation of plasma insulin level. Furthermore, in a current study, we found that sulfonylureas administered supraspinally increase the blood insulin level. However, immobilization stress-induced insulin is attenuated by the supraspinal administration of sulfonylureas.

Thus, it is speculated that the attenuation of the insulin level by sulfonylureas in stress model may be primarily due to the reduction of immobilization stress-induced blood glucose level by sulfonylureas.

We conclude in the present study that the supraspinal treatment with sulfonylureas attenuates the immobilization stress induced blood glucose level. Moreover, we found that corticosterone and insulin levels were enhanced by immobilization stress. In addition, immobilization stress-induced insulin level was attenuated by the supraspinal pretreatment with sulfonylureas, whereas immobilization stress-induced corticosterone level is further enhanced by sulfonylureas, suggesting that lowering effect of sulfonylureas administered supraspinally against immobilization stress-induced increase of the blood glucose level appears to be primarily mediated via elevation of the plasma insulin level.

## ACKNOWLEDGEMENTS

This research was supported by Priority Research Centers (NRF-2009-0094071). Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology.

## REFERENCES

1. **Chen Y, Kramár EA, Chen LY, Babayan AH, Andres AL, Gall CM, Lynch G, Baram TZ.** Impairment of synaptic plasticity by the stress mediator CRH involves selective destruction of thin dendritic spines via RhoA signaling. *Mol Psychiatry*. 2013;18:485-496.
2. **Glavin GB, Paré WP, Sandbak T, Bakke HK, Murison R.** Restraint stress in biomedical research: an update. *Neurosci Biobehav Rev*. 1994;18:223-249.
3. **Honkaniemi J.** Colocalization of peptide- and tyrosine hydroxylase-like immunoreactivities with Fos-immunoreactive neurons in rat central amygdaloid nucleus after immobilization stress. *Brain Res*. 1992;598:107-113.
4. **Honkaniemi J, Kainu T, Ceccatelli S, Rechart L, Hökfelt T, Pelto-Huikko M.** Fos and jun in rat central amygdaloid nucleus and paraventricular nucleus after stress. *Neuroreport*. 1992;3:849-852.
5. **Seo YJ, Kwon MS, Shim EJ, Park SH, Choi OS, Suh HW.** Changes in pain behavior induced by formalin, substance P, glutamate and pro-inflammatory cytokines in immobilization-induced stress mouse model. *Brain Res Bull*. 2006;71:279-286.
6. **Surwit RS, Schneider MS, Feinglos MN.** Stress and diabetes mellitus. *Diabetes Care*. 1992;15:1413-1422.
7. **Gądek-Michalska A, Tadeusz J, Rachwalska P, Spyrczka J, Bugajski J.** Effect of prior stress on interleukin-1 $\beta$  and HPA axis responses to acute stress. *Pharmacol Rep*. 2011;63:1393-1403.
8. **Uresin Y, Erbas B, Ozek M, Ozkök E, Gürol AO.** Losartan may prevent the elevation of plasma glucose, corticosterone and catecholamine levels induced by chronic stress. *J Renin Angiotensin Aldosterone Syst*. 2004;5:93-96.
9. **Kalantaridou SN, Zoumakis E, Makrigiannakis A, Lavasidis LG, Vrekoussis T, Chrousos GP.** Corticotropin-releasing hormone, stress and human reproduction: an update. *J Reprod Immunol*. 2010;85:33-39.
10. **Fagerholm V, Haaparanta M, Scheinin M.**  $\alpha$ 2-adrenoceptor regulation of blood glucose homeostasis. *Basic Clin Pharmacol Toxicol*. 2011;108:365-370.
11. **Groop LC.** Sulfonylureas in NIDDM. *Diabetes Care*. 1992;15:737-754.
12. **Lebovitz HE, Feinglos MN.** Sulfonylurea drugs: mechanism of

- antidiabetic action and therapeutic usefulness. *Diabetes Care*. 1978;1:189-198.
13. **Groop L, Wählin-Boll E, Groop PH, Tötterman KJ, Melander A, Tolppanen EM, Fyhrqvist F.** Pharmacokinetics and metabolic effects of glibenclamide and glipizide in type 2 diabetics. *Eur J Clin Pharmacol*. 1985;28:697-704.
  14. **Skillman TG, Feldman JM.** The pharmacology of sulfonylureas. *Am J Med*. 1981;70:361-372.
  15. **Chari M, Lam CK, Wang PY, Lam TK.** Activation of central lactate metabolism lowers glucose production in uncontrolled diabetes and diet-induced insulin resistance. *Diabetes*. 2008;57:836-840.
  16. **Park SH, Sim YB, Kang YJ, Kim SM, Lee JK, Jung JS, Suh HW.** Characterization of blood glucose level regulation in mouse opioid withdrawal models. *Neurosci Lett*. 2010;476:119-122.
  17. **Sim YB, Park SH, Kang YJ, Kim SS, Kim CH, Kim SJ, Lim SM, Jung JS, Ryu OH, Choi MG, Suh HW.** Repaglinide, but not nateglinide administered supraspinally and spinally exerts an anti-diabetic action in d-glucose fed and streptozotocin-treated mouse models. *Korean J Physiol Pharmacol*. 2013;17:493-497.
  18. **Haley TJ.** Pharmacological effects produced by intracerebral administration of drugs of unrelated structure to conscious mice. *Arch Int Pharmacodyn Ther*. 1957;110:239-244.
  19. **Suh HW, Song DK, Huh SO, Kim YH.** Involvement of dynorphin in immobilization stress-induced antinociception in the mouse. *Eur Neuropsychopharmacol*. 2000;10:407-413.
  20. **Glick D, Vonredlich D, Levine S.** Fluorometric determination of corticosterone and cortisol in 0.02-0.05 milliliters of plasma or submilligram samples of adrenal tissue. *Endocrinology*. 1964;74:653-655.
  21. **Kim JG, Jung HS, Kim KJ, Min SS, Yoon BJ.** Basal blood corticosterone level is correlated with susceptibility to chronic restraint stress in mice. *Neurosci Lett*. 2013;555:137-142.
  22. **Nooshinfar E, Akbarzadeh-Baghdan A, Meisami E.** Effects of increasing durations of immobilization stress on plasma corticosterone level, learning and memory and hippocampal BDNF gene expression in rats. *Neurosci Lett*. 2011;500:63-66.
  23. **Pitman DL, Ottenweller JE, Natelson BH.** Plasma corticosterone levels during repeated presentation of two intensities of restraint stress: chronic stress and habituation. *Physiol Behav*. 1988;43:47-55.
  24. **Magariños AM, Verdugo JM, McEwen BS.** Chronic stress alters synaptic terminal structure in hippocampus. *Proc Natl Acad Sci U S A*. 1997;94:14002-14008.
  25. **Ricart-Jané D, Rodríguez-Sureda V, Benavides A, Peinado-Onsurbe J, López-Tejero MD, Llobera M.** Immobilization stress alters intermediate metabolism and circulating lipoproteins in the rat. *Metabolism*. 2002;51:925-931.
  26. **Müller G, Wied S.** The sulfonylurea drug, glimepiride, stimulates glucose transport, glucose transporter translocation, and dephosphorylation in insulin-resistant rat adipocytes in vitro. *Diabetes*. 1993;42:1852-1867.
  27. **Bernardi H, De Weille JR, Epelbaum J, Mourre C, Amoroso S, Slama A, Fosset M, Lazdunski M.** ATP-modulated K<sup>+</sup> channels sensitive to antidiabetic sulfonylureas are present in adenohipophysis and are involved in growth hormone release. *Proc Natl Acad Sci U S A*. 1993;90:1340-1344.
  28. **Levin BE, Brown KL, Dunn-Meynell AA.** Differential effects of diet and obesity on high and low affinity sulfonylurea binding sites in the rat brain. *Brain Res*. 1996;739:293-300.
  29. **Amoroso S, Schmid-Antomarchi H, Fosset M, Lazdunski M.** Glucose, sulfonylureas, and neurotransmitter release: role of ATP-sensitive K<sup>+</sup> channels. *Science*. 1990;247:852-854.