ORIGINAL RESEARCH

∂ OPEN ACCESS

Phase II study of the GPC3-derived peptide vaccine as an adjuvant therapy for hepatocellular carcinoma patients

Yu Sawada^{a,b}, Toshiaki Yoshikawa^a, Kazuya Ofuji^a, Mayuko Yoshimura^a, Nobuhiro Tsuchiya^{a,b}, Mari Takahashi^a, Daisuke Nobuoka^{a,c}, Naoto Gotohda^d, Shinichiro Takahashi^d, Yuichiro Kato^d, Masaru Konishi^d, Taira Kinoshita^e, Masafumi Ikeda^f, Kohei Nakachi^f, Naoya Yamazaki^g, Shoichi Mizuno^a, Tadatoshi Takayama^h, Kenji Yamaoⁱ, Katsuhiko Uesaka^j, Junji Furuse^k, Itaru Endo^b, and Tetsuya Nakatsura^a

^aDivision of Cancer Immunotherapy, Exploratory Oncology Research and Clinical Trial Center, National Cancer Center, Kashiwa, Chiba, Japan; ^bDepartment of Gastroenterological Surgery, Yokohama City University Graduate School of Medicine, Kanazawa-ku, Yokohama, Japan; ^cDepartment of Gastroenterological Surgery, Okayama University Graduate School of Medicine, Kita-ku, Okayama, Japan; ^dDivision of Surgery, National Cancer Center, Hospital East, Kashiwa, Chiba, Japan; ^eAichi Cancer Center Hospital, Chikusa-ku, Nagoya, Japan; ^fDivision of Hepatobiliary & Pancreatic Medical Oncology, National Cancer Center Hospital East, Kashiwa, Chiba, Japan; ^gDepartment of Dermatology, National Cancer Center Hospital, Chuo-ku, Tokyo, Japan; ^hDepartment of Digestive Surgery, Nihon University School of Medicine, Itabashi-ku, Tokyo, Japan; ⁱDepartment of Gastroenterology, Aichi Cancer Center Hospital, Chikusa-ku, Nagoya, Japan; ^jDivision of Hepato-Biliary-Pancreatic Surgery, Shizuoka Cancer Center Hospital, Sunto- Nagaizumi, Shizuoka, Japan; ^kDepartment of Medical Oncology, Kyorin University School of Medicine, Mitaka-shi, Tokyo, Japan

ABSTRACT

The recurrence rates of Hepatocellular carcinoma (HCC) are high, necessitating novel and effective adjuvant therapies. Therefore, we conducted a phase II study of glypican-3 (GPC3) peptide vaccine as an adjuvant therapy for HCC patients. Forty-one patients with initial HCC who had undergone surgery or radiofrequency ablation (RFA) were analyzed in this phase II, open-label, single-arm trial. Ten vaccinations were performed for 1 y after curative treatment. We also investigated case-control subjects, where selected patients treated surgically during the same period were analyzed. The expression of GPC3 in the available primary tumors was determined by immunohistochemical analysis. Six patients received RFA therapy while 35 received surgery. The recurrence rate tended to be lower in the 35 patients treated with surgery plus vaccination compared to 33 patients who underwent surgery alone (28.6% vs. 54.3% and 39.4% vs. 54.5% at 1 and 2 y, respectively; p = 0.346, 0.983). Twenty-five patients treated with surgery and vaccination had GPC3-positive tumors; the recurrence rate in this group was significantly lower compared to that in 21 GPC3-positive patients who received surgery only (24% vs. 48% and 52.4% vs. 61.9% at 1 and 2 y, respectively; p = 0.047, 0.387). The GPC3 peptide vaccine improved the 1-y recurrence rate in patients with GPC3-positive tumors. This study demonstrated that GPC3 expression by the primary tumor may be used as a biomarker in a putative larger randomized clinical trial to determine the efficacy of the GPC3-derived peptide vaccine.

Abbreviations: AFP, α -fetoprotein; CTL, cytotoxic T lymphocyte; GPC3, glypican-3; HCC, hepatocellular carcinoma; HLA, human leukocyte antigen; IFA, incomplete Freund's adjuvant; IFN γ , interferon- γ ; MHC, major histocompatibility complex; PBMC, peripheral blood mononuclear cell; RFA, radiofrequency ablation

Introduction

HCC is one of the most common forms of cancer. The prognosis for patients with HCC is very poor, and it is the third leading cause of cancer mortality worldwide.^{1,2} The 5-y recurrence rate of HCC exceeds 70% following surgery or RFA due to a high risk of metastasis and the development of de novo tumors.³⁻⁵ One of the major reasons for such a poor prognosis of HCC patients is the limited availability of treatment options to prevent recurrence. In recent years, clinical trials with sorafenib, such as the STORM trial, showed that this kinase inhibitor

was not an effective intervention in the adjuvant setting for HCC following resection or ablation⁶, although the recurrencefree survival rate could be improved by adjuvant therapy with vitamin K2⁷, retinoid⁸, or interferon.⁹ However, immunotherapy using activated lymphocytes or tumor vaccines was shown to reduce the risk of cancer recurrence in randomized controlled trials.^{10,11} As these data are derived from clinical trials, and no standard adjuvant therapy has been established, the development of an effective method of inhibiting the occurrence and recurrence of HCC remains a necessity.

CONTACT Tetsuya Nakatsura 🖾 tnakatsu@east.ncc.go.jp

ARTICLE HISTORY

Received 28 October 2015 Revised 30 November 2015 Accepted 30 November 2015

KEYWORDS

Adjuvant therapy; CTL; glypican-3; HCC; peptide vaccine

Supplemental data for this article can be accessed on the publisher's website.

Published with license by Taylor & Francis Group, LLC © Yu Sawada, Toshiaki Yoshikawa, Kazuya Ofuji, Mayuko Yoshimura, Nobuhiro Tsuchiya, Mari Takahashi, Daisuke Nobuoka, Naoto Gotohda, Shinichiro Takahashi, Yuichiro Kato, Masaru Konishi, Taira Kinoshita, Masafumi Ikeda, Kohei Nakachi, Naoya Yamazaki, Shoichi Mizuno, Tadatoshi Takayama, Kenji Yamao, Katsuhiko Uesaka, Junji Furuse, Itaru Endo, and Tetsuya Nakatsura.

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The carcinoembryonic antigen GPC3 is an ideal target for antigen-specific immunotherapy against HCC, as it is specifically overexpressed in HCC.^{12,13} Our previous study demonstrated that the tumor expression profile of GPC3 was not correlated with hepatic virus infection. Furthermore, the prognosis of patients with GPC3-positive HCC, as determined by immunohistochemical analysis, was found to be poor following initial hepatectomy.¹⁴ We therefore anticipate that GPC3-targeted therapy will prevent hepatocarcinogenesis following surgery in patients with GPC3-positive HCC.

A phase I clinical trial of a GPC3-derived peptide vaccine for advanced HCC patients was conducted in Japan. This trial demonstrated that the vaccine was well tolerated and that the frequency of GPC3-derived, peptide-specific cytotoxic T lymphocytes (CTLs) in peripheral blood mononuclear cells (PBMCs) was correlated with overall survival.¹⁵ The GPC3₂₉₈₋₃₀₆ peptide (EYILSLEEL) was used in human leukocyte antigen (HLA)-A24-positive patients and the GPC3₁₄₄₋₁₅₂ peptide (FVGEFFTDV) in HLA-A2-positive patients, as HLA-A2 and A24 are common HLA class I alleles within the Japanese population. Furthermore, we reported remarkable tumor lysis in advanced HCC patients immediately following GPC3derived peptide vaccination ¹⁶, as well as significant clinical response of progressive recurrent ovarian clear cell carcinoma to GPC3-derived peptide vaccine.¹⁷

RFA induced or enhanced T-cell responses specific for HCC-associated antigens in PBMCs derived from patients with HCC.^{18,19} Similarly, we demonstrated that RFA induced GPC3-specific CTLs in HCC patients and tumor-bearing mice.²⁰ Therefore, we hypothesized that use of an adjuvant GPC3-peptide vaccine would benefit patients with initial HCC who had previously undergone surgery or RFA.

To test this hypothesis, we conducted a single-arm phase II study of the GPC3-derived peptide vaccine as an adjuvant therapy for HCC patients in an effort to provide a larger randomized clinical trial that would determine the efficacy of the GPC3-derived peptide vaccine.

Results

Patient characteristics

Fifty-seven patients were screened in this trial. Twelve patients were excluded from the study because of HLA-A status (nine patients), non-curative treatment (one patient), or withdrawal of consent (one patient). Forty-five patients received the vaccine; however, four discontinued injections due to protocol violation, including having double cancers (two patients), not having HCC (one patient), and undergoing percutaneous ethanol injection therapy (PEIT) (one patient). Forty-one patients were ultimately analyzed in this study (Fig. 1). None of the patients dropped out due to adverse events caused by peptide vaccination. Nine patients discontinued the regimen due to recurrence within 1 y, after receiving five or more vaccinations. Patient characteristics are presented in Table 1. The median age of the patients was 64.0 y; 30 of 41 were males. The performance status was one in a single patient and 0 in the remainder. All patients had Child-Pugh class A disease. Thirty-two patients (78%) had a hepatic virus infection, 11 with HBV, and 21 with HCV. Twenty-six patients received the HLA-A*24:02-



Figure 1. Trial profile.

restricted GPC3₂₉₈₋₃₀₆ peptide, and 15 patients received the HLA-A2-restricted GPC3₁₄₄₋₁₅₂ (FVGEFFTDV) peptide.

Toxicity

One patient received PEIT therapy before vaccination, and was dropped from the study after discontinuing vaccinations owing to protocol violations immediately after the first vaccination. This patient did not receive adequate follow-up to monitor toxicity. The remaining 44 patients received adequate follow-up.

All adverse events observed in this trial are listed in detail in Table 2. All patients experienced a grade 1 or 2 local skin reaction at the injection site, and six patients experienced transient fever. These immune-related adverse events were well-tolerated. The treatment did not affect the reactivation of viral hepatitis.

Other adverse effects are unlikely to be attributable to the treatment. Grade 3 laboratory adverse events were observed in eight patients; one patient experienced a central nervous system

Table 1	١.	Patient	charact	teristics.
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Age, median (range)	64 (40–82)
Sex (M/F)	30/11
Hepatic virus infection(+/-)	78% (32/9)
AFP (ng/mL), median (range)	27.7 (1.9–59992)
PIVKA-II (mAU/mL)	51.5 (7.0-35000)
Maximum tumor diameter (mm)	25 (7–150)
Number of tumors (multiple/solitary)	12/29
Stage [*] (I/II/III)	20/17/4
Surgery/RFA	35/6
HLA-A2/A24 (vaccine type)	15/26

*Stage: Staging was performed according to the TNM classification for HCC (Union for International Cancer Control). AFP, α-fetoprotein; PIVKA-II, prothrombin induced by vitamin K absence-II; RFA, radiofrequency ablation.

Table 2. Adverse events.

Adverse events	То	tal (%)	Gr	ade 1	Gr	ade 2	G	rade 3	G	rade 4
Any events	44	(100.0)	14	(31.8)	22	(50.0)	8	(18.2)	0	(0.0)
Any immune-related event	44	(100.0)	37	(84.1)	7	(15.9)	0	(0.0)	0	(0.0)
Drug fever	6	(13.6)	4	(9.1)	2	(4.5)	0	(0.0)	0	(0.0)
Rash or Flushing	44	(100.0)	42	(95.5)	2	(4.5)	0	(0.0)	0	(0.0)
Injection site reaction	44	(100.0)	40	(90.9)	4	(9.1)	0	(0.0)	0	(0.0)
Pruritus	13	(29.5)	13	(29.5)	0	(0.0)	0	(0.0)	0	(0.0)
Any non-immune-related events	44	(100.0)	16	(36.4)	20	(45.5)	8	(18.2)	0	(0.0)
Leukopenia	20	(45.5)	13	(29.5)	7	(15.9)	0	(0.0)	0	(0.0)
Neutropenia	17	(38.6)	11	(25.0)	5	(11.4)	1	(2.3)	0	(0.0)
Lymphopenia	15	(34.1)	10	(22.7)	5	(11.4)	0	(0.0)	0	(0.0)
Anemia	16	(36.4)	10	(22.7)	5	(11.4)	1	(2.3)	0	(0.0)
Thrombopenia	13	(29.5)	11	(25.0)	2	(4.5)	0	(0.0)	0	(0.0)
Hyperbilirubinemia	9	(20.5)	7	(15.9)	2	(4.5)	0	(0.0)	0	(0.0)
Increase in aspartate aminotransferase	19	(43.2)	11	(25.0)	5	(11.4)	3	(6.8)	0	(0.0)
Increase in alanine aminotransferase	17	(38.6)	9	(20.5)	7	(15.9)	1	(2.3)	0	(0.0)
Increase in creatinine	3	(6.8)	2	(4.5)	1	(2.3)	0	(0.0)	0	(0.0)
Proteinuria	9	(20.5)	5	(11.4)	4	(9.1)	0	(0.0)	0	(0.0)
Increase in PT-INR*	9	(20.5)	9	(20.5)	0	(0.0)	0	(0.0)	0	(0.0)
Increase in blood sugar	29	(65.9)	17	(38.6)	8	(18.2)	4	(9.1)	0	(0.0)
Sugar in urine	9	(20.5)	2	(4.5)	5	(11.4)	2	(4.5)	0	(0.0)
Increase in alkaline phosphatase	14	(31.8)	11	(25.0)	3	(6.8)	0	(0.0)	0	(0.0)
Hypoalbuminemia	16	(36.4)	14	(31.8)	2	(4.5)	0	(0.0)	0	(0.0)
Hyponatremia	17	(38.6)	17	(38.6)	0	(0.0)	0	(0.0)	0	(0.0)
Hyperkalemia	2	(4.5)	2	(4.5)	0	(0.0)	0	(0.0)	0	(0.0)
CNS hemorrhage	1	(2.3)	0	(0.0)	1	(2.3)	0	(0.0)	0	(0.0)

*PT-INR, prothrombin time-international normalized ratio.

hemorrhage (grade 2), and two developed interstitial pneumonia 2 and 10 mo after the observation period, respectively. and 53.7% vs. 42.5% and 66.2% at 1 and 2 y, respectively; p = 0.054, 0.198) (Fig. S1). The primary endpoint was not reached for 1- and 2-y recurrence rates.

Recurrence rate

The median follow-up period was 40.4 mo (range 24–60 mo). All patients were followed for 2 y or more. The median overall survival rate was 20.1 mo (95% confidence interval, 14.7–25.5) (Fig. 2). HCC recurred in 31 patients, most commonly within the liver (28/31) during follow-up. Two metastases were detected in the lung, as were two in the lymph node and one in bone. The 1- and 2-y recurrence rates for the 41 patients who received the vaccine were 24.4% and 53.7%, respectively (Fig. 2); the expected corresponding recurrence rates were 20% and 45%, respectively, in this single-arm trial (p = 0.635, 0.436). The recurrence rates tended to be lower in the 41 patients treated with surgery or RFA together with vaccination compared to the 80 patients who received surgery only (24.4%)

Immunological analysis of PBMCs and recurrent tumor

To determine whether the GPC3 peptide vaccine could induce a specific immune response, PBMCs obtained from all patients before and after vaccination were subjected to an *ex vivo* interferon γ (IFN γ) ELISPOT assay as described previously.²¹ Prior to the vaccination, the number of GPC3 peptide-specific CTLs in 5 × 10⁵ PBMCs was below 10 in all patients. However, a number greater than 10 was achieved in 35 of the 41 patients (85.4%) after vaccination. Furthermore, we found that induction of GPC3-specific CTL tended to correlate with GPC3 expression in the primary tumor (p = 0.224, Student's *t*-test; Fig. S2A). However, there was no statistically significant correlation between the maximum number of GPC3-peptide-specific



Figure 2. Kaplan–Meier curves for recurrence-free survival and overall survival in 41 patients who received vaccinations in conjunction with surgery or radiofrequency ablation.



Figure 3. Cytotoxic T lymphocyte (CTL) analysis in peripheral blood mononuclear cells (PBMCs) and recurrent tumors. (A) Immunohistochemical staining for glypican-3 (GPC3) showed positivity in the primary tumor. The recurrent tumor appeared to lack GPC3 expression. Change in the expression of GPC3 in cases 42 and 53 after GPC3 peptide vaccination. Magnification = 200X. (B) *Ex vivo* interferon_{*Y*} (IFN_{*Y*}) ELISPOT assays for GPC3 in 5 × 10⁵ peripheral blood mononuclear cells were performed before and after vaccination. Δ spot number indicates the number of GPC3-peptide-specific CTLs. The number of IFN_{*Y*}-positive spots increased in the wells pre-incubated with GPC3 peptide.

CTLs and the recurrence-free or overall survival (p = 0.136 and p = 0.415 [Spearman's test], respectively; Fig. S2B).

In 11 patients, GPC3 expression in recurrent tumors was subjected to immunohistochemical analysis. Two of the 11 cases appeared to lack GPC3 expression in the recurrent tumor, although GPC3 was expressed in the primary HCC tissue prior to GPC3-peptide vaccination (cases 42 and 53) (Fig. 3A). In both these cases, the spot number of GPC3-specific CTLs after vaccination was high, as determined by an *ex vivo* IFN γ ELI-SPOT assay (Case 42: 239; Case 53: 379) (Fig. 3B). These results indicate that GPC3-peptide-specific CTLs were present in these patients after having undergone vaccination. In the other nine patients whose recurrent tumors did express GPC3, the spot numbers of GPC3-specific CTLs tended to be lower than the aforementioned two patients (Case 4: 42, 11: 56, 13: 21, 21: 56, 38: 22, 45: 2, 50: 20, 51: 130, 52: 353; *p* = 0.059, Mann–Whitney U test). These results suggested that GPC3-peptide-specific CTLs induced by vaccination prevented the recurrence of GPC3 expressing tumors in patients 42 and 53. Although peptide-specific CTLs were induced by vaccination, the reduction of recurrence due to the peptide vaccine might be limited.

We established GPC3-peptide-specific CTL lines from needle-biopsy specimens of the recurrent tumor of Case 52. These CTL lines recognized GPC3 peptide presented by target cells. In this patient, we were able to compare the expression of the inhibitory molecules PD-1 and Tim-3 in CD8-positive T-cells among PBMCs and in recurrent tumor tissues obtained by needle biopsy using flow cytometry. The expression of PD-1 was higher in CD8-positive T-cells in the recurrent tumor then in PBMCs (data not shown).

Discussion

The primary endpoint was not attained for 1- and 2-y recurrence rates in this study. However, there were significant differences in the clinical backgrounds of the 41 patients who received the vaccine compared to the historical control group of 80 patients who underwent surgery between 2001 and 2002 (Table S1). Hence, it was difficult to evaluate the reduction in the risk of post-operative recurrence by vaccination.

Therefore, we analyzed the case-control study, and selected those HCC patients who had undergone curative resection at National Cancer Center Hospital East during the same period as the vaccine trial and had provided written informed consent. The recurrence rate tended to be lower in the 35 patients treated with surgery plus vaccination compared to 33 patients who underwent surgery alone (28.6% vs. 54.3% and 39.4% vs. 54.5% at 1 and 2 y, respectively; p = 0.346, 0.983) (Fig. 4A). There were no significant differences in the patients' clinical backgrounds (Table S2). We evaluated GPC3 expression by immunohistochemical analysis of specimens from 31 of the 35 patients treated with surgery plus vaccination whose tissues were available; GPC3 expression was detected in 25 of these 31 patients. On the other hand, GPC3 expression was detected in 21 of the 33 patients in the surgery-only group. We previously reported that the prognosis of patients with GPC3-positive HCC was poor following initial hepatectomy.¹⁴ We compared the 25 surgery plus vaccine-treated patients who were GPC3-positive in their primary tumors to the 21 GPC3positive patients who underwent surgery only, and found that the recurrence rate was significantly lower in the former group than in the latter (24% vs. 48% and 52.4% vs. 61.9% at 1 and 2 y, respectively; p = 0.047, 0.387) (Fig. 4B). There were no significant differences in the patients' clinical backgrounds (Table 3).

Several clinical trials have reported that immunotherapy improves the prognosis of postoperative HCC patients, and is thus a promising adjuvant therapy.^{10,11,22} However, immunotherapy for HCC is still in the preclinical and clinical trial phases of development. Analysis of the correlation between clinical and immunological responses is required to improve the efficacy of immunotherapy.²³ To our knowledge, no clinical trial that included proper immunological analysis, especially for immune-effector cells, has been conducted for HCC in an adjuvant setting. In this trial, we analyzed vaccine-induced CTLs in PBMCs as well as in the recurrent tumors. Furthermore, we performed immunohistochemical analysis of the primary and recurrent tumors to evaluate the target cancer.

Early recurrence corresponds to intrahepatic metastasis undetected at the time of curative treatment.²⁴ In this study, the 1-y recurrence rate was lower in the vaccination group than in the control group. There was a limitation in our ability to evaluate the efficacy of the vaccine, as this was not a randomized-controlled study, but rather a case-controlled study. However, our phase II study, which included immunological analysis to examine the role of tumor-antigen-specific CTLs in



Figure 4. Kaplan–Meier curves for recurrence-free and overall survival. (A) Thirty-five patients treated with surgery and vaccination did not have significantly longer recurrence-free survival or overall survival rates than the 33 patients who underwent surgery only. (B) Among patients with glypican-3 (GPC3)-positive tumors, the recurrence rate was significantly lower in the 25 patients treated with surgery and vaccination compared to the 21 patients who underwent surgery only (24% vs. 48% and 52.4% vs. 61.9% at 1 and 2 y, respectively; p = 0.047, 0.387). The 25 patients treated with surgery and vaccination tended to have longer recurrence-free and overall survival rates compared to the 21 patients who underwent surgery only.

immunotherapy against GPC3, was necessary prior to conducting a larger randomized clinical trial.

In this vaccination trial, marked antigen-specific immune responses were observed in PBMCs by *ex vivo* IFN γ ELISPOT assays. Prior to vaccination, the number of GPC3-peptide-specific CTLs in 5 × 10⁵ PBMCs was below 10 in all patients. In contrast, the number of GPC3-peptide-specific CTLs before vaccination in a phase I trial for advanced HCC was greater than 10 in 5 of the 33 patients.¹⁵ Differences in the presence of tumors may account for the varied spontaneous GPC3-peptide-specific CTL responses between the two vaccination trials. Although the GPC3-specific CTL frequency after vaccination was correlated with overall survival in the previous phase I trial, we found no correlation between GPC3-peptide-specific CTL frequency and recurrence-free survival in this study. In a clinical trial of MART-1 peptide vaccine for resected melanoma, recurrence-

Table 3. Patient characteristics.

	Vaccine $n = 25$	Control $n = 21$	p value
Age (median, range)	63.0 (47–78)	67 (41–80)	0.651
Sex (M/F)	17/8	17/4	0.502
Hepatic virus infection (+/-)	20/5	20/1	0.198
AFP (ng/mL) (median, range)	24.2 (1.9–59992)	36.9 (1.4–48960)	0.842
PIVKA-2 (median, range)	71 (15–14358)	197 (10–29498)	0.246
Maximum tumor diameter (mm)	24 (7–150)	44 (20–130)	0.126
Number of tumor (multiple/solitary)	5/20	5/16	1.000
Stage [*] (I/II/III)	14/7/4	11/6/4	0.956

*Stage: Staging was performed according to the TNM classification for HCC (Union for International Cancer Control). AFP, α-fetoprotein; PIVKA-II, prothrombin induced by vitamin K absence-II.

free survival was not correlated with an ELISPOT response after *in vitro* sensitization via cytokine release ELISA.²⁵ In a randomized trial of advanced melanoma treatment with 12 major histocompatibility complex (MHC) class I peptides alone or in combination with either a T-helper tetanus peptide or a mixture MHC class II peptide, an ELISPOT response after *in vitro* sensitization to MHC class II peptides was associated with clinical response and overall survival.²⁶ Further studies are necessary to determine whether an antigen-specific immune response in PBMCs was associated with the clinical outcome.

We demonstrated that GPC3-peptide-specific CTLs are present in PBMCs and recurrent tumors, which suggests that the GPC3-peptide-specific CTLs induced by the peptide vaccine had a limited effect on recurrence. Fourcade et al. reported that tumor-antigen-specific CTLs were detected ex vivo in PBMCs from patients immunized with incomplete Freund's adjuvant (IFA), CpG oligodeoxynucleotide, and HLA-A2restricted analog NY-ESO-1 peptide and Pan-DR epitope peptides.²⁷ However, this trial did not demonstrate dramatic antitumor efficacy in patients with advanced melanoma. To explain the limitation of the cancer vaccine, the authors showed that the expansion of antigen-specific CTLs was associated with the upregulation of PD-1 and Tim-3 at the time of immunization, and that PD-1 and Tim-3 blockade further augmented the expansion of, and cytokine production by, CTLs. In a mouse model, expression of the death receptor was increased on the surface of CTLs at the vaccination site after peptide/IFA vaccination. Furthermore, peptide/IFA vaccination induced higher antigen-driven expression of inhibitory receptor PD-1, lymphocyte activating gene 3, CTLA-4, and Tim-3 in CTLs, suggesting partial exhaustion.²⁸ In our study, the expression of PD-1 among CD8-positive T-cells was higher in the recurrent tumor compared to PBMCs. These results suggest that the expression of inhibitory receptors in humans induced by vaccination is similar to that in the mouse model.

The previous study using mice showed that peptide/IFA vaccination primed tumor-specific CD8⁺ T-cells, which accumulated not in the tumors but rather at the persisting, antigenrich vaccination site.²⁸ In contrast, T-cell recruitment to the site of injection with IFA alone was observed in a randomized clinical trial in advanced melanoma that aimed to assess the cellular and molecular events at the site of immunization with IFA alone or a peptide in IFA.²⁹ Further studies are necessary to determine whether the peptide vaccine induces antigen-specific T-cell recruitment to the site of vaccination in humans.

We found no GPC3 expression in the recurrent tumor following GPC3 peptide vaccination. In another GPC3 peptide vaccine trial for ovarian clear-cell carcinoma, one case demonstrated a partial response using the RECIST criteria, but later appeared to lack GPC3 expression in the resected right inguinal lymph node metastasis following vaccination.¹⁷ These results suggest the activation of immune-escape mechanisms following GPC3 peptide vaccination, including the downregulation of cancer-specific antigens by tumor cells.

We clearly demonstrated the presence of GPC3-peptide-specific CTLs in peripheral blood, and that GPC3-peptide-specific CTLs had infiltrated into the recurring tumor following GPC3 peptide vaccination. Questions remain as to why many vaccinated patients show increased numbers of circulating tumor-specific T-cells in the absence of a clinical response.³⁰ Our data suggest that vaccine-

induced CTLs are partially exhausted. The challenge is to overcome T-cell exhaustion by developing new adjuvants, novel vaccine formulations, or combination therapies with immune-modulating antibodies.³¹ Recently, we reported that a PD-1-blocking antibody augmented GPC3-specific CTL clones that degranulate against liver tumor cells evaded by the CTLs because of PDL1 expression *in vitro*.³² This should be investigated further to enhance the antitumor effect of the GPC3 peptide vaccine. The immunological analysis in this study may lead to improvement of the antitumor effect of GPC3 peptide vaccines.

In this case-control study, patients treated with surgery and vaccination did not have longer recurrence-free or overall survival than patients who underwent surgery only. However, GPC3 peptide vaccination in patients with GPC3-positive tumors did improve 1-y recurrence rates. This study demonstrated that GPC3 expression by the primary tumor could be a potential biomarker for a future clinical trial of a GPC3-derived peptide vaccine. We are planning another randomized clinical trial with eligibility criteria that include GPC3 expression in the primary tumor to determine the efficacy of a GPC3-derived peptide vaccine.

Materials and methods

Patient eligibility

This clinical trial was approved by the Ethics Committee of the National Cancer Center and was performed between September 2009 and August 2014. Patients with initial HCC who had undergone surgery or RFA were enrolled after providing written informed consent. The following eligibility criteria were employed: initial HCC, had undergone curative surgery or RFA (curability was evaluated by computed tomography [CT] or magnetic resonance imaging [MRI] before vaccination); HLA-A24- or HLA-A2-positive status, as determined by commercially available genomic DNA typing tests (Mitsubishi Chemical Medience, Tokyo, Japan); age between 20 and 85 y; an Eastern Cooperative Oncology Group performance status of 0-2; Child-Pugh liver function class A-B; and adequate organ function (total bilirubin \leq 3.0 mg/dL, aspartate aminotransferase \leq 200 IU/L, alanine aminotransferase \leq 200 IU/L, and serum creatinine ≤ 1.5 mg/dL). The following exclusion criteria were applied: massive vascular invasion before curative surgery or RFA; extensive ascites; other active malignancy; serious infection; severe cardiac insufficiency; clinically serious comorbidity; pregnancy or lactation; and concurrent treatments unsuitable for the trial based on clinical judgment.

Study design and endpoints

This study was an open-label, single-arm, phase II clinical trial of the GPC3 peptide vaccine as an adjuvant therapy for initial HCC patients. The HLA-A*24:02-restricted GPC3₂₉₈₋₃₀₆ peptide (EYILSLEEL) (American Peptide Company, Sunnyvale, CA) was used in HLA-A24-positive patients, and the HLA-A2restricted GPC3₁₄₄₋₁₅₂ peptide (FVGEFFTDV) (American Peptide Company) was used in HLA-A2-positive patients. GPC3 peptide (3.0 mg) was emulsified with IFA (Montanide ISA-51VG, SEPPIC) and administered by intradermal injection. Peptides and IFA were synthesized according to Good Manufacturing Practice guidelines. The first vaccine was administered within 4 weeks after curative surgery or RFA. Ten vaccinations were performed for 1 y after curative treatment; initially six times every 2 weeks followed by four times every 2 mo. Vaccine treatment was discontinued in cases in which patients experienced recurrence within 1 y following curative treatment. Primary endpoints were 1- and 2-y recurrence rates; secondary endpoints were safety and immunological responses. In this single-arm trial, the expected 1- and 2-y recurrence rates were 20% and 45%, respectively. We estimated that we would require the enrollments of 40 patients to justify proceeding with a follow-up randomized controlled trial. We noted that the 1and 2- y recurrence rates of 80 patients who had undergone surgery between 2001 and 2002 were 42.5% and 66.2%, respectively (the estimated rates were 40.5% and 58.7%, respectively) at the National Cancer Center Hospital East. The study would have at least 80% power to detect a relative reduction in the risk of recurrence compared to a historical control, using a two-sided α level of 0.05. This study was approved by the Ethics Committee of the National Cancer Center, and conformed to the ethical guidelines of the 1975 Declaration of Helsinki. The trial has been registered with the University Hospital Medical Information Network Clinical Trials Registry (UMIN-CTR number, 000002614).

Evaluation of toxicity and recurrence-free survival

Patients were examined for signs of toxicity during and after vaccination. Adverse events were graded according to the Common Terminology Criteria for Adverse Events v3.0. Hematological examinations were conducted prior to each vaccination. Contrastenhanced CT was performed every 8–12 weeks. Contrastenhanced MRI was performed, as needed, to diagnose suspicious recurrent lesions. We defined recurrence as the appearance of new lesions with radiological features typical of HCC. All time estimates were recorded with the date of curative treatment as the baseline. All patients received follow-up for at least 2 y, or until death.

Measurement of immunological response

PBMCs and ex vivo IFN γ enzyme-linked immunospot (ELISPOT) assay

An ex vivo IFNy ELISPOT assay was performed to evaluate the antigen-specific CTL response, as described previously.²¹ Peripheral blood (30 mL) was obtained for 2 y; initially every 2 weeks before each vaccination during the initial 3 mo, then every month for 9 mo (pre-vaccination), and finally every 2 mo during the second year. Samples were centrifuged with a Ficoll-Paque gradient and PBMCs were frozen prior to immunological analysis. All PBMCs obtained from an individual patient were incubated in the same plate and analyzed by an ex vivo IFNy ELISPOT assay at the same time. Non-cultured PBMCs (5 \times 10⁵/well) were added to plates in the presence of peptide antigens (10 μ g/mL) and incubated for 20 h at 37°C in 5% CO₂. The antigen for GPC3 was either the HLA-A2-restricted GPC3₁₄₄₋₁₅₂(FVGEFFTDV) peptide or the HLA-A*24:02-restricted GPC3₂₉₈₋₃₀₆ peptide (EYILSLEEL). PBMCs plus HLA-A2-restricted HIV₁₉₋₂₇ (TLNAWVKVV) peptide (ProImmune) or HLA-A*24:02-restricted HIV₅₈₃₋₅₉₁ (RYLKDQQLL) (ProImmune) were used as negative controls. All analyses were performed in duplicate.

Immunohistochemical analysis

Biopsy or resected specimens were taken from some of the vaccinated patients, each of whom provided informed consent. Specimens were stained with hematoxylin and eosin or monoclonal antibodies against GPC3 (clone 1G12; dilution 1:300; BioMosaics), CD8 (clone 1A5; dilution 1:80; Novocastra), and HLA class I (clone EMR8/5; dilution 1:2500; Hokudo), according to the manufacturers' directions.

Statistical analysis

All statistical analyses were performed using the PASW Statistics software, version 18.0 (SPSS Inc..). Survival rates were analyzed by the Kaplan–Meier method. Comparisons of patient characteristics were done using the Pearson χ^2 , Student's *t*, and Mann–Whitney *U* tests. Statistical significance was defined by a value of p < 0.05.

Disclosure of potential conflicts of interest

T. N. is a scientific advisor for Ono Pharmaceutical co, Ltd. T. N. and Y. S. are supported by a fundamental research funding from Ono Pharmaceutical co, Ltd. The other authors have no potential conflicts of interest to declare with regard to this study.

Acknowledgments

Y.S. would like to thank the Foundation for Promotion of Cancer Research (Japan) for the Third-Term Comprehensive Control Research for Cancer for the award of a research resident fellowship. We also thank Manami Shimomura, Kayoko Shoda, and Yukiko Kozaki for technical assistance.

Funding

This study was supported in part by the National Cancer Center Research and Development Fund (25-A-7), as well as Health and Labor Science Research Grants for Research on Hepatitis and for Clinical Research and Third Term Comprehensive Control Research for Cancer from the Ministry of Health, Labor and Welfare, Japan. This study was performed as a part of a research program of the Project for Development of Innovative Research on Cancer Therapeutics (P-Direct), Ministry of Education, Culture, Sports, Science and Technology of Japan.

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