GENOME SEQUENCES





Draft Genome Sequence of *Rhodococcus aetherivorans* JCM 14343^T, a Bacterium Capable of Degrading Recalcitrant Noncyclic and Cyclic Ethers

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ABSTRACT Here, we report the draft genome sequence of *Rhodococcus aetheriv*orans JCM 14343^T, which possesses the versatile ability to degrade recalcitrant noncyclic and cyclic ether compounds. The 4.2-Mbp genome of this bacterium contains alkane hydroxylase and propane monooxygenase genes involved in the degradation of noncyclic and cyclic ethers, respectively.

Members of the genus *Rhodococcus* possess the versatile ability to catabolize a variety of xenobiotic organic pollutants and are useful for the bioremediation of polluted environments (1, 2). Among the members of this genus, *Rhodococcus aetherivorans* is a species of special interest in the biodegradation of various recalcitrant organic pollutants, such as noncyclic and cyclic ethers (3–5), petroleum compounds (6), and chlorinated organic compounds (7). The type strain of this species, *R. aetherivorans* JCM 14343 (originally named strain 10bc312), was originally isolated from petrochemical biotreater sludge as a methyl *tert*-butyl ether (MTBE)-degrading strain (3). Our recent study revealed that this strain is also capable of degrading recalcitrant cyclic ethers, such as 1,4-dioxane and tetrahydrofuran (4). To further understand the genetic basis of the degradation of ethers and other abilities, genome sequence analysis of *R. aetherivorans* JCM 14343^T was performed.

The sequenced strain, R. aetherivorans JCM 14343^T, was provided by Riken BRC through the National Bio-Resource Project of the Ministry of Education, Culture, Sports, Science, and Technology (MEXT), Japan. After cultivation in malt extract-glucose-yeast extract (MGY) medium (8), total genomic DNA (gDNA) of R. aetherivorans JCM 14343^T was extracted using a NucleoSpin tissue kit (Macherey-Nagel, Düren, Germany) and purified with ethanol precipitation and using the NucleoSpin gDNA cleanup kit (Macherey-Nagel). The purified DNA was fragmented and prepared into a sequence library using the TruSeg Nano DNA LT library preparation kit (Illumina, San Diego, CA, USA). This library was sequenced with 101-bp paired-end sequencing using the Illumina HiSeq 2500 system, which produced 22,532,896 reads with a yield of 2,276 Mbp. Unless otherwise noted, default parameters were used for all software tools. From the obtained sequence reads, the adapter sequences were trimmed using Cutadapt v. 1.1, and low-quality-value regions were further trimmed using Trimmomatic v. 0.32 (http://www .usadellab.org/cms/?page=trimmomatic). The trimmed reads were assembled onto the draft genome sequence using Velvet v. 1.2.08 (https://www.ebi.ac.uk/~zerbino/velvet/). The draft genome sequence had a total length of 6,442,200 bp with a G+C content of 70.2% and a genome coverage of 353 \times and is based on 265 contigs with an N_{so} value of 159,427 bp and a maximum contig length of 504,978 bp.

The genome was annotated with Rapid Annotations using Subsystems Technology (9). Totals of 5,988 coding sequences, 49 tRNAs, and 6 rRNAs were predicted. The

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Received 25 October 2019 Accepted 22 December 2019 Published 16 January 2020 genome of *R. aetherivorans* JCM 14343^T contains alkane hydroxylase genes (an alkane monooxygenase gene, two rubredoxin genes, and a rubredoxin reductase gene) identical to those of *Rhodococcus* spp. The alkane hydroxylase is involved in the initial MTBE oxidation (10). Additionally, the genome contains propane monooxygenase genes (a type of soluble di-iron monooxygenase), which are similar to those of *Mycobacterium* spp. and are considered to be associated with the initial hydroxylation of cyclic ethers (e.g., 1,4-dioxane and tetrahydrofuran), leading to the cleavage of their high-energy C-O bond (11).

Data availability. The whole-genome shotgun project of *R. aetherivorans* JCM 14343^T has been deposited at DDBJ/EMBL/GenBank under the accession number BLAH00000000. The version described in this paper is the first version, BLAH01000000, and consists of sequences BLAH01000001 to BLAH01000265. The raw sequencing reads were deposited under accession number DRR201558. The BioProject and BioSample numbers are PRJDB8788 and SAMD00186790, respectively, and the assembled and annotated set of contigs is publicly available under assembly accession number GCA_009176285.

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D. Inoue performed the experiments, interpreted the data, and drafted and revised the manuscript. M. Nakazawa, N. Yamamoto, and K. Sei interpreted the data and revised the manuscript. M. Ike interpreted the data, revised the manuscript, and supervised the project.

REFERENCES

- 1. Larkin MJ, Kulakov LA, Allen CC. 2005. Biodegradation and *Rhodococcus*: masters of catabolic versatility. Curr Opin Biotechnol 16:282–290. https://doi.org/10.1016/j.copbio.2005.04.007.
- Martínková L, Uhnáková B, Pátek M, Nesvera J, Kren V. 2009. Biodegradation potential of the genus *Rhodococcus*. Environ Int 35:162–177. https://doi.org/10.1016/j.envint.2008.07.018.
- Goodfellow M, Jones AL, Maldonado LA, Salanitro J. 2004. *Rhodococcus aetherivorans* sp. nov., a new species that contains methyl *t*-butyl ether-degrading actinomycetes. Syst Appl Microbiol 27:61–65. https://doi.org/10.1078/0723-2020-00254.
- Inoue D, Tsunoda T, Yamamoto N, Ike M, Sei K. 2018. 1,4-Dioxane degradation characteristics of *Rhodococcus aetherivorans* JCM 14343. Biodegradation 29:301–310. https://doi.org/10.1007/s10532-018-9832-2.
- Tajima T, Hayashida N, Matsumura R, Omura A, Nakashimada Y, Kato J. 2012. Isolation and characterization of tetrahydrofuran-degrading *Rhodococcus aetherivorans* strain M8. Process Biochem 47:1665–1669. https://doi.org/10.1016/j.procbio.2011.08.009.
- Auffret M, Labbé D, Thouand G, Greer CW, Fayolle-Guichard F. 2009. Degradation of a mixture of hydrocarbons, gasoline, and diesel oil additives by *Rhodococcus aetherivorans* and *Rhodococcus wratislaviensis*. Appl Environ Microbiol 75:7774–7782. https://doi.org/10.1128/AEM.01117-09.
- 7. Frascari D, Pinelli D, Nocentini M, Fedi S, Pii Y, Zannoni D. 2006. Chlo-

roform degradation by butane-grown cells of *Rhodococcus aetherivorans* BCP1. Appl Microbiol Biotechnol 73:421–428. https://doi.org/10.1007/s00253-006-0433-3.

- Inoue D, Tsunoda T, Sawada K, Yamamoto N, Saito Y, Sei K, Ike M. 2016. 1,4-Dioxane degradation potential of members of the genera *Pseudono-cardia* and *Rhodococcus*. Biodegradation 27:277–286. https://doi.org/10 .1007/s10532-016-9772-7.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Ferreira NL, Malandain C, Fayolle-Guichard F. 2006. Enzymes and genes involved in the aerobic biodegradation of methyl *tert*-butyl ether (MTBE). Appl Microbiol Biotechnol 72:252–262. https://doi.org/10.1007/ s00253-006-0494-3.
- He Y, Mathieu J, Yang Y, Yu P, da Silva ML, Alvarez PJJ. 2017. 1, 4-Dioxane biodegradation by *Mycobacterium dioxanotrophicus* PH-06 is associated with a group-6 soluble di-iron monooxygenase. Environ Sci Technol Lett 4:494–499. https://doi.org/10.1021/acs.estlett.7b00456.