

1 **IVF under COVID-19: treatment outcomes of fresh ART cycles**

2 **Running title:** SARS-CoV-2 infection and ART treatment outcomes

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23 **Abstract**

24 **Study question:** Does prior SARS-CoV-2 infection in women undergoing fertility treatments  
25 affect the outcomes of fresh ART cycles?

26 **Summary answer:** SARS-CoV-2 infection does not affect fresh ART treatment outcomes,  
27 except for a possible long term negative effect on oocyte yield (>180 days post infection).

28 **What is known already:** A single previous study suggested no evidence that a history of  
29 asymptomatic or mild SARS-CoV-2 infection in females caused impairment of fresh ART  
30 treatment outcomes.

31 **Study design, size, duration:** Retrospective cohort study, including all SARS-CoV-2  
32 infected women who underwent fresh ART cycles within a year from infection (the first cycle  
33 post infection), between October 2020 and June 2021, matched to non-diagnosed controls.

34 **Participants/materials, setting, methods:** Patients from two large IVF units in Israel who  
35 were infected with SARS-CoV-2 and later underwent fresh ART cycles were matched by age  
36 to non-diagnosed, non-vaccinated controls. Demographics, cycle characteristics and cycle  
37 outcomes, including oocyte yield, maturation rate, fertilization rate, number of frozen  
38 embryos per cycle, and clinical pregnancy rates, were compared between groups.

39 **Main results and the role of chance:** One hundred and twenty-one infected patients and  
40 121 controls who underwent fresh ART cycles were included. Oocyte yield (12.50 versus  
41 11.29;  $p=0.169$ ) and mature oocyte rate (78% versus 82%;  $p=0.144$ ) in all fresh cycles were  
42 similar between groups, as were fertilization rates, number of frozen embryos per cycle and  
43 clinical pregnancy rates (43% versus 40%;  $p=0.737$ ) in fresh cycles with an embryo transfer.  
44 In a logistic regression model, SARS-CoV-2 infection more than 180 days prior to retrieval  
45 had a negative effect on oocyte yield ( $p=0.018$ , Slope=-4.08, 95% CI -7.41 – -0.75), although  
46 the sample size was small.

47 **Limitations, reasons for caution:** A retrospective study with data that was not uniformly  
48 generated under a study protocol, no antibody testing for the control group.

49 **Wider implications of the findings:** The study findings suggest that SARS-CoV-2 infection  
50 does not affect treatment outcomes, including oocyte yield, fertilization and maturation rate,  
51 number of good quality embryos, and clinical pregnancy rates, in fresh ART cycles, except  
52 for a possible long term negative effect on oocyte yield when retrieval occurs > 180 days  
53 post SARS-CoV-2 infection. Further studies are warranted to support these findings.

54 **Study funding/competing interest(s):** None.

55 **Trial registration number:** 0010-21-HMC, 0094-21-ASF

56 **Key words:** IVF, COVID-19, infertility, SARS-CoV-2, pregnancy, oocytes

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## 71 Introduction

72 Corona Virus disease 19 (COVID-19) is caused by severe acute respiratory  
73 syndrome coronavirus-2 (SARS-CoV-2). SARS-CoV-2 enters target host cells via the cellular  
74 receptor angiotensin-converting enzyme 2 (ACE2) and the cellular transmembrane protease  
75 serine-2 (TMPRSS2) (Lukassen *et al.*, 2020). In theory, organs with a high expression of  
76 ACE2 or TMPRSS2 are more vulnerable to infection (Zou *et al.*, 2020). The COVID-19  
77 pandemic has raised concerns regarding the possible effect on human fertility, especially for  
78 couples undergoing fertility treatment. The male component has been the focus of most  
79 studies investigating the virus' effect on fertility, given the abundance of ACE2 receptors and  
80 TMPRSS2 in the testis tissue (Anifandis *et al.*, 2020; Jing *et al.*, 2020; Li *et al.*, 2020a,  
81 2020b; Gacci *et al.*, 2021; Guo *et al.*, 2021).

82 There is evidence that the renin–angiotensin–aldosterone system (RAS) is involved  
83 in female reproductive processes such as folliculogenesis, steroidogenesis, oocyte  
84 maturation and ovulation. The existence of the ACE2 axis and ACE2 markers were  
85 confirmed in all stages of follicular maturation in the human ovary, including the granulosa  
86 cells and follicular fluid (Reis *et al.*, 2011; Jing *et al.*, 2020; Anifandis *et al.*, 2021; Choi *et al.*,  
87 2021). ACE2 and TMPRSS2 are also expressed in the endometrium, possibly affecting  
88 implantation (Vaz-Silva *et al.*, 2009; Henarejos-Castillo *et al.*, 2020).

89 Furthermore, as with other viral infections, it can be assumed that SARS-CoV-2 may  
90 promote oxidative stress through oxidant-sensitive pathways, leading to activation of  
91 pathogenic mechanisms (Barzon *et al.*, 2017; Liu *et al.*, 2017; Khomich *et al.*, 2018).  
92 Increased oxidative stress may affect male fertility through reduction in motility and an  
93 increase in sperm DNA fragmentation (Bisht *et al.*, 2017; Agarwal *et al.*, 2018; Homa *et al.*,  
94 2019). Similarly, SARS-CoV-2 may affect oocyte performance through mechanisms that  
95 increase oxidative stress which has been associated with alterations in DNA methylation  
96 (Menezo *et al.*, 2016).

97           Given these considerations, it is reasonable to suspect that COVID-19 may affect  
98 oocyte performance or early implantation. Nevertheless, to date, the possible effects of  
99 COVID-19 on female fertility are largely unknown, and the effects on IVF outcomes has yet  
100 to be elucidated (Setti *et al.*, 2021; Wang *et al.*, 2021). In this study we aimed to evaluate  
101 the effect of female SARS-CoV-2 infection on the outcomes of IVF treatments in fresh  
102 cycles.

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## 105 **Materials and methods**

106 A retrospective cohort study, including all SARS-CoV-2 infected women aged 20-42  
107 years that underwent fresh IVF treatment cycles between January 1, 2021, and June 31,  
108 2021, at Shamir Medical Center and Herzliya medical center, Israel (COVID group). To be  
109 included in the study, maximal time from SARS-CoV-2 infection to treatment was defined as  
110 one year. Only the first cycle following recovery was included. The study was approved by  
111 the Institutional Review Board of both participating medical centers.

112 The study group was matched by age to the first following non-vaccinated patient  
113 with no history of past infection, who underwent IVF treatments at the same time period  
114 (October 2020- June 2021) (control group). Stimulation protocols, fertilization methods and  
115 embryo transfer parameters were individually tailored by the treating team, as per usual  
116 institutional routine. Demographic characteristics (including age, partner's age and COVID  
117 status, smoking status, number of previous pregnancies, deliveries and IVF treatments and  
118 infertility cause) as well as cycle characteristics (treatment protocol, overall gonadotropins  
119 (GT) administered, Estrogen levels on day of ovulation triggering (maximal E2), fertilization  
120 method and endometrial thickness) were recorded. Primary outcome measures were the  
121 mean number of retrieved oocytes per cycle and clinical pregnancy rates (defined as an  
122 intrauterine gestational sac on ultrasound imaging). Secondary outcomes included MII  
123 (mature oocyte) rates (MII/oocytes retrieved – in ICSI only cycles), fertilization rates  
124 (2PN/oocytes retrieved), and mean number of vitrified embryos. As varying time from  
125 infection to retrieval may have a different pathophysiologic effect on cycle outcomes, further  
126 stratification by time from SARS-CoV-2 infection to retrieval into groups of  $\leq 90$ , 90-180 and  
127  $> 180$  days was performed. For the purpose of pregnancy rates, fertilization rates and  
128 number of vitrified embryos, only women undergoing embryo transfer were included. Embryo  
129 grading was based on the Istanbul consensus workshop parameters (Balaban *et al.*, 2011).

## 130 **Data analysis**

131 Shapiro & Wilk test was used to test for normality of distribution. Continuous variables were  
132 summarized with mean and 95% CI and compared between groups using the Mann-Whitney  
133 test. Categorical variables were summarized using counts and percentages. The Fisher  
134 Exact Test or Chi-square test were used to compare differences between groups.

135 A logistic regression model was applied to identify factors associated with clinical pregnancy  
136 rates. Backwards elimination was applied to select the optimal model, while the age &  
137 COVID group were forced to be included in the model. To confirm the adequacy of the  
138 model we have applied the models including the minimal selected variables with similar  
139 results.

140 A linear regression model was applied to identify factors related to the total number of  
141 oocytes retrieved. No imputations for missing data were applied.

142 A two-sided  $P < 0.05$  was considered significant. R core team (2021). R: A language and  
143 environment for statistical computing. R foundation for statistical computing, Vienna, Austria.

144 Multivariate analyses were conducted using SPSS-27 software, IBM, Armonk, NY, USA.

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## 146 **Results**

### 147 **All cycles**

148 One hundred and twenty-one women in the study group and 121 women in the  
149 control group were included (Table I). The mean time from SARS-CoV-2 infection to oocyte  
150 retrieval was 84.5 days (SD 78.02; range 8-348). Mean age was similar in the study and  
151 control groups (33.3 versus 33.2 years respectively), as were mean partner's age, smoking  
152 rates and BMI. No differences were observed in the obstetrical history, infertility cause and  
153 number of prior IVF treatments. Patients in the study group and the control group had similar  
154 cycle characteristics in terms of stimulation protocol, total GT dosage, maximal E2 levels and  
155 endometrial thickness. The mean number of oocytes retrieved per cycle (12.50 versus  
156 11.29;  $p=0.169$ ) and the rate of mature oocytes in ICSI cycles (78 versus 82;  $p=0.144$ ) were  
157 similar between groups.

158 Similarly, a univariate analysis, with stratification by time from SARS-CoV-2 infection  
159 to treatment ( $\leq 90$  day, 90-180 days and  $> 180$  days), revealed no differences between  
160 groups in any of the parameters (Supplementary Table SI).

161 A linear regression model for oocyte yield in all patients including patients' age,  
162 previous transfer and past SARS-CoV-2 infection, demonstrated no effect of COVID-19  
163 status on oocyte yield ( $p=0.104$ ), while age remained a significant factor, reducing the  
164 number of oocytes by 0.64 for every additional year ( $p<0.001$ ) (Table II). In a sub analysis of  
165 the linear regression model according to time from SARS-CoV-2 infection (Table II), while  
166 age remained a significant factor, the COVID status was not significant in the first two  
167 groups ( $\leq 90$  day, 90-180 days). In the small subgroup (29 patients) with a past infection  $>$   
168 180 days, a negative effect on oocyte yield was observed ( $p=0.018$ , slope=-4.08, 95% CI -  
169 7.41 – -0.75). A Bonferroni correction for multiple comparisons attenuated this result  
170 ( $p=0.054$ ).

### 171 **Cycles with an embryo transfer**



172 Ninety-one of 121 women in the COVID group and 94 of 121 women in the control group  
173 underwent embryo transfer and were included in the pregnancy rate analysis (Table III). Of  
174 the 57 patients who did not undergo embryo transfer and were excluded from this analysis  
175 the majority were treated for fertility preservation (medical or social), underwent genetic  
176 testing, or had a hyper-response preventing embryo transfer. Only one patient from each  
177 group, both with an infertility diagnosis of premature ovarian insufficiency, did not undergo  
178 embryo transfer, without a pre-planned indication.

179 Demographic characteristics were similar in both groups (Supplementary Table SII).  
180 Partners' COVID status significantly differed between groups with higher rates of recovered  
181 and vaccinated partners in the COVID group ( $p < 0.001$ ). Cycle characteristics were similar  
182 between groups except for the fertilization method, with a higher intra-cytoplasmic sperm  
183 injection (ICSI) rate in the COVID group (70% versus 50%;  $p = 0.009$ ). Number of oocytes  
184 retrieved, mature oocytes, fertilization rates and number of vitrified embryos were similar  
185 between groups. Number of embryos transferred, and the day of transfer did not differ, but  
186 significantly more embryos graded C were transferred ( $p = 0.007$ ) in the control group with no  
187 difference in grade A and B embryos. Clinical pregnancy rates were similar between groups  
188 (43% versus 40%;  $p = 0.737$ ).

189 Stratifying by time from SARS-CoV-2 infection to treatment  $\leq 90$  day, 90-180 days  
190 and  $> 180$  days (Supplementary Table SIII), pregnancy rates (41% versus 30%,  $p = 0.19$ ;  
191 38% versus 67%,  $p = 0.063$ ; 58% versus 46%,  $p = 0.54$  respectively), mature oocytes,  
192 fertilization rate and number of vitrified embryos were similar between the COVID and  
193 control groups.

194 A backwards multivariate logistic regression model for pregnancy rate (Table IV)  
195 (including age, previous transfers, number of embryos transferred, day of transfer, embryo  
196 grade, endometrial thickness, number of oocytes retrieved, and fertilization method) was  
197 performed showing no effect of past SARS-CoV-2 infection on pregnancy rates ( $p = 0.889$ ).  
198 Patient age and endometrial thickness were the only significant variables. The same model

199 was applied for patients having an embryo transfer within 90 days of SARS-CoV-2 infection,  
200 with patient age being the only significant variable (Table IV). The groups of patients with  
201 SARS-CoV-2 infection 90-180 days and > 180 days before transfer were too small for  
202 inclusion in the model.

203

## 204 Discussion

205 In this retrospective cohort study, past infection with SARS-CoV-2 had no impact on  
206 fresh IVF treatment outcomes in terms of oocyte yield, maturation rate, fertilization rate,  
207 number of vitrified embryos and clinical pregnancy rates, except for a possible long-term  
208 effect on oocyte yield (retrieval > 180 days post infection).

209 The COVID-19 pandemic has had a profound psychosocial impact on fertility  
210 patients, which was especially apparent at the beginning of the pandemic, when fertility  
211 treatments were suspended in many countries (Ben-Kimhy *et al.*, 2020; Boivin *et al.*, 2020;  
212 Marom Haham *et al.*, 2021). The purpose of our study was to examine whether, in addition,  
213 there was a measurable effect of SARS-CoV-2 infection on fertility treatment outcomes. To  
214 the best of our knowledge, this is the largest study to date reporting the effect of prior SARS-  
215 CoV-2 infection on fertility treatment outcomes in fresh ART cycles.

216 SARS-CoV2 enters cells via the ACE-2 cellular receptor and the TMPRSS2 cellular  
217 protease. Those are expressed in all stages of follicular maturation in the human ovary, in  
218 the granulosa cells and in the endometrium (Reis *et al.*, 2011; Jing *et al.*, 2020; Anifandis *et*  
219 *al.*, 2021; Choi *et al.*, 2021). In this study, the effect on the follicular development, maturation  
220 and ovulation was evaluated by oocyte yield, maturation and number of good quality  
221 embryos (vitrified). Further effects on embryo development and implantation were evaluated  
222 by pregnancy rates.

223 We found that recent past infection with SARS-CoV-2 (<180 days) had no influence  
224 on treatment outcomes in terms of oocyte yield and maturation. This result is in line with the  
225 limited literature published to date (Setti *et al.*, 2021; Wang *et al.*, 2021). As the dominant  
226 follicle originates from a primordial follicle that has been recruited up to one year earlier  
227 (Gougeon, 1986; Erickson and Shimasaki, 2001), we stratified the participants by time from  
228 infection to evaluate different possible mechanisms of influence. Shortly after the infection, in  
229 addition to direct viral cell invasion, possible oxidative stress may affect ovarian function,

230 compromising antral and preovulatory follicular growth and development. Furthermore,  
231 endometrial cellular damage seems more likely in proximity to the acute infection. We  
232 hypothesized that a possible differential effect in oocyte yield and maturation would be  
233 apparent for more than 90 days after infection in case the growing follicle was damaged in its  
234 earlier developmental stages. Nevertheless, no difference was observed in either outcome  
235 when the infection occurred up to six months prior to treatment (sub-groups ( $\leq 90$  and 90-  
236 180 days). These results were consistent in all regression models performed, including the  
237 linear model for oocyte yield and the logistic regression model for pregnancy rates, providing  
238 reassurance that recent SARS-CoV-2 infection does not compromise IVF treatment  
239 outcomes.

240 Further analysis of women infected more than 180 days prior to treatment was  
241 performed to assess for long-term effects as a result of possible damage to the primordial  
242 follicles or during the initial recruitment process. In this subgroup of patients, lower oocyte  
243 yields were observed, while all other parameters were unaffected. It should be noted  
244 however that the sample size was small, thus cautious interpretation of the results is  
245 warranted.

246 In univariate analyses of cycles with embryo transfer, a significant difference was  
247 observed in the fertilization method. ICSI was more commonly used in the COVID group  
248 whereas IVF was more commonly used in the control group, despite the fact that there was  
249 no difference in infertility causes. This may possibly be explained by the lab evaluation of  
250 partner sperm prior to fertilization. Partners' COVID status was, as expected, more likely to  
251 show past infection in the COVID group. Prior studies have reported a decrease in sperm  
252 parameters, potentially explaining the higher ICSI rates in the COVID group. However,  
253 fertilization method was not found to be significantly associated with the pregnancy rate in  
254 the multivariate regression model.

255 The strength of our study is our relatively large sample of women that tested positive  
256 for SARS-CoV-2, allowing us to evaluate different stages of the IVF procedure, including

257 oocyte development, maturation and embryo implantation, at different time points after the  
258 acute infection. The main limitation of the study is its retrospective nature, with the inherent  
259 biases of collecting data that was not uniformly generated under a study protocol. Another  
260 caveat is the lack of sperm analyses, which, especially given the possible effect of SARS-  
261 CoV-2 infection on sperm parameters and significant difference in rates of recovered  
262 partners between groups, may explain the significant difference in the fertilization method  
263 utilized. Even so, fertilization rates and embryo grade were similar between groups. Another  
264 limitation is the fact that women in the control group were chosen based on medical history  
265 and did not undergo antibody testing.

266           In conclusion, recent past SARS-CoV-2 infection, less than 180 days prior to IVF  
267 treatment, did not affect oocyte yield, fertilization, maturation and clinical pregnancy rates in  
268 fresh IVF cycles. A possible long-term effect (>180 days) compromising oocyte yield was  
269 observed. Further studies are warranted in order to support these findings with special  
270 attention to the long-term effects.

271

272 **Data availability:** The data underlying this article will be shared on reasonable request to  
273 the corresponding author.

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277 collection, M.Y, A.H, A.K drafted the first version of the manuscript. M.Y, A.H, G.Y, RS  
278 contributed to data analysis and interpretation. All authors revised the manuscript and  
279 approved the final submitted version.

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373 **Table I:** Demographic and cycle characteristics and outcomes of COVID versus control group in fresh  
 374 cycles. Data are presented as mean and (SD) and [range] or counts and (percentage)

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Group	COVID-19 (N=121)	Non COVID-19 (N=121)	p value
Patient age (y)	33.3 (5.37) [21-42]	33.23 (5.33) [22-42]	0.896
Partners age (y)	35.78 (6.90) [22-55]	34.39 (5.45) [21-48]	0.2
Smoker	14 (12%)	17 (15%)	0.445
Previous retrievals	1.07 (1.60) [0- 8]	1.19 (1.48) [0.00 - 8.00]	0.156
Previous transfers	1.11 (2.18) [0 - 12]	1.25 (1.92)	0.087
BMI	25.25 (5.55) [16.23 - 42.97]	25.48 (5.86) [16.53 - 42.45]	0.959
Infertility cause (N)	110	102	0.209
Age related	14 (13%)	19 (19%)	
Male factor	32 (29%)	35 (34%)	
Ovulation	6 (5%)	11 (11%)	
Mechanical	11 (10%)	4 (4%)	
Unexplained	27 (25%)	17 (16%)	
Fertility preservation	12 (11%)	8 (8%)	
Other	8 (7%)	8 (8%)	
Parity (N)	104	95	0.519
0	71 (68%)	66 (70%)	
1	19 (18%)	21 (22%)	

>=2	14 (14%)	8 (8%)	
<b>Gravidity (N)</b>	105	93	0.993
0	65 (62%)	55 (59%)	
1	18 (17%)	21 (23%)	
>=2	22 (21%)	17 (18%)	
<b>Days from COVID to oocyte retrieval</b>	84.54 (78.02) [8-348]	NA	
<=90	77 (64%)	NA	
>90-180	29 (24%)	NA	
>180	15 (12%)	NA	
<b>Protocol</b>			0.177
Antagonist	106 (88%)	104 (87%)	
Long luteal	6 (5%)	6 (5%)	
MNC	1 (1%)	6 (5%)	
Short (flare)	8 (6%)	4 (3%)	
<b>Gonadotropins dosage (IU)</b>	2524 (1317)600- 7800	2335 (1220)348 - 6600	0.255
<b>Max. E2 (pmol/L)</b>	8584 (6191) [1337–31650]	8842 (6415) [456–35898]	0.824
<b>Endometrial thickness (mm)</b>	10.53 (2.29) [4.5 - 17.1]	9.97 (2.29) [4.6 – 16]	0.108
<b>Oocytes retrieved</b>	12.50 (7.83) [0 – 40]	11.29 (7.60) [1 – 39]	0.169
<b>Fertilization method (N)</b>	112	113	0.004
ICSI	82 (73%)	60 (53%)	0.002*
ICSI/IVF	25 (22%)	39 (34%)	0.043*
IVF	5 (5%)	14 (12%)	0.033*
<b>MII /oocytes%(ICSI only)(N)</b>	80	60	0.144
	78 (18.03) [25 – 100]	82 (18.86) [33.33- 100]	
<b>Total available embryos</b>	3.41 (2.71) [0-13]	3.72 (2.77) [0-16]	0.398
<b>Partner COVID status (N)</b>	76	84	<0.001
Recovered	48 (63%)	0	
Vaccinated	17 (22%)	9 (11%)	0.046*
Non	11 (15%)	75 (89%)	<0.001*

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377 \*Post hoc analysis

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380 **Table II:** Linear Regression model for number of oocytes retrieved – total sample and  
 381 subdivided by time from COVID-19  
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Patients included	N	Variables	P value	Slope	Lower 95% CI	Upper 95% CI
<b>All</b>	227	Group	0.164	1.285	-0.53	3.10
		Patient age	<0.001	-0.638	-0.82	-0.46
		Previous retrievals	0.810	-0.075	-0.69	0.54
<b>Days ≤ 90</b>	142	Group COVID versus control	0.172	1.70	-0.75	4.14
		Patient age	<0.001	-0.61	-0.83	-0.39
		Previous retrievals	0.815	-0.10	-0.98	0.77
<b>90-180</b>	56	Group COVID versus control	0.125	2.81	-0.805	6.432
		Patient age	0.001	-0.69	-1.086	-0.302
		Previous retrievals	0.362	0.54	-0.638	1.721
<b>&gt;180</b>	29	Group COVID versus. control	0.018	-4.08	-7.41	-0.75
		Patient age	0.055	-0.51	-1.04	0.01
		Previous retrievals	0.061	-0.96	-1.97	0.05

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400 **Table III:** Cycle characteristics and outcomes of COVID versus control group - fresh embryo  
 401 transfer cycle. Data are presented as mean and (SD) and [range] or counts and (percentage).  
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Group	COVID-19 (N=91)	Non COVID-19 (N=94)	p value
<b>Gonadotropin dosage (IU)</b>	2529.42 (1418) [600-7800]	2334.70 (1269) [600-6600]	0.365
<b>Max. E2 (pmol/L)</b>	7598 (5375) [1337-28382]	7510 (5151) [456-25022]	0.939
<b>Endometrial thickness (mm)</b>	10.75 (2.25) [6-17]	9.99 (2.22) [5.4-15]	0.062
<b>Oocytes retrieved</b>	11.26 (6.19) [1-33]	10.04 (6.90) [1-31]	0.085
<b>Fertilization method</b>	91	93	0.009
ICSI	64 (70%)	46 (50%)	
ICSI/IVF	22 (24%)	33 (35%)	
IVF	5 (6%)	14 (15%)	
<b>Percent MII /oocytes (ICSI) (N)</b>	64	46	0.072
	77.60 (18.87) [25-100]	83.08 (19.98) [33-100]	
<b>Fertilization rate</b>	0.59 (0.24) [0.07-1]	0.62 (0.26) [0-1]	0.365
<b>Total frozen embryos</b>	1.71 (2.40) [0-15]	2.12 (2.34) [0-11]	0.168
<b>No. of embryos transferred</b>			0.545
1	57 (63%)	63 (67%)	
2	30 (34%)	30 (32%)	
3	3 (3%)	1 (1%)	
<b>Day of transfer</b>			0.252
2	16 (17%)	26 (28%)	
3	57 (63%)	53 (56%)	
5	18 (20%)	15 (16%)	
<b>Embryo grade</b>			0.015
A	52 (58%)	54 (57%)	
B	35 (39%)	26 (28%)	
C	3 (3%)	14 (15%)	0.007*
<b>Clinical pregnancy</b>	39 (43%)	38 (40%)	0.737
<b>Partner COVID status (N)</b>	63	68	<0.001
Recovered	40 (63%)	0	
Vaccinated	13 (21%)	7 (10%)	
Non	10 (16%)	61 (90%)	

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\*Post hoc analysis

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410 **Table IV:** Logistic regression model for clinical pregnancy rates in fresh embryo transfer –  
 411 Whole group and subgroup of patients with a recent infection  
 412

Patients included	N	%	Variables	P value	Odds Ratio	Lower 95% CI	Upper 95% CI
All	154	83	Previous transfers	0.302	0.895	0.724	1.105
			Embryo grade	0.226			
			Embryo grade B versus A	0.267	0.659	0.315	1.377
			Embryo grade C versus A	0.135	0.330	0.077	1.414
			Endometrial thickness	0.031	1.194	1.017	1.402
			Group COVID vs control	0.889	1.052	0.517	2.140
			Patient age >39y versus <=39y	0.040	0.249	0.066	0.939
			Number of embryos transferred (1 versus 2+3)	0.351	1.457	0.661	3.215
≤90 days from COVID	95	81	Embryo grade	0.260			
			Embryo grade B versus A	0.960	0.975	0.366	2.602
			Embryo grade C versus A	0.104	0.162	0.018	1.452
			Endometrial thickness	0.122	1.173	0.958	1.435
			Group COVID versus control	0.320	1.596	0.636	4.008
			Patient age >39y versus <=39y	0.037	0.179	0.036	0.903
			Number of embryos transferred (1 versus. 2+3)	0.933	1.045	0.376	2.904

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