





Draft Genome Sequence of "Candidatus Arthromitus" UMNCA01, a Suspected Commensal Isolated from the Gut Microbiome of Commercial Turkey

Grant A. Hedblom,^a Kamal Dev,^{a,b} Steven D. Bowden,^{a,d} Bonnie Weber,^e Sally Noll,^f David J. Baumler,^{a,c,d} Timothy J. Johnson^e

ABSTRACT "Candidatus Arthromitus" UMNCA01 was recovered from ileal samples of commercial turkey poults and may have probiotic capabilities. The complete genome was determined using the Illumina MiSeq and HiSeq sequencing platforms. The complete genome consists of 1,631,326 bp and has a G+C content of 26.14%, 1,540 coding sequences (CDS), and 37 RNA coding genes.

candidate genus of segmented filamentous bacteria, "Candidatus Arthromitus" belongs to the family Clostridiaceae. These commensal organisms promote adaptive and innate immune responses in murine models in a host-specific manner, prevent diseases, and promote animal growth (1, 2). Different strains of "Candidatus Arthromitus" inhabit the ileum region in many vertebrate animals, such as cattle, pigs, chickens, humans, and, as shown more recently, turkeys (3). In an attempt to discern the microbial basis of light turkey syndrome (LTS), a condition where commercial turkey flocks fail to meet their genetic potential weights despite standardized diets and growth conditions (4), Danzeisen et al. performed 165 rRNA sequencing of intestinal microbiome analysis of high-performing and low-performing (based upon flock weights) turkey flocks (5). This analysis revealed that at the age of 2 to 3 weeks, high-performing turkey flocks harbored significantly higher proportions of "Candidatus Arthromitus" bacteria than their low-performing counterparts (5). In this study, the genome of a turkey-specific strain of "Candidatus Arthromitus" was sequenced from the gut microbiome.

"Candidatus Arthromitus" UMNCA01, a Gram-positive bacterium, was recovered from ileal samples harvested from 2-week-old turkey poults from a research turkey flock in barns at the University of Minnesota. The sample was identified for shotgun sequencing by previous 16S rRNA amplicon profiling indicating a high relative abundance of "Candidatus Arthromitus" bacteria and light microscopy confirming the presence of high levels of segmented filamentous bacteria. For metagenomic shotgun sequencing, the total genomic DNA was isolated using a Qiagen stool kit (Hilden, Germany). The quantity of the genomic DNA was determined by measuring A_{260} using a UV-visible (UV-Vis) spectrophotometer ($A_{260} = 1$ corresponds to $50 \, \text{ng}/\mu\text{l}$ of double-stranded DNA [dsDNA]). The quality of the genomic DNA was determined by measuring the A_{260}/A_{280} ratio, and a value of 1.8 indicated pure DNA preparation as described (6). Twenty micrograms of metagenomic DNA was used to prepare a paired-end (PE) sequencing library (Nextera XT, Illumina, San Diego, CA), and a PCR amplified library was sequenced using the Illumina MiSeq and HiSeq platforms. The shotgun data were assembled using CLC Genomics Workbench v. 9.0/APRIL-2016, with default parameters, and then contigs were mapped to an existing mouse

Citation Hedblom GA, Dev K, Bowden SD, Weber B, Noll S, Baumler DJ, Johnson TJ. 2020. Draft genome sequence of "Candidatus Arthromitus" UMNCA01, a suspected commensal isolated from the gut microbiome of commercial turkey. Microbiol Resour Announc 9:e01143-19. https://doi.org/10.1128/

Editor David Rasko, University of Maryland School of Medicine

Copyright © 2020 Hedblom et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to David J. Baumler, dbaumler@umn.edu.

Received 13 September 2019 **Accepted** 26 December 2019 **Published** 23 January 2020

^aDepartment of Food Science and Nutrition, University of Minnesota—Twin Cities, St. Paul, Minnesota, USA

^bFaculty of Applied Sciences and Biotechnology, Shoolini University, Solan, Himachal Pradesh, India

^cMicrobial and Plant Genomics Institute, University of Minnesota—Twin Cities, St. Paul, Minnesota, USA

^dBiotechnology Institute, University of Minnesota—Twin Cities, St. Paul, Minnesota, USA

eDepartment of Veterinary and Biomedical Sciences, University of Minnesota—Twin Cities, St. Paul, Minnesota, USA

^fDepartment of Animal Science, University of Minnesota—Twin Cities, St. Paul, Minnesota, USA

Hedblom et al.

♠ Microbiology

TABLE 1 Global statistics of the "Candidatus Arthromitus" UMNCA01 genome

Parameter ^a	Value
Total sequence length (bp)	1,631,326
No. of genes (total)	1,577
No. of CDS (total)	1,540
No. of genes (coding)	1,480
No. of CDS (coding)	1,480
No. of genes (RNA)	37
No. of rRNAs (16S)	1
No. of partial rRNAs (16S)	1
No. of tRNAs	33
No. of ncRNAs	3
No. of pseudogenes	60
No. of scaffolds	41
Scaffold N_{50} (bp)	68,513
Scaffold L_{50} (bp)	9
No. of contigs	44
Contig N ₅₀ (bp)	57,760
Contig L_{50} (bp)	10

^a CDS, coding DNA sequences; ncRNAs, noncoding RNAs.

"Candidatus Arthromitus" genome using Mauve (7) to retrieve and arrange "Candidatus Arthromitus" sequences (sourced from turkeys) that mapped to those genomes. Following manual curation, unmapped contigs were then filtered from the metagenomic assembly. The final "Candidatus Arthromitus" assembly resulted in an average $100 \times \text{genome}$ coverage with a total number of 1,631,326 bp arranged into 41 contigs. The G+C content of these contigs was 26.14%, with an average contig size of 39,788 bp and an N_{50} value of 57,760 bp. The draft genome contains 1,480 protein coding sequences, 37 RNA genes, and 60 pseudogenes. The genome sequence of "Candidatus Arthromitus" UMNCA01 was annotated using the National Center for Biological Information (NCBI) Prokaryotic Genome Annotation Pipeline and the best-placed reference protein set of GeneMarkS+ (annotation software v. 4.6) as described (8, 9); the results are summarized in Table 1.

Data availability. This "Candidatus Arthromitus" UMNCA01 whole-genome shotgun (WGS) project has the GenBank accession number NZ_LXFF00000000. The version of this project is NZ_LXFF01000000 and consists of sequences LXFF01000001 through LXFF010000041. The filtered assembly and raw sequencing reads can be accessed through BioProject accession number PRJNA319431 and BioSample accession numbers SAMN04889864 and SAMN13392129, respectively.

ACKNOWLEDGMENTS

We acknowledge Holly Reiland for assistance in surveying the genome contents. This work was funded by an Agriculture and Food Research Initiative competitive grant (2016-67015-24911) from the USDA National Institute of Food and Agriculture and by the University of Minnesota through Global Food Ventures.

REFERENCES

- Hedblom GA, Reiland HA, Sylte MJ, Johnson TJ, Baumler DJ. 2018. Segmented filamentous bacteria—metabolism meets immunity. Front Microbiol 9:1991. https://doi.org/10.3389/fmicb.2018.01991.
- Schnupf P, Gaboriau-Routhiau V, Sansonetti PJ, Cerf-Bensussan N. 2017. Segmented filamentous bacteria, Th17 inducers and helpers in a hostile world. Curr Opin Microbiol 35:100–109. https://doi.org/10.1016/j.mib.2017 .03.004.
- Ericsson AC, Hagan CE, Davis DJ, Franklin CL. 2014. Segmented filamentous bacteria: commensal microbes with potential effects on research. Comp Med 64:90–98.
- Mor SK, Sharafeldin TA, Abin M, Kromm M, Porter RE, Goyal SM, Patnayak DP. 2013. The occurrence of enteric viruses in light turkey syndrome. Avian Pathol 42:497–501. https://doi.org/10.1080/03079457.2013.832145.
- Danzeisen JL, Calvert AJ, Noll SL, McComb B, Sherwood JS, Logue CM, Johnson TJ. 2013. Succession of the turkey gastrointestinal bacterial micro-

- biome related to weight gain. PeerJ 1:e237. https://doi.org/10.7717/peerj 237
- Sambrook J, Fritsch FF, Maniatis T. 1989. Molecular cloning: a laboratory manual, 3rd ed. Cold Spring Harbor Laboratory, Cold Spring Harbor, NY.
- Darling AC, Mau B, Blattner FR, Perna NT. 2004. Mauve: multiple alignment of conserved genomic sequence with rearrangements. Genome Res 14:1394–1403. https://doi.org/10.1101/gr.2289704.
- Ewing B, Hillier L, Wendl MC, Green P. 1998. Base-calling of automated sequencer traces using Phred. I. Accuracy assessment. Genome Res 8:175–185. https://doi.org/10.1101/gr.8.3.175.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44:6614–6624. https://doi.org/10.1093/nar/gkw569.

Volume 9 lssue 4 e01143-19 mra.asm.org **2**