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Association between single nucleotide polymorphisms in the mu opioid receptor gene (*OPRM1*) and self-reported responses to alcohol in American Indians

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Abstract

Background: Variation in response to the hedonic and adverse effects of a substance is in part an inherited factor that may influence its use, abuse and dependence. The mu opioid receptor is the primary site of action for opiates and individuals with polymorphisms in this receptor appear to have variation in the CNS effects of opiates. Several studies have suggested that this receptor may also mediate some of the effects of non-opioid drugs, such as alcohol. The purpose of this study was to investigate associations between 13 single nucleotide polymorphisms in the mu opioid receptor gene (*OPRM1*) with self-reported responses to alcohol, an endophenotype associated with the development of alcohol dependence, in American Indians living on eight contiguous reservations.

Methods: Each participant gave a blood sample and completed a structured diagnostic interview. Additionally, response to alcohol was indexed using the expectation version of the subjective high assessment scale (SHAS-E). SNPs were genotyped in 251 participants and data analyses were conducted using SOLAR.

Results: The estimated heritability (h^2) for the SHAS-E phenotypes ranged from 0.01 to 0.28. Endorsing the expectation of a *more* intense response on one or more of the following items from the SHAS-E: buzzed, clumsy, dizzy, drunk, effects, high, nausea, sleepy, talkative, terrible, and/or uncomfortable after imbibing 2–3 drinks was significantly associated with having at least one minor allele for at least one of 7 SNPs ($p < 0.01$) in the *OPRM1* receptor gene.

Conclusion: These studies provide data to suggest that the minor allele, for most of the polymorphisms in the *OPRM1* receptor gene investigated, was found to be associated with a more intense, and/or more adverse, response to alcohol, traits that are significantly correlated with lowered quantity of alcohol consumption and less susceptibility to dependence in this Indian population. These data further suggest that making conclusions on the role of the mu opioid receptor gene in the development of alcohol dependence may be limited if only one polymorphism in the gene is evaluated in isolation.

Background

A number of studies have documented that the dosage requirements for targeted effects of CNS drugs can vary widely [1]. For example, in a study of over 3,000 patients experiencing pain following postoperative hip replacement, the therapeutic morphine dosage requirements varied almost 40-fold [2]. Wide inter-patient variability in response to, and therefore in the dosage requirement for morphine have been demonstrated in cancer patients receiving morphine for pain control [3].

It appears that a number of genetic and environmental factors can lead to significant variation in the doses of a drug necessary to produce therapeutic, hedonic and/or adverse effects. However, there is increasing evidence that gene polymorphisms may be an important factor in determining a person's sensitivity and tolerance to a drug. The mu opioid receptor (*OPRM1*) is the primary site of action for opiates; about 20 variants in the mu opioid receptor gene (*OPRM1*) have been identified with amino acid substitutions that have polymorphic frequencies over 1% [4-11]. The most common single nucleotide polymorphism (SNP) reported on is A118G (rs1799971), which encoded the Asp40Asn codon change with most data suggesting that it is a functional variant [4,12].

There have been a series of studies in both healthy volunteers and in clinical patients suggesting that, the A118G variant may alter response to opioid drugs (see [1] for review). Lotsch and colleagues reported the 118G allele conferred smaller analgesic effects and produced less pupillary constriction during morphine and morphine-6-glucuronide (MG6) infusion [13-15]. In an experiment using a measure of pain tolerance to electrical stimulation, higher MG6 concentrations were associated with a 25% increase in current (C25) participants with the 118G allele [16,17]. Similar findings have been found for alfentanil [18] and levomethadone [19]. In clinical studies, data from patients with the 118G polymorphism tend to confirm data from experimental pain studies where those patients with the variant required higher alfentanil doses for analgesia or more morphine during colorectal surgery [20] or for pain/toxicity associated with morphine use in renal failure [13,14]. However, it appears that the effects may be drug or disease specific owing to presumed variation in environmental and/or other uncontrolled variables [1,21,22].

Several studies have suggested that the mu receptor may also mediate some of the hedonic and/or addictive effects of non-opioid drugs, such as alcohol [23,24]. Indirect support for this hypothesis is provided by studies demonstrating the efficacy of naltrexone for the treatment of alcohol dependence [25-31]. Further support is provided by studies evaluating associations between response to

naltrexone pharmacotherapy for alcohol dependence and the presence of the A118G variant. In a study that combined data from three different clinical trials, Oslin and colleagues [32] demonstrated that carriers of the 118G allele had a significantly lower rate of relapse and a longer time to a return to heavy drinking when compared to those individuals who were homozygous for the 118A allele. This finding was not supported in the Veterans Affairs (VA) Cooperative Study where no significant interactions were found between naltrexone treatment response and any polymorphic variants at each of the three opioid receptor genes [33]. More recently, data from the Study for the Combined Pharmacotherapies and Behavioral Interventions for Alcohol Dependence (COMBINE) study demonstrated that treatment with naltrexone produced a significantly improved clinical global outcome in alcohol dependent participants with the 118G allele, as compared to those with the 118A allele [34]. The A118G polymorphism has also been associated with an individual's response to a naloxone challenge with subjects with the 118G allele showing higher plasma cortisol concentrations [35,36].

There has been a plethora of studies that have investigated the relationship between a diagnosis of drug and/or alcohol dependence and the A118G polymorphism. The results have been conflicting and inconsistent. In a recent meta-analysis of 28 different studies, including over 8000 subjects, the conclusion was that the *OPRM1* A118G variant did not appear to affect risk for substance dependence. However, the authors further speculated that additional research would be needed to determine whether another polymorphism in the gene might influence receptor function and thus risk for substance dependence [37]. An additional feature of these studies, that may have weakened the results, is the use of a dichotomous phenotype, drug dependence, a diagnosis that is made based on both heritable and non-heritable factors [38]. Town and colleagues [39] suggested that genetic studies on the influence of mu opioid receptors polymorphisms be viewed within the broader context of alcoholism where the opioid receptor genes are taken to be partial, rather than complete, risk factors for the disorder. Thus, it may be that polymorphisms in *OPRM1* encode for a variant that influences a more narrowly defined risk factor for alcoholism. This risk factor is envisioned to partially influence the development of the disorder, but may or may not ultimately be associated with the diagnosis depending on the age of the participant, presence of other risk factors and environmental variables.

Individual sensitivity to alcohol represents such an inherited factor that affects the likelihood of drinking and mediates the disposition for developing alcoholism [40], and has a strong genetic basis [41]. In general, people at

higher genetic risk for alcoholism are less sensitive to the effects of alcohol and people at lower genetic risk for alcoholism are more sensitive. Support for this theory is provided by many, but not all, studies examining the reaction to alcohol among children of alcoholics, who are at greatly elevated risk for developing alcoholism [42]. Results have indicated that at moderate doses of alcohol, subjects who are family history positive for alcoholism and subjects who are family history negative for alcoholism attain equivalent blood alcohol concentrations, but most studies have found that subjects with a positive family history rate themselves as significantly less intoxicated than control subjects with a negative family history [43-46]. Although not all studies agree [47], a meta-analysis focusing on subjective level of intoxication confirmed a diminished response to alcohol as a characteristic more frequently seen in subjects with a positive family history than in those with a negative family history [48]. In addition, an 8-year follow-up of previously studied men with positive and negative family histories found that both a family history of alcoholism and a low response to alcohol were related to the development of alcohol-related problems [49].

Studies using similar methodologies among groups at lower risk for alcoholism have provided additional support for the idea that individual sensitivity to alcohol might also mediate protection from developing alcoholism. Individuals of Asian heritage, who have mutations in the aldehyde dehydrogenase gene (ALDH2) [50-53], and individuals of Jewish descent [54], two groups with low rates of alcoholism, were found to have more intense, although not necessarily more negative, responses to alcohol than matched control subjects of average alcoholism risk.

Genetic studies of complex phenotypes, such as sensitivity to alcohol, often have advantages when they are conducted in well-defined populations such as Native American tribes living on reservations [55]. A once popular notion, called the firewater myth, proposed that Native American Indians are constitutionally predisposed to an altered response to drinking alcohol [56]. In one empirical study, Native American Indians, like Caucasian sons of alcoholics, were found to have less intense objective and subjective effects of alcohol in an alcohol challenge paradigm. Additionally, participants with at least 50% Native American heritage reported less intense effects of alcohol than did those with less than 50% Native American heritage, despite equivalent blood alcohol concentrations [57-59].

The present report is part of a larger study exploring risk factors for substance dependence among Native American Indians [57-68]. The lifetime prevalence of substance

dependence in this Indian population is high and evidence for heritability and linkage to specific chromosome locations has been demonstrated [65,69-72]. The purpose of the present set of analyses was to determine if a significant association could be detected between 13 SNPs in the *OPRM1* receptor gene and self-report of subjective response to alcohol in this population.

Methods

Participants, who were of mixed heritage but at least one-sixteenth Native American, were recruited from eight geographically contiguous reservations with a total population of about 3,000 individuals. They were recruited using a combination of a venue-based method [73,74], and a respondent-driven procedure [75], as described previously [64,76]. To be included in the study a participant had to be an American Indian of one of four tribal groups between the age of 18 and 70 without major medical problems that would preclude mobility.

Potential participants gave written informed consent using a protocol approved for the study by The Institutional Review Board (IRB) of The Scripps Research Institute, the Scientific Advisory Committee of the GCRC, and the Indian Health Council, a tribal review group overseeing health issues for the reservations where recruitment was undertaken. They also responded to a screening questionnaire that was used to gather information on demographics, personal medical history, ethnicity and detailed measures of substance abuse history [77] and weight & height. Each participant also completed an interview with the Semi-Structured Assessment for the Genetics of Alcoholism (SSAGA) [78], which was used to make diagnoses [79]. Response to alcohol was assessed using the subjective high assessment scale-expectations version (SHAS-E). This scale consists of 14 items rated on Likert scales ranging from 0 (normal) to 36 (extreme effect). The participants indicated how intoxicated they felt after drinking 2-3 drinks for the following items: buzzed, clumsy, dizzy, drunk, effects of alcohol, energy, good, high, nausea, sleepy, talkative, uncomfortable, terrible overall and great overall. A total score was also calculated for the first 12 SHAS-E items. The intersession reliability of the SHAS, from which the SHAS-E was constructed, is approximately 0.80 with a Cronbach alpha of 0.96 overall [80]. The items cluster together with an overall item-to-total correlation of 0.80 or higher and a Cronbach alpha of 0.96. The Cronbach alpha for the SHAS-E is also 0.96 and values on the SHAS-E have been demonstrated to significantly ($p < 0.0001$) correlate with responses on the alcohol challenge SHAS at 30 and 60 minutes ($r^2 = 0.49$, 30 min, $r^2 = 0.51$, 60 min) (Schuckit and Smith, personal communication).

Two hundred (251) individuals have both genotype and phenotype data for this analyses. Power analyses revealed

that for a medium effect size (0.5) that power at this n would be equal to 0.976. DNA was isolated from whole blood using an automated DNA extraction procedure. All primers, probes and reagents were purchased from ABI (Applied Biosystems, Foster City, CA). SNPs were genotyped using TaqMan™ fluorescence 5' exonuclease technology. Each 5 microL reaction contained 25 ng genomic DNA, 1.6× TaqMan assay primer/probe mix, 1× PCR Buffer A, 2.5 mM MgCl₂, 250 microM dNTPs, and 0.5 U AmpliTaq Gold polymerase. Thermocycling was performed as recommended by ABI. Genotypes were determined on an ABI 7900 HT Fast Real-Time PCR System using the allelic discrimination mode. Hardy-Weinberg equilibrium analyses were completed in Haploview (version 4.0) [81].

Since genotype data was not available from the International HapMap Project public database[82] at the time this study was conceived (October 2004), seventeen single nucleotide polymorphisms (SNPs) in or near the *OPRM1* locus were selected from the Applied Biosystems SNP database [83]. SNPs were initially chosen to be evenly distributed across *OPRM1* with an average intermarker spacing of 5,133 bp. Assays for three SNPs (rs561720, hCV32237184, rs3798687) failed and were excluded from analyses. One SNP, rs12333298 in intron 1, showed significant ($p = 0.018$) deviations from Hardy-Weinberg equilibrium (HWE) at a $p < 0.05$ level and was also dropped from further analyses. The locations of the thirteen remaining SNPs typed in the study are shown in Figure 1 and SNP information, including the observed minor allele frequency (MAF), is described in Table 1.

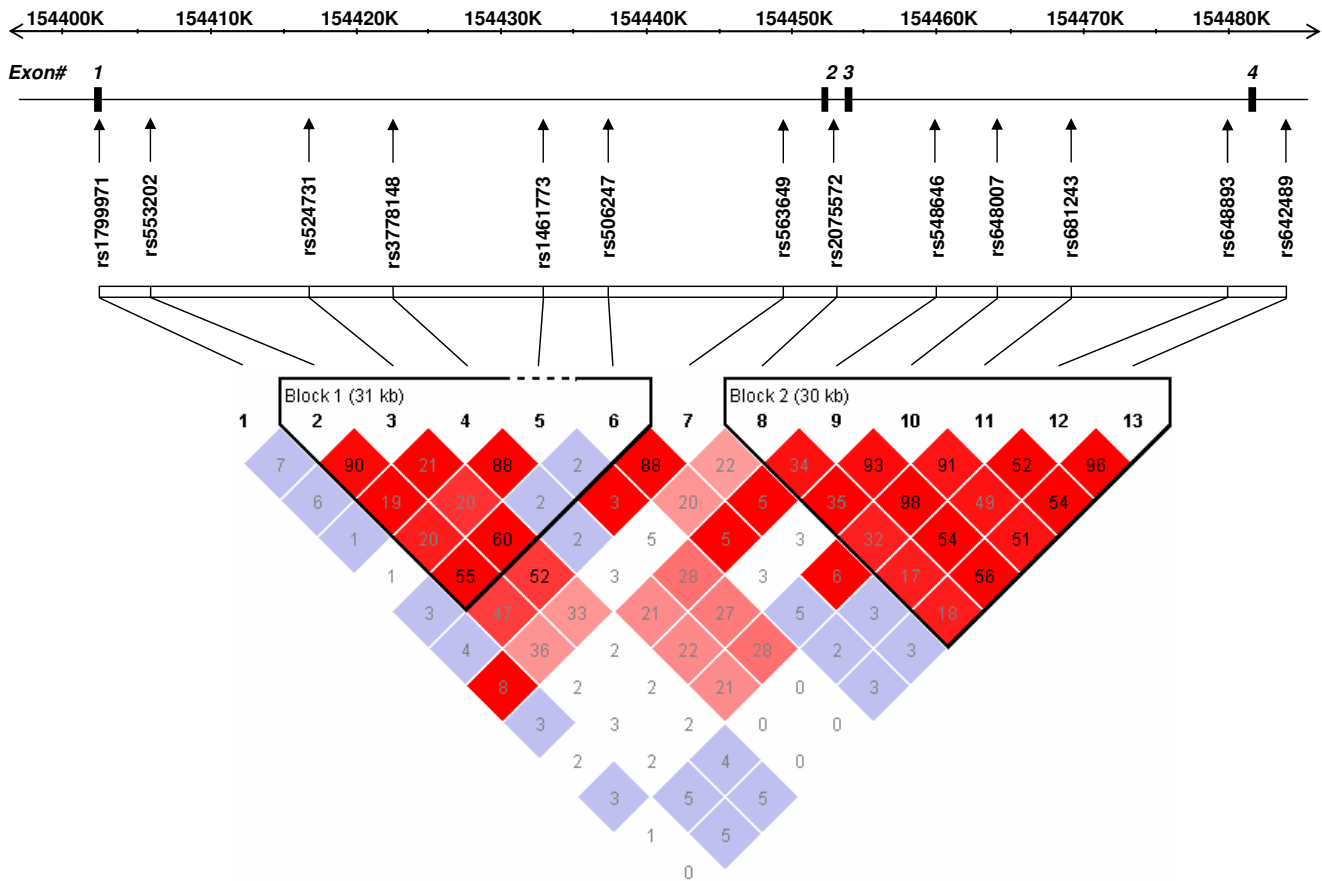


Figure 1

A schematic representation of *OPRM1* Gene Structure, Linkage Disequilibrium and Genotyped SNPs. The gene structure of *OPRM1* is shown with exons numbered from 1 to 4 and relative exon size denoted by the width of the vertical bars. Thirteen SNPs analyzed in this study are shown in relation to their location across *OPRM1*. Linkage disequilibrium (LD; shown below the gene structure) data, as measured by the correlation coefficient r^2 statistic, was generated using Haploview [81]. LD causes tightly linked genetic variants to be highly correlated. Shading represents correlation magnitudes between low r^2 (white) and high r^2 (red).

Table 1: OPRM1 marker information, including genetic map position, location within OPRM1 and minor allele frequency.

Marker Name		Gene	Chromosomal	Functional	Alleles		MAF ^d					References ^e
dbSNP	Celera	Location	Location (bp) ^a	Location (bp) ^b	Minor ^c	Major	Indian	CEPH	China	Japan	Yoruba	
rs1799971	hCV8950074	Exon 1	154,402,490	+118	G	A	0.13	0.17	0.36	0.49	0.01	1,2,3,4,5,6,7,8,9,10
rs553202	hCV809975	Intron 1	154,406,510	+4,138	A	G	0.33	0.20	nd	nd	0.41	
rs524731	hCV809963	Intron 1	154,416,785	+14,413	A	C	0.32	0.18	0.04	0.02	0.05	1,3,4
rs3778148	hCV27499812	Intron 1	154,422,705	+20,333	T	G	0.08	0.14	0.04	0.02	0.04	
rs1461773	hCV8949980	Intron 1	154,433,062	+30,690	T	C	0.16	0.14	0.04	0.03	0.08	
rs506247	hCV27335981	Intron 1	154,437,620	+35,248	G	T	0.21	0.04	0.00	0.00	0.01	
rs563649	hCV809947	Intron 1	154,449,660	+47,288	A	G	0.24	0.08	0.13	0.06	0.16	1
rs2075572	hCV1691815	Intron 2	154,453,697	+51,325	G	C	0.39	0.43	0.20	0.21	0.62	1,2,3,4
rs548646	hCV3073603	Intron 3	154,459,840	+57,468	T	C	0.18	0.33	0.07	0.13	0.46	1,3
rs648007	hCV1691794	Intron 3	154,464,304	+61,932	T	C	0.18	0.33	0.07	0.13	0.46	1
rs681243	hCV3073596	Intron 3	154,469,434	+67,062	A	G	0.18	0.24	0.06	0.14	0.43	
rs648893	hCV3073587	Intron 3	154,480,321	+77,949	C	T	0.11	0.20	0.05	0.10	0.04	1,3,4,5
rs642489	hCV3073582	3' UTR	154,484,368	+81,996	A	C	0.11	0.21	0.04	0.10	0.13	

^abp = base-pair position on NCBI Genome Build 127.

^bPosition relative to transcription start site at 154,402,372 on Chromosome 6 (NCBI Genome Build 127).

^cThe allele with the lowest frequency.

^dMAF = Minor Allele Frequency for the Indian tribes studied (Indians) and the reference populations CEPH (European), China, Japan, Yoruba (African). *nd* = no data.

^eReports where each SNP has previously been genotyped: 1. Xuei et al., 2007 [100]; 2. D. Zhang et al., 2007 [98]; 3. H. Zhang et al., 2006 [99]; 4. L. Zhang et al., 2006 [101]; 5. Gelernter et al., 2007 [33]; 6. Luo et al., 2003 [97]; 7. Smith et al., 2005 [102]; 8. Crowley et al., 2003 [103]; 9. Shi et al., 2002 [104]; 10 Y. Zhang et al. 2005 [12].

The OPRM1 region spanned by the 13 SNPs analyzed in this study is 81.8 kb. This region includes 140 SNPs that were typed by the HapMap project. Because the allele frequencies observed in this Indian population for each of the SNPs is intermediate between the allele frequencies observed for the European (CEU) and Asian (JC) HapMap populations, we investigated the linkage disequilibrium structure in these populations. The inferred CEU and JP population haplotypes were used estimate the consensus phylogenetic tree based on 500 bootstrapped trees produced with Neighbor-Joining method based on Kimura distance matrix as implemented in PHYLIP [84].

The inferred haplotypes for the 140 available SNPs for CEU and JP populations can be divided into four major clades, which can be further subdivided into related clades some of which may arise from recombination of the primary clades. Each of the subclades characteristically is relatively specific to either the CEU or JP population based on additional SNPs that were typed in by the HapMap project. It was found using haplotype analysis that the SNPs typed in the present study represented the major haplotypes identified in HapMap but they were unable to identify the large number of minor haplotypes. Ultimately, a more complete analysis depends on resequencing the OPRM1 gene and determining which sequence variants have functional significance.

The total additive genetic variance (heritability, h^2) and its standard error were estimated for the SHAS-E phenotypes using SOLAR [85]. A genetic association analyses was conducted where the number of copies of the minor allele of each individual was used as a covariate in a variance component analysis as implemented in SOLAR v2.0.4 [86], and the statistical significance of the ability of the covariate to explain phenotypic variance was determined. Age and sex were also accounted for in the analyses. To account for multiple comparisons, in these exploratory analyses, nominal significance was set at the $p < 0.01$ level.

Results

The demographic characteristics of the sample are virtually equivalent to the U.S. census data for these tribes and have been presented previously [65,87]. The mean age of the sample was 30.2 (± 0.7) yrs, there were 110 males and 141 females with a mean of 11.5(0.1) yrs of education, 60% of the sample was over 50% Native American heritage as estimated by their federal Indian blood quantum and 55% reported income of less than \$20,000 per annum. Within the sample of 251 participants 160 (64%) were found to have a lifetime DSM-III-R diagnosis of alcohol dependence and an additional 47(18.7%) were found to have alcohol abuse. One hundred seventy-three (69%) were current drinkers who reported a mean of 13 years of alcohol use. The mean number of drinking occasions

reported per month was 10 and their mean drinks per occasion were nine.

In total, 13 *OPRM1* SNPs spanning a region of 81.9 Kb were genotyped in 251 individuals. DNA from these individuals had been previously genotyped for linkage analyses [64,70,71]. The 251 individuals originated from a total of 41 families, comprising of 1 to 4 generations, with an average number of seven members per family (range, 1–30). The 251 individuals within the 41 families that were genetically informative includes: 77 parent-child, 212 sibling, 26 half sibling, 8 grandparent-grandchild, 151 avuncular, and 245 cousin relative pairs [65]. Marker information including genetic map position, location within *OPRM1*, and minor allele frequencies within this Indian population (as well as four reference populations for comparison) are listed in Table 1. Mendelian inconsistencies were identified using PEDSTATS [88] and made up 0.03% of the data. The physical locations of and pattern of linkage disequilibrium (LD) between the 13 SNPs typed across the *OPRM1* gene are schematically presented in Figure 1.

The estimated heritability (h^2) for the SHAS-E phenotypes ranged from near to zero for "energy" to .28 for "terrible" (see Table 2). The only two phenotypes with significant heritability were talkative and terrible ($p < 0.01$). Table 2 also gives values for the mean \pm S.D. for each of the SHAS-E items as well as the total for this population. As seen in Table 3, endorsing a more intense response on one or more of the following SHAS-E items: dizzy, drunk, high, nausea, talkative, and/or uncomfortable after imbibing 2–3 drinks was significantly associated with having at least one minor allele for 7 SNPs ($p < 0.01$) in or near the *OPRM1* receptor gene. Whereas, for the 118G allele, the

most commonly genotyped Asn40Asp polymorphism, there was only a trend for an association with reporting a less intense response to alcohol for the items: dizzy ($p < 0.02$) and sleepy ($p < 0.02$).

Discussion

The CNS effects of alcohol range from mild euphoria (high), to impaired coordination, to ataxia, decreased mentation, labile mood, to poor judgment, slurred speech, nausea and vomiting, and finally to respiratory failure, coma and death, depending on the dose imbibed [89]. The final level of impairment appears to depend on a number of factors including a persons' gender, age, weight, prior experience with alcohol and level of tolerance [40]. Another source of variation in response to alcohol is individual variation in alcohol metabolism. Some individuals, particularly East Asians who are homozygous for the *ALDH2*2* allele, are intolerant of alcohol and report intense facial flushing, tachycardia, hypotension, headache, nausea and vomiting following drinking more than one drink [67]. African Americans, with at least one *ADH1B*3*, also report expecting to have a more intense response to a standard dose of alcohol when compared to African Americans who are homozygous for the *ADH1B*1* allele [90].

Other sources of the genetic variation in sensitivity and tolerance to alcohol not attributed to differences in alcohol metabolism are less well understood. Several studies have found moderate heritability for level of response to alcohol. In one study, heritability was found to be 60% for a composite sensitivity measure that was used during an alcohol challenge in twins [91]. Correlations of level of response to alcohol using body sway and the SHAS in an alcohol challenge paradigm using sibling pairs as partici-

Table 2: Estimated heritability (h^2) for the Subjective High Assessment Scale-Expectations (SHAS-E) phenotypes.

Item	Trait	Mean	Std Dev	Range	Heritability	Std. Err.	p
1	Buzzed	12.32	11.48	0–36	0.09	0.10	0.18
2	Clumsy	9.52	10.19	0–36	0.01	0.09	0.45
3	Dizzy	7.75	10.44	0–36	0.09	0.10	0.18
4	Drunk	8.27	10.73	0–36	0.14	0.10	0.06
5	Effects of Alcohol	12.15	11.67	0–36	0.15	0.11	0.07
6	Energy	9.24	9.45	0–35	0		0.50
7	Good	12.33	10.40	0–36	0.05	0.10	0.29
8	Great	11.54	10.57	0–36	0.06	0.11	0.29
9	High	11.65	10.44	0–36	0.01	0.09	0.44
10	Nausea	6.0	9.85	0–36	0.12	0.11	0.11
11	Sleepy	7.20	9.99	0–36	0.23	0.12	0.02
12	Talkative	13.31	11.22	0–36	0.26	0.12	0.0089
13	Terrible	7.95	11.03	0–36	0.29	0.11	0.002
14	Uncomfortable	9.67	10.06	0–36	0.05	0.11	0.31
Total		118.80	98.7	0–377	0.11	0.11	0.14

Mean, standard deviation and range of values are given. Significant values ($p < 0.01$) are highlighted in bold.

Table 3: Association of OPRM1 SNPs with response to alcohol, as measured by the Subjective High Assessment Scale-Expectations (SHAS-E) questionnaire. Significant values (p < 0.01) are highlighted in bold.

Marker	Minor		Subjective High Assessment Scale-Expectations Item														
	Allele	Allele	Buzzed	Clumsy	Dizzy	Drunk	Effects	Energy	Good	Great	High	Nausea	Sleepy	Talk	Terrible	Uncom	Total
rs1799971	G	A	0.26	0.84	0.02	0.23	0.67	0.64	0.98	0.49	0.20	0.32	0.02	0.67	0.71	0.07	0.24
rs553202	A	A	0.03	0.10	0.01	0.001	0.10	0.48	0.32	0.64	0.001	0.01	0.15	0.21	0.008	0.008	0.02
rs524731	A	A	0.03	0.10	0.01	0.001	0.09	0.56	0.26	0.94	0.002	0.06	0.13	0.16	0.01	0.07	0.02
rs3778148	T	T	0.02	0.02	0.01	0.01	0.02	0.92	0.39	0.52	0.000	0.003	0.09	0.001	0.11	0.000	0.003
rs1461773	T	T	0.02	0.03	0.01	0.03	0.03	0.94	0.67	0.60	0.003	0.01	0.07	0.005	0.08	0.009	0.01
rs506247	G	G	0.40	0.95	0.51	0.06	0.91	0.58	0.63	0.55	0.35	0.44	0.47	0.48	0.07	0.83	0.64
rs563649	A	A	0.73	0.77	0.56	0.12	0.70	0.91	0.73	0.39	0.84	0.62	0.58	0.38	0.06	0.57	0.91
rs2075572	G	G	0.12	0.04	0.01	0.003	0.31	0.69	0.76		0.21	0.002	0.05	0.74	0.07	0.17	0.06
rs548646	T	T	0.01	0.01	0.001	0.02	0.02	0.12	0.77	0.91	0.06	0.02	0.04	0.03	0.33	0.02	0.005
rs648007	T	T	0.09	0.02	0.01	0.08	0.10	0.21	0.85	0.95	0.12	0.01	0.23	0.09	0.82	0.05	0.03
rs681243	A	A	0.02	0.01	0.001	0.02	0.03	0.15	0.90	0.91	0.05	0.006	0.06	0.06	0.37	0.02	0.007
rs648893	C	C	0.51	0.34	0.37	0.62	0.45	0.09	0.85	0.82	0.78	0.82	0.60	0.28	0.42	0.50	0.41
rs642489	A	A	0.43	0.25	0.31	0.55	0.40	0.09	0.83	0.80	0.79	0.71	0.45	0.28	0.50	0.55	0.36

The increaser allele for the TOTAL phenotype (i.e., the allele associated with a higher TOTAL score)
 TOTAL = sum of scores reported for the first 12 SHAS-E items.

pants was reported to be 0.36 [41]. Lower correlations were found in first-degree relatives using a retrospective self-report measure to assess level of response to alcohol (0.12–0.22). In the present study, values for the heritability of the SHAS-E were found to range from near to zero (for energy) to 0.28 (for terrible).

Further evidence for a genetic component to level of response to alcohol was provided by a genome-wide segregation analysis that evaluated subjective response to alcohol challenge in sibpairs. In that study, nine chromosome regions with LOD scores between 2.2 and 3.2 suggesting potential regions of interest in the genome that may contribute to the variance in alcohol responsivity [41]. An expanded dataset, collected in the same laboratory, also identified five areas of the genome with LOD scores between 2.2 and 2.6 for level of response to alcohol in sibpairs [92]. None of the locations identified in those studies were on chromosome 6 (6q24q25) near the location of the mu opioid receptor. However, a previous study in this Indian population found suggestion for linkage on chromosome 6q24q25 for several substance dependence phenotypes, as well as Body Mass Index, suggesting genes in that location may be associated with risk for substance dependence and other consumption-related phenotypes [71].

In the present study, evidence was obtained for an association between expectations of the effects of a standard dose of alcohol and polymorphisms in the OPRM1 receptor gene. Participants with at least one 118G allele for the Asp40Asn polymorphism reported that they expected to feel a less intense response to alcohol for the items: dizzy (p < 0.02) and sleepy (p < 0.02) when compared to individuals without any 118G alleles, findings that were not

significant in these analyses when multiple comparisons were taken into account. These data are, however, consistent with data from Kim and colleagues [93], who found that alcoholics with two copies of the 118G allele spent more days drinking than those who were heterozygous or homozygous for the 118A allele, perhaps suggesting a less intense response to alcohol. Assuming that alcohol may act as a partial agonist at the mu opioid receptor, the findings in the present study of a trend for reduced effect of alcohol in participants with the 118G allele, are also consistent with studies that evaluated response to opioid agonists where a reduced response to drug challenge (pupillary diameter, pain, respiratory depression) and/or increased dosage requirements are seen in those individuals with the 118G allele (see [1] for review).

Few studies have evaluated whether an association exists between response to alcohol and polymorphisms in the OPRM1 gene. In one study, the ability of naltrexone to blunt an alcohol-induced high was found to be greater in those participants with the 118G allele [94]. The finding of a more intense response to a mu opioid receptor antagonist found by Ray and Hutchinson [94] is consistent with previous studies that have demonstrated that subjects with the 118G allele that were given naloxone had higher cortisol concentrations [36]. It is also consistent with the finding that naltrexone may be more efficacious for the treatment of alcoholism in those with at least one 118G allele [32]. However, Ray and Hutchinson [94,95] have also reported that young participants in an IV alcohol challenge, with one 118G allele, reported feeling more subjective feelings of "high" across rising breath alcohol concentrations, as compared to those participants homozygous for the 118A allele. These findings are not consistent with the findings in the present study for the

118G allele, nor are they particularly consistent with studies that have found a less intense response to opioid agonists. However, the findings of Ray and Hutchinson [94,95] are consistent with the findings in the present study of an association between expecting to experience a more intense response to alcohol and carrying at least one minor allele for eight other SNPs in the opioid receptor gene. Since, in the study of Ray and Hutchinson [94,95], only one SNP was genotyped and the ethnic characteristics of the sample were not specified, it is possible that the findings reflected stratification of the sample or that the A118G variant was in linkage disequilibrium with several other alleles that may encode for a more intense response to alcohol. These data further suggest that making conclusions on the role of the mu opioid receptor gene in the development of alcohol-related behaviors may be limited if only one polymorphism in the gene is evaluated in isolation.

Several alcohol- or drug-related association studies [96-99] have expanded their investigations to include up to 20 SNPs in or near *OPRM1*, although all include the A118G variant. Ide and colleagues [96] genotyped 20 SNPs including 10 SNPs in the 3'UTR region among Japanese subjects meeting ICD-10 criteria for methamphetamine (MAP) dependence/psychosis and controls. Four SNPs (including the A118G and rs2075572 variants that were genotyped in the present study) representing the major haplotypes observed in the study sample were tested for association with four features of MAP dependence/psychosis. While A118G and two other SNPs were not associated with MAP dependence/psychosis, the rs2075572 G-allele was significantly associated with increased risk for a diagnosis of MAP dependence/psychosis ($p = 0.011$), as well as four aspects/symptoms of the disorder ($p < 0.01$). Interestingly, within this Indian population, the rs2075572 G-allele was related to expecting to feel a more intense response to alcohol in four of the 14 items of the SHAS-E, an indication that carriers of this allele may be protected from developing alcohol dependence. Zhang and colleagues [98] investigated the relationship between heroin-induced subjective responses in a Chinese population and ten SNPs selected throughout *OPRM1*. They found three SNPs in intron 1 were associated with an increased risk of positive responses on first use of heroin and were likely contributing to further heroin consumption. However, A118G and rs2075572 were not associated with any differences in heroin-induced subjective responses. In another study, Luo and colleagues [97] typed eight variants in alcohol, cocaine and opioid and poly-substance dependent European Americans (EA) and African Americans (AA). They found that the A-allele of the -2044C/A polymorphism was a susceptibility allele for the combination of alcohol and opioid dependence in the EA sample, but not the AA sample. Once again, A118G

was not associated with any of the substance dependent phenotypes. Finally, Zhang and colleagues [99] studied the role of *OPRM1* genetic variation in a large case-control sample of alcohol dependent and/or drug (cocaine and/or opioid) dependent European Americans. Thirteen SNPs, five of which were typed in the present study, were genotyped representing the major haplotypes observed in HapMap. Seven SNPs (but not A118) were associated with alcohol, cocaine, opioid plus opioid/cocaine dependency. Zhang and colleagues [99] found that the frequencies of the rs524731 A-allele and rs648893 T-allele were significantly higher among the dependent EA subjects. Within this Indian population, the rs524731 A-allele and rs648893 T-allele were generally associated with a more intense response to alcohol.

Conclusion

In conclusion, these data represent the first association analysis of a level of response to alcohol phenotype with multiple SNPs in the *OPMR1* receptor gene in American Indians. SNPs highlighted in prior studies of substance dependence phenotypes were also identified as well as new SNPs of potential importance to substance dependence research. The results of this study should, however, be interpreted in the context of several limitations. A more conservative approach to multiple comparisons would have led to fewer significant effects. Level of response to alcohol was evaluated using the SHAS-E, and a more direct measure of intoxication using the SHAS or body sway may produce more reliable results. Haplotype analysis using the SNPs typed in this study was unable to specially tag all of the clades observed in the HapMap population. Ultimately, a more complete analysis depends on resequencing the *OPRM1* and determining which sequence variants have functional significance. Additionally, the findings may not generalize to other Native Americans or represent all Indians of the tribes studied, and comparisons of association findings to non-Indian populations may be limited by differences in a host of potential genetic and environmental variables. Finally, because this population has significant admixture estimates of allele frequencies may produce biased results. Despite these limitations, this report represents an important step in an ongoing investigation to understand the genetic determinants associated with the development of substance use disorders in this high risk and understudied ethnic group.

List of abbreviations

CNS: Central Nervous System; DNA: Deoxyribonucleic Acid; *ADH1B*3*: Alcohol Dehydrogenase 1B* 3; *ADH1B*1*: Alcohol Dehydrogenase 1B*1.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

CLE contributed to the recruitment, collection and analysis of the clinical and genetic data on the subjects. KCW contributed to the genetic and heritability analyses. PAL did the genotyping and its analysis. All authors contributed to writing the paper.

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References

- Somogyi AA, Barratt DT, Collier JK: **Pharmacogenetics of opioids.** *Clin Pharmacol Ther* 2007, **81**:429-444.
- Aubrun F, Langeron O, Quesnel C, Coriat P, Riou B: **Relationships between measurement of pain using visual analog score and morphine requirements during postoperative intravenous morphine titration.** *Anesthesiology* 2003, **98**:1415-1421.
- Ashby M, Fleming B, Wood M, Somogyi A: **Plasma morphine and glucuronide (M3G and M6G) concentrations in hospice inpatients.** *J Pain Symptom Manage* 1997, **14**:157-167.
- Bond C, LaForge KS, Tian M, Melia D, Zhang S, Borg L, Gong J, Schluger J, Strong JA, Leal SM, Tischfield JA, Kreek MJ, Yu L: **Single-nucleotide polymorphism in the human mu opioid receptor gene alters beta-endorphin binding and activity: possible implications for opiate addiction.** *Proc Natl Acad Sci USA* 1998, **95**:9608-9613.
- Conne B, Stutz A, Vassalli JD: **The 3' untranslated region of messenger RNA: A molecular 'hotspot' for pathology?** *Nat Med* 2000, **6**:637-641.
- Hoehe MR, Kopke K, Wendel B, Rohde K, Flachmeier C, Kidd KK, Berrettini WH, Church GM: **Sequence variability and candidate gene analysis in complex disease: association of mu opioid receptor gene variation with substance dependence.** *Hum Mol Genet* 2000, **9**:2895-2908.
- Ide S, Kobayashi H, Tanaka K, Ujike H, Sekine Y, Ozaki N, Inada T, Harano M, Komiyaama T, Yamada M, Iyo M, Ikeda K, Sora I: **Gene polymorphisms of the mu opioid receptor in methamphetamine abusers.** *Ann N Y Acad Sci* 2004, **1025**:316-324.
- Ikeda K, Ide S, Han W, Hayashida M, Uhl GR, Sora I: **How individual sensitivity to opiates can be predicted by gene analyses.** *Trends Pharmacol Sci* 2005, **26**:311-317.
- Lotsch J, Geisslinger G: **Are mu-opioid receptor polymorphisms important for clinical opioid therapy?** *Trends Mol Med* 2005, **11**:82-89.
- Lotsch J, Geisslinger G: **Relevance of frequent mu-opioid receptor polymorphisms for opioid activity in healthy volunteers.** *Pharmacogenomics J* 2006, **6**:200-210.
- Wang JB, Johnson PS, Persico AM, Hawkins AL, Griffin CA, Uhl GR: **Human mu opiate receptor. cDNA and genomic clones, pharmacologic characterization and chromosomal assignment.** *FEBS Lett* 1994, **338**:217-222.
- Zhang Y, Wang D, Johnson AD, Papp AC, Sadee W: **Allelic expression imbalance of human mu opioid receptor (OPRM1) caused by variant A118G.** *J Biol Chem* 2005, **280**:32618-32624.
- Lotsch J, Skarke C, Grosch S, Darimont J, Schmidt H, Geisslinger G: **The polymorphism A118G of the human mu-opioid receptor gene decreases the pupil constrictory effect of morphine-6-glucuronide but not that of morphine.** *Pharmacogenetics* 2002, **12**:3-9.
- Lotsch J, Zimmermann M, Darimont J, Marx C, Dudziak R, Skarke C, Geisslinger G: **Does the A118G polymorphism at the mu-opioid receptor gene protect against morphine-6-glucuronide toxicity?** *Anesthesiology* 2002, **97**:814-819.
- Skarke C, Darimont J, Schmidt H, Geisslinger G, Lotsch J: **Analgesic effects of morphine and morphine-6-glucuronide in a transcutaneous electrical pain model in healthy volunteers.** *Clin Pharmacol Ther* 2003, **73**:107-121.
- Romberg R, Olofsen E, Sarton E, den HJ, Taschner PE, Dahan A: **Pharmacokinetic-pharmacodynamic modeling of morphine-6-glucuronide-induced analgesia in healthy volunteers: absence of sex differences.** *Anesthesiology* 2004, **100**:120-133.
- Romberg RR, Olofsen E, Bijl H, Taschner PE, Teppema LJ, Sarton EY, van Kleef JW, Dahan A: **Polymorphism of mu-opioid receptor gene (OPRM1:c.118A>G) does not protect against opioid-induced respiratory depression despite reduced analgesic response.** *Anesthesiology* 2005, **102**:522-530.
- Oertel BG, Schmidt R, Schneider A, Geisslinger G, Lotsch J: **The mu-opioid receptor gene polymorphism 118A>G depletes alfentanil-induced analgesia and protects against respiratory depression in homozygous carriers.** *Pharmacogenet Genomics* 2006, **16**:625-636.
- Lotsch J, Skarke C, Wieting J, Oertel BG, Schmidt H, Brockmoller J, Geisslinger G: **Modulation of the central nervous effects of levomethadone by genetic polymorphisms potentially affecting its metabolism, distribution, and drug action.** *Clin Pharmacol Ther* 2006, **79**:72-89.
- Klepstad P, Ravvag TT, Kaasa S, Holthe M, Dale O, Borchgrevink PC, Baar C, Vikan T, Krokan HE, Skorpen F: **The 118A > G polymorphism in the human mu-opioid receptor gene may increase morphine requirements in patients with pain caused by malignant disease.** *Acta Anaesthesiol Scand* 2004, **48**:1232-1239.
- Coulbault L, Beaussier M, Verstuyft C, Weickmans H, Dubert L, Tregouet D, Descot C, Parc Y, Lienhart A, Jaillon P, Becquemont L: **Environmental and genetic factors associated with morphine response in the postoperative period.** *Clin Pharmacol Ther* 2006, **79**:316-324.
- Janicki PK, Schuler G, Francis D, Bohr A, Gordin V, Jarzembowski T, Ruiz-Velasco V, Mets B: **A genetic association study of the functional A118G polymorphism of the human mu-opioid receptor gene in patients with acute and chronic pain.** *Anesth Analg* 2006, **103**:1011-1017.
- Herz A: **Endogenous opioid systems and alcohol addiction.** *Psychopharmacology (Berl)* 1997, **129**:99-111.
- Kreek MJ: **Opioid receptors: some perspectives from early studies of their role in normal physiology, stress responsivity, and in specific addictive diseases.** *Neurochem Res* 1996, **21**:1469-1488.
- Anton RF, Moak DH, Waid LR, Latham PK, Malcolm RJ, Dias JK: **Naltrexone and cognitive behavioral therapy for the treatment of outpatient alcoholics: results of a placebo-controlled trial.** *Am J Psychiatry* 1999, **156**:1758-1764.
- Anton RF, O'Malley SS, Ciraulo DA, Cisler RA, Couper D, Donovan DM, Gastfriend DR, Hosking JD, Johnson BA, LoCastro JS, Longabaugh R, Mason BJ, Mattson ME, Miller WR, Pettinati HM, Randall CL, Swift R, Weiss RD, Williams LD, Zweben A: **Combined pharmacotherapies and behavioral interventions for alcohol dependence: the COMBINE study: a randomized controlled trial.** *JAMA* 2006, **295**:2003-2017.
- Balldin J, Berglund M, Borg S, Mansson M, Bendtsen P, Franck J, Gustafsson L, Halldin J, Nilsson LH, Stolt G, Willander A: **A 6-month controlled naltrexone study: combined effect with cognitive behavioral therapy in outpatient treatment of alcohol dependence.** *Alcohol Clin Exp Res* 2003, **27**:1142-1149.
- Kiefer F, Jahn H, Tarnaske T, Helwig H, Briken P, Holzbach R, Kampf P, Stracke R, Baehr M, Naber D, Wiedemann K: **Comparing and combining naltrexone and acamprosate in relapse prevention of alcoholism: a double-blind, placebo-controlled study.** *Arch Gen Psychiatry* 2003, **60**:92-99.
- Monti PM, Rohsenow DJ, Swift RM, Gulliver SB, Colby SM, Mueller TI, Brown RA, Gordon A, Abrams DB, Niaura RS, Asher MK: **Naltrexone and cue exposure with coping and communication skills training for alcoholics: treatment process and 1-year outcomes.** *Alcohol Clin Exp Res* 2001, **25**:1634-1647.
- O'Malley SS, Jaffe AJ, Chang G, Schottenfeld RS, Meyer RE, Rounsaville B: **Naltrexone and coping skills therapy for alcohol dependence. A controlled study.** *Arch Gen Psychiatry* 1992, **49**:881-887.
- Volpicelli JR, Alterman AI, Hayashida M, O'Brien CP: **Naltrexone in the treatment of alcohol dependence.** *Arch Gen Psychiatry* 1992, **49**:876-880.
- Oslin DW, Berrettini W, Kranzler HR, Pettinati H, Gelernter J, Volpicelli JR, O'Brien CP: **A functional polymorphism of the mu-opi-**

- oid receptor gene is associated with naltrexone response in alcohol-dependent patients. *Neuropsychopharmacology* 2003, **28**:1546-1552.
33. Gelernter J, Gueorguieva R, Kranzler HR, Zhang H, Cramer J, Rosenheck R, Krystal JH: **Opioid receptor gene (OPRM1, OPRK1, and OPRD1) variants and response to naltrexone treatment for alcohol dependence: results from the VA Cooperative Study.** *Alcohol Clin Exp Res* 2007, **31**:555-563.
 34. Anton RF, Oroszi G, O'Malley S, Couper D, Swift R, Pettinati H, Goldman D: **An evaluation of mu-opioid receptor (OPRM1) as a predictor of naltrexone response in the treatment of alcohol dependence: results from the Combined Pharmacotherapies and Behavioral Interventions for Alcohol Dependence (COMBINE) study.** *Arch Gen Psychiatry* 2008, **65**:135-144.
 35. Hernandez-Avila CA, Wand G, Luo X, Gelernter J, Kranzler HR: **Association between the cortisol response to opioid blockade and the Asn40Asp polymorphism at the mu-opioid receptor locus (OPRM1).** *Am J Med Genet B Neuropsychiatr Genet* 2003, **118**:60-65.
 36. Wand GS, McCaul M, Yang X, Reynolds J, Gotjen D, Lee S, Ali A: **The mu-opioid receptor gene polymorphism (A118G) alters HPA axis activation induced by opioid receptor blockade.** *Neuropsychopharmacology* 2002, **26**:106-114.
 37. Arias A, Feinn R, Kranzler HR: **Association of an Asn40Asp (A118G) polymorphism in the mu-opioid receptor gene with substance dependence: a meta-analysis.** *Drug Alcohol Depend* 2006, **83**:262-268.
 38. Slutske WS, True WR, Scherrer JF, Heath AC, Bucholz KK, Eisen SA, Goldberg J, Lyons MJ, Tsuang MT: **The heritability of alcoholism symptoms: "indicators of genetic and environmental influence in alcohol-dependent individuals" revisited.** *Alcohol Clin Exp Res* 1999, **23**:759-769.
 39. Town T, Schinka J, Tan J, Mullan M: **The opioid receptor system and alcoholism: a genetic perspective.** *Eur J Pharmacol* 2000, **410**:243-248.
 40. Schuckit MA: **A clinical model of genetic influences in alcohol dependence.** *J Stud Alcohol* 1994, **55**:5-17.
 41. Wilhelmsen KC, Schuckit M, Smith TL, Lee JV, Segall SK, Feiler HS, Kalmijn J: **The search for genes related to a low-level response to alcohol determined by alcohol challenges.** *Alcohol Clin Exp Res* 2003, **27**:1041-1047.
 42. Schuckit MA: **Subjective responses to alcohol in sons of alcoholics and control subjects.** *Arch Gen Psychiatry* 1984, **41**:879-884.
 43. Moss HB, Yao JK, Maddock JM: **Responses by sons of alcoholic fathers to alcoholic and placebo drinks: perceived mood, intoxication, and plasma prolactin.** *Alcohol Clin Exp Res* 1989, **13**:252-257.
 44. O'Malley SS, Maisto SA: **Effects of family drinking history and expectancies on responses to alcohol in men.** *J Stud Alcohol* 1985, **46**:289-297.
 45. Pollock VE, Teasdale TW, Gabrielli WF, Knop J: **Subjective and objective measures of response to alcohol among young men at risk for alcoholism.** *J Stud Alcohol* 1986, **47**:297-304.
 46. Savoie TM, Emory EK, Moody-Thomas S: **Acute alcohol intoxication in socially drinking female and male offspring of alcoholic fathers.** *J Stud Alcohol* 1988, **49**:430-435.
 47. Newlin DB, Thomson JB: **Alcohol challenge with sons of alcoholics: a critical review and analysis.** *Psychol Bull* 1990, **108**:383-402.
 48. Pollock VE: **Meta-analysis of subjective sensitivity to alcohol in sons of alcoholics.** *Am J Psychiatry* 1992, **149**:1534-1538.
 49. Schuckit MA, Smith TL: **An 8-year follow-up of 450 sons of alcoholic and control subjects.** *Arch Gen Psychiatry* 1996, **53**:202-210.
 50. Wall TL, Ehlers CL: **Acute effects of alcohol on P300 in Asians with different ALDH2 genotypes.** *Alcohol Clin Exp Res* 1995, **19**:617-622.
 51. Wall TL, Thomasson HR, Schuckit MA, Ehlers CL: **Subjective feelings of alcohol intoxication in Asians with genetic variations of ALDH2 alleles.** *Alcohol Clin Exp Res* 1992, **16**:991-995.
 52. Wall TL, Gallen CC, Ehlers CL: **Effects of alcohol on the EEG in Asian men with genetic variations of ALDH2.** *Biol Psychiatry* 1993, **34**:91-99.
 53. Wall TL, Nemeroff CB, Ritchie JC, Ehlers CL: **Cortisol responses following placebo and alcohol in Asians with different ALDH2 genotypes.** *J Stud Alcohol* 1994, **55**:207-213.
 54. Monteiro MG, Klein JL, Schuckit MA: **High levels of sensitivity to alcohol in young adult Jewish men: a pilot study.** *J Stud Alcohol* 1991, **52**:464-469.
 55. Lander ES, Schork NJ: **Genetic dissection of complex traits.** *Science* 1994, **265**:2037-2048.
 56. Leland J: *Firewater myths: North American Indian drinking and alcohol addiction* New Brunswick, NJ: Publications Division, Rutgers Center of Alcohol Studies; 1976.
 57. Ehlers CL, Garcia-Andrade C, Wall TL, Sobel DF, Phillips E: **Determinants of P3 amplitude and response to alcohol in Native American Mission Indians.** *Neuropsychopharmacology* 1998, **18**:282-292.
 58. Ehlers CL, Garcia-Andrade C, Wall TL, Cloutier D, Phillips E: **Electroencephalographic responses to alcohol challenge in Native American Mission Indians.** *Biol Psychiatry* 1999, **45**:776-787.
 59. Garcia-Andrade C, Wall TL, Ehlers CL: **The firewater myth and response to alcohol in Mission Indians.** *Am J Psychiatry* 1997, **154**:983-988.
 60. Ehlers CL, Wall TL, Garcia-Andrade C, Phillips E: **Auditory P3 findings in Mission Indian youth.** *J Stud Alcohol* 2001, **62**:562-570.
 61. Ehlers CL, Wall TL, Garcia-Andrade C, Phillips E: **Visual P3 findings in Mission Indian youth: relationship to family history of alcohol dependence and behavioral problems.** *Psychiatry Res* 2001, **105**:67-78.
 62. Ehlers CL, Wall TL, Garcia-Andrade C, Phillips E: **Effects of age and parental history of alcoholism on EEG findings in Mission Indian children and adolescents.** *Alcohol Clin Exp Res* 2001, **25**:672-679.
 63. Ehlers CL, Wall TL, Garcia-Andrade C, Phillips E: **EEG asymmetry: relationship to mood and risk for alcoholism in Mission Indian youth.** *Biol Psychiatry* 2001, **50**:129-136.
 64. Ehlers CL, Wall TL, Betancourt M, Gilder DA: **The clinical course of alcoholism in 243 Mission Indians.** *Am J Psychiatry* 2004, **161**:1204-1210.
 65. Ehlers CL, Gilder DA, Wall TL, Phillips E, Feiler H, Wilhelmsen KC: **Genomic screen for loci associated with alcohol dependence in Mission Indians.** *Am J Hum Genet* 2004, **129**:110-115.
 66. Ehlers CL, Slutske WS, Gilder DA, Lau P, Wilhelmsen KC: **Age at first intoxication and alcohol use disorders in Southwest California Indians.** *Alcohol Clin Exp Res* 2006, **30**:1856-1865.
 67. Wall TL, Garcia-Andrade C, Thomasson HR, Carr LG, Ehlers CL: **Alcohol dehydrogenase polymorphisms in Native Americans: identification of the ADH2*3 allele.** *Alcohol Alcohol* 1997, **32**:129-132.
 68. Wall TL, Carr LG, Ehlers CL: **Protective association of genetic variation in alcohol dehydrogenase with alcohol dependence in Native American Mission Indians.** *Am J Psychiatry* 2003, **160**:41-46.
 69. Ehlers CL, Wilhelmsen KC: **Genomic scan for alcohol craving in Mission Indians.** *Psychiatr Genet* 2005, **15**:71-75.
 70. Ehlers CL, Wilhelmsen KC: **Genomic screen for loci associated with tobacco usage in Mission Indians.** *BMC Med Genet* 7:9. 2006, **Feb 10**;
 71. Ehlers CL, Wilhelmsen KC: **Genomic screen for substance dependence and body mass index in Southwest California Indians.** *Genes Brain Behav* 2007, **6**:184-191.
 72. Wilhelmsen KC, Ehlers C: **Heritability of substance dependence in a Native American population.** *Psychiatr Genet* 2005, **15**:101-107.
 73. Kalton G, Anderson DW: **Sampling rare populations.** *J Roy Stat Soc* 1986, **49**:65-82.
 74. Muhib FB, Lin LS, Stueve A, Miller RL, Ford WL, Johnson WD, Smith PJ: **A venue-based method for sampling hard-to-reach populations.** *Public Health Rep* 2001, **116**(Suppl 1):216-222.
 75. Heckathorn DD: **Respondent-driven sampling: a new approach to the study of hidden populations.** *Soc Probl* 1997, **44**:174-199.
 76. Ehlers CL, Spence JP, Wall TL, Gilder DA, Carr LG: **Association of ALDH1 promoter polymorphisms with alcohol-related phenotypes in Southwest California Indians.** *Alcohol Clin Exp Res* 2004, **28**:1481-1486.
 77. Schuckit MA: **Genetics and the risk for alcoholism.** *JAMA* 1985, **254**:2614-2617.
 78. Bucholz KK, Cadoret R, Cloninger CR, Dinwiddie SH, Hesselbrock VM, Nurnberger Jr J, Reich T, Schmidt I, Schuckit MA: **A new, semi-structured psychiatric interview for use in genetic linkage**

- studies: a report on the reliability of the SSAGA. *J Stud Alcohol* 1994, **55**:149-158.
79. American Psychiatric Association, Task Force on DSM-IV: *Diagnostic and statistical manual of mental disorders (DSM-IV)* Washington, DC: American Psychiatric Association; 1994.
 80. Schuckit MA, Smith TL, Kalmijn J, Tsuang J, Hesselbrock V, Bucholz K: **Response to alcohol in daughters of alcoholics: a pilot study and a comparison with sons of alcoholics.** *Alcohol Alcohol* 2000, **35**:242-248.
 81. Barrett JC, Fry B, Maller J, Daly MJ: **Haploview: analysis and visualization of LD and haplotype maps.** *Bioinformatics* 2005, **21**:263-265.
 82. **International HapMap Public Database** [<http://www.hapmap.org>]
 83. **Applied Biosystems SNP Database** [<http://www.appliedbiosystems.com>]
 84. **PHYLIP PHYLogeny Inference Package** [<http://evolution.gs.washington.edu/phylip/phylip.html>]. Department of Genome Sciences, University of Washington, Seattle, WA
 85. **SOLAR. Sequential Oligogenic Linkage Analysis Routines** [<http://www.sfbr.org/solar/>]. Southwest Foundation for Biomedical Research, San Antonio, TX
 86. Almasy L, Blangero J: **Multipoint quantitative-trait linkage analysis in general pedigrees.** *Am J Hum Genet* 1998, **62**:1198-1211.
 87. Ehlers CL, Wall TL, Corey L, Lau P, Gilder DA, Wilhelmsen K: **Heritability of illicit drug use and transition to dependence in Southwest California Indians.** *Psych Genet* 2007, **17**:171-176.
 88. Wigginton JE, Abecasis GR: **PEDSTATS: descriptive statistics, graphics and quality assessment for gene mapping data.** *Bioinformatics* 2005, **21**:3445-3447.
 89. Schuckit MA: *Drug and alcohol abuse: a clinical guide to diagnosis and treatment* 4th edition. New York: Plenum Medical Book Co; 1995.
 90. Ehlers CL, Phillips E, Sweeny A, Slaweki CJ: **Event-related potential responses to alcohol-related stimuli in African-American young adults: relation to family history of alcoholism and drug usage.** *Alcohol Alcohol* 2003, **38**:332-338.
 91. Heath AC, Madden PA, Bucholz KK, Dinwiddie SH, Slutske WS, Bierut LJ, Rohrbach JW, Statham DJ, Dunne MP, Whitfield JB, Martin NG: **Genetic differences in alcohol sensitivity and the inheritance of alcoholism risk.** *Psychol Med* 1999, **29**:1069-1081.
 92. Schuckit MA, Wilhelmsen K, Smith TL, Feiler HS, Lind P, Lange LA, Kalmijn J: **Autosomal linkage analysis for the level of response to alcohol.** *Alcohol Clin Exp Res* 2005, **29**:1976-1982.
 93. Kim SG, Kim CM, Kang DH, Kim YJ, Byun WT, Kim SY, Park JM, Kim MJ, Oslin DW: **Association of functional opioid receptor genotypes with alcohol dependence in Koreans.** *Alcohol Clin Exp Res* 2004, **28**:986-990.
 94. Ray LA, Hutchison KE: **Effects of naltrexone on alcohol sensitivity and genetic moderators of medication response: a double-blind placebo-controlled study.** *Arch Gen Psychiatry* 2007, **64**:1069-1077.
 95. Ray LA, Hutchison KE: **A polymorphism of the mu-opioid receptor gene (OPRM1) and sensitivity to the effects of alcohol in humans.** *Alcohol Clin Exp Res* 2004, **28**:1789-1795.
 96. Ide S, Kobayashi H, Ujike H, Ozaki N, Sekine Y, Inada T, Harano M, Komiyama T, Yamada M, Iyo M, Iwata N, Tanaka K, Shen H, Iwahashi K, Itokawa M, Minami M, Satoh M, Ikeda K, Sora I: **Linkage disequilibrium and association with methamphetamine dependence/psychosis of mu-opioid receptor gene polymorphisms.** *Pharmacogenomics J* 2006, **6**:179-188.
 97. Luo X, Kranzler HR, Zhao H, Gelernter J: **Haplotypes at the OPRM1 locus are associated with susceptibility to substance dependence in European-Americans.** *Am J Med Genet B Neuropsychiatr Genet* 2003, **120**:97-108.
 98. Zhang D, Shao C, Shao M, Yan P, Wang Y, Liu Y, Liu W, Lin T, Xie Y, Zhao Y, Lu D, Li Y, Jin L: **Effect of mu-opioid receptor gene polymorphisms on heroin-induced subjective responses in a Chinese population.** *Biol Psychiatry* 2007, **61**:1244-1251.
 99. Zhang H, Luo X, Kranzler HR, Lappalainen J, Yang BZ, Krupitsky E, Zvartau E, Gelernter J: **Association between two mu-opioid receptor gene (OPRM1) haplotype blocks and drug or alcohol dependence.** *Hum Mol Genet* 2006, **15**:807-819.
 100. Xuei X, Flury-Wetherill L, Bierut L, Dick D, Nurnberger J Jr, Foroud T, Edenberg HJ: **The opioid system in alcohol and drug dependence: family-based association study.** *Am J Med Genet B Neuropsychiatr Genet* 2007, **144**:877-884.
 101. Zhang L, Kendler KS, Chen X: **The mu-opioid receptor gene and smoking initiation and nicotine dependence.** *Behav Brain Funct* 2006, **2**:28.
 102. Smith RJ, Doyle GA, Han AM, Crowley JJ, Oslin DW, Patkar AA, Mannelli P, Demaria PA Jr, O'Brien CP, Berrettini WH: **Novel exonic mu-opioid receptor gene (OPRM1) polymorphisms not associated with opioid dependence.** *Am J Med Genet B Neuropsychiatr Genet* 2005, **133**:105-109.
 103. Crowley JJ, Oslin DW, Patkar AA, Gotthel E, Demaria PA Jr, O'Brien CP, Berrettini WH, Grice DE: **A genetic association study of the mu opioid receptor and severe opioid dependence.** *Psychiatr Genet* 2003, **13**:169-173.
 104. Shi J, Hui L, Xu Y, Wang F, Huang W, Hu G: **Sequence variations in the mu-opioid receptor gene (OPRM1) associated with human addiction to heroin.** *Hum Mutat* 2002, **19**:459-460.

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