

Review

Major Tumor Suppressor and Oncogenic Non-Coding RNAs: Clinical Relevance in Lung Cancer

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Academic Editor: Alexander E. Kalyuzhny

Received: 20 April 2017; Accepted: 5 May 2017; Published: 9 May 2017

Abstract: Lung cancer is the leading cause of cancer deaths worldwide, yet there remains a lack of specific and sensitive tools for early diagnosis and targeted therapies. High-throughput sequencing techniques revealed that non-coding RNAs (ncRNAs), e.g., microRNAs and long ncRNAs (lncRNAs), represent more than 80% of the transcribed human genome. Emerging evidence suggests that microRNAs and lncRNAs regulate target genes and play an important role in biological processes and signaling pathways in malignancies, including lung cancer. In lung cancer, several tumor suppressor/oncogenic microRNAs and lncRNAs function as biomarkers for metastasis and prognosis, and thus may serve as therapeutic tools. In this review, recent work on microRNAs and lncRNAs is introduced and briefly summarized with a focus on potential biological and therapeutic applications.

Keywords: adenocarcinoma; carcinoma; carcinogenesis; histology; long non-coding RNA; miRNA; molecular pathology; oncology

1. Introduction

Lung cancer is the leading cause of cancer deaths among males in both developing and developed countries, and it has outpaced breast cancer as the leading cause of cancer deaths in females in developed countries [1]. Lung cancers are categorized into two main histological groups: small cell lung cancer (SCLC, 15% of all lung cancers) and non-SCLC (NSCLC, 85% of all lung cancers). NSCLCs are further subcategorized into three major histological subtypes: adenocarcinoma, squamous cell carcinoma (SqCC), and large cell carcinoma. Lung cancers are heterogeneous diseases characterized by distinct epigenetic and genetic abnormalities, even within the same histological type [2–18]. Although extensive efforts to develop molecular-targeted drugs are ongoing [19–27], no targeted therapies are available for a number of lung cancer patients. Although techniques and strategies for the early detection of lung cancer have improved, most lung cancers are still diagnosed at advanced stages. Therefore, the identification of novel diagnostic biomarkers and therapeutic strategies is a prerequisite for the management of lung cancer.

In recent years, the dysregulation of microRNAs and long non-coding RNAs (lncRNAs) has been highlighted as potential diagnostic and therapeutic tools in malignancies, including lung cancer [28–49]. Indeed, microRNAs and lncRNAs regulate a number of target genes, play an important role in lung carcinogenesis, and serve as potential diagnostic and therapeutic tools in lung cancer [37–49].

In this review, recent work on lncRNAs and microRNAs in lung cancer is summarized with a focus on their biological and therapeutic applications.

2. Non-Coding RNA

Emerging evidence from the human genome sequencing projects suggests that more than 80% of the human genome is actively transcribed into RNA, even though less than 3% of the human

genome encodes translated proteins [50,51]. RNAs that do not yield coding proteins are collectively referred to as non-coding RNAs (ncRNA). These ncRNAs are divided into housekeeping ncRNAs and regulatory ncRNAs. Housekeeping ncRNAs include transfer RNAs (tRNAs) and ribosomal RNAs (rRNAs). Regulatory ncRNAs are generally transcribed in a location- and time-dependent fashion. Regulatory ncRNAs can be further divided into two groups based on their size: small ncRNAs (shorter than 200 nucleotides) and lncRNAs (200 nucleotides or longer). Small ncRNAs contain microRNAs, small nucleolar RNAs (snoRNAs), small interfering RNAs (siRNAs), small nuclear RNAs (snRNAs), and PIWI-interacting RNAs (piRNAs) (Figure 1) [49–51].

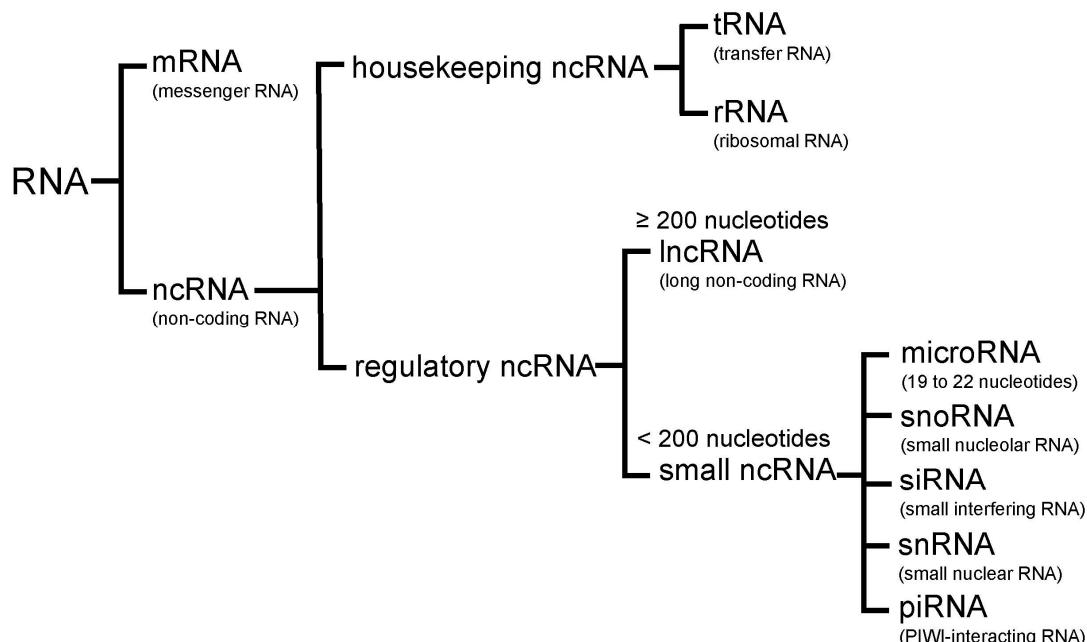


Figure 1. RNA categories. RNAs are divided into two major classes: messenger RNA (mRNA) and non-coding RNA (ncRNA). NcRNAs include housekeeping ncRNA, which consists of transfer RNA (tRNA) and ribosomal RNA (rRNA), and regulatory ncRNA. Regulatory ncRNAs are classified into long ncRNA (lncRNA) and small ncRNA. Small ncRNAs are subclassified into microRNA, small nucleolar RNA (snoRNA), small interfering RNA (siRNA), small nuclear RNA (snRNA), and PIWI-interacting RNA (piRNA).

2.1. MicroRNAs

MicroRNAs are small single stranded ncRNAs (19 to 22 nucleotides in length) that are highly conserved among different organisms. MicroRNAs play important regulatory roles in animals and plants by targeting messenger RNAs (mRNAs) for translational repression or degradation. MicroRNAs comprise one of the most abundant classes of gene regulatory molecules and therefore influence the output of many protein-coding genes [52]. MicroRNAs have the potential to serve as biomarkers and therapeutic tools for different cancer subtypes, as classified by origin, histology, aggressiveness, or chemosensitivity [37–40,45–49]. Importantly, in comparison to mRNAs, microRNAs are markedly less degraded in formalin-fixed paraffin-embedded (FFPE) samples, which are typically collected and stored in hospitals. Therefore, the easy availability of archived FFPE samples and the ability to accurately measure microRNA expression enables translation studies on microRNAs. Unlike mRNA, microRNAs exist in tissues and body fluids, such as blood and sputum. Thus, the characteristics of microRNAs support the development of liquid biopsies.

2.2. LncRNAs

LncRNAs are largely polyadenylated and comprise more than 200-nucleotide or longer transcripts. LncRNAs are also engaged in gene expression via transcriptional and epigenetic regulation, imprinting, splicing, and subcellular transport. Although the major mechanism of lncRNAs is the regulation of expression of neighboring genes, lncRNAs also serve as scaffold for protein–protein interactions or decoys to proteins [53]. Additionally, lncRNAs can regulate kinase functions. LncRNA *NBR2* engages a metabolic checkpoint by regulating AMP-activated protein kinase (AMPK) under energy stress [54]. Accumulating evidence suggests that lncRNAs play a role in fundamental biological functions, and dysregulation of lncRNAs contributes to cancer development, progression, and metastasis in many malignancies, including lung cancer [41–44]. Therefore, lncRNAs can be used as therapeutic targets.

Notably, lncRNAs can serve as minimally invasive and sensitive molecular markers for the screening and early diagnosis of lung cancer [55]. Peng et al. constructed a serum ncRNA panel (*miR-1254*, *miR-485-5p*, *miR-574-5p*, and lncRNA *MALAT1*), and tested whether the ncRNA panel could distinguish NSCLC patient samples from controls. Their results indicated that the four ncRNA panel can serve as a convenient tool for early NSCLC diagnosis [55]. LncRNAs are stable even in body fluids and show tissue-specific expression. These characteristic features make lncRNAs attractive as potential biomarkers in liquid biopsies.

3. Experimental Techniques to Detect MicroRNAs and LncRNAs

The golden standard for experimental techniques to detect microRNAs and lncRNAs is considered to be reverse transcriptase quantitative polymerase chain reaction (RT-qPCR). However, regarding the extraction methods, kits, controls, and quantification methods, no standards exist, which can affect the results [56]. Nonetheless, the traditional amplification method, RT-qPCR, has been well established and widely used because of its higher sensitivity, wider dynamic range, and higher precision [57]. Compared to the methods of quantifying lncRNAs, those of quantifying microRNAs have advanced. Because the traditional amplification-based method is not enough to fulfill the various requests of laboratory and clinical applications, RT-free qPCR, isothermal amplification methods, and some cross-platforms combined with amplification-based methods have been developed. Among them, the traditional RT-qPCR and cross-platforms including PCR-based arrays and next generation sequencing have been well designed and are useful [57].

4. MicroRNAs and Lung Cancer

A number of studies have identified aberrant microRNA expression in lung cancer, thus supporting microRNAs involvement in lung carcinogenesis [37–40]. MicroRNAs play important regulatory roles in lung cancer, including both tumor suppression and oncogenesis. Table 1 and Figure 2 show the major tumor suppressor and oncogenic microRNAs in lung cancer. In this section, tumor suppressor and oncogenic microRNAs are introduced with suggestions on their potential utility in the diagnosis and treatment of lung cancer.

4.1. Tumor Suppressor MicroRNAs

4.1.1. *Let-7*

Let-7 is one of the earliest identified tumor suppressor microRNAs in lung cancer, and its decreased expression is associated with poor prognosis [58]. *Let-7* expression is reduced in adenocarcinoma in situ (AIS) when compared with adjacent normal lung tissue, indicating that *let-7* expression is decreased during the early phase of lung carcinogenesis [59]. *Let-7* inhibits the expression of oncogenes involved in cell proliferation, such as *MYC*, *RAS*, and *HMGA2* [60,61]. Interestingly, *let-7* directly down-regulates *DICER1* expression, indicating that the global production of microRNAs may be regulated by *let-7* [62]. Furthermore, *let-7* regulates the cell cycle by inhibiting *CDK6* expression [61]. Recently, Zhou et al. showed that *miR-203* played an important role in promoting the apoptosis and inhibiting the cell

proliferation of lung cancer by downregulating LIN28B and upregulating *let-7* biogenesis. Their results demonstrate a novel regulatory network among *miR-203*, LIN28B, and *let-7* in lung cancer [63].

4.1.2. *MiR-34*

MiR-34 is directly transcribed by TP53, responding to DNA damage and oncogenic stress. *MiR-34* is an important component of TP53 tumor suppressor function [30]. Decreased expression of *miR-34* in lung cancer induces increased-expression of *miR-34* target genes, such as *BCL2*, *MET*, *PDGFRA*, and *PDGFRB*, which leads to TNF-related apoptosis-inducing ligand (TRAIL)-induced cell death. The down-regulation of *miR-34* upregulates *MET* and *BCL2*, which leads to cell proliferation [64–66]. A recent study found that tumor PD-L1 expression is regulated by TP53 via *miR-34* [67]. *MiR-34*, which is transcribed by TP53, directly binds to the PD-L1 3' untranslated region and downregulates it. The identified TP53/*miR-34*/PD-L1 axis deserves consideration for the improvement of emerging immunotherapy.

4.1.3. *MiR-200*

MiR-200 plays a critical role in the induction of epithelial-mesenchymal transition (EMT). Decreased expression of *miR-200* up-regulates its target genes, namely *CDH1* (also known as *E-cadherin*), *VIM* (also known as *vimentin*), *ZEB1*, and *ZEB2*, which leads to EMT as lung cancer progresses [68,69]. Zhang et al. evaluated the effect of decitabine, a DNA methyltransferase inhibitor, on TGF- β 1-induced EMT in NSCLC cells, considering the involvement of the *miR-200/ZEB* axis. Decitabine reversed TGF- β 1-induced EMT in NSCLC cells by downregulating *ZEB1* and *ZEB2* epigenetically by *miR-200*. They found that epigenetic regulation of the *miR-200/ZEB* axis is responsible for EMT induction by TGF- β 1 in NSCLC cells, and that decitabine inhibits EMT in NSCLC cells via *miR-200* [70].

4.1.4. *MiR-126*

The decreased expression of *miR-126* in lung cancer reduces its target *PIK3R2* and the PTEN/PI3K/AKT signaling pathway is therefore influenced, leading to the suppression of cell growth, migration, and invasion [71]. Recently, An et al. found that Matrine, which is an active component of traditional Chinese medicine, induces cell cycle arrest and apoptosis by upregulating *miR-126* in NSCLC cells [72].

4.1.5. *MiR-195*

MiR-195 expression is lower in NSCLC than in non-cancerous normal tissues and low *miR-195* expression has been associated with unfavorable overall survival of patients with NSCLC. *MiR-195* suppresses cancer growth and is associated with lower mortality in several cancers, including NSCLC. *CHEK1* is a direct target of *miR-195*, which results in decreased *CHEK1* expression in lung cancer. Up-regulation of *CHEK1* by reduced expression of *miR-195* promotes cell proliferation, migration, and invasion, and is associated with a higher overall mortality in lung cancer [73].

4.2. Oncogenic MicroRNAs

4.2.1. *MiR-21*

MiR-21 represents one of the most famous oncogenic microRNAs, and it is over-expressed in a number of malignancies, including lung cancer. High expression of *miR-21* predicts recurrence and higher mortality in NSCLC [74]. Furthermore, abundant *miR-21* exists in body fluids, being one of the most promising microRNAs in patients with lung cancer [49]. *MiR-21* promotes carcinogenesis via inhibition of negative regulators of RAS/MEK/ERK signaling pathway and apoptosis suppression. Over-expressed *miR-21* down-regulates the expression of *PDCD4*, *PTEN*, *SOCS1*, *SOCS6*, and *TPM1*, thus promoting cell proliferation and migration and inhibiting apoptosis [75–78]. *MiR-21* has been used as a biomarker to predict therapeutic responses to cisplatin [79]. Xu et al. conducted in vitro

and in vivo experiments and demonstrated that downregulation of *miR-21* suppression increased the cisplatin sensitivity of NSCLC. Regarding radiosensitivity, downregulation of *miR-21* sensitized radio-resistant NSCLC A549 cells to ionizing radiation through the inhibition of the PI3K/AKT signaling pathway [80].

In addition, certain drugs were reported to downregulate *miR-21*. Solasodine is an aglycone of solamargine and solasonine, which are the major solasodine glycosides in eggplant. Shen et al. demonstrated that solasodine inhibited the invasion of NSCLC A549 cells via the downregulation of *miR-21* and matrix metalloprotease (MMP) expression [81]. Triptolide is isolated from *Tripterygium wilfordii* plants and used in traditional Chinese medicine. Lie et al. demonstrated that triptolide reduced proliferation and enhanced the apoptosis of NSCLC PC-9 cells in a *PTEN*-dependent manner by downregulating *miR-21* [82].

4.2.2. *MiR-155*

As with *miR-21*, *miR-155* is an important oncogenic microRNA that has been suggested as a therapeutic target in NSCLC [74,78,83–85]. As with *miR-21*, *miR-155* is also a promising circulating microRNA in patients with lung cancer [49]. *MiR-155* directly targets *TP53*, whereas *TP53* directly regulates the expression of *miR-155*. This *miR-155/TP53* feedback loop is involved in chemotherapy resistance [85]. Xue et al. reported that *miR-21* and *miR-155* promote the progression of NSCLCs, in part by downregulating *SOCS1*, *SOCS6*, and *PTEN*, all of which are tumor suppressor genes. Thus, the combined inhibition of *miR-21* and *miR-155* may improve the treatment of NSCLCs [78].

4.2.3. *MiR-17-92*

The *miR-17-92* intronic cluster comprises seven different microRNAs (*miR-17-5p*, *miR-17-3p*, *miR-18a*, *miR-19a*, *miR-19b-1*, *miR-20a*, and *miR-92*) and resides in intron 3 of the *C13orf25* gene at 13q31.3 [86]. Overexpression of the *miR-17-92* cluster with occasional gene amplification plays a role in the lung carcinogenesis, especially SCLC [86]. The *MiR-17-92* cluster down-regulates *HIF1A*, *E2F1*, and *PTEN*, which leads to cell proliferation and cancer progression [87,88]. Matsubara et al. reported that antisense oligonucleotides against *miR-17-5p* and *miR-20a* within the *miR-17-92* cluster induced apoptosis in lung cancers overexpressing *miR-17-92* [87]. Recently, Li et al. found that *FLI1*, which is an Ets transcription factor family member and known as a major driver of hematological malignancies, plays an important role in tumor progression in SCLC. Furthermore, they uncovered *FLI1* as a critical driving factor that promotes cancer growth in SCLC through the *miR-17-92* pathway [89]. *FLI1* and *miR-17-92* may serve as promising therapeutic targets to improve the treatment of SCLCs.

4.2.4. *MiR-221/222*

Both *miR-221* and *miR-222* have an identical seed sequence and are predicted to have overlapping targets. *MiR-221/222* is associated with tyrosine kinase inhibitor (TKI)-resistant NSCLCs. Garofalo et al. reported that TKI-resistance was overcome by anti-*miR-221/222* and anti-*miR-30c*, which recovered *BCL2L11* expression and increased the gefitinib sensitivity of NSCLCs, thus providing a microRNA-mediated therapeutic approach for TKI-resistant NSCLCs [90]. *MiR-221/222* influences lung carcinogenesis by down-regulating *TIMP3* and *PTEN* tumor suppressor genes [91,92]. Up-regulated *miR-221/222* promotes cell migration and suppresses apoptosis by targeting *TIMP3* and *PTEN*.

4.2.5. *MiR-31*

MiR-31 targets *BAP1*, which is a necessary nuclear-located deubiquitinating enzyme that acts as a tumor suppressor in lung cancer. Increased expression of *miR-31* reduced the *BAP1* expression, leading to cell proliferation and suppressed apoptosis [93]. Edmonds et al. reported that *miR-31* directly downregulated six negative regulators of the RAS/MAPK signaling pathway (*SPRED1*, *SPRED2*, *SPRY1*, *SPRY3*, *SPRY4*, and *RASA1*) and promoted mutant *KRAS*-mediated oncogenesis [94].

Although many studies suggest that *miR-31* is an oncogenic microRNA, there are reports suggesting that *miR-31* functions as a tumor suppressor in lung cancer [95,96]. Xu et al. reported that the downregulation of *miR-31* enhanced lung cancer proliferation and migration by upregulating HuR, an RNA binding protein [95]. Okudela et al. reported that restoration and knockdown of *miR-31* in lung cancer cell lines attenuated their growth activities and enhanced oncogenic phenotypes, respectively, suggesting that *miR-31* acts as a tumor suppressor [95,96]. Further research is required to determine whether *miR-31* plays a pleiotropic role in individual tumors.

Table 1. Tumor suppressor and oncogenic microRNAs in lung cancer.

MicroRNA	Expression	Affected Biological Processes and Target Genes
Tumor suppressor microRNAs		
<i>let-7</i>	Decreased	i) Cell proliferation (<i>MYC, RAS, HMGA2</i>) [60,61] ii) MicroRNA biogenesis (<i>DICER1</i>) [62] iii) Cell cycle regulation (<i>CDK6</i>) [61]
<i>miR-34</i>	Decreased	TRAIL-induced cell death and cell proliferation (<i>BCL2, MET, PDGFRA, PDGFRB</i>) [64–66]
<i>miR-200</i>	Decreased	Promotion of EMT and metastasis (<i>CDH1, VIM, ZEB1, ZEB2</i>) [68,69]
<i>miR-126</i>	Decreased	Cell proliferation, migration, and invasion through PTEN/PI3K/AKT pathway (<i>PIK3R2</i>) [71]
<i>miR-195</i>	Decreased	Cell proliferation, migration, and invasion (<i>CHEK1</i>) [73]
Oncogenic microRNAs		
<i>miR-21</i>	Increased	Cell proliferation, migration, and apoptosis (<i>PDCD4, PTEN, SOCS1, SOCS6, TPM1</i>) [75–78]
<i>miR-155</i>	Increased	Resistance to chemotherapy (<i>TP53</i>) [85] Cell proliferation and apoptosis (<i>SOCS1, SOCS6, PTEN</i>) [78]
<i>miR-17-92</i>	Increased	Carcinogenesis and cell proliferation (<i>HIF1A, E2F1, PTEN</i>) [87,88]
<i>miR-221/222</i>	Increased	Cell migration and apoptosis (<i>PTEN, TIMP3</i>) [91,92]
<i>miR-31</i>	Increased	Cell proliferation and apoptosis (<i>BAP1</i>) [93] Promotion of KRAS/MAPK signaling (<i>SPRED1, SPRED2, SPRY1, SPRY3, SPRY4, RASA1</i>) [94]

EMT, epithelial-mesenchymal transition. TRAIL, TNF-related apoptosis-inducing ligand.

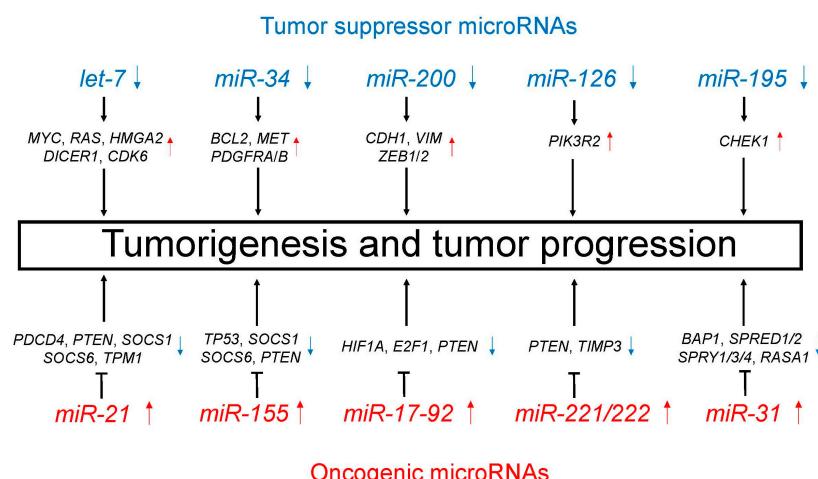


Figure 2. Tumor suppressor (blue)/oncogenic (red) microRNAs regulate unique target genes. This regulation by tumor suppressor/oncogenic microRNAs leads to tumorigenesis and tumor progression.

5. LncRNAs and Lung Cancer

Emerging evidence suggests the aberrant expression of lncRNAs in lung cancer, thus indicating a role for lncRNAs in lung carcinogenesis [41–44]. Many lncRNAs play critical regulatory roles in lung cancer as tumor suppressor or oncogenic lncRNAs. Table 2 and Figure 3 summarize major tumor suppressor and oncogenic lncRNAs in lung cancer.

5.1. Tumor Suppressor LncRNAs

5.1.1. MEG3

MEG3, which is located on 14q32.2, is a maternally expressed imprinted lncRNA found in a variety of normal tissues. MEG3 expression was reduced in several cancers, and up-regulation of MEG3 inhibits tumor growth [97]. Lu et al. reported that the expression of MEG3 was down-regulated in NSCLCs when compared to adjacent normal lung tissues; furthermore, decreased expression of MEG3 was associated with a relatively poor prognosis [98]. MEG3 inhibits NSCLC cell proliferation and induces apoptosis by up-regulating TP53 expression [97,98]. As with NSCLC, downregulated MEG3 in breast cancer also regulates proliferation, migration, and invasion by depending on the transcriptional activity of TP53 [99].

5.1.2. GAS6-AS1

GAS6-AS1, which is located on chromosome 13q34, is transcribed antisense to the GAS6 gene. Down-regulation of GAS6-AS1 promotes cancer progression in several malignancies, including lung cancer [100]. Han et al. demonstrated that GAS6-AS1 expression is lower in NSCLC than adjacent non-cancerous normal tissue and suggested downregulation of GAS6-AS1 as an independent biomarker for higher overall mortality in NSCLC patients [100]. Because the molecular mechanisms of GAS6-AS1-mediated NSCLC progression still remain elusive, further mechanical studies should be required.

5.1.3. BANCR

BANCR mediates cell growth by regulating cell-growth arrest, leading to the reduction of cancer incidence. The expression of BANCR is significantly down-regulated in NSCLC tissues when compared to adjacent non-cancerous normal lung tissues, and the lower BANCR expression has been associated with higher mortality in NSCLC patients [101]. Although the mechanical explanation why BANCR suppresses invasiveness and metastasis of NSCLCs remains unclear, it is likely associated with EMT inhibition. Loss of BANCR expression reduces CDH1 (also known as E-cadherin) expression and induces CDH2 (also known as N-cadherin), VIM (also known as vimentin), and MMPs [101]. Thus, BANCR is also a potential target of lncRNA-mediated therapeutics.

5.1.4. PANDAR

PANDAR, which is located on chromosome 6q21.2, is a tumor suppressor lncRNA. Han et al. reported that decreased expression of PANDAR was associated with higher overall mortality in NSCLC patients. A direct transcription target of PANDAR includes TP53 in NSCLC cells, and PANDAR affects cell apoptosis by regulating BCL2 [102]. Because PANDAR is a direct transcriptional target of TP53 in NSCLC, overexpression of PANDAR could inhibit the proliferation of NSCLC cells.

5.2. Oncogenic LncRNAs

5.2.1. MALAT1

MALAT1, also known as NEAT2, is a promising lncRNA, and is a candidate of biomarker in liquid biopsy for the diagnosis of NSCLCs. Furthermore, high MALAT1 expression in FFPE specimens indicates a higher mortality in NSCLCs and experimentally promotes cell proliferation,

migration, metastasis, and EMT [103–105]. One possible mechanistic explanation for the oncogenic activity of *MALAT1* is that *MALAT1* participates in aberrant alternative splicing, which results in the dysregulated expression of genes, including B-MYB transcription factor [106]. Recently, Wang et al. conducted a follow-up study for 538 patients of NSCLC, and genetic variant rs3200401 in *MALAT1* was then genotyped among this population. They demonstrated that the rs3200401 T allele located on the lncRNA *MALAT1* was associated with lower mortality for patients with advanced lung adenocarcinoma [107].

5.2.2. *HOTAIR*

HOTAIR is a lncRNA, which is located downstream, in the antisense direction, of *HOXC12* gene [108]. *HOTAIR* promotes the invasiveness and metastasis of several cancer types by recruiting PRC2 or chromatin reorganization [109]. Additionally, *HOTAIR* has been reported to participate in chemoresistance to cisplatin in lung adenocarcinoma [110]. Liu et al. also reported that elevated *HOTAIR* expression was associated with cisplatin resistance in NSCLC, and showed that *HOTAIR* expression was directly related to KLF4 expression, suggesting a new therapeutic target for drug-resistance patients with NSCLC [111].

5.2.3. *CCAT2*

CCAT2 is a novel lncRNA and its overexpression is associated with a number of cancers, including NSCLC. *CCAT2* is highly expressed in NSCLCs, especially adenocarcinoma. According to a study using NSCLC cell lines by Qiu et al. [112], reducing *CCAT2* expression by siRNA inhibits tumor proliferation and invasiveness in NSCLC cells. *CCAT2* promotes cell migration by up-regulating *MYC*, *miR-20a*, and *miR-17-5p* through *TCF7L2*-mediated transcription in cell lines [113]. Recently, Zhao et al. showed that *CCAT2* promotes tumorigenesis by overexpressing of Pokemon (also known as ZBTB7A) and suggested that the potential mechanism might relate to the Pokemon-related gene *CDKN1A* (also known as p21) [114].

Table 2. Tumor suppressor and oncogenic long non-coding RNAs (lncRNAs) in lung cancer.

MicroRNA	Chromosome	Biological Processes
Tumor suppressor lncRNAs		
MEG3	14q32.2	TP53 up-regulation [97,98]
GAS6-AS1	13q34	Unknown
BANCR	9q21.12	EMT inhibition [101]
PANDAR	6q21.2	Apoptosis by BCL2 down-regulation [102]
Oncogenic lncRNAs		
<i>MALAT1</i>	11q13.1	Cell proliferation, migration, metastasis, and EMT [105]
<i>HOTAIR</i>	12q13.13	Invasiveness and metastasis by recruiting PRC2 or reorganizing chromatin [109]
<i>CCAT2</i>	8q24.21	Cell migration by up-regulating of <i>MYC</i> , <i>miR-17-5p</i> , and <i>miR-20a</i> through <i>TCF7L2</i> -mediated transcription [113]

EMT, epithelial-mesenchymal transition; lncRNA, long non-coding RNA.

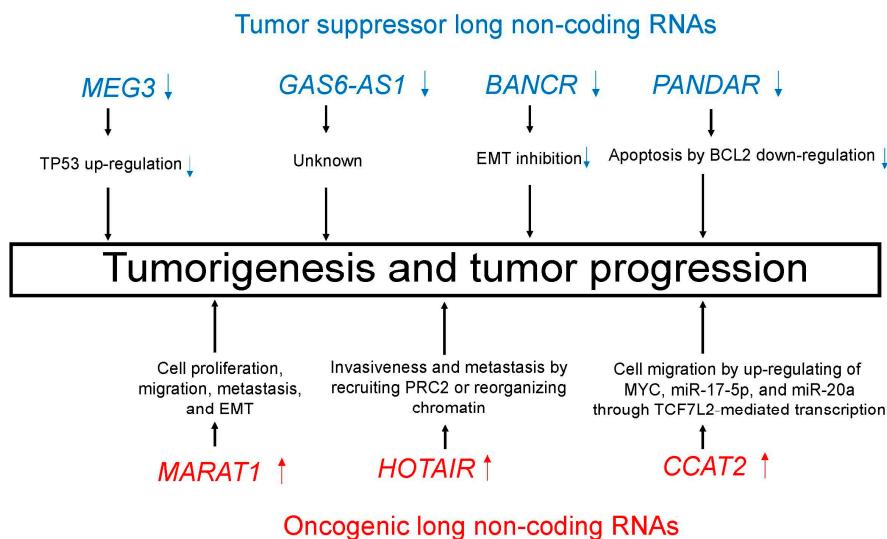


Figure 3. Tumor suppressor (blue)/oncogenic (red) long non-coding RNAs (lncRNAs) induce important processes that lead to tumorigenesis and tumor progression.

6. Conclusions and Future Directions

In this review, recent studies of microRNAs and lncRNAs in lung cancer are introduced with a focus on potential biological and therapeutic applications. Accumulating evidence suggests that microRNAs and lncRNAs represent very promising biomarkers in patients with NSCLC and will be markedly useful in non-invasive screening methods. MicroRNA- or lncRNA-mediated therapy for patients with lung cancer is also very promising. Further studies and clinical trials are needed to assess microRNA or lncRNA profiles as diagnostic markers and conduct microRNA- and lncRNA-based therapies in clinical practice.

Acknowledgments: This study was supported by JSPS KAKENHI Grant Number JP16K08679; and the Ministry of the Environment, Japan.

Author Contributions: Kentaro Inamura conceived the review, wrote the text, and created the tables and figure.

Conflicts of Interest: The author declares no conflicts of interest.

References

1. Torre, L.A.; Bray, F.; Siegel, R.L.; Ferlay, J.; Lortet-Tieulent, J.; Jemal, A. Global cancer statistics, 2012. *CA Cancer J. Clin.* **2015**, *65*, 87–108. [[CrossRef](#)] [[PubMed](#)]
2. Travis, W.D.; Brambilla, E.; Burke, A.P.; Marx, A.; Nicholson, A.G. *WHO Classification of Tumours of the Lung, Pleura, Thymus and Heart*, 4th ed.; IARC: Lyon, France, 2015.
3. The Cancer Genome Atlas Research Network. Comprehensive molecular profiling of lung adenocarcinoma. *Nature* **2014**, *511*, 543–550.
4. The Cancer Genome Atlas Research Network. Comprehensive genomic characterization of squamous cell lung cancers. *Nature* **2012**, *489*, 519–525.
5. George, J.; Lim, J.S.; Jang, S.J.; Cun, Y.; Ozretic, L.; Kong, G.; Leenders, F.; Lu, X.; Fernandez-Cuesta, L.; Bosco, G.; et al. Comprehensive genomic profiles of small cell lung cancer. *Nature* **2015**, *524*, 47–53. [[CrossRef](#)] [[PubMed](#)]
6. Rudin, C.M.; Durinck, S.; Stawiski, E.W.; Poirier, J.T.; Modrusan, Z.; Shames, D.S.; Bergbower, E.A.; Guan, Y.; Shin, J.; Guillory, J.; et al. Comprehensive genomic analysis identifies SOX2 as a frequently amplified gene in small-cell lung cancer. *Nat. Genet.* **2012**, *44*, 1111–1116. [[CrossRef](#)] [[PubMed](#)]

7. Inamura, K.; Fujiwara, T.; Hoshida, Y.; Isagawa, T.; Jones, M.H.; Virtanen, C.; Shimane, M.; Satoh, Y.; Okumura, S.; Nakagawa, K.; et al. Two subclasses of lung squamous cell carcinoma with different gene expression profiles and prognosis identified by hierarchical clustering and non-negative matrix factorization. *Oncogene* **2005**, *24*, 7105–7113. [[CrossRef](#)] [[PubMed](#)]
8. Inamura, K.; Satoh, Y.; Okumura, S.; Nakagawa, K.; Tsuchiya, E.; Fukayama, M.; Ishikawa, Y. Pulmonary adenocarcinomas with enteric differentiation: Histologic and immunohistochemical characteristics compared with metastatic colorectal cancers and usual pulmonary adenocarcinomas. *Am. J. Surg. Pathol.* **2005**, *29*, 660–665. [[CrossRef](#)] [[PubMed](#)]
9. Shaw, A.T.; Yeap, B.Y.; Mino-Kenudson, M.; Digumarthy, S.R.; Costa, D.B.; Heist, R.S.; Solomon, B.; Stubbs, H.; Admire, S.; McDermott, U.; et al. Clinical features and outcome of patients with non-small-cell lung cancer who harbor EML4-ALK. *J. Clin. Oncol.* **2009**, *27*, 4247–4253. [[CrossRef](#)] [[PubMed](#)]
10. Takeuchi, K.; Soda, M.; Togashi, Y.; Suzuki, R.; Sakata, S.; Hatano, S.; Asaka, R.; Hamanaka, W.; Ninomiya, H.; Uehara, H.; et al. RET, ROS1 and ALK fusions in lung cancer. *Nat. Med.* **2012**, *18*, 378–381. [[CrossRef](#)] [[PubMed](#)]
11. Inamura, K.; Yokouchi, Y.; Sakakibara, R.; Kobayashi, M.; Subat, S.; Ninomiya, H.; Nagano, H.; Nomura, K.; Okumura, S.; Ishikawa, Y. Relationship of tumor PD-L1 expression with EGFR wild-type status and poor prognosis in lung adenocarcinoma. *Jpn. J. Clin. Oncol.* **2016**, *46*, 935–941. [[CrossRef](#)] [[PubMed](#)]
12. Brambilla, E.; Le Teuff, G.; Marguet, S.; Lantuejoul, S.; Dunant, A.; Graziano, S.; Pirker, R.; Douillard, J.Y.; Le Chevalier, T.; Filipits, M.; et al. Prognostic Effect of Tumor Lymphocytic Infiltration in Resectable Non-Small-Cell Lung Cancer. *J. Clin. Oncol.* **2016**, *34*, 1223–1230. [[CrossRef](#)] [[PubMed](#)]
13. Awad, M.M.; Oxnard, G.R.; Jackman, D.M.; Savukoski, D.O.; Hall, D.; Shviddasani, P.; Heng, J.C.; Dahlberg, S.E.; Janne, P.A.; Verma, S.; et al. MET Exon 14 Mutations in Non-Small-Cell Lung Cancer Are Associated With Advanced Age and Stage-Dependent MET Genomic Amplification and c-Met Overexpression. *J. Clin. Oncol.* **2016**, *34*, 721–730. [[CrossRef](#)] [[PubMed](#)]
14. Inamura, K.; Ninomiya, H.; Nomura, K.; Tsuchiya, E.; Satoh, Y.; Okumura, S.; Nakagawa, K.; Takata, A.; Kohyama, N.; Ishikawa, Y. Combined effects of asbestos and cigarette smoke on the development of lung adenocarcinoma: Different carcinogens may cause different genomic changes. *Oncol. Rep.* **2014**, *32*, 475–482. [[CrossRef](#)] [[PubMed](#)]
15. Celiktas, M.; Tanaka, I.; Chandra Tripathi, S.; Fahrmann, J.F.; Aguilar-Bonavides, C.; Villalobos, P.; Delgado, O.; Dhillon, D.; Dennison, J.B.; Ostrin, E.J.; et al. Role of CPS1 in Cell Growth, Metabolism, and Prognosis in LKB1-Inactivated Lung Adenocarcinoma. *J. Natl. Cancer Inst.* **2017**, *109*, 1–9. [[CrossRef](#)] [[PubMed](#)]
16. Inamura, K.; Yokouchi, Y.; Kobayashi, M.; Sakakibara, R.; Ninomiya, H.; Subat, S.; Nagano, H.; Nomura, K.; Okumura, S.; Shibutani, T.; et al. Tumor B7-H3 (CD276) expression and smoking history in relation to lung adenocarcinoma prognosis. *Lung Cancer* **2017**, *103*, 44–51. [[CrossRef](#)] [[PubMed](#)]
17. Zhang, Y.; Li, Z.Y.; Hou, X.X.; Wang, X.; Luo, Y.H.; Ying, Y.P.; Chen, G. Clinical significance and effect of AEG-1 on the proliferation, invasion, and migration of NSCLC: A study based on immunohistochemistry, TCGA, bioinformatics, in vitro and in vivo verification. *Oncotarget* **2017**, *8*, 16531–16552. [[CrossRef](#)] [[PubMed](#)]
18. Inamura, K.; Yokouchi, Y.; Kobayashi, M.; Ninomiya, H.; Sakakibara, R.; Subat, S.; Nagano, H.; Nomura, K.; Okumura, S.; Shibutani, T.; et al. Association of tumor TROP2 expression with prognosis varies among lung cancer subtypes. *Oncotarget* **2017**, *8*, 28725–28735. [[CrossRef](#)] [[PubMed](#)]
19. Lynch, T.J.; Bell, D.W.; Sordella, R.; Gurubhagavatula, S.; Okimoto, R.A.; Brannigan, B.W.; Harris, P.L.; Haserlat, S.M.; Supko, J.G.; Haluska, F.G.; et al. Activating mutations in the epidermal growth factor receptor underlying responsiveness of non-small-cell lung cancer to gefitinib. *N. Engl. J. Med.* **2004**, *350*, 2129–2139. [[CrossRef](#)] [[PubMed](#)]
20. Kwak, E.L.; Bang, Y.J.; Camidge, D.R.; Shaw, A.T.; Solomon, B.; Maki, R.G.; Ou, S.H.; Dezube, B.J.; Janne, P.A.; Costa, D.B.; et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N. Engl. J. Med.* **2010**, *363*, 1693–1703. [[CrossRef](#)] [[PubMed](#)]
21. Shaw, A.T.; Ou, S.H.; Bang, Y.J.; Camidge, D.R.; Solomon, B.J.; Salgia, R.; Riely, G.J.; Varella-Garcia, M.; Shapiro, G.I.; Costa, D.B.; et al. Crizotinib in ROS1-rearranged non-small-cell lung cancer. *N. Engl. J. Med.* **2014**, *371*, 1963–1971. [[CrossRef](#)] [[PubMed](#)]

22. Garon, E.B.; Rizvi, N.A.; Hui, R.; Leighl, N.; Balmanoukian, A.S.; Eder, J.P.; Patnaik, A.; Aggarwal, C.; Gubens, M.; Horn, L.; et al. Pembrolizumab for the treatment of non-small-cell lung cancer. *N. Engl. J. Med.* **2015**, *372*, 2018–2028. [CrossRef] [PubMed]
23. Goldenberg, D.M.; Cardillo, T.M.; Govindan, S.V.; Rossi, E.A.; Sharkey, R.M. Trop-2 is a novel target for solid cancer therapy with sacituzumab govitecan (IMMU-132), an antibody-drug conjugate (ADC). *Oncotarget* **2015**, *6*, 22496–22512. [CrossRef] [PubMed]
24. Shaw, A.T.; Hsu, P.P.; Awad, M.M.; Engelman, J.A. Tyrosine kinase gene rearrangements in epithelial malignancies. *Nat. Rev. Cancer* **2013**, *13*, 772–787. [CrossRef] [PubMed]
25. Boolell, V.; Alamgeer, M.; Watkins, D.N.; Ganju, V. The Evolution of Therapies in Non-Small Cell Lung Cancer. *Cancers (Basel)* **2015**, *7*, 1815–1846. [CrossRef] [PubMed]
26. Rothschild, S.I. Targeted Therapies in Non-Small Cell Lung Cancer-Beyond EGFR and ALK. *Cancers (Basel)* **2015**, *7*, 930–949. [CrossRef] [PubMed]
27. Manchado, E.; Weissmueller, S.; Morris, J.P.T.; Chen, C.C.; Wullenkord, R.; Lujambio, A.; de Stanchina, E.; Poirier, J.T.; Gainor, J.F.; Corcoran, R.B.; et al. A combinatorial strategy for treating KRAS-mutant lung cancer. *Nature* **2016**, *534*, 647–651. [CrossRef] [PubMed]
28. Lu, J.; Getz, G.; Miska, E.A.; Alvarez-Saavedra, E.; Lamb, J.; Peck, D.; Sweet-Cordero, A.; Ebert, B.L.; Mak, R.H.; Ferrando, A.A.; et al. MicroRNA expression profiles classify human cancers. *Nature* **2005**, *435*, 834–838. [CrossRef] [PubMed]
29. Pasquinelli, A.E.; Reinhart, B.J.; Slack, F.; Martindale, M.Q.; Kuroda, M.I.; Maller, B.; Hayward, D.C.; Ball, E.E.; Degnan, B.; Muller, P.; et al. Conservation of the sequence and temporal expression of let-7 heterochronic regulatory RNA. *Nature* **2000**, *408*, 86–89. [PubMed]
30. He, L.; He, X.; Lim, L.P.; de Stanchina, E.; Xuan, Z.; Liang, Y.; Xue, W.; Zender, L.; Magnus, J.; Ridzon, D.; et al. A microRNA component of the p53 tumour suppressor network. *Nature* **2007**, *447*, 1130–1134. [CrossRef] [PubMed]
31. Takahashi, R.U.; Miyazaki, H.; Ochiya, T. The Roles of MicroRNAs in Breast Cancer. *Cancers (Basel)* **2015**, *7*, 598–616. [CrossRef] [PubMed]
32. Takahashi, R.U.; Miyazaki, H.; Takeshita, F.; Yamamoto, Y.; Minoura, K.; Ono, M.; Kodaira, M.; Tamura, K.; Mori, M.; Ochiya, T. Loss of microRNA-27b contributes to breast cancer stem cell generation by activating ENPP1. *Nat. Commun.* **2015**, *6*, 7318. [CrossRef] [PubMed]
33. Orellana, E.A.; Kasinski, A.L. MicroRNAs in Cancer: A Historical Perspective on the Path from Discovery to Therapy. *Cancers (Basel)* **2015**, *7*, 1388–1405. [CrossRef] [PubMed]
34. Frixa, T.; Donzelli, S.; Blandino, G. Oncogenic MicroRNAs: Key Players in Malignant Transformation. *Cancers (Basel)* **2015**, *7*, 2466–2485. [CrossRef] [PubMed]
35. Vaca, L. Point-of-care diagnostic tools to detect circulating microRNAs as biomarkers of disease. *Sensors (Basel)* **2014**, *14*, 9117–9131. [CrossRef] [PubMed]
36. Ajit, S.K. Circulating microRNAs as biomarkers, therapeutic targets, and signaling molecules. *Sensors (Basel)* **2012**, *12*, 3359–3369. [CrossRef] [PubMed]
37. Yanaihara, N.; Caplen, N.; Bowman, E.; Seike, M.; Kumamoto, K.; Yi, M.; Stephens, R.M.; Okamoto, A.; Yokota, J.; Tanaka, T.; et al. Unique microRNA molecular profiles in lung cancer diagnosis and prognosis. *Cancer Cell* **2006**, *9*, 189–198. [CrossRef] [PubMed]
38. Yu, S.L.; Chen, H.Y.; Chang, G.C.; Chen, C.Y.; Chen, H.W.; Singh, S.; Cheng, C.L.; Yu, C.J.; Lee, Y.C.; Chen, H.S.; et al. MicroRNA signature predicts survival and relapse in lung cancer. *Cancer Cell* **2008**, *13*, 48–57. [CrossRef] [PubMed]
39. Inamura, K.; Ishikawa, Y. MicroRNA In Lung Cancer: Novel Biomarkers and Potential Tools for Treatment. *J. Clin. Med.* **2016**, *5*, 36. [CrossRef] [PubMed]
40. MacDonagh, L.; Gray, S.G.; Finn, S.P.; Cuffe, S.; O’Byrne, K.J.; Barr, M.P. The emerging role of microRNAs in resistance to lung cancer treatments. *Cancer Treat. Rev.* **2015**, *41*, 160–169. [CrossRef] [PubMed]
41. Chen, J.; Wang, R.; Zhang, K.; Chen, L.B. Long non-coding RNAs in non-small cell lung cancer as biomarkers and therapeutic targets. *J. Cell Mol. Med.* **2014**, *18*, 2425–2436. [CrossRef] [PubMed]
42. Wei, M.M.; Zhou, G.B. Long Non-coding RNAs and Their Roles in Non-small-cell Lung Cancer. *Genom. Proteom. Bioinform.* **2016**, *14*, 280–288. [CrossRef] [PubMed]
43. Xu, Y.J.; Du, Y.; Fan, Y. Long noncoding RNAs in lung cancer: What we know in 2015. *Clin. Transl. Oncol.* **2016**, *18*, 660–665. [CrossRef] [PubMed]

44. Ricciuti, B.; Mencaroni, C.; Paglialunga, L.; Paciullo, F.; Crino, L.; Chiari, R.; Metro, G. Long noncoding RNAs: New insights into non-small cell lung cancer biology, diagnosis and therapy. *Med. Oncol.* **2016**, *33*, 18. [[CrossRef](#)] [[PubMed](#)]
45. Booton, R.; Lindsay, M.A. Emerging role of MicroRNAs and long noncoding RNAs in respiratory disease. *Chest* **2014**, *146*, 193–204. [[CrossRef](#)] [[PubMed](#)]
46. Matikas, A.; Syrigos, K.N.; Agelaki, S. Circulating Biomarkers in Non-Small-Cell Lung Cancer: Current Status and Future Challenges. *Clin. Lung Cancer* **2016**, *17*, 507–516. [[CrossRef](#)] [[PubMed](#)]
47. Murlidhar, V.; Ramnath, N.; Nagrath, S.; Reddy, R.M. Optimizing the Detection of Circulating Markers to Aid in Early Lung Cancer Detection. *Cancers (Basel)* **2016**, *8*, 61. [[CrossRef](#)] [[PubMed](#)]
48. Kunz, M.; Wolf, B.; Schulze, H.; Atlan, D.; Walles, T.; Walles, H.; Dandekar, T. Non-Coding RNAs in Lung Cancer: Contribution of Bioinformatics Analysis to the Development of Non-Invasive Diagnostic Tools. *Genes (Basel)* **2017**, *8*, 8. [[CrossRef](#)] [[PubMed](#)]
49. Zhao, C.; Lu, F.; Chen, H.; Zhao, F.; Zhu, Z.; Zhao, X.; Chen, H. Clinical significance of circulating miRNA detection in lung cancer. *Med. Oncol.* **2016**, *33*, 41. [[CrossRef](#)] [[PubMed](#)]
50. ENCODE Project Consortium. An integrated encyclopedia of DNA elements in the human genome. *Nature* **2012**, *489*, 57–74.
51. Djebali, S.; Davis, C.A.; Merkel, A.; Dobin, A.; Lassmann, T.; Mortazavi, A.; Tanzer, A.; Lagarde, J.; Lin, W.; Schlesinger, F.; et al. Landscape of transcription in human cells. *Nature* **2012**, *489*, 101–108. [[CrossRef](#)] [[PubMed](#)]
52. Bartel, D.P. MicroRNAs: Genomics, biogenesis, mechanism, and function. *Cell* **2004**, *116*, 281–297. [[CrossRef](#)]
53. Xiao, Z.D.; Zhuang, L.; Gan, B. Long non-coding RNAs in cancer metabolism. *Bioessays* **2016**, *38*, 991–996. [[CrossRef](#)] [[PubMed](#)]
54. Liu, X.; Xiao, Z.D.; Han, L.; Zhang, J.; Lee, S.W.; Wang, W.; Lee, H.; Zhuang, L.; Chen, J.; Lin, H.K.; et al. LncRNA NBR2 engages a metabolic checkpoint by regulating AMPK under energy stress. *Nat. Cell Biol.* **2016**, *18*, 431–442. [[CrossRef](#)] [[PubMed](#)]
55. Peng, H.; Wang, J.; Li, J.; Zhao, M.; Huang, S.K.; Gu, Y.Y.; Li, Y.; Sun, X.J.; Yang, L.; Luo, Q.; et al. A circulating non-coding RNA panel as an early detection predictor of non-small cell lung cancer. *Life Sci.* **2016**, *151*, 235–242. [[CrossRef](#)] [[PubMed](#)]
56. Shi, T.; Gao, G.; Cao, Y. Long Noncoding RNAs as Novel Biomarkers Have a Promising Future in Cancer Diagnostics. *Dis. Markers* **2016**, *2016*, 9085195. [[CrossRef](#)] [[PubMed](#)]
57. Shen, Y.; Tian, F.; Chen, Z.; Li, R.; Ge, Q.; Lu, Z. Amplification-based method for microRNA detection. *Biosens. Bioelectron.* **2015**, *71*, 322–331. [[CrossRef](#)] [[PubMed](#)]
58. Takamizawa, J.; Konishi, H.; Yanagisawa, K.; Tomida, S.; Osada, H.; Endoh, H.; Harano, T.; Yatabe, Y.; Nagino, M.; Nimura, Y.; et al. Reduced expression of the let-7 microRNAs in human lung cancers in association with shortened postoperative survival. *Cancer Res.* **2004**, *64*, 3753–3756. [[CrossRef](#)] [[PubMed](#)]
59. Inamura, K.; Togashi, Y.; Nomura, K.; Ninomiya, H.; Hiramatsu, M.; Satoh, Y.; Okumura, S.; Nakagawa, K.; Ishikawa, Y. let-7 microRNA expression is reduced in bronchioloalveolar carcinoma, a non-invasive carcinoma, and is not correlated with prognosis. *Lung Cancer* **2007**, *58*, 392–396. [[CrossRef](#)] [[PubMed](#)]
60. Johnson, S.M.; Grosshans, H.; Shingara, J.; Byrom, M.; Jarvis, R.; Cheng, A.; Labourier, E.; Reinert, K.L.; Brown, D.; Slack, F.J. RAS is regulated by the let-7 microRNA family. *Cell* **2005**, *120*, 635–647. [[CrossRef](#)] [[PubMed](#)]
61. Johnson, C.D.; Esquela-Kerscher, A.; Stefani, G.; Byrom, M.; Kelnar, K.; Ovcharenko, D.; Wilson, M.; Wang, X.; Shelton, J.; Shingara, J.; et al. The let-7 microRNA represses cell proliferation pathways in human cells. *Cancer Res.* **2007**, *67*, 7713–7722. [[CrossRef](#)] [[PubMed](#)]
62. Tokumaru, S.; Suzuki, M.; Yamada, H.; Nagino, M.; Takahashi, T. let-7 regulates Dicer expression and constitutes a negative feedback loop. *Carcinogenesis* **2008**, *29*, 2073–2077. [[CrossRef](#)] [[PubMed](#)]
63. Zhou, Y.; Liang, H.; Liao, Z.; Wang, Y.; Hu, X.; Chen, X.; Xu, L.; Hu, Z. miR-203 enhances let-7 biogenesis by targeting LIN28B to suppress tumor growth in lung cancer. *Sci. Rep.* **2017**, *7*, 42680. [[CrossRef](#)] [[PubMed](#)]
64. Bommer, G.T.; Gerin, I.; Feng, Y.; Kaczorowski, A.J.; Kuick, R.; Love, R.E.; Zhai, Y.; Giordano, T.J.; Qin, Z.S.; Moore, B.B.; et al. p53-mediated activation of miRNA34 candidate tumor-suppressor genes. *Curr. Biol.* **2007**, *17*, 1298–1307. [[CrossRef](#)] [[PubMed](#)]

65. Kasinski, A.L.; Slack, F.J. miRNA-34 prevents cancer initiation and progression in a therapeutically resistant K-ras and p53-induced mouse model of lung adenocarcinoma. *Cancer Res.* **2012**, *72*, 5576–5587. [CrossRef] [PubMed]
66. Garofalo, M.; Jeon, Y.J.; Nuovo, G.J.; Middleton, J.; Secchiero, P.; Joshi, P.; Alder, H.; Nazaryan, N.; Di Leva, G.; Romano, G.; et al. MiR-34a/c-Dependent PDGFR-alpha/beta Downregulation Inhibits Tumorigenesis and Enhances TRAIL-Induced Apoptosis in Lung Cancer. *PLoS ONE* **2013**, *8*, e67581. [CrossRef] [PubMed]
67. Cortez, M.A.; Ivan, C.; Valdecana, D.; Wang, X.; Peltier, H.J.; Ye, Y.; Araujo, L.; Carbone, D.P.; Shilo, K.; Giri, D.K.; et al. PDL1 Regulation by p53 via miR-34. *J. Natl. Cancer Inst.* **2015**, *108*. [CrossRef] [PubMed]
68. Ceppi, P.; Mudduluru, G.; Kumarswamy, R.; Rapa, I.; Scagliotti, G.V.; Papotti, M.; Allgayer, H. Loss of miR-200c expression induces an aggressive, invasive, and chemoresistant phenotype in non-small cell lung cancer. *Mol. Cancer Res.* **2010**, *8*, 1207–1216. [CrossRef] [PubMed]
69. Takeyama, Y.; Sato, M.; Horio, M.; Hase, T.; Yoshida, K.; Yokoyama, T.; Nakashima, H.; Hashimoto, N.; Sekido, Y.; Gazdar, A.F.; et al. Knockdown of ZEB1, a master epithelial-to-mesenchymal transition (EMT) gene, suppresses anchorage-independent cell growth of lung cancer cells. *Cancer Lett.* **2010**, *296*, 216–224. [CrossRef] [PubMed]
70. Zhang, N.; Liu, Y.; Wang, Y.; Zhao, M.; Tu, L.; Luo, F. Decitabine reverses TGF-beta1-induced epithelial-mesenchymal transition in non-small-cell lung cancer by regulating miR-200/ZEB axis. *Drug Des. Devel. Ther.* **2017**, *11*, 969–983. [CrossRef] [PubMed]
71. Song, L.; Li, D.; Gu, Y.; Wen, Z.M.; Jie, J.; Zhao, D.; Peng, L.P. MicroRNA-126 Targeting PIK3R2 Inhibits NSCLC A549 Cell Proliferation, Migration, and Invasion by Regulation of PTEN/PI3K/AKT Pathway. *Clin. Lung Cancer* **2016**, *17*, e65–e75. [CrossRef] [PubMed]
72. An, Q.; Han, C.; Zhou, Y.; Li, F.; Li, D.; Zhang, X.; Yu, Z.; Duan, Z.; Kan, Q. Matrine induces cell cycle arrest and apoptosis with recovery of the expression of miR-126 in the A549 non-small cell lung cancer cell line. *Mol. Med. Rep.* **2016**, *14*, 4042–4048. [CrossRef] [PubMed]
73. Liu, B.; Qu, J.; Xu, F.; Guo, Y.; Wang, Y.; Yu, H.; Qian, B. MiR-195 suppresses non-small cell lung cancer by targeting CHEK1. *Oncotarget* **2015**, *6*, 9445–9456. [CrossRef] [PubMed]
74. Yang, M.; Shen, H.; Qiu, C.; Ni, Y.; Wang, L.; Dong, W.; Liao, Y.; Du, J. High expression of miR-21 and miR-155 predicts recurrence and unfavourable survival in non-small cell lung cancer. *Eur. J. Cancer* **2013**, *49*, 604–615. [CrossRef] [PubMed]
75. Zhang, J.G.; Wang, J.J.; Zhao, F.; Liu, Q.; Jiang, K.; Yang, G.H. MicroRNA-21 (miR-21) represses tumor suppressor PTEN and promotes growth and invasion in non-small cell lung cancer (NSCLC). *Clin. Chim. Acta* **2010**, *411*, 846–852. [CrossRef] [PubMed]
76. Bhatti, I.; Lee, A.; James, V.; Hall, R.I.; Lund, J.N.; Tufarelli, C.; Lobo, D.N.; Larvin, M. Knockdown of microRNA-21 inhibits proliferation and increases cell death by targeting programmed cell death 4 (PDCD4) in pancreatic ductal adenocarcinoma. *J. Gastrointest. Surg.* **2011**, *15*, 199–208. [CrossRef] [PubMed]
77. Zhu, S.; Wu, H.; Wu, F.; Nie, D.; Sheng, S.; Mo, Y.Y. MicroRNA-21 targets tumor suppressor genes in invasion and metastasis. *Cell Res.* **2008**, *18*, 350–359. [CrossRef] [PubMed]
78. Xue, X.; Liu, Y.; Wang, Y.; Meng, M.; Wang, K.; Zang, X.; Zhao, S.; Sun, X.; Cui, L.; Pan, L.; et al. MiR-21 and MiR-155 promote non-small cell lung cancer progression by downregulating SOCS1, SOCS6, and PTEN. *Oncotarget* **2016**, *7*, 84508–84519. [CrossRef] [PubMed]
79. Wei, J.; Gao, W.; Zhu, C.J.; Liu, Y.Q.; Mei, Z.; Cheng, T.; Shu, Y.Q. Identification of plasma microRNA-21 as a biomarker for early detection and chemosensitivity of non-small cell lung cancer. *Chin. J. Cancer* **2011**, *30*, 407–414. [CrossRef] [PubMed]
80. Ma, Y.; Xia, H.; Liu, Y.; Li, M. Silencing miR-21 sensitizes non-small cell lung cancer A549 cells to ionizing radiation through inhibition of PI3K/Akt. *Biomed. Res. Int.* **2014**, *2014*, 617868. [CrossRef] [PubMed]
81. Shen, K.H.; Hung, J.H.; Chang, C.W.; Weng, Y.T.; Wu, M.J.; Chen, P.S. Solasodine inhibits invasion of human lung cancer cell through downregulation of miR-21 and MMPs expression. *Chem. Biol. Interact.* **2017**, *268*, 129–135. [CrossRef] [PubMed]
82. Li, X.; Zang, A.; Jia, Y.; Zhang, J.; Fan, W.; Feng, J.; Duan, M.; Zhang, L.; Huo, R.; Jiao, J.; et al. Triptolide reduces proliferation and enhances apoptosis of human non-small cell lung cancer cells through PTEN by targeting miR-21. *Mol. Med. Rep.* **2016**, *13*, 2763–2768. [CrossRef] [PubMed]

83. Lv, X.; Yao, L.; Zhang, J.; Han, P.; Li, C. Inhibition of microRNA-155 sensitizes lung cancer cells to irradiation via suppression of HK2-modulated glucose metabolism. *Mol. Med. Rep.* **2016**, *14*, 1332–1338. [CrossRef] [PubMed]
84. Lv, L.; An, X.; Li, H.; Ma, L. Effect of miR-155 knockdown on the reversal of doxorubicin resistance in human lung cancer A549/dox cells. *Oncol. Lett.* **2016**, *11*, 1161–1166. [CrossRef] [PubMed]
85. Van Roosbroeck, K.; Fanini, F.; Setoyama, T.; Ivan, C.; Rodriguez-Aguayo, C.; Fuentes-Mattei, E.; Xiao, L.; Vannini, I.; Redis, R.; D'Abundo, L.; et al. Combining anti-miR-155 with chemotherapy for the treatment of lung cancers. *Clin. Cancer Res.* **2016**. [CrossRef] [PubMed]
86. Hayashita, Y.; Osada, H.; Tatematsu, Y.; Yamada, H.; Yanagisawa, K.; Tomida, S.; Yatabe, Y.; Kawahara, K.; Sekido, Y.; Takahashi, T. A polycistronic microRNA cluster, miR-17-92, is overexpressed in human lung cancers and enhances cell proliferation. *Cancer Res.* **2005**, *65*, 9628–9632. [CrossRef] [PubMed]
87. Matsubara, H.; Takeuchi, T.; Nishikawa, E.; Yanagisawa, K.; Hayashita, Y.; Ebi, H.; Yamada, H.; Suzuki, M.; Nagino, M.; Nimura, Y.; et al. Apoptosis induction by antisense oligonucleotides against miR-17-5p and miR-20a in lung cancers overexpressing miR-17-92. *Oncogene* **2007**, *26*, 6099–6105. [CrossRef] [PubMed]
88. Osada, H.; Takahashi, T. let-7 and miR-17-92: Small-sized major players in lung cancer development. *Cancer Sci.* **2011**, *102*, 9–17. [CrossRef] [PubMed]
89. Li, L.; Song, W.; Yan, X.; Li, A.; Zhang, X.; Li, W.; Zhou, X.W.L.; Yu, D.; Hu, J.F.; Cui, J. Friend leukemia virus integration 1 promotes tumorigenesis of small cell lung cancer cells by activating the miR-17-92 pathway. *Oncotarget* **2017**, in press. [CrossRef] [PubMed]
90. Garofalo, M.; Romano, G.; Di Leva, G.; Nuovo, G.; Jeon, Y.J.; Ngankeu, A.; Sun, J.; Lovat, F.; Alder, H.; Condorelli, G.; et al. EGFR and MET receptor tyrosine kinase-altered microRNA expression induces tumorigenesis and gefitinib resistance in lung cancers. *Nat. Med.* **2011**, *18*, 74–82. [CrossRef] [PubMed]
91. Garofalo, M.; Quintavalle, C.; Di Leva, G.; Zanca, C.; Romano, G.; Taccioli, C.; Liu, C.G.; Croce, C.M.; Condorelli, G. MicroRNA signatures of TRAIL resistance in human non-small cell lung cancer. *Oncogene* **2008**, *27*, 3845–3855. [CrossRef] [PubMed]
92. Garofalo, M.; Di Leva, G.; Romano, G.; Nuovo, G.; Suh, S.S.; Ngankeu, A.; Taccioli, C.; Pichiorri, F.; Alder, H.; Secchiero, P.; et al. miR-221&222 regulate TRAIL resistance and enhance tumorigenicity through PTEN and TIMP3 downregulation. *Cancer Cell* **2009**, *16*, 498–509. [PubMed]
93. Yu, M.; Liang, H.; Fu, Z.; Wang, X.; Liao, Z.; Zhou, Y.; Liu, Y.; Wang, Y.; Hong, Y.; Zhou, X.; et al. BAP1 suppresses lung cancer progression and is inhibited by miR-31. *Oncotarget* **2016**, *7*, 13742–13753. [PubMed]
94. Edmonds, M.D.; Boyd, K.L.; Moyo, T.; Mitra, R.; Duszynski, R.; Arrate, M.P.; Chen, X.; Zhao, Z.; Blackwell, T.S.; Andl, T.; et al. MicroRNA-31 initiates lung tumorigenesis and promotes mutant KRAS-driven lung cancer. *J. Clin. Investig.* **2016**, *126*, 349–364. [CrossRef] [PubMed]
95. Xu, H.; Ma, J.; Zheng, J.; Wu, J.; Qu, C.; Sun, F.; Xu, S. MiR-31 Functions as a Tumor Suppressor in Lung Adenocarcinoma Mainly by Targeting HuR. *Clin. Lab.* **2016**, *62*, 711–718. [CrossRef] [PubMed]
96. Okudela, K.; Suzuki, T.; Umeda, S.; Tateishi, Y.; Mitsui, H.; Miyagi, Y.; Ohashi, K. A comprehensive search for microRNAs with expression profiles modulated by oncogenic KRAS: Potential involvement of miR-31 in lung carcinogenesis. *Oncol. Rep.* **2014**, *32*, 1374–1384. [CrossRef] [PubMed]
97. Zhou, Y.; Zhong, Y.; Wang, Y.; Zhang, X.; Batista, D.L.; Gejman, R.; Ansell, P.J.; Zhao, J.; Weng, C.; Klibanski, A. Activation of p53 by MEG3 non-coding RNA. *J. Biol. Chem.* **2007**, *282*, 24731–24742. [CrossRef] [PubMed]
98. Lu, K.H.; Li, W.; Liu, X.H.; Sun, M.; Zhang, M.L.; Wu, W.Q.; Xie, W.P.; Hou, Y.Y. Long non-coding RNA MEG3 inhibits NSCLC cells proliferation and induces apoptosis by affecting p53 expression. *BMC Cancer* **2013**, *13*, 461. [CrossRef] [PubMed]
99. Sun, L.; Li, Y.; Yang, B. Downregulated long non-coding RNA MEG3 in breast cancer regulates proliferation, migration and invasion by depending on p53's transcriptional activity. *Biochem. Biophys. Res. Commun.* **2016**, *478*, 323–329. [CrossRef] [PubMed]
100. Han, L.; Kong, R.; Yin, D.D.; Zhang, E.B.; Xu, T.P.; De, W.; Shu, Y.Q. Low expression of long noncoding RNA GAS6-AS1 predicts a poor prognosis in patients with NSCLC. *Med. Oncol.* **2013**, *30*, 694. [CrossRef] [PubMed]
101. Sun, M.; Liu, X.H.; Wang, K.M.; Nie, F.Q.; Kong, R.; Yang, J.S.; Xia, R.; Xu, T.P.; Jin, F.Y.; Liu, Z.J.; et al. Downregulation of BRAF activated non-coding RNA is associated with poor prognosis for non-small cell lung cancer and promotes metastasis by affecting epithelial-mesenchymal transition. *Mol. Cancer* **2014**, *13*, 68. [CrossRef] [PubMed]

102. Han, L.; Zhang, E.B.; Yin, D.D.; Kong, R.; Xu, T.P.; Chen, W.M.; Xia, R.; Shu, Y.Q.; De, W. Low expression of long noncoding RNA PANDAR predicts a poor prognosis of non-small cell lung cancer and affects cell apoptosis by regulating Bcl-2. *Cell Death Dis.* **2015**, *6*, e1665. [CrossRef] [PubMed]
103. Weber, D.G.; Johnen, G.; Casjens, S.; Bryk, O.; Pesch, B.; Jockel, K.H.; Kollmeier, J.; Bruning, T. Evaluation of long noncoding RNA MALAT1 as a candidate blood-based biomarker for the diagnosis of non-small cell lung cancer. *BMC Res. Notes* **2013**, *6*, 518. [CrossRef] [PubMed]
104. Ji, P.; Diederichs, S.; Wang, W.; Boing, S.; Metzger, R.; Schneider, P.M.; Tidow, N.; Brandt, B.; Buerger, H.; Bulk, E.; et al. MALAT-1, a novel noncoding RNA, and thymosin beta4 predict metastasis and survival in early-stage non-small cell lung cancer. *Oncogene* **2003**, *22*, 8031–8041. [CrossRef] [PubMed]
105. Schmidt, L.H.; Spieker, T.; Koschmieder, S.; Schaffers, S.; Humberg, J.; Jungen, D.; Bulk, E.; Hascher, A.; Wittmer, D.; Marra, A.; et al. The long noncoding MALAT-1 RNA indicates a poor prognosis in non-small cell lung cancer and induces migration and tumor growth. *J. Thorac. Oncol.* **2011**, *6*, 1984–1992. [CrossRef] [PubMed]
106. Tripathi, V.; Shen, Z.; Chakraborty, A.; Giri, S.; Freier, S.M.; Wu, X.; Zhang, Y.; Gorospe, M.; Prasanth, S.G.; Lal, A.; et al. Long noncoding RNA MALAT1 controls cell cycle progression by regulating the expression of oncogenic transcription factor B-MYB. *PLoS Genet.* **2013**, *9*, e1003368. [CrossRef] [PubMed]
107. Wang, J.Z.; Xiang, J.J.; Wu, L.G.; Bai, Y.S.; Chen, Z.W.; Yin, X.Q.; Wang, Q.; Guo, W.H.; Peng, Y.; Guo, H.; et al. A genetic variant in long non-coding RNA MALAT1 associated with survival outcome among patients with advanced lung adenocarcinoma: A survival cohort analysis. *BMC Cancer* **2017**, *17*, 167. [CrossRef] [PubMed]
108. Rinn, J.L.; Kertesz, M.; Wang, J.K.; Squazzo, S.L.; Xu, X.; Brugmann, S.A.; Goodnough, L.H.; Helms, J.A.; Farnham, P.J.; Segal, E.; et al. Functional demarcation of active and silent chromatin domains in human HOX loci by noncoding RNAs. *Cell* **2007**, *129*, 1311–1323. [CrossRef] [PubMed]
109. Gupta, R.A.; Shah, N.; Wang, K.C.; Kim, J.; Horlings, H.M.; Wong, D.J.; Tsai, M.C.; Hung, T.; Argani, P.; Rinn, J.L.; et al. Long non-coding RNA HOTAIR reprograms chromatin state to promote cancer metastasis. *Nature* **2010**, *464*, 1071–1076. [CrossRef] [PubMed]
110. Liu, Z.; Sun, M.; Lu, K.; Liu, J.; Zhang, M.; Wu, W.; De, W.; Wang, Z.; Wang, R. The long noncoding RNA HOTAIR contributes to cisplatin resistance of human lung adenocarcinoma cells via downregulation of p21(WAF1/CIP1) expression. *PLoS ONE* **2013**, *8*, e77293.
111. Liu, M.Y.; Li, X.Q.; Gao, T.H.; Cui, Y.; Ma, N.; Zhou, Y.; Zhang, G.J. Elevated HOTAIR expression associated with cisplatin resistance in non-small cell lung cancer patients. *J. Thorac. Dis.* **2016**, *8*, 3314–3322. [CrossRef] [PubMed]
112. Qiu, M.; Xu, Y.; Yang, X.; Wang, J.; Hu, J.; Xu, L.; Yin, R. CCAT2 is a lung adenocarcinoma-specific long non-coding RNA and promotes invasion of non-small cell lung cancer. *Tumour. Biol.* **2014**, *35*, 5375–5380. [CrossRef] [PubMed]
113. Ling, H.; Spizzo, R.; Atlasi, Y.; Nicoloso, M.; Shimizu, M.; Redis, R.S.; Nishida, N.; Gafa, R.; Song, J.; Guo, Z.; et al. CCAT2, a novel noncoding RNA mapping to 8q24, underlies metastatic progression and chromosomal instability in colon cancer. *Genome Res.* **2013**, *23*, 1446–1461. [CrossRef] [PubMed]
114. Zhao, Z.; Wang, J.; Wang, S.; Chang, H.; Zhang, T.; Qu, J. LncRNA CCAT2 promotes tumorigenesis by over-expressed Pokemon in non-small cell lung cancer. *Biomed. Pharmacother.* **2017**, *87*, 692–697. [CrossRef] [PubMed]



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