Identifying patterns differing between high-dimensional datasets with generalized contrastive PCA

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- Abstract High-dimensional data have become ubiquitous in the biological sciences, and it is
- ¹⁴ often desirable to compare two datasets collected under different experimental conditions to
- 15 extract low-dimensional patterns enriched in one condition. However, traditional dimensionality
- ¹⁶ reduction techniques cannot accomplish this because they operate on only one dataset.
- ¹⁷ Contrastive principal component analysis (cPCA) has been proposed to address this problem, but
- it has seen little adoption because it requires tuning a hyperparameter resulting in multiple
- ¹⁹ solutions, with no way of knowing which is correct. Moreover, cPCA uses foreground and
- ²⁰ background conditions that are treated differently, making it ill-suited to compare two
- ²¹ experimental conditions symmetrically. Here we describe the development of generalized
- ²² contrastive PCA (gcPCA), a flexible hyperparameter-free approach that solves these problems.
- ²³ We first provide analyses explaining why cPCA requires a hyperparameter and how gcPCA avoids
- ²⁴ this requirement. We then describe an open-source gcPCA toolbox containing Python and
- ²⁵ MATLAB implementations of several variants of gcPCA tailored for different scenarios. Finally, we
- ²⁶ demonstrate the utility of gcPCA in analyzing diverse high-dimensional biological data, revealing
- ²⁷ unsupervised detection of hippocampal replay in neurophysiological recordings and
- heterogeneity of type II diabetes in single-cell RNA sequencing data. As a fast, robust, and
- ²⁹ easy-to-use comparison method, gcPCA provides a valuable resource facilitating the analysis of
- ³⁰ diverse high-dimensional datasets to gain new insights into complex biological phenomena.
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2 Introduction

- ³³ Investigators in the biological sciences are increasingly collecting high-dimensional datasets that
- ³⁴ are challenging to analyze, with modalities ranging from imaging to electrophysiology to single-cell
- ³⁵ RNA sequencing. Dimensionality reduction algorithms such as principal components analysis (PCA)
- and its many variants (Pearson, 1901; Hotelling, 1933; Zou et al., 2006; Zass and Shashua, 2006; Tip-
- ³⁷ ping and Bishop, 1999) are used widely to help simplify these datasets and facilitate analysis. PCA
- ³⁸ examines the covariance structure of the data to find dimensions that account for more variance
- ³⁹ than chance; these constitute patterns that are overrepresented in the data, such as assemblies of
- ⁴⁰ neurons whose activity fluctuates up and down together across time in a neural recording (*Chapin*

- and Nicolelis, 1999; Peyrache et al., 2010; Lopes-dos Santos et al., 2013; Sjulson et al., 2018), or
- networks of genes that are up- or down-regulated together across cells in a single-cell RNAseq
- dataset (Chung and Storey, 2015; Li et al., 2016). However, in many cases, the goal is to compare
- data collected under two different experimental conditions, which we refer to here as datasets.
- 45 Since PCA and other dimensionality reduction techniques operate on only one dataset, they can-
- ⁴⁶ not take experimental conditions into account.

The most common approach for comparing two high-dimensional datasets is linear discriminant analysis (*Izenman, 2008*) or its multidimensional analog, partial least squares discriminant analysis (*Brereton and Lloyd, 2014*). These methods find dimensions that optimally distinguish one dataset from the other, which could correspond to which neurons fire more, or which genes are upregulated, in condition *A* vs. condition *B*. However, an analogous method to compare the covariance structure of two datasets is not as well established. This addresses more subtle and detailed questions, such as which subsets of neurons exhibit increased temporal correlations in

- ⁵⁴ condition *A* than *B*, or which subsets of genes are more likely to be up- or downregulated together
- in individual cells in condition A than B. Mathematically, answering these questions corresponds
- ⁵⁶ to finding dimensions that account for more or less variance in *A* than *B*.

⁵⁷ Recently, contrastive PCA (cPCA) was proposed as a method to address this problem (*Abid et al.*, ⁵⁸ **2018**). Although cPCA is an important first step, it requires a hyperparameter α , which controls how ⁵⁹ much covariance from the second condition to subtract from the first. The algorithm must there-⁶⁰ fore iterate over multiple choices of α with no objective criteria to determine which value of α yields ⁶¹ the correct answer. Moreover, cPCA is asymmetric, identifying the most enriched dimensions in ⁶² the first condition after subtracting out the second condition as background; it cannot treat the ⁶³ two experimental conditions equally.

⁶⁴ Here we propose a novel solution to these problems we call generalized contrastive PCA (gcPCA). ⁶⁵ We first demonstrate the role the α hyperparameter plays in cPCA, then explain our strategy for ⁶⁶ eliminating it. We then describe an open source toolbox for Python and MATLAB implementing ⁶⁷ several versions of gcPCA with different objective functions that are either asymmetric or symmet-

- ric, orthogonal or non-orthogonal, or sparse or dense, tailored to suit the specific application at
- ⁶⁹ hand. Finally, we demonstrate the utility of gcPCA in the analysis of diverse biological datasets.

70 Results

The cPCA hyperparameter α compensates for bias toward high-variance dimenra sions in noisy, finitely-sampled datasets

To explain the need for the hyperparameter α in cPCA and how we avoid it in gcPCA, we will describe 73 the objective function of each method and show how they perform in generated synthetic data. For 74 illustration purposes, we generated synthetic data for two experimental conditions containing two-75 dimensional manifolds on a background of high-variance shared dimensions. The generated data 76 consisted of condition A, with a manifold (additional variance) in the 71st and 72nd dimensions 77 (ranked in order of descending variance), and condition B, with a manifold in the 81st and 82nd 78 dimensions (Fig. 1A). The manifold dimensions contained less total variance than most of the other 70 dimensions in the dataset, but their variance is two-fold higher in one condition relative to the other 80 (i.e., the 81st and 82nd dimensions have twice as much variance in condition *B* than condition *A*). 81 An important property of real-world biological datasets is that they are noisy and finitely sam-82 pled. We aimed to model the finite data regime by comparing 1×10^3 samples (finite data) to 83 1×10^{5} samples, which approximates infinite data. To inspect the effects of finite sampling on es-84 timated variance, we projected the "finite" and "infinite" data onto the ground truth dimensions 85 and calculated the variance explained by each (see methods). Our results (Fig. 1B) reveal that finite sampling yields noisy estimates of the true variance, with greater noise in high-variance di-87 mensions. 88

⁸⁹ To understand the practical consequences of this, it is helpful to start by reviewing traditional



Figure 1. Generalized contrastive PCA avoids cPCA's need for the hyperparameter α **in noisy, finitely-sampled data. A** We generated two conditions in noisy synthetic data that each contain low-variance manifolds that are not present in the other. These manifolds have lower overall variance than many other dimensions and are not trivially discoverable. **B** Eigenvalue spectra for each condition estimated from finite (dark line) or infinite (light line) data. Note the sampling error in the finite date case. C With infinite data, eigendecomposition of $(C_A - C_B)$ suffices to extract the correct answers (dimensions 71-72 and 81-82, light lines). However, with finite data, these peaks are smaller than the sampling error in high-variance dimensions, creating a bias toward high-variance dimensions being selected. **D** cPCA uses the hyperparameter α to adjust how much influence C_B has on C_A . As α increases, the bias toward high-variance dimensions decreases until it becomes negative with $\alpha > 1$, eventually exposing the differences in lower-variance dimensions. Importantly, there is no way to know which value of α yields the correct solution. **E** Using gcPCA, the dimensions most changed in each condition are identified correctly, even with finitely-sampled data. **F** cPCA with the optimal choice of α does not extract the correct dimensions in *B*. **G** gcPCA identifies the enriched dimensions in each condition and correctly returns the low-variance manifolds. Because gcPCA is symmetric, it extracts the correct dimensions in both *A* and *B*.

- PCA. With PCA, the principal components of a data matrix **D**, of size $n \times p$ (samples \times features), are
- the dimensions explaining the most variance. These can be identified by estimating the covariance
- matrix $\mathbf{C} = \mathbf{D}^{\mathsf{T}} \mathbf{D}/(n-1)$, then solving the following quadratic optimization problem in equation 1:

$$\underset{\mathbf{x}: \mathbf{x}^{\mathsf{T}} \mathbf{x}=1}{\arg \max} \quad \mathbf{x}^{\mathsf{T}} \mathbf{C} \mathbf{x} \tag{1}$$

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This problem can be solved by eigendecomposition of C, yielding the matrix of eigenvectors Xknown as principal components (PCs).

- To extend this to two datasets, a logical strategy involves formulating an objective function to describe the difference in variances between the two conditions, enabling us to extract dimensions
- ⁹⁷ describe the difference in variances between the two conditions, enabling us to extract dimensions ⁹⁸ that show the greatest increase in variance in *A* relative to *B*. We now have two covariance matrices.
- that show the greatest increase in variance in A relative to B. We now have two covariance matrices, C_A and C_B , and the contrastive PCs (cPCs) are the vectors that maximize the objective function 2:

$$\underset{\mathbf{x}: \mathbf{x}^{\mathsf{T}} \mathbf{x}=1}{\operatorname{arg max}} \quad \mathbf{x}^{\mathsf{T}} (\mathbf{C}_A - \mathbf{C}_B) \mathbf{x}$$
(2)

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Analogous to traditional PCA, this problem can be solved by eigendecomposition of $(C_4 - C_B)$. This 101 vields cPCs that account for more variance in either condition A or B, corresponding to the eigen-102 vectors with the largest or smallest eigenvalues, respectively. With infinite data, this approach 103 correctly finds the dimensions enriched in conditions A and B (Fig. 1C, light line), but with finite 104 data, it fails to do so because the sampling error in higher-variance dimensions is larger than the 105 true signal in the lower-variance dimensions (Fig. 1C, dark line). In other words, it has a system-106 atic bias toward high-variance dimensions. To compensate for this effect, cPCA Abid et al. (2018) 107 introduces the hyperparameter α , changing the following objective function to 3: 108

$$\underset{\mathbf{x}: \mathbf{x}^{\mathsf{T}} \mathbf{x}=1}{\operatorname{arg\,max}} \quad \mathbf{x}^{\mathsf{T}} (\mathbf{C}_A - \alpha \mathbf{C}_B) \mathbf{x}$$
(3)

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As discussed in *Abid et al.* (2018), α represents a trade-off determining the extent to which C_p influ-110 ences the identification of enriched vectors in C_4 . In our synthetic data, we can visually appreciate 111 the effect of different values of α in the resulting cPCA objective function value (Fig. 1D). In effect, 112 α tunes the amount of bias toward high-variance dimensions in the cPCA calculation. with $\alpha < 1$ 113 biasing toward high-variance dimensions and $\alpha > 1$ biasing against them. In this case, $\alpha = 2$ yields 114 the correct solution that dimensions 71-72 are enriched in A (Fig. 1F), but other values of α yield 115 equally plausible, but incorrect, solutions. Importantly, we can only determine which solution is 116 correct because we knew the answer in advance, which is not typically the case for experimental 117 data. Further, negative values in cPCA are generally interpreted as dimensions enriched in condi-118 tion B (Abid et al., 2018; Boileau et al., 2020), but our simulation shows that values of α larger than 119 1 bias the highest-variance dimensions to be negative (Fig. 1D). This creates the illusion that these 120 dimensions are enriched in condition B, even though the correct answer is that only dimensions 121 81-82 are enriched in B. Similar to the situation of finding dimensions enriched in A, the results 122 depend on the choice of α , with no way to determine which solution is correct (Fig. 1F). Importantly, 123 the range of α 's yielding the correct solution can be incredibly narrow, as in Fig. S1, where $\alpha = 2.6$ 124 vields the correct solution, but 2.2 or 3.0 do not. 125

₁₂₆ gcPCA avoids hyperparameters by including a normalization factor

¹²⁷ Our goal for gcPCA was to eliminate the need for hyperparameters and provide unique, correct so-

- ¹²⁸ lutions. To mitigate the bias toward high-variance dimensions, we introduce a normalization factor, ¹²⁹ such as the total variance in both conditions, which can be calculated by summing the covariance
- matrices ($C_A + C_B$). The objective function then becomes (eq. 8):

$$\underset{\mathbf{x}: \mathbf{x}^{\mathsf{T}} \mathbf{x}=1}{\operatorname{arg\,max}} \quad \frac{\mathbf{x}^{\mathsf{T}} (\mathbf{C}_{A} - \mathbf{C}_{B}) \mathbf{x}}{\mathbf{x}^{\mathsf{T}} (\mathbf{C}_{A} + \mathbf{C}_{B}) \mathbf{x}} \tag{4}$$

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- ¹³² Solving this problem is slightly more complicated than with PCA or cPCA, but ultimately it can also
- ¹³³ be reduced to a computationally efficient eigenvalue problem (see Methods). The resulting general-
- ized contrastive PCs (gcPCs) maximize relative, rather than absolute, changes in variance between
- $_{135}$ conditions A and B. This effectively handles the bias toward high-variance dimensions and suc-
- cessfully extracts the ground truth dimensions in our synthetic data, even with finite sampling (Fig.
- 137 1E, G), and even when the range of acceptable α is narrow (Fig. S1).
- ¹³⁸ This creates two minor complications to be aware of: first, unlike PCA or cPCA, gcPCs are not or-¹³⁹ thogonal by default. If orthogonality is important for a particular application, we have implemented
- ¹³⁹ thogonal by default. If orthogonality is important for a particular application, we have implemented ¹⁴⁰ versions of gcPCA with an orthogonality constraint (see Methods). Second, the normalization fac-
- tor can create numerical instability if the data are rank-deficient, yielding dimensions with zero or
- near-zero variance in the denominator. Our implementation of gcPCA prevents this by detecting
- ¹⁴³ and excluding those dimensions before performing the calculation (see Methods).

The open-source gcPCA toolbox contains multiple gcPCA variants enabling optimal handling of diverse use cases

- ¹⁴⁶ We developed an open-source gcPCA toolbox with implementations in Python and MATLAB of sev-¹⁴⁷ eral different variants of gcPCA. This toolbox is freely available at:
- 148 https://github.com/SjulsonLab/generalized contrastive PCA. Here we will present the different vari-
- ¹⁴⁹ ants of gcPCA and their use cases.

¹⁵⁰ gcPCA v1.0: traditional cPCA

¹⁵¹ For version 1.0, we include an implementation of the original cPCA algorithm that finds cPCs max-

imizing the objective function in Eqn. 3.

¹⁵³ gcPCA v2.0: gcPCA maximizing A/B

¹⁵⁴ Here we include an implementation that finds gcPCs maximizing the ratio of variance in A to B:

$$\underset{\mathbf{x}: \mathbf{x}^{\mathsf{T}} \mathbf{x}=1}{\operatorname{arg\,max}} \quad \frac{\mathbf{x}^{\mathsf{T}} \mathbf{C}_{A} \mathbf{x}}{\mathbf{x}^{\mathsf{T}} \mathbf{C}_{B} \mathbf{x}}$$
(5)

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Like cPCA, this method is asymmetrical, meaning it is suitable for situations in which A is a fore-

- 157 ground condition and *B* is a background condition; in other words, *A* is presumed to be equal to *B*
- ¹⁵⁸ with a low-dimensional pattern added, and the goal is to extract that pattern. The resulting eigen-
- values are the ratio of the variance a given gcPC accounts for in A to the variance it accounts for in
- B. Thus, they fall in the range $[0, \infty)$, with gcPCs enriched in A having eigenvalues > 1.
- ¹⁶¹ gcPCA v3.0: gcPCA maximizing (A-B)/B
- ¹⁶² The second method developed is also asymmetrical but based on a relative change:

$$\underset{\mathbf{x}: \mathbf{x}^{\mathsf{T}} \mathbf{x}=1}{\operatorname{arg\,max}} \quad \frac{\mathbf{x}^{\mathsf{T}} (\mathbf{C}_{A} - \mathbf{C}_{B}) \mathbf{x}}{\mathbf{x}^{\mathsf{T}} (\mathbf{C}_{B}) \mathbf{x}}$$
(6)

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¹⁶⁴ This method is closely-related to v2.0 and is suitable for scenarios in which the investigator wishes ¹⁶⁵ to define the gcPCs based on a relative change to a background condition (i.e., finding a 30% in-

 $_{166}$ crease in the neural activity in condition A relative to B). The eigenvalues returned are in the range

¹⁶⁷ $[-1, \infty)$, with gcPCs enriched in A having eigenvalues > 0.

- 168 gcPCA v4.0: gcPCA maximizing (A-B)/(A+B)
- ¹⁶⁹ The last of the three methods is based on a relative change:

$$\underset{\mathbf{x}: \mathbf{x}^{\mathsf{T}} \mathbf{x}=1}{\operatorname{arg\,max}} \quad \frac{\mathbf{x}^{\mathsf{T}} (\mathbf{C}_{A} - \mathbf{C}_{B}) \mathbf{x}}{\mathbf{x}^{\mathsf{T}} (\mathbf{C}_{A} + \mathbf{C}_{B}) \mathbf{x}}$$
(7)

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- ¹⁷¹ This method is symmetrical, treating conditions *A* and *B* equally, and is appropriate for contrasting
- conditions in which *B* is a distinct condition and not merely a background to be removed, for exam-
- ple comparing neural data in sleep vs. wakefulness. The eigenvalues are in the range [-1, 1] and
- are easily interpretable as a traditional index of the form (A B)/(A + B): 1 means that a gcPC only
- accounts for variance in A, -1 means it only accounts for variance in B, and 0 means it accounts for
- equal variance in both. This method is fully symmetrical in the sense that switching A and B will
- 177 yield the same gcPCs with the signs of the eigenvalues reversed.

gcPCA v2.1, v3.1, and v4.1: Orthogonal gcPCA

- ¹⁷⁹ Unlike PCs or cPCs, gcPCs are not orthogonal by default. Because orthogonality may be important
- 180 for some applications, we also include versions of gcPCA with an orthogonality constraint (see
- ¹⁸¹ Methods). gcPCA v2.1 is the orthogonal version of 2.0, and v3.1 is the orthogonal version of v3.0,
- and v4.1 is the orthogonal version of v4.0.
- ¹⁸³ Sparse vs. dense gcPCA
- ¹⁸⁴ In high-dimensional datasets, it is often desirable to perform feature selection for easy interpre-
- tation of the results. With that in mind, we have also developed sparse versions of gcPCA. This
- 186 method works by first finding the gcPCs based on the objective function selected, then performing
- 187 feature selection using an L1 lasso penalty (see Methods). All non-orthogonal versions of gcPCA
- can be run in either sparse or dense mode, but sparsification cannot be used with the orthogonality
- 189 constraint (*Zou et al., 2006*).

| gcPCA version | Symmetric | Orthogonal | Sparse solution | Note |
|---------------|--|--|-----------------|----------------------|
| v1 | × | 1 | 1 | Equivalent to cPCA |
| v2 | × | × | 1 | Objective function 5 |
| v2.1 | × | 1 | × | Objective function 5 |
| v3 | × | × | 1 | Objective function 6 |
| v3.1 | × | 1 | × | Objective function 6 |
| v4 | 1 | × | 1 | Objective function 7 |
| v4.1 | Image: A second s | Image: A second s | × | Objective function 7 |

Table 1. gcPCA variants in the gcPCA toolbox

¹⁹⁰ Successful extraction of facial expression features with gcPCA

To illustrate the utility of gcPCA with real-world datasets, we first use the Chicago Face Dataset (*Ma et al., 2015*), which contains faces with different facial expressions. Here we used happy and angry expressions as condition *A* and neutral faces as condition *B* (Fig. 2A). This dataset is useful for two reasons: 1) the categorical separation of facial expressions allows an easy evaluation of the contrastive methods, and 2) the dimensions can be visually inspected for features that are being discovered by the method.

¹⁹⁷ We first applied cPCA to the two conditions and used its automatic α selection algorithm to ¹⁹⁸ pick two different α values. The automatic α selection algorithm developed by *Abid et al.* (2018) ¹⁹⁹ finds representative α 's that yield different cPC embeddings so that the investigator can choose ²⁰⁰ the appropriate α value. The algorithm is explained fully in the original paper (see supplementary



Figure 2. gcPCA correctly extracts contrastive facial features in the Chicago Face Dataset. A For this test, contrastive methods were applied to a set of happy and angry face images (Condition *A*) versus neutral face images from the same subjects (Condition *B*). Facial expression changes along the happy-angry axis were therefore the low-dimensional pattern that is enriched in condition *A*. **B** The first α identified by cPCA's algorithm, $\alpha = 0.2$, yields loadings similar to the first PCA dimensions, *i.e.* eigenfaces. **C** Projecting the faces onto the cPCs reveals clusters unrelated to the happy and angry facial expressions in condition *A*, indicating an incorrect solution. **D** cPCA with $\alpha = 1.7$ correctly reveals features associated with the expected facial expressions in condition *A*, such as furrowed eyebrows and the region around the mouth and nose. **E** Projecting the faces onto these cPCs reveals the separation of happy and angry faces along the first cPC. Importantly, it was only possible to determine this answer was correct because we knew the labels in advance. **F** Dimensions identified by gcPCA correctly reflect features related to the facial expressions in condition *A*. **G** Projecting the faces onto the first two gcPCs also reveals the separation of happy and angry faces along the first gcPC.

methods - algorithm 2 *Abid et al. (2018)*). Briefly, it calculates cPCA for an array of different α 's (default: 40 different α values ranging from 0.01 to 1000 spaced on a log scale), defining a subspace with the top *k* cPCs (default: 2 dimensions), and calculating an affinity matrix between subspaces of different α 's. This affinity matrix is then clustered to find *p* clusters, and the medoid of each cluster is a candidate α .

The first value returned is $\alpha = 0.2$, and we can see that the first two cPC loadings resemble 206 "eigenfaces" (*Turk and Pentland, 1991*), the largest principal components of facial images (Fig. 2B), 207 This suggests that this α is too small, leading cPCA to extract the highest-variance components in 208 condition A. Looking at the individual faces projected on these cPCs, two clusters can be identified 200 that separate cleanly along cPC1 (Fig. 2C). However, this solution is incorrect because these clusters 210 do not reflect the two facial expressions comprising condition A. Instead, they represent skin color. 211 which should account for equal variance in both datasets because images of the same subjects 212 are present in both conditions. For the second α value returned ($\alpha = 1.7$), the cPC loadings exhibit 213 features specific to condition A (Fig. 2D), and data projected onto these cPCs recovers the different 214 expressions (Fig. 2F). It is important to note that if we did not have the class labels a priori, we 215 could easily believe $\alpha = 0.2$ was the correct answer because it produces better clustering than for 216 $\alpha = 1.7$ (Fig. 2C.F). Using gcPCA v3.0 (asymmetric, non-orthogonal) also reveals features specific to 217 condition A (Fig. 2F), and the two expressions in the dataset can be distinguished by their projection 218 onto gcPC, (Fig. 2G), without the requirement of fine-tuning a hyperparameter. 210

Applying gcPCA to neurophysiological recordings reveals hippocampal replay without *a priori* knowledge of replay content

A key application for gcPCA is neuronal recordings, which frequently contain hundreds of iso-222 lated single units (de Oliveira et al., 2022; Jun et al., 2017). A well-studied neurophysiological 223 phenomenon is hippocampal replay, in which hippocampal neurons encoding spatial trajectories 224 traversed during a behavioral task "replay" the same activity patterns in post-task periods (*Wil*-225 son and McNaughton, 1994). It is important to note that spatial location is a continuous variable. 226 so we are not expecting gcPCA or cPCA to find dimensions that cluster the activity into different 227 groups, as with the facial expression data. Instead, we are hoping to see replay of the firing pat-228 terns that encode the linear track the animal recently explored (Wilson and McNaughton. 1994: 220 Foster, 2017). For our analysis, we used a previously published dataset recorded from hippocam-230 pal CA1 Girardeau et al. (2017) where rats learn the location of an aversive air puff on a linear track. 231 The air puff is only delivered when the rat is running in one direction, called the danger run, and 232 the other direction is the safe run (Fig. 3A). Using previously-established methods (Kudrimoti et al., 233 1999), Girardeau et al. (2017) found that hippocampal neurons exhibit reactivation of the task rep-234 resentation in post-task activity when compared to pre-task activity. We tested whether cPCA or 235 gcPCA could extract hippocampal replay directly from neuronal activity by contrasting post-task 236 activity (condition A) with pre-task activity (condition B) (Fig. 3B). For this, we used cPCA and gcPCA 237 to extract cPCs or gcPCs from the pre- and post-task data, then projected the during-task data onto 238 the cPCs/gcPCs to test whether any spatial structure was discernible. 239

Using cPCA, it was not straightforward to detect replay. When we requested the cPCA algorithm 240 evaluate all the dimensions (k = 48), the α values identified by the automatic selection did not reveal 241 any obvious task-related spatial structure (Fig. 3C – left column, α values 0.44 and 701.70). When we 242 requested a smaller set of dimensions (k = 2), the automatic α selection returned several different 243 values, with one of them ($\alpha = 1.43$) revealing spatial structure in the task data (Fig. 3C - right column). 244 This reveals that although the only hyperparameter for cPCA is α in theory, the automatic alpha 245 selection algorithm depends on k, the number of components requested, constituting a second de 246 facto hyperparameter. 247

Applying gcPCA v4 (symmetrical, non-orthogonal), we readily recovered replay of task-related spatial structure (Fig. 3D, top). Importantly, traditional replay analyses require knowledge of the firing patterns during the task to test whether they were overrepresented in post-task sleep (*Kud*-



Figure 3. gcPCA v4.0 correctly identifies hippocampal replay in neurophysiological data without prior knowledge of replay content. A In *Girardeau et al. (2017*), rats were trained to traverse a linear track where one direction has an aversive air puff and the other does not. Rats learned the location of the air puff and which direction was dangerous or safe. **B** *Girardeau et al. (2017*) recorded hippocampal neurons in preand post-task periods and used activity recorded during the task as a template to determine that that activity was "replayed" during post-task. We reanalyzed their published data using post-task activity as condition *A* and pre-task activity as condition *B* to identify the dimensions most enriched in post-task activity without taking task-related activity into account. **C** We first performed cPCA, then projected task-related neuronal data onto the cPCs to determine whether task-related data was enriched in the post-task period. The automatic α selection algorithm from cPCA returns various α values depending on the number of cPCs requested (parameter *k*). *Left Column* Representative α values returned with *k*=48 cPCs. cPC₁₋₂ are the dimensions most enriched in post-task, and cPC_{last} and cPC_{last-1} are the most enriched in pre-task. No discernible spatial structure is identified, indicating that replay was not detected. *Right Column* With *k*=2, the α values returned are of different magnitudes, and one of them ($\alpha = 1.43$) reveals spatial structure related to the task, indicative of hippocampal replay. **D** gcPCA readily identifies the replay of the spatial task structure (left) with no parameter search.



Figure 4. gcPCA v4.0 reveals possible disease heterogeneity in type II diabetes. A *Martínez-López et al. (2023*) performed scRNA-seq on isolated pancreatic cells from type II diabetes (T2D) patients and healthy controls. We used gcPCA v4.0 to compare the two conditions and found clustering of beta cells by donor identity (each color is a donor) in gcPC_{1,2} (top left panel). Such clustering was not found in the last gcPCs (top right panel) or in the control donors (bottom left and right panels) **B** The list of 40 genes with the highest loadings on gcPC₁ includes several previosly linked to T2D (red). Notably, the top two hits, *TMEM176A* and *TMEM176B*, were shown by *Martínez-López et al. (2023*) to play functional roles in beta cell function.

rimoti et al., 1999; Wilson and McNaughton, 1994; Foster, 2017). gcPCA was able to extract signa tures of replay without prior knowledge of the task-related activity. This may prove useful in many

analogous situations in which the experimenter does not have prior knowledge of the pattern they
 are searching for.

We also took advantage of the symmetric nature of gcPCA and investigated the patterns enriched in pre-task activity, gcPC_{last} and gcPC_{last-1}. As expected, they did not exhibit task-related spatial structure (Fig. 3D, top), suggesting they contain replay of environments other than the linear track.

Applying gcPCA to scRNA-seq data prioritizes disease genes and reveals disease heterogeneity in type II diabetes

Another key application of gcPCA is high-dimensional omics datasets, which often reflect a com-261 parison of two conditions (e.g. disease vs. healthy). For this analysis, we used published single-cell 262 RNA sequencing data from pancreatic beta cells taken from healthy controls or patients with type 263 Il diabetes (T2D)Martínez-López et al. (2023)). We used T2D beta cells as condition A and healthy 264 control beta cells as condition B to test whether gcPCA 4.0 could identify groups of genes that 265 vary more among T2D patients or controls, $gcPC_1$ and $gcPC_2$ therefore represent axes along which cells from patients vary more and the gcPC_{last} and gcPC_{last1} represent axes along which control 267 cells vary more. We found that T2D and control data had similar levels of variability (Fig. 4A), but 268 in T2D patients this variability exhibited clear donor-based clustering (Fig. 4A, top left) that was 260 not observed in controls (Fig. 4A, bottom right). Several of the genes with the highest loadings 270 on gcPC, have been previously implicated in T2D (IMMP2L (Diabetes Genetics Initiative of Broad 271 Institute of Harvard and MIT. Lund University, and Novartis Institutes of BioMedical Research 272 et al., 2007; Greenwald et al., 2019), STMN1 (Horn et al., 2016), PDHA1 (Srinivasan et al., 2010), CLU 273 (Kim et al., 2001, 2006), DDIT3 (Li et al., 2022; Yong et al., 2021), SSTR5-AS1, (Jian and Felsenfeld, 27/ 2018), TFF3 (Fueger et al., 2008), Fig. 4B, red), notably the top two hits, which were the TMEM176A/B 275 genes that Martínez-López et al. (2023) demonstrated are functionally important for T2D-related 276 beta-cell function. The fact that gcPCA identifies known T2D-related genes and that clustering is 277 specific to cases suggests that gcPCA is revealing disease heterogeneity (Ahlqvist et al., 2020). 278

279 **Discussion**

Discovering low-dimensional patterns that vary between conditions in high-dimensional datasets 280 is a crucial analysis in many research contexts. Here we present gcPCA, a method that achieves this 281 by examining the covariance structure of the datasets to find dimensions that exhibit the largest 282 relative changes in variance between conditions. This work builds on the pioneering insights in 283 the development of cPCA Abid et al. (2018) but solves cPCA's key problem, the requirement for 28/ the hyperparameter α . Here we showed that the function of α is to compensate for bias toward 285 high-variance dimensions in noisy, finitely-sampled data. Further, we showed how this can be cir-286 cumvented by introducing a normalization factor. Previous work Abid et al. (2018): Boileau et al. 287 (2020) has focused on developing and improving methods to find appropriate choices for α , but 288 with gcPCA we chose instead to eliminate the α hyperparameter entirely. Importantly, the advan-289 tage of our approach is not merely that it is computationally cheaper than scanning a range of 290 α 's; it is that in most real-world cases there is no way to know whether a given choice of α yields a 291 correct solution. 292

We wish to address a common point of confusion by reiterating how gcPCA differs from LDA or PLS. These are methods that find patterns optimally distinguishing two datasets, but gcPCA finds patterns that exhibit more within-dataset variability in one dataset than another. As a fictitious example, LDA might find a height/weight dimension distinguishing a university rugby team from the general population because rugby players are taller and heavier. In contrast, gcPCA would be more likely to find an age/education-level dimension because those features exhibit more variability in the general population than in a university team. Despite the simplicity of this analysis, it reveals interesting phenomena in high-dimensional biological data such as hippocampal replay (Fig. 3) or

³⁰¹ transcriptomic heterogeneity in disease states (Fig. 4).

gcPCA has a few caveats: first, unlike ordinary PCs or cPCs, gcPCs are not orthogonal by default. 302 Our toolbox includes versions of gcPCA with an orthogonality constraint (v2.1, v3.1, and v4.1), which 303 comes at increased computational cost because a new eigendecomposition must be performed 304 for each gcPC. Second, the normalization factor introduces the possibility of numerical instability 305 if the denominator matrix is rank-deficient, meaning it has dimensions with zero (or near-zero) 306 variance that create a "divide-by-zero" situation. However, the implementation of gcPCA in the 307 toolbox automatically excludes these dimensions if they exist. Finally, there may be situations in 308 which cPCA's α could be a feature, rather than a bug, if the investigator has prior knowledge that 309 the patterns of interest will lie in high- or low-variance dimensions. Choosing an appropriate α 310 could then intentionally bias the analysis in favor of the results of interest. In such cases, it would 311 be relatively straightforward to extend gcPCA by adding a parameter that accomplishes a similar 312 result by adjusting the eigenspectrum of the denominator matrix in the objective function, e.g. 313 $(\mathbf{C}_{A} - \mathbf{C}_{P})$. There are also other extensions that could be added, such as contrasting more than two 314 conditions or incorporating nonlinearity, which can be relevant to specific data problems. However, 315 we leave the development of such tools for future efforts. 316

The biological sciences are currently undergoing an explosion of technologies that produce high-dimensional datasets, including novel forms of microscopy and neuroimaging, high-speed video tracking, Neuropixels recordings, -omics approaches with single-cell resolution, and many others. In addition to the analyses of electrophysiological recordings or single-cell RNA sequencing data demonstrated here, gcPCA could be applied to any of these experimental modalities. We thus anticipate that the open-source gcPCA toolbox will provide a valuable resource facilitating a broad range of biological investigations that require contrasting two experimental conditions.

324 Methods

325 Generalized contrastive PCA

³²⁶ Our motivation for the following method stems from eliminating the necessity of the free param-³²⁷ eter α in the contrastive PCA method. To accomplish this, we introduce a normalization factor ³²⁸ to mitigate the bias toward high-variance dimensions. We will summarize the process of calculat-³²⁹ ing the gcPCs using gcPCA v4.0 as an example, but v2 and v3 are analogous. gcPCA v4.0 has the ³³⁰ following objective function, as shown in equation (eq. 8):

$$\underset{\mathbf{x}: \mathbf{x}^{\mathsf{T}} \mathbf{x}=1}{\arg \max} \frac{\mathbf{x}^{\mathsf{T}} (\mathbf{C}_{A} - \mathbf{C}_{B}) \mathbf{x}}{\mathbf{x}^{\mathsf{T}} (\mathbf{C}_{A} + \mathbf{C}_{B}) \mathbf{x}}$$
(8)

331

³³² A potential problem of this objective function is the denominator creating numerical instability if ³³³ there are vectors that have eigenvalues approaching zero in the denominator covariance matrix. ³³⁴ To address this, we consider only the principal components (*J*) of that matrix that have non-zero ³³⁵ eigenvalues. In this case, the matrix *J* is composed of the principal components of the row-wise ³³⁶ concatenated datasets *A* and *B* that have non-zero eigenvalues. We then substitute for **x** using ³³⁷ **x** = **J** γ , yielding:

$$\underset{\gamma: \gamma^{\mathsf{T}}\gamma=1}{\operatorname{arg\,max}} \quad \frac{\gamma^{\mathsf{T}}\mathbf{J}^{\mathsf{T}}(\mathbf{C}_{A}-\mathbf{C}_{B})\mathbf{J}\gamma}{\gamma^{\mathsf{T}}\mathbf{J}^{\mathsf{T}}(\mathbf{C}_{A}+\mathbf{C}_{B})\mathbf{J}\gamma} \tag{9}$$

338

The matrix $(\mathbf{J}^{\mathsf{T}}(\mathbf{C}_A + \mathbf{C}_B)\mathbf{J})$ in the denominator is guaranteed to be positive definite, allowing us to find a symmetric matrix **M** that is its square root, yielding equation (eq. 10):

arg max

 $\mathbf{y} : \mathbf{y}^{\mathsf{T}} \mathbf{y} = 1$

$$\underset{\gamma: \gamma^{\mathsf{T}} \gamma=1}{\operatorname{arg\,max}} \quad \frac{\gamma^{\mathsf{T}} \mathbf{J}^{\mathsf{T}} (\mathbf{C}_{A} - \mathbf{C}_{B}) \mathbf{J} \gamma}{\gamma^{\mathsf{T}} \mathbf{M}^{\mathsf{T}} \mathbf{M} \gamma} \tag{10}$$

341

Let $\mathbf{y} = \mathbf{M}\boldsymbol{\gamma}$, yielding:

343

This optimization problem can be solved with the eigendecomposition of the numerator matrix. The vectors in \mathbf{X} are then calculated using equation (eq. 12):

 $\frac{\mathbf{y}^{\mathsf{T}}\mathbf{M}^{-1}\mathbf{J}^{\mathsf{T}}(\mathbf{C}_{A}-\mathbf{C}_{B})\mathbf{J}\mathbf{M}^{-1}\mathbf{y}}{\mathbf{y}^{\mathsf{T}}\mathbf{y}}$

$$\mathbf{X} = \mathbf{J}\mathbf{M}^{-1}\mathbf{y} \tag{12}$$

(11)

346

³⁴⁷ The column vectors in **X** are referred to as gcPCs, following the term cPCs used in *Abid et al.* (2018).

The other versions of gcPCA can be solved in the exact same way, by replacing $C_A - C_B$ with C_A (v2) and replacing $C_A + C_B$ with C_B (v2 and v3).

 $_{
m 350}$ In our case, the gcPCs (X) are not guaranteed to be orthogonal due to the presence of the ${f M}^{-1}$

matrix. By default, gcPCA returns non-orthogonal gcPCs. If orthogonality is desired (as in gcPCA

 $_{_{352}}$ v4.1), we iteratively shrink matrix J to remove the subspace spanned by the gcPCs (*i.e.* columns of

 $_{_{353}}$ X) that have already been found. At each step, we compute the largest eigenvector x of equation

11, then project it into the feature space with equation 12 and concatenate it column-wise into the

 $_{355}$ growing matrix **X**. To shrink **J**, we first regress out the **x** from **J**, as shown in equation 13:

$$\hat{\mathbf{J}} = \mathbf{J} - \mathbf{x}\mathbf{x}^{\mathsf{T}}\mathbf{J} \tag{13}$$

356

³⁵⁷ We then use SVD to get the left singular vectors of $\hat{\mathbf{J}}$, and we define $\tilde{\mathbf{J}}$ as the first n - i of these (on ³⁵⁸ the *i*-th iteration). $\tilde{\mathbf{J}}$ serves as an orthonormal basis for the subspace of \mathbf{J} that is orthogonal to \mathbf{X} ,

and we use $\tilde{\mathbf{J}}$ as the new \mathbf{J} in eq. 9 for the next iteration. This process continues until *n* gcPCs are

³⁶⁰ found, which can be the number of features in the dataset, the minimum rank of the conditions,

³⁶¹ or a number specified by the user for the gcPCs to be extracted.

362 Sparse gcPCA

³⁶³ We developed an extension for sparse gcPCA using a similar approach as sparse PCA (Zou et al.,

³⁶⁴ 2006) and sparse cPCA (Boileau et al., 2020). Here we will first review the sparse PCA framework,

then explain how we adapt it for gcPCA.

366 Sparse PCA method

³⁶⁷ Sparse PCA was first proposed as a reinterpretation of PCA as a regression problem. In brief, given

a matrix \mathbf{X}_{nxk} , where the first k ordinary PCs are organized column-wise and are orthonormal, PCA

³⁶⁹ can be seen as minimizing the following objective:

$$\underset{\mathbf{X}_{n \times k}: \mathbf{X}^{\mathsf{T}} \mathbf{X} = I}{\arg \min} ||\mathbf{D} - \mathbf{D} \mathbf{X} \mathbf{X}^{\mathsf{T}}||^2$$
(14)

³⁷⁰ Where **D** is the data matrix of size features \times samples. To achieve sparse loadings in the first *k* PCs,

371 Zou et al. (2006) proposes the use of elastic net regularization, as shown in the following objective

372 function:

$$\underset{\mathbf{Y}_{n\times k},\mathbf{B}_{n\times k}:\mathbf{Y}^{\mathsf{T}}\mathbf{Y}=I}{\arg\min} ||\mathbf{D} - \mathbf{D}\mathbf{B}\mathbf{Y}^{\mathsf{T}}||^{2} + \lambda \sum_{j=1}^{k} ||\boldsymbol{\beta}_{j}||^{2} + \sum_{j=1}^{k} \lambda_{1,j} ||\boldsymbol{\beta}_{j}||_{1}}$$
(15)

- ³⁷³ Where **B** is the matrix containing the sparse PCs $\beta_{j'}$ and **Y** is a column-wise matrix that projects
- data from the sparse PC space to the feature space. The elastic net is the combination of the ridge
- penalty ($\lambda \sum_{j=1}^{k} ||\boldsymbol{\beta}_{j}||^{2}$) and the lasso penalty ($\sum_{j=1}^{k} \lambda_{1,j} ||\boldsymbol{\beta}_{j}||_{1}$). The ridge penalty is used to correct a
- rank-deficient matrix **D** for numerical purposes. In **Zou et al.** (2006), the same λ was used for all
- $_{377}$ k components while using a different $\lambda_{1,j}$ for every component. To numerically solve equation 15,
- 378 (Zou et al., 2006) use an alternating algorithm in which Y is held constant as we solve for B, then B
- ³⁷⁹ is held constant while we update **Y**, and this is repeated until the algorithm converges. **Y** is initially
- $_{\scriptscriptstyle 380}$ set to be equal to the ordinary PCs (X), then we can find each column of **B** by the following elastic
- ³⁸¹ net regression:

$$\hat{\boldsymbol{\beta}}_{j} = \underset{\boldsymbol{\beta}_{j}}{\arg\min} ||\mathbf{D}\mathbf{y}_{j} - \mathbf{D}\boldsymbol{\beta}_{j}||^{2} + \lambda ||\boldsymbol{\beta}_{j}||^{2} + \lambda_{1,j} ||\boldsymbol{\beta}_{j}||_{1}$$
(16)

³⁸² Next, **B** is fixed, meaning the penalty terms can be ignored, and the new **Y** is defined as:

$$\underset{\mathbf{Y} \in \mathbf{Y} = I_{k \times k}}{\arg \min} ||\mathbf{D} - \mathbf{D} \mathbf{B} \mathbf{Y}^{\mathsf{T}}||^2$$
(17)

³⁸³ The solution to 17 can be found by a reduced rank form of Procrustes rotation. Using SVD, we find

$$(\mathbf{D}^{\mathsf{T}}\mathbf{D})\mathbf{B} = \mathbf{U}\mathbf{S}\mathbf{V}^{\mathsf{T}}$$
(18)

³⁸⁴ We then set $\hat{\mathbf{Y}} = \mathbf{U}\mathbf{V}^{\mathsf{T}}$. To solve eq. 16, *Zou et al.* (2006) has shown that is only necessary to know ³⁸⁵ the Gram matrix **D**^T**D**. For a fixed **Y**, finding $\boldsymbol{\beta}_i$ is equivalent to minimizing:

$$||\mathbf{D}\mathbf{y}_{j} - \mathbf{D}\boldsymbol{\beta}_{j}|| + \lambda ||\boldsymbol{\beta}_{j}||^{2} + \lambda_{1,j}||\boldsymbol{\beta}_{j}||_{1}$$

= $(\mathbf{y}_{j} - \boldsymbol{\beta}_{j})^{\mathsf{T}}\mathbf{D}^{\mathsf{T}}\mathbf{D}(\mathbf{y}_{j} - \boldsymbol{\beta}_{j}) + \lambda ||\boldsymbol{\beta}_{j}||^{2} + \lambda_{1,j}||\boldsymbol{\beta}_{j}||_{1}$ (19)

- If the covariance matrix of **D** is known (denoted below as Σ), the term **D**^T**D** can be replaced with
- ₃₈₇ Σ . For solving the eq. 16, the **D** matrix can be replaced by $\Sigma^{\frac{1}{2}}$, which is the square root matrix of Σ .
- 388 Resulting in the updated equation 20

$$\hat{\boldsymbol{\beta}}_{j} = \underset{\boldsymbol{\beta}_{j}}{\arg\min} ||\boldsymbol{\Sigma}^{\frac{1}{2}} \mathbf{y}_{j} - \boldsymbol{\Sigma}^{\frac{1}{2}} \boldsymbol{\beta}_{j}||^{2} + \lambda ||\boldsymbol{\beta}_{j}||^{2} + \lambda_{1,j} ||\boldsymbol{\beta}_{j}||_{1}$$
(20)

389 Sparse gcPCA method

We can implement sparse gcPCA by adapting the sparse PCA method presented and following sim-390 ilar steps proposed in **Boileau et al. (2020**). For sparse gcPCA, the covariance matrix Σ is replaced 391 with the matrix Θ which reflects the appropriate objective function of the version used. Following 392 gcPCA objective function 11, $\Theta = \mathbf{M}^{-1} (\mathbf{C}_A - \mathbf{C}_B) \mathbf{M}^{-1}$, where **M** is the square root matrix of $\mathbf{C}_A + \mathbf{C}_B$. 393 For version 1 (equivalent to cPCA), we instead use $\Theta = C_A - \alpha C_B$, and for versions 2 and 3 we 394 change Θ to match their respective objective functions, as mentioned previously. We removed the 395 I matrix from the objective function so the sparsity is enforced in the features and not in the principal components. The components \mathbf{X} are the gcPCs identified by the ordinary gcPCA algorithm. 397 Following the numerical solution for sparse PCA presented before, the sparse gcPCA is obtained 398 by the following alternating algorithm until convergence: 300 **B** given Y: Each sparse gcPC (denoted here as β_i) was found according to the following elastic net 400

401 solution:

$$\hat{\boldsymbol{\beta}}_{j} = \underset{\boldsymbol{\beta}_{j}}{\arg\min} ||\boldsymbol{\Theta}^{\frac{1}{2}} \mathbf{y}_{j} - \boldsymbol{\Theta}^{\frac{1}{2}} \boldsymbol{\beta}_{j}||^{2} + \lambda ||\boldsymbol{\beta}_{j}||^{2} + \lambda_{1,j} ||\boldsymbol{\beta}_{j}||_{1}$$
(21)

⁴⁰² Where \mathbf{y}_j is the *j*th gcPC, and $\boldsymbol{\beta}$ is the sparse gcPC. The ridge penalty λ is used to fix rank-deficient ⁴⁰³ matrices. To simplify our approach, we used the same λ_1 for all components instead of a different ⁴⁰⁴ $\lambda_{1,j}$ for every *j*th component. Therefore, the eq. 21 is reduced to a lasso regression and is solved ⁴⁰⁵ through least angle regression, similar to **Zou et al. (2006)**. 406 **Y given B:** Using fixed **B**, we can find a new $\hat{\mathbf{Y}}$ with a Procrustes rotation using SVD:

$$\boldsymbol{\Theta}^{\frac{1}{2}}\boldsymbol{\Theta}^{\frac{1}{2}}\mathbf{B} = \mathbf{U}\mathbf{S}\mathbf{V}^{\mathsf{T}}$$
(22)

We can then determine $\hat{\mathbf{Y}} = \mathbf{U}\mathbf{V}^{\mathsf{T}}$. These steps are repeated until loadings converge. The main caveat with this approach is that for gcPCA v3 or v4, the matrix $\boldsymbol{\Theta}$ can have negative eigenvalues, which prevents the calculation of the square root matrix $\boldsymbol{\Theta}^{\frac{1}{2}}$. To overcome this problem we follow

similar steps to **Boileau et al. (2020**), which we briefly replicate here. In gcPCA v3 or v4, positive

and negative eigenvalues have a clear interpretation: for the matrix Θ , positive eigenvalues denote

vectors with larger variance in condition A, while negative eigenvalues denote vectors with larger

variance in condition *B*. We can then perform eigendecomposition of Θ and replace any negative

 $_{\scriptscriptstyle 414}$ eigenvalue with zeros to make Θ_+ positive semi-definite:

$$\Theta = \mathbf{V} \mathbf{\Lambda} \mathbf{V}^{\mathsf{T}}$$

$$\Theta_{+} = \mathbf{V} \mathbf{S} \mathbf{V}^{\mathsf{T}}$$

where $\mathbf{S}_{i} = \begin{cases} \mathbf{\Lambda}_{i} & \text{if } \mathbf{\Lambda}_{i} > 0 \\ 0 & \text{otherwise} \end{cases}$
(23)

⁴¹⁵ We can then define the square root matrix $\Theta_{+} = \Theta_{+}^{\frac{1}{2}} \Theta_{+}^{\frac{1}{2}}$. We note that this matrix is only able to ⁴¹⁶ solve sparse gcPCA for vectors enriched in condition *A*. Here we propose a solution for finding ⁴¹⁷ the vectors also in condition *B*. As mentioned previously, the sign of the eigenvalues of matrix Θ ⁴¹⁸ indicates whether they are more expressed in condition *A*(+) or *B*(-). To solve the sparse gcPCA for ⁴¹⁹ condition *B*, we turn any positive eigenvalue to zero and switch the sign of negative eigenvalues to ⁴²⁰ positive:

$$\Theta = \mathbf{V} \mathbf{\Lambda} \mathbf{V}^{\mathsf{T}}$$

$$\Theta_{-} = \mathbf{V} \mathbf{D} \mathbf{V}^{\mathsf{T}}$$
where $\mathbf{D}_{i} = \begin{cases} -\mathbf{\Lambda}_{i} & \text{if } \mathbf{\Lambda}_{i} < 0 \\ 0 & \text{otherwise} \end{cases}$
(24)

Although Θ_{-} does not contain negative eigenvalues, it still represents the dimensions most expressed in condition *B*. This procedure is equivalent to switching the order of conditions *A* and *B* as the eigenvalues would be flipped in sign. The sparse gcPCs are found separately for Θ_{+} and Θ_{-} and are later concatenated for the final sparse gcPCs.

425 Synthetic data generation

In the synthetic data, we generated two conditions with 1×10^5 samples and 100 dimensions. The 426 dimensions were sampled from a Gaussian distribution (mu = 0 and sigma = 1) and then orthogo-427 nalized using singular value decomposition and picking the left singular vectors. In each condition, 428 we created a pattern in the samples that was to be discovered. In condition A, we took dimensions 429 71 and 72 and drew the samples from a uniform distribution ([0, 1]). In dimension 71 we replaced 430 any value from 0.3 and 0.7 with a different uniform distribution ([0, 0,4]). In dimension 72 all the 431 values between 0.4 and 0.6 were replaced with another uniform distribution ([0, 0,4]). This created 432 the square with a square hole in the middle in Fig. 1 A. The values were then offset by 0.5, the sam-433 ples were sorted by the angle they formed in each dimension, calculated by the inverse tangent 434 (tan inverse $\frac{X_{71}}{x}$). The samples in each dimension were normalized by their l_2 -norm. In condition B, 435 we generated the samples of dimensions 81 and 82 from a uniform distribution ([0, 1]), sorted the 436 sample values based on dimension 81, and rotated both dimensions by 45 degrees. This created 437

the diamond shape seen in Fig. 1 A. The sample values were later normalized by their l₂-norm. 438 The samples for all the other dimensions were drawn from a Gaussian distribution ($\mu = 0$ and 439 $\sigma = 1$), and then normalized by their l_2 -norm. The magnitude of each dimension was established 440 as a line with a negative slope, starting at value 10 in the 1st dimension and ending at 0.001 in 441 the 100th dimension. For condition A, we doubled the magnitude in dimensions 71 and 72, while 442 in condition B we doubled the magnitude in dimensions 81 and 82. Identifying these changes in 443 magnitude is the goal of contrastive methods. Because the samples were drawn from a normal 444 distribution, the dimensions will display correlations among them. To estimate the total variance 445 explained by each dimension, we use a OR decomposition approach described in *Zou et al.* (2006). 44F In brief, let Z be a matrix containing scores of each dimension generated, the variance is usually 447 calculated through tr($Z^{T}Z$), where tr is the trace of the matrix. However, in correlated scores, this 448 estimate is too optimistic. Using regression projection, it is possible to find the linear relationships 110 of the dimensions and correct to find the adjusted total variance. Zou et al. (2006) shows that this 450 is equivalent to using QR decomposition in Z, such that Z = OR where O is orthonormal and R is 451 upper triangular, and calculating the adjusted variance as follows: 452

adjusted variance
$$_{i} = \sum_{j=1}^{k} R_{ij}^{2}$$
 (25)

453

454 Face dataset

For the facial expression analysis, we used the Chicago Face Database *Mg et gl. (2015)*. This 455 database consists of neutral and emotional expression faces, and for the analysis we used a sub-456 set of samples that had happy and angry faces alongside neutral ones. We used only male faces 457 to reduce variability in feature positioning. The images used were cropped by an ellipse (length 458 of 75 pixels and width of 45 pixels) centered in the face to focus on the facial expression rather 450 than other features such as hairstyle or shoulders. Each sample image was flattened from a two-460 dimensional matrix to a vector, and all the flattened samples were then concatenated, resulting in 461 a matrix of samples x features. Each feature was z-scored and normalized by its l^2 -norm. Condition 463 A consisted of all the samples of happy and angry facial expressions, while condition B samples 463 were neutral expressions. 464

465 Hippocampal electrophysiology data

We used a previously published hippocampal electrophysiology dataset, with the experimental 466 details listed in the original publication Girardeau et al. (2017). In brief, Long-Evans rats were im-467 planted with silicon probes in the dorsal hippocampus CA1 region (either left or right hemisphere). 468 and neuronal activity was isolated through automatic spike sorting and manually curated. Animals 469 were trained to collect water rewards at the end of a linear track, and an air puff was introduced 470 at a fixed location for every lap, in only one of the directions. Recordings consisted of task, where 471 the animal learned the air puff location, and periods of pre- and post-task activity. For testing the 472 contrastive methods, we used pre-task recordings as condition B and post-task recordings as con-473 dition A. For our analysis, we only used neurons that had a minimum firing rate of 0.01 spikes/s 474 during the task. We binned the neural data using a bin size of 10 ms and smoothed using a rolling 475 average with a Gaussian window of size 5 bins. The data was then z-scored and normalized by the 476 norm before testing the contrastive methods. The task data was then projected on the contrastive 477 dimensions for evaluation. 478

479 Pancreatic single-cell RNA sequencing

For the single-cell RNA sequencing data analysis, we used a previously published dataset *Martínez-López et al.* (*2023*), available at GEO accession GSE153855, consisting of scRNA-seq data from human pancreatic islet cells from patients with type II diabetes and healthy controls. We used

- ⁴⁸³ the annotated dataset to identify the beta cells, which were identified previously by the authors
- 484 *Martínez-López et al. (2023*). For condition *A* we used the beta cells from subjects that had type II
- $_{435}$ diabetes, and for condition *B* we used the beta cells from healthy patients. We used the expres-
- sion values in reads per kilobase of the gene model and million mappable reads (RPKMs). The
- values were log-transformed, and all the features were centered before the analysis. gcPCA was
- ⁴⁸⁸ performed using the same set of genes used in the analysis by *Martínez-López et al.* (2023).

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495 Author contributions

- 496 L.S. and E.F.O. conceived the project and developed the mathematical framework. E.F.O., P.G.,
- and L.S. wrote the toolbox code and performed data analysis. L.S. supervised the project with
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499 **References**

- Abid A, Zhang MJ, Bagaria VK, Zou J. Exploring patterns enriched in a dataset with contrastive principal compo nent analysis. Nat Commun. 2018 May; 9(1):2134.
- Ahlqvist E, Prasad RB, Groop L. Subtypes of type 2 diabetes determined from clinical parameters. Diabetes.
 2020 Oct; 69(10):2086–2093.
- **Boileau P**, Hejazi NS, Dudoit S. Exploring high-dimensional biological data with sparse contrastive principal component analysis. Bioinformatics. 2020 Jun; 36(11):3422–3430.
- Brereton RG, Lloyd GR. Partial least squares discriminant analysis: taking the magic away: PLS-DA: taking the
 magic away. J Chemom. 2014 Apr; 28(4):213–225.
- Chapin JK, Nicolelis MA. Principal component analysis of neuronal ensemble activity reveals multidimensional
 somatosensory representations. J Neurosci Methods. 1999 Dec; 94(1):121–140.
- ⁵¹⁰ Chung NC, Storey JD. Statistical significance of variables driving systematic variation in high-dimensional data.
 ⁵¹¹ Bioinformatics. 2015 Feb; 31(4):545–554.
- 512 Diabetes Genetics Initiative of Broad Institute of Harvard and MIT, Lund University, and Novartis Insti-
- tutes of BioMedical Research, Saxena R, Voight BF, Lyssenko V, Burtt NP, de Bakker PIW, Chen H, Roix JJ,
 Kathiresan S, Hirschhorn IN, Daly MI, Hughes TE, Groop L, Altshuler D, Almgren P, Florez IC, Mever I, Ardlie K.
- Bengtsson Boström K, Isomaa B, et al. Genome-wide association analysis identifies loci for type 2 diabetes
- and triglyceride levels. Science. 2007 Jun; 316(5829):1331–1336.
- **Foster DJ**. Replay Comes of Age. Annu Rev Neurosci. 2017 Jul; 40(1):581–602.
- Fueger PT, Schisler JC, Lu D, Babu DA, Mirmira RG, Newgard CB, Hohmeier HE. Trefoil factor 3 stimulates human and rodent pancreatic islet beta-cell replication with retention of function. Mol Endocrinol. 2008
- 520 May; 22(5):1251–1259.
- Girardeau G, Inema I, Buzsáki G. Reactivations of emotional memory in the hippocampus–amygdala system
 during sleep. Nat Neurosci. 2017 Sep; 20(11):1634–1642.
- 523 Greenwald WW, Chiou J, Yan J, Qiu Y, Dai N, Wang A, Nariai N, Aylward A, Han JY, Kadakia N, Regue L, Okino
- 524 ML, Drees F, Kramer D, Vinckier N, Minichiello L, Gorkin D, Avruch J, Frazer KA, Sander M, et al. Pancreatic 525 islet chromatin accessibility and conformation reveals distal enhancer networks of type 2 diabetes risk. Nat
- islet chromatin accessibility and conformation reve
- 526 Commun. 2019 May; 10(1):2078.

- 527 Horn S, Kirkegaard JS, Hoelper S, Seymour PA, Rescan C, Nielsen JH, Madsen OD, Jensen JN, Krüger M, Grønborg
- M, Ahnfelt-Rønne J. Research resource: A dual proteomic approach identifies regulated islet proteins during
 β-cell mass expansion in vivo. Mol Endocrinol. 2016 Ian: 30(1):133–143.
- β -cell mass expansion in vivo. Mol Endocrinol. 2016 Jan; 30(1):133–143.
- Hotelling H. Analysis of a complex of statistical variables into principal components. J Educ Psychol. 1933 Sep;
 24(6):417–441.
- Izenman AJ. Linear Discriminant Analysis. In: Izenman AJ, editor. *Modern Multivariate Statistical Techniques: Regression, Classification, and Manifold Learning* New York, NY: Springer New York; 2008.p. 237–280.
- Jian X, Felsenfeld G. Insulin promoter in human pancreatic β cells contacts diabetes susceptibility loci and regulates genes affecting insulin metabolism. Proc Natl Acad Sci U S A. 2018 May; 115(20):E4633–E4641.
- 536 Jun JJ, Steinmetz NA, Siegle JH, Denman DJ, Bauza M, Barbarits B, Lee AK, Anastassiou CA, Andrei A, Aydın
- ⁵³⁷ Ç, Barbic M, Blanche TJ, Bonin V, Couto J, Dutta B, Gratiy SL, Gutnisky DA, Häusser M, Karsh B, Ledochow-⁵³⁸ itsch P, et al. Fully integrated silicon probes for high-density recording of neural activity. Nature. 2017 Nov;
- 539 551(7679):232-236.
- 540 Kim BM, Han YM, Shin YJ, Min BH, Park IS. Clusterin expression during regeneration of pancreatic islet cells in
 541 streptozotocin-induced diabetic rats. Diabetologia. 2001 Dec; 44(12):2192–2202.
- 542 Kim BM, Kim SY, Lee S, Shin YJ, Min BH, Bendayan M, Park IS. Clusterin induces differentiation of pancreatic
 543 duct cells into insulin-secreting cells. Diabetologia. 2006 Feb; 49(2):311–320.
- Kudrimoti HS, Barnes CA, McNaughton BL. Reactivation of hippocampal cell assemblies: effects of behavioral
 state, experience, and EEG dynamics. J Neurosci. 1999 May; 19(10):4090–4101.
- Li J, Klughammer J, Farlik M, Penz T, Spittler A, Barbieux C, Berishvili E, Bock C, Kubicek S. Single-cell transcriptomes reveal characteristic features of human pancreatic islet cell types. EMBO Rep. 2016 Feb; 17(2):178–187.
- ⁵⁴⁸ Li J, Inoue R, Togashi Y, Okuyama T, Satoh A, Kyohara M, Nishiyama K, Tsuno T, Miyashita D, Kin T, Shapiro ⁵⁴⁹ AMJ, Chew RSE, Teo AKK, Oyadomari S, Terauchi Y, Shirakawa J. Imeglimin ameliorates β -cell apoptosis by ⁵⁵⁰ modulating the endoplasmic reticulum homeostasis pathway. Diabetes. 2022 Mar; 71(3):424–439.
- Ma DS, Correll J, Wittenbrink B. The Chicago face database: A free stimulus set of faces and norming data.
 Behav Res Methods. 2015 Dec; 47(4):1122–1135.
- Martínez-López JA, Lindqvist A, Lopez-Pascual A, Chen P, Shcherbina L, Chriett S, Skene NG, Prasad RB, Lancien
 M, Johnson PF, Eliasson P, Louvet C, Muñoz-Manchado AB, Sandberg R, Hjerling-Leffler J, Wierup N. Single-cell
 mRNA-regulation analysis reveals cell type-specific mechanisms of type 2 diabetes: 2023.
- de Oliveira EF, Kim S, Qiu TS, Peyrache A, Batista-Brito R, Sjulson L. Off-manifold coding in visual cortex revealed
 by sleep; 2022.
- Pearson K. LIII. On lines and planes of closest fit to systems of points in space. The London, Edinburgh, and Dublin
 Philosophical Magazine and Journal of Science. 1901 Nov; 2(11):559–572.
- Peyrache A, Benchenane K, Khamassi M, Wiener SI, Battaglia FP. Principal component analysis of ensemble
 recordings reveals cell assemblies at high temporal resolution. J Comput Neurosci. 2010 Aug; 29(1-2):309–
- ⁵⁶² 325.
- Lopes-dos Santos V, Ribeiro S, Tort ABL. Detecting cell assemblies in large neuronal populations. J Neurosci
 Methods. 2013 Nov; 220(2):149–166.
- Sjulson L, Peyrache A, Cumpelik A, Cassataro D, Buzsáki G. Cocaine Place Conditioning Strengthens Location Specific Hippocampal Coupling to the Nucleus Accumbens. Neuron. 2018 Jun; 98(5):926–934.e5.
- Srinivasan M, Choi CS, Ghoshal P, Pliss L, Pandya JD, Hill D, Cline G, Patel MS. ß-Cell-specific pyruvate dehydro genase deficiency impairs glucose-stimulated insulin secretion. Am J Physiol Endocrinol Metab. 2010 Dec;
 299(6):E910–7.
- Tipping ME, Bishop CM. Probabilistic Principal Component Analysis. J R Stat Soc Series B Stat Methodol. 1999
 Sep; 61(3):611–622.
- Turk M, Pentland A. Eigenfaces for recognition. J Cogn Neurosci. 1991; 3(1):71–86.
- Wilson MA, McNaughton BL. Reactivation of hippocampal ensemble memories during sleep. Science. 1994
 Jul; 265(5172):676–679.

- 575 Yong J, Parekh VS, Reilly SM, Nayak J, Chen Z, Lebeaupin C, Jang I, Zhang J, Prakash TP, Sun H, Murray S, Guo
- 576 S, Ayala JE, Satin LS, Saltiel AR, Kaufman RJ. Chop/Ddit3 depletion in β cells alleviates ER stress and corrects
- hepatic steatosis in mice. Sci Transl Med. 2021 Jul; 13(604):eaba9796.
- 578 Zass R, Shashua A. Nonnegative Sparse PCA. Adv Neural Inf Process Syst. 2006 Dec; p. 1561–1568.
- 579 Zou H, Hastie T, Tibshirani R. Sparse Principal Component Analysis. J Comput Graph Stat. 2006; 15(2):265–286.



Figure 1—figure supplement 1. The range of cPCA α values yielding correct solutions can be very narrow. A We generated synthetic data with enriched variance in both high and low variance dimensions simultaneously. In condition A we enriched the variance of two dimensions (dimensions 19 and 92), and condition B in two other dimensions with high and low variance (dimensions 31 and 82). This panel shows the finite and infinite data results for $C_A - C_B$. Stars represents the finite data value in the enriched dimensions. Even though the high variance dimensions are easy to detect with this method, the low variance ones are still occluded by spurious variability in high variance dimensions are hard to identify. C gcPCA can find all enriched dimensions simultaneously for conditions A and B. D The range of α values yielding the correct solution becomes narrow because the enriched dimensions have different absolute variance. E gcPCA correctly identifies all enriched dimensions in both conditions.