Review Article

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Regulatory T Cells in Tumor Microenvironment and Approach for Anticancer Immunotherapy

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OPEN ACCESS

Received: Jan 19, 2020 Revised: Jan 30, 2020 Accepted: Feb 2, 2020

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Conflict of Interest

The authors declare no potential conflicts of interest.

ABSTRACT

Tregs have a role in immunological tolerance and immune homeostasis by suppressing immune reactions, and its therapeutic potential is critical in autoimmune diseases and cancers. There have been multiple studies conducted on Tregs because of their roles in immune suppression and therapeutic potential. In tumor immunity, Tregs can promote the development and progression of tumors by preventing effective anti-tumor immune responses in tumor-bearing hosts. High infiltration of Tregs into tumor tissue results in poor survival in various types of cancer patients. Identifying factors specifically expressed in Tregs that affect the maintenance of stability and function of Tregs is important for understanding cancer pathogenesis and identifying therapeutic targets. Thus, manipulation of Tregs is a promising anticancer strategy, but finding markers for Treg-specific depletion and controlling these cells require fine-tuning and further research. Here, we discuss the role of Tregs in cancer and the development of Treg-targeted therapies to promote cancer immunotherapy.

Keywords: T-lymphocytes, regulatory (Treg cells); Tumor microenvironment; Immunotherapy

INTRODUCTION

Tregs have been known to function as suppressors of immune responses to self- or foreign-Ags in order to maintain immune homeostasis (1). Tregs are characterized by the expression of a master transcription factor, forkhead box P3 (FOXP3), which is critical for Treg differentiation and function, including secretion of suppressive cytokines and expression of inhibitory surface molecules (1-3). Severe autoimmune-related diseases leading to scurfy phenotype develop in mice that have the transcription factor *FOXP3* gene deleted, and humans with impaired FOXP3 suffer from immune-dysregulation, poly-endocrinopathy, enteropathy, and X-linked syndrome (IPEX), which is characterized by the development of multiple autoimmune disorders (4). Therefore, FOXP3⁺ Tregs have attracted tremendous interest because of their essential role in maintaining immune tolerance and their therapeutic potential.

In cancer, a large population of CD4⁺FOXP3⁺ T cells infiltrates into several tumor tissues to suppress the effector functions of tumor-specific T cells (5). Therefore, the depletion of Tregs

Abbreviations

A2AR, A2A receptor; APC, Ag-presenting cell: BACH2. broad complex-tramtrackbric a brac and Cap'n'collar homology 2; CCR, C-C motif chemokine receptor; CNS1, conserved non-coding sequence 1; CRC, colorectal cancer; DC, dendritic cell; eTreg, effector Treg; FOXP3, Forkhead box p 3; FR. folate receptor: GITR. glucocorticoidinduced TNFR-related protein; ICI, immune checkpoint inhibitor; ICOS, inducible T-cell costimulatory; IDO, indoleamine 2,3-dioxygenase; IPEX, immune-dysregulation, poly-endocrinopathy, enteropathy, and X-linked syndrome; iTreg, induced Treg; LAG3, lymphocyte-activation gene 3 protein; LAG-3, lymphocyte activation gene-3; PTEN, phosphatase and tensin homolog; pTreg Peripheral Treg; Teff, effector T; TIGIT, T cell immunoreceptor with Ig and ITIM domains; TIL, tumor-infiltrating lymphocyte; TIM3, T cell immunoglobulin mucin receptor 3; TME, tumor microenvironment; tTreg, thymusderived Treg; VEGFR, VEGF receptor

Author Contributions

Conceptualization: Lee SK, Kim JH, Kim BS; Investigation: Kim JH, Kim BS; Project administration: Lee SK; Supervision: Lee SK; Writing - original draft: Kim JH, Kim BS; Writing - review & editing: Lee SK, Kim JH, Kim BS. in the tumor microenvironment (TME) leads to anti-tumor effects via the reactivation of effector T (Teff) cells (6). Indeed, in cancer patients, FOXP3⁺ Tregs migrate into the TME and suppress various types of effector lymphocytes, including CD4⁺ Th cells and CD8⁺ CTLs (7,8).

Anticancer immunotherapy, especially immune checkpoint inhibitors (ICIs), can reverse the effects of immunosuppression and revitalize dysfunctional or "exhausted" CTLs, enabling them to attack cancer cells (9,10). mAbs targeting PD-1, PD-L1, and CTLA-4 have exceptional clinical efficacy against various types of cancer (11-13). However, the efficacy of ICIs proved to be unsatisfactory in most patients, and more effective therapies are required, including combination immunotherapy.

Here, we discuss the roles Tregs play in cancer and how cancer immunotherapy can be developed by targeting Tregs for immune precision medicine.

ONTOGENIC CLASSIFICATION AND DEVELOPMENT OF TREGS

Tregs can be classified into 2 subtypes depending on the site of development (14,15). Thymus-derived Tregs (tTregs) comprise the immunosuppressive subpopulation that originates from the thymus. tTregs develop by strong interactions between the TCR of CD4/CD8 double-positive or CD4 single-positive thymocytes and self-peptide-MHC complexes in the thymus, resulting in the suppression of autoimmune reactions directed against self-Ags (16,17). Whereas thymic selection leads to differentiation of self-Agspecific tTregs, peripheral Tregs (pTregs) induced in peripheral tissues mediate tolerance to innocuous foreign Ags not encountered in the thymus (18). Consequently, pTregs prevent inflammation directed against innocuous Ags, which are expressed by commensal microflora or dietary components. In certain environments, such as a TME, some Teff cells turn into FOXP3⁺ Tregs in the periphery, which are termed induced Tregs (iTregs). These different subtypes of Tregs share significant similarities, such as their dependence on the activity of the transcription factors FOXP3 and broad complex-tramtrack-bric a brac and Cap'n'collar homology 2 (BACH2); however, some distinguishable features exist (19-22). tTregs overexpress helios (a member of the Ikaros family of transcription factors) and neurophilin1 (a type 1 transmembrane protein), which are involved in the immunosuppressive activity and dominant Ag recognition, whereas iTregs frequently lack or express less of these proteins (23-25). On the other hand, an intronic FOXP3 cis-regulatory element, conserved non-coding sequence 1, harboring SMAD3 binding sites, is necessary for pTreg differentiation but is dispensable for tTreg differentiation (26). Additionally, the TCR specificity of tTregs and pTregs is distinct in many ways (18,27).

THE SUBTYPE OF TREGS CLASSIFIED BY SUPPRESSIVE FUNCTION

Tregs were initially defined as CD4⁺ T cells with high expression of CD25, an α -subunit of IL-2 receptor. However, CD25 is a general marker of T cell activation and not exclusive to Tregs, thus emphasizing the need for additional Treg-specific markers. Although FOXP3 expression is mostly restricted to the Treg population in mice, FOXP3⁺ T cells in humans possess heterogeneous properties in terms of their phenotype and immunosuppressive



Figure 1. Classification of human CD4⁺FOXP3⁺ T cells. In humans, CD4⁺FOXP3⁺ T cells can be classified into three subsets: naïve Tregs (Fr.1), eTregs (Fr.2), and non-Tregs (Fr.3). These three fractions can be distinguished based on the expression of CD45RA, cell surface markers of naive T cells, and the transcription factor FOXP3. Moreover, these subpopulations are functionally different in terms of their suppressive activity. Effector Tregs harbor strong immune suppressive activity, but non-Tregs do not possess immune suppressive activity. In the majority of cancer, eTregs predominantly infiltrate into tumor tissues. In general, the frequency of eTregs in cancer patients is 2-5% in peripheral blood but approximately 10-50% in the tumor tissues. In contrast, naïve Tregs and FOXP3⁺ non-Tregs are insufficient or absent altogether.

functions, despite the high expression level of FOXP3 upon TCR stimulation of Teff cells (28). CD4⁺CD25⁺ Tregs expressing low levels of CD127 (the α -chain of the IL-7 receptor) are regarded as functional Tregs with suppressive activities (29,30). However, TCR stimulation of naïve T cells transiently induces FOXP3 expression along with the downregulation of CD127. Given this fact, CD4⁺CD25⁺CD127¹⁰ T cells may contain some activated non-Tregs in their population. Therefore, the expression levels of CD45RA, a marker of naïve T cells, have been previously proposed as a complementary marker, as well as CD25 and FOXP3, for alternative classification of Tregs (14,15,31). According to this classification, CD4+CD25+FOXP3+ T cells can be categorized into three fractions: naïve Tregs (CD4+CD25loFOXP3loCD45RA+); effector Tregs (eTregs) (CD4⁺CD25^{hi}FOXP3^{hi}CD45RA⁻); and non-Tregs (CD4⁺CD25^{hi}FOXP3^{lo}CD45RA⁻) (Figure 1). Naïve Tregs are separated from the thymus but have not yet been stimulated in the periphery, and barely possess any immunosuppressive function. After TCR stimulation, naïve Tregs differentiate into eTregs and thus display highly immunosuppressive activities. However, FOXP3⁺ non-Tregs are not immunosuppressive but rather are immunostimulatory, providing inflammatory cytokines, such as IFN- γ and IL-17 (31). Therefore, the features of these types of CD4+FOXP3+T cells are closely connected to human autoimmune and inflammatory diseases. Specifically, eTregs have been referred to as the dominant CD4⁺FOXP3⁺ T cell subpopulation in patients with inflammatory diseases (including sarcoidosis), whereas FOXP3⁺ non-Tregs have been implicated as the predominant subpopulation for those with autoimmune diseases, such as lupus erythematosus (31).

MECHANISMS OF IMMUNOSUPPRESSION FOR TREGS

Tregs exert their immunosuppressive function through various modes of action. The first suppressive mechanism is associated with cytokines, and include the expenditure of IL-2 by Tregs with high levels of CD25 expression (32,33), and suppression by inhibitory cytokines (such as TGF- β , IL-10, and IL-35) (34-37). Metabolite-related suppressive mechanisms include

conversion of ATP into adenosine that can prevent optimal T cell activation (38,39), as well as the expression of indoleamine 2,3-dioxygenase (IDO) in dendritic cells (DCs), which results in T cell exhaustion by depleting amino acids essential for survival (40). Other important suppressive mechanisms involving immune checkpoint-related pathways include the disruption of Teff cells by the lymphocyte activation gene-3 (LAG-3)-MHC class II interaction, the inducible T-cell costimulator (ICOS)-ICOS ligand (ICOSL)-mediated T cell activation, and the interaction between PD-1/PD-L1 (41). Impairment of Ag-presenting cell (APC) maturation is considered as a crucial mode of action for immune suppression through the binding of CTLA-4 expressed in eTregs, which causes downregulation of CD80/86 expression. Moreover, APCs are directly eliminated by Fas/Fas ligand, perforin, and granzyme B signaling (42). The majority of observations seem to indicate that, CTLA-4-dependent and/or highaffinity IL-2R-dependent suppression of T cell activity is an especially crucial process for immunosuppression by Tregs: mice specifically lacking CTLA-4 in Tregs have impaired Tregmediated immunosuppression (43); heterozygous CTLA4 mutations have been described in patients with multiple autoimmune symptoms, and are associated with impairments in the immunosuppressive activity of Tregs (44,45); treating CTLA-4-immunoglobulin fusion protein leads to the conversion of Teff cells into an anergic state (46); high-dose IL-2 neutralizes Treg-mediated suppression of T cell activation and proliferation in vitro (32,33). Through these mechanisms, Tregs can suppress Ag-specific Teff cells.

TREG AND CANCER

Tregs in the TME

The association between Tregs and tumors in the TME has been studied for decades. The involvement of Tregs in anti-tumor immunity was initially reported in 1999 (47,48). It is demonstrated that anti-CD25 Ab depleting CD4+CD25+ Tregs retarded tumor growth in T cell-deficient mice transplanted with CD25⁺ cell-depleted splenocytes. Tregs accumulate at tumor sites and in the peripheral circulation of patients with cancer, and their immunosuppressive function, as well as their number, are increased compared to those found in healthy donors (47,48). Tregs that have infiltrated into human tumors account for 10%–50% of CD4⁺ T cells in tumors, which is more abundant relative to the 2%–5% of CD4⁺ T cells found in the peripheral blood of individuals without cancer. Furthermore, higher levels of tumor-infiltrating Tregs and Treg/Teff cell ratio indicate poor prognosis in patients with various types of cancers, such as non-small cell lung carcinoma (NSCLC), melanoma, and gastric cancer (49,50). The accumulation of Tregs in tumors are well-studied in the previous reports, elucidating their ability to effectively migrate into tissue sites depending on the expression of multiple chemokine receptors; for example, CXCR5 in Tregs from the lymph node of patients with lung cancer (51). C-C motif chemokine receptor (CCR) 4 with CCL12, CCR4 with CCL17, CCR10 with CCL28, and CXCR4 with CXCL1 have been reported as other chemokine receptors on Tregs with their partner chemokines (51-56). Treg infiltration into tumor tissues has been extensively investigated in the context of recent "immune-oncology" researches (57,58). These studies confirmed the conspicuous presence of Tregs among tumor-infiltrating lymphocytes, especially in tumors harboring large immune cell infiltrates (59). Also, based on numerous articles, it is demonstrated that Tregs block antitumor immunity, and thus enhance tumor progression, and their presence in the TME is profoundly linked with unfavorable prognosis, resulting in short OS (60,61). Notably, Tregs that directly interact with the tumor are more essential for the study of immune evasion by the tumor, because the peripheral Tregs do not always represent immune-tolerant TME (49). Recently,



Figure 2. Role of Tregs in immune-evasion of cancer after differentiation from the thymus. Natural Tregs, generated in the thymus, are initially differentiated from the thymocytes by using thymic "positive selection" based on the binding affinity of TCR to the self- peptide-MHC complexes expressed on thymic APCs. The CD4⁺ T cells which bind to self-peptide-MHC complexes with the highest affinity are removed through apoptosis, and those that cannot bind at all with the complexes will also be removed because of the absence of TCR stimulation. After strong TCR stimulation, these immature precursor cells undergo IL-2-mediated signaling, thus expressing the master transcription factor FOXP3, which orchestrates the differentiation of these cells into Tregs. By contrast, immature T cells with lower affinity for self-peptide-MHC complexes are also positively selected but differentiate into Teff cells. Even though some Teff cells are auto-reactive, Tregs can block the autoimmunity of Teff cells owing to their higher affinity. These immune cells that have departed from the thymus travel through the blood vessels and move wherever they are needed. In the tumor microenvironment, especially, Tregs expressing the chemokine receptors, such as CCR4, CCR5, CCR8, and CCR10, are recruited to and around the tumors by binding to chemokines including CCL1, CCL5, CCL22, and CCL28 that are secreted from various kinds of tumors. Moreover, Tregs constitutively express the IL-2 receptor subunit- α (also known as CD25) that binds to IL-2 with higher affinity, resulting in the depletion of IL-2 from their surroundings. This leads to the reduction of the availability of this cytokine to Teff cells and secrete granzymes and perforins that can directly kill these cells. Moreover, abundant adenosine is produced by Tregs via nucleotidase activity of CD39 and CD73, which provides immunosuppressive signals to Teff cells and secrete granzymes and perforins that can directly kill these cells. Ancover, abundant adenosine A2AR.

compelling evidence suggests that colorectal cancer (CRC) abundantly infiltrated with the FOXP3^{hi} subset of suppression-competent eTregs lead to poor prognosis, while the presence of pro-inflammatory cytokine-secreting CD4⁺CD45RA^{lo}FOXP3^{lo} T cells (non-Tregs) in tumor tissues is associated with favorable outcomes (50). Therefore, especially in cancer patients with high numbers of tumor-infiltrating Tregs, further analysis needs to be conducted in order to distinguish FOXP3⁺ non-Tregs from FOXP3^{hi} eTregs in tumors. This will help evaluate the clinical importance of FOXP3⁺ cells in tumor tissues. In summary, Tregs, particularly in the TME, are a key factor of hindrance in anti-tumor immunity in various types of cancer patients, resulting in the initiation of tumor progression or resistance against cancer immunotherapy (**Figure 2**).

Molecular and cellular characteristics of Tregs in the tumor

On the basis of the functional classification of Tregs described above, Tregs in the TME is mostly composed of bona fide Treg (eTreg) cells that overexpress immunosuppressive molecules including CTLA-4 and T cell immunoreceptor with Ig and ITIM domains (TIGIT), which are not expressed much in naïve Tregs (14,62,63). Also, transcriptome analysis on human cancer specimens shows that tumor-infiltrating Tregs have high expression levels of Treg-activation surface markers, such as glucocorticoid-induced TNFR-related protein (GITR; also known as TNFRSF18), lymphocyte-activation gene 3 protein (LAG3), T cell immunoglobulin mucin receptor 3 (TIM3; also known as HAVCR2), OX40 (also known as TNFRSF4), and ICOS (64). These phenotypes, distinct from peripheral Tregs, indicate that Tregs in the TME show potent immunosuppressive activities in terms of function and number. One possible mechanism that has been suggested is that proliferating and dying

tumor cells produce a large number of self-Ags, which are recognized by Tregs, thereby inducing the activation of Tregs in the TME (65). As part of the mechanism mentioned above, whether Tregs recognize Ags exclusively or share Ags with Th cells remains unclear at this stage (65,66). Nevertheless, Tregs usually possess a higher binding affinity to TCRs than does Teff cells, resulting in the predominant activation of Tregs in the TME, even in the presence of competition with Teff cells. Furthermore, tumors can harbor some immature dendritic cells, which drive the activation and/or proliferation of Tregs in a TGF- β -dependent manner in animal models. In contrast to the abundant animal studies regarding iTregs, the existence of TGF- β -iTreg cells in humans have not been elucidated clearly; accordingly, investigations on human tumor specimens are crucial to understanding the phenotypes and origins of tumor-infiltrating Tregs.

Regulation of tumor Ag-specific T cells by Tregs

Generally, 2 different types of Ags can exist in tumor cells. First, 'neoantigens' are nonself-Ags derived from either oncogenic viral proteins or abnormal self- proteins caused by somatic mutations. Second, self-Ags that arise from the aberrant overexpression of endogenous proteins are categorized as 'shared antigens.' How CD8⁺ T cells distinguish each of these 2 types of Ag for anti-tumor immunity remains unclear. Therefore, the different immunosuppressive mechanisms of Tregs against CD8⁺ T cells specific for shared Ags versus neoantigens need to be resolved through further research. Interestingly, in some animal models, it is suggested that Tregs select for non-self-Ag specific CD8⁺ T cells harboring high-affinity TCRs by manipulating co-stimulatory signaling (67). In particular, CD8⁺ T cells targeting self-Ags are more susceptible to Tregs due to the APCs that provide limited co-stimulatory signals (68). By contrast, non-self-specific CD8⁺ T cells are resistant to suppression by Tregs in humans (68). These results demonstrate that CD8⁺ T cells specific for neoantigens are more resistant to Treg-mediated immunosuppression, and given this fact, tumors that express shared Ags can serve as more vulnerable targets for cancer immunotherapy.

TARGETING TREGS FOR CANCER IMMUNOTHERAPY

Tregs, which express the transcription factor FOXP3, are indispensable for immunological self-tolerance and immune homeostasis. They also disturb tumor immunity and can, therefore, be targeted to elicit an anti-tumor immune response by depleting them or diminishing their suppressive capabilities (69).

FOXP3, a well-characterized Treg-specific marker and the key phenotype of Tregs to function as suppressive cells, is a transcription factor expressed in the nucleus and is therefore hard to detect for clinical use.

Therapies targeting Tregs are not likely to be effective against all types of tumors. For example, Treg depletion in animal models led to the regression of tumors from certain cell lines, such as RL-male1 or MethA cells, but did not in other cell lines like AKSL2 or RL-female8 cells (70).

Thus, the identification of novel and specific biomarkers that distinguish Tregs from other cells in the TME is essential for increasing the possibility of successfully developing effective cancer therapies targeting Tregs.

Specific surface molecules on Tregs

Depletion of Tregs or attenuation of their suppressive activity can enhance tumor immunity. Tregs in the TME reveal several cell surface markers, including CD25, CTLA-4, GITR, OX40, ICOS, PD-1, LAG3, TIM3, TIGIT, CCR4, folate receptor (FR) 4 (71) and CD15s (72), and specific mAbs for these cell surface marker can be used to deplete Tregs or hinder their function (**Table 1**).

CD25

Several studies show that the removal of CD25⁺CD4⁺Tregs by anti-CD25 mAb or toxinconjugated anti-IL-2 (Denileukin diftitox) facilitates the activation of Teff cells, which greatly inhibit tumor growth in rodents (47,48,73,74). Treg depletion using an anti-CD25 mAb has been evaluated in clinical trials. When patients with breast cancer were vaccinated with various tumor-associated peptides followed by a treatment with daclizumab—an anti-CD25 mAb—to deplete Tregs, there was robust T cell priming with prolonged stable disease for 6 out of 10 patients and a median progression-free survival of 4.8 months (75). By contrast, another study showed that the administration of daclizumab depleted Teff cells as well as Tregs in patients with melanoma, but neither an antitumor immune response nor Ab production was observed (76). Because activation of Teff cells induces CD25 expression, Treg depletion by targeting CD25 can be accompanied by a deficiency in Teff cells. Thus, anti-CD25 mAb administration may lead to limited efficacy in increasing antitumor T cell responses.

CTLA-4

CTLA-4, an immune-checkpoint molecule, is expressed by tumor-infiltrating CD4⁺ and CD8⁺ Teff cells and FOXP3⁺CD4⁺ Tregs (77). The anti-tumor activity of anti-CTLA-4 mAb was originally thought to be dependent on the reinvigoration of exhausted Teff cells expressing CTLA-4 (78). However, several preclinical studies indicate that the anti-tumor activity of anti-CTLA-4 mAb is instead dependent on the depletion of CTLA-4-expressing Tregs in the TME through Ab-dependent cellular cytotoxicity, thereby increasing the Teff cell to Treg ratio. Consequently, disrupting the function of the Fc portion of the Ab completely abrogated the anti-tumor activity of the anti-CTLA-4 mAb (79-82). Therefore, further research to address the relative roles of CTLA-4 in Teff cells and Tregs in the TME of various cancers is needed.

Co-stimulatory molecules (GITR, OX40, and ICOS)

Co-stimulatory receptors, such as GITR, OX40, and ICOS, highly expressed by Tregs can be candidates for Treg depletion and functional modulation.

GITR is expressed at a high level by Tregs but at a low level by resting CD4⁺ and CD8⁺ T cells, and they play an important role in Treg expansion (83). Activation of GITR signaling

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Treg markers	Representative drugs	Function of Ab	Tumor types	Development stage
CD25	Daclizumab	Depletion	Leukemia/lymphoma	Phase 2
CTLA-4	Ipilimumab	Antagonist	Melanoma	Commercialized
PD-1	Nivolumab	Antagonist	Melanoma, lymphoma	Commercialized
GITR	TRX518	Agonist	Melanoma	Phase 1
CCR4	Mogamulizumab	Antagonist	CCR4⁺ adult T-cell leukemia/lymphoma	Commercialized
OX40	PF-04518600	Agonist	Advanced malignant cancer	Phase 2
ICOS	JTX-2011	Agonist	Advanced/refractory solid cancer	Phase 1/2
LAG3	Sym-2011	Antagonist	Solid tumor, lymphoma	Phase 1
TIM-3	Sym-023	Antagonist	Solid tumor, lymphoma	Phase 1
TIGIT	BMS-986207	Antagonist	Multiple myeloma	Phase 1/2

Table 1. Ab-drug development status of Treg-targeting therapy

through an agonistic anti-GITR mAb inhibits the suppression activity of Tregs and induces Treg-resistant Teff cells (84). The GITR agonists are now being investigated in patients with advanced solid cancer.

OX40 is constitutively expressed by a subset of Tregs, but is also found on Teff cells (85). Although OX40-agonists are used to stimulate anti-tumor responses of Teff cells, the effect on Tregs in cancer is not well understood. OX40 agonists are being investigated alone or in combination with other immunotherapies in patients with solid cancer or melanoma (86).

ICOS is important in Treg function and homeostasis (87,88), and is highly expressed by activated Tregs in tumor-infiltrating lymphocyte (TIL) of gastric cancer patients (89). Agonistic anti-ICOS mAbs, like OX40 and GITR agonists, are expected to have a dual-mode of action involving activation of Ag-specific CD4⁺ Teff cells and selective depletion of Tregs (90).

Co-inhibitory molecules (TIGIT, LAG3, and TIM3)

Immune co-inhibitory receptors predominantly expressed by Tregs are also being explored as Treg-targeted immunotherapies. TIGIT marks a population of Tregs with an enhanced suppressive capacity in the TME (91,92). TIGIT Tregs have a highly suppressive activity and they express more co-inhibitory molecules, such as LAG3, TIM3, and PD-1 compared to TIGIT-Tregs (91). In contrast, another study showed that TIGIT expression correlated with CD8⁺ Teff cell exhaustion, and TIGIT blockade increased the production of effector cytokines, such as IFN- γ and TNF- α , by CD8⁺ Teff cells in a Treg-independent manner (93). Thus, TIGIT blockade may promote anti-tumor immunity through both Treg dependent and independent mechanisms. LAG3 is expressed on TILs, especially on Tregs. Interestingly, CD4⁺CD25⁺FOXP3⁻LAG3⁺ T cell population from colorectal cancer patients produce immunosuppressive cytokines, such as IL-10 and TGF- β , and show 50% more suppressive activity than FOXP3⁺ Tregs (94). The humanized LAG3 Ab is under phase I and phase II clinical trials in patients with various solid cancers. TIM3 is expressed on activated T cells and certain subsets of Tregs and binds to several identified ligands (i.e. galectin-9, HMGB1, caecam, phosphatidyl serine) (95,96). The co-inhibitory function of TIM3 is implicated in tumor evasion and TIM3⁺ Tregs have an increased suppressive function (97,98). Co-inhibitory receptors such as LAG3, TIM3, and TIGIT seem to offer an advantage as they are dominantly overexpressed on tumor-infiltrating Tregs. However, broader studies need to be conducted in order to determine their safety and efficacy.

CCR4

Chemokine receptors, which allow Tregs to migrate to the TME site, can be a candidate molecule for Treg depletion (99). Tumor-infiltrating macrophages and tumor cells produce the CCL22, which chemoattracts Tregs expressing CCR4 (52,100,101). CCR4 is highly expressed by eTregs but not by naive Tregs or most Teff cells, except for some Th2 and Th17 cells in peripheral blood (102). *In vitro* or *in vivo* anti-CCR4 mAb treatment selectively depleted eTregs and efficiently induced tumor-specific effector CD4⁺ and CD8⁺ T cells (63). Additionally, the administration of an anti-CCR4 mAb (mogamulizumab) on advanced solid cancer patients significantly reduces eTregs in peripheral blood (70). Additional clinical trials are underway with immune checkpoint blockades.

Treg and Immune checkpoint inhibition

Immune checkpoint molecules, including CTLA-4 and PD-1, are highly expressed by activated Tregs and Teff cells (49,77). The role of CTLA-4 in Tregs is mentioned above. The role of the

inhibitory receptor PD-1 on Teff cells is well established, but its function in Tregs is less clear. Tregs in the TME show comparable levels of PD-1 expression with that of Teff cells. Because PD-1 signaling in Treg reduces its immunosuppressive activity, PD-1-deficient Tregs might potentiate the activation and immunosuppressive function of Tregs (103). Various studies reported that the anti-PD-1 mAb, nivolumab, reduced the immune-suppressive activity of Tregs (104). However, another research maintains that PD-1 inhibition induced the immune-suppressive activity mediated by Tregs in some cancer patients (105). Therefore, more research is needed to investigate the role of PD-1 in Teff cells and the role of Tregs in the TME.

Treg modulation factor in the TME

Cytokines

The TGF- β and IL-2 signaling pathways are essential to maintain the differentiation and survival of Tregs in the thymus and peripheral tissues. The effect of cancer therapy by IL-2 blockade is still unclear. In particular, hyperactivation of the TGF- β pathway in the TME enhances tumor progression by stimulating angiogenesis and inhibiting innate and adaptive anti-tumor immune responses (106). A type I TGF- β receptor serine/threonine kinase inhibitor (galunisertib) increased the ratio of CD8⁺ T cells to Tregs in melanoma animal models in a combination treatment with an anti-CTLA-4 mAb (107). In addition, a combination therapy of galunisertib with an anti-PD-1 or anti-PD-L1 mAb is currently underway in clinical trials (108). Thus, the regulation of TGF- β signaling pathways can be a noteworthy candidate for Treg control.

Targeting intracellular signaling in Tregs

PI3K signaling pathway, which is crucial for Treg maintenance and function, is a promising target for Treg-directed therapy (109). Inhibitors of PI3K effectively reduced immune suppression by Tregs in mouse models. In particular, selective inactivation of PI3Kδ in Tregs increases the activity of CD8⁺ T cells, preventing or slowing tumor development, progression, and metastasis (110). Specific ablation of the PI3K-phosphatase and tensin homolog (PTEN)-mTOR pathway in Tregs impairs mitochondrial fitness, upregulates glycolysis, leads to the loss of FOXP3 expression in Tregs, and induces Teff cell activity (111,112). Combination treatment of pembrolizumab and PI3Kδ inhibitors is currently being explored at an early stage of phase I trial in patients with advanced solid tumors. Also, tyrosine kinase inhibitors, including imatinib and dasatinib, which are known to target specific TCR signaling molecules, have been shown to reduce Treg survival and function through off-target effects (113,114). In the discontinued clinical trial for dasatinib, Treg reduction was observed and showed favorable clinical outcomes in patients with chronic myeloid leukemia (113).

CD39 and CD73

Tregs produce extracellular adenosine by the activity of CD39 and CD73 on their cell surface. Tregs express high levels of CD39 and CD73 and directly inhibit T cell activation via interaction with adenosine A2A receptor (A2AR). Moreover, adenosine increases tolerogenic APCs and enhances the immunosuppressive activity of Tregs (115). Therefore, CD39 and CD73, which are important for adenosine metabolism, can be promising therapeutic targets.

VEGF signaling

VEGF receptor (VEGFR) 2 plays an important role in tumor angiogenesis, and this signaling pathway has been shown to increase the infiltration of Tregs into tumors in animal models (116,117). In addition, blockade of VEGF-VEGFR2 signaling has been reported to inhibit tumor growth by reducing the accumulation of immunosuppressive cells, including Tregs, myeloid-

derived suppressor cells, and M2 macrophages in the TME (118). Furthermore, researchers have established that treatment of a humanized anti-VEGFR2 mAb, ramucirumab, led to a decrease in PD-1 expression in CD8⁺ T cells and a reduction in eTreg infiltration into the TME (49,119). Thus, targeting VEGFR2 molecules expressed by activated Tregs or blocking the VEGF-VEGFR2 signaling may contribute to cancer therapy through Treg inhibition.

CONCLUSION

Tregs serve as a specialized cell lineage that plays an essential role in the immunological tolerance of immune homeostasis through their immune suppressive activity. High levels of Treg infiltration in the TME lead to an undesirable prognosis in patients with various types of cancers. Depleting Tregs and regulating their function in the TME may be potential strategies for cancer therapy. Several Treg-targeted therapies are under investigation, but the lack of specific markers for Tregs has limited their clinical application. Since drugs that selectively deplete Tregs in the TME of cancer patients have not been developed at present, identification of specific targets for disrupting and depleting Tregs is important for the success of cancer immunotherapy. In the future, the development of Treg-targeted therapies based on the TME's comprehensive immune profiling may lead to new therapies and immune precision for individual cancer patients.

ACKNOWLEDGEMENTS

This work was supported by the National Research Foundation of Korea (NRF) grant funded by the Korea government, Ministry of Science and ICT (MSIT) (NRF-2017R1A2A1A17069807); Global Research Laboratory (GRL) Program through the NRF funded by the MSIT (NRF-2016K1A1A2912755).

REFERENCES

- Josefowicz SZ, Lu LF, Rudensky AY. Regulatory T cells: mechanisms of differentiation and function. *Annu Rev Immunol* 2012;30:531-564.
 PUBMED | CROSSREF
- Fontenot JD, Gavin MA, Rudensky AY. Foxp3 programs the development and function of CD4⁺CD25⁺ regulatory T cells. *Nat Immunol* 2003;4:330-336.
 PUBMED | CROSSREF
- Hori S, Nomura T, Sakaguchi S. Control of regulatory T cell development by the transcription factor Foxp3. Science 2003;299:1057-1061.
 PUBMED | CROSSREF
- 4. Bennett CL, Christie J, Ramsdell F, Brunkow ME, Ferguson PJ, Whitesell L, Kelly TE, Saulsbury FT, Chance PF, Ochs HD. The immune dysregulation, polyendocrinopathy, enteropathy, X-linked syndrome (IPEX) is caused by mutations of FOXP3. *Nat Genet* 2001;27:20-21.
 PUBMED | CROSSREF
- 5. Motz GT, Coukos G. Deciphering and reversing tumor immune suppression. *Immunity* 2013;39:61-73. PUBMED | CROSSREF
- Baecher-Allan C, Brown JA, Freeman GJ, Hafler DA. CD4*CD25^{high} regulatory cells in human peripheral blood. *J Immunol* 2001;167:1245-1253.
 PUBMED | CROSSREF
- 7. Spranger S, Spaapen RM, Zha Y, Williams J, Meng Y, Ha TT, Gajewski TF. Up-regulation of PD-L1, IDO, and Tregs in the melanoma tumor microenvironment is driven by CD8* T cells. *Sci Transl Med* 2013;5:200ra116. PUBMED | CROSSREF



- Williams JB, Horton BL, Zheng Y, Duan Y, Powell JD, Gajewski TF. The EGR2 targets LAG-3 and 4-1BB describe and regulate dysfunctional antigen-specific CD8⁺ T cells in the tumor microenvironment. *J Exp Med* 2017;214:381-400.
 PUBMED | CROSSREF
- 9. Pardoll DM. The blockade of immune checkpoints in cancer immunotherapy. *Nat Rev Cancer* 2012;12:252-264. PUBMED | CROSSREF
- Zou W, Wolchok JD, Chen L. PD-L1 (B7-H1) and PD-1 pathway blockade for cancer therapy: mechanisms, response biomarkers, and combinations. *Sci Transl Med* 2016;8:328rv4.
- Brahmer JR, Tykodi SS, Chow LQ, Hwu WJ, Topalian SL, Hwu P, Drake CG, Camacho LH, Kauh J, Odunsi K, et al. Safety and activity of anti-PD-L1 antibody in patients with advanced cancer. *N Engl J Med* 2012;366:2455-2465.
 PUBMED | CROSSREF
- 12. Hodi FS, O'Day SJ, McDermott DF, Weber RW, Sosman JA, Haanen JB, Gonzalez R, Robert C, Schadendorf D, Hassel JC, et al. Improved survival with ipilimumab in patients with metastatic melanoma. *N Engl J Med* 2010;363:711-723.

PUBMED | CROSSREF

- Topalian SL, Hodi FS, Brahmer JR, Gettinger SN, Smith DC, McDermott DF, Powderly JD, Carvajal RD, Sosman JA, Atkins MB, et al. Safety, activity, and immune correlates of anti-PD-1 antibody in cancer. *N Engl J Med* 2012;366:2443-2454.
 PUBMED | CROSSREF
- 14. Togashi Y, Nishikawa H. Regulatory T cells: molecular and cellular basis for immunoregulation. *Curr Top Microbiol Immunol* 2017;410:3-27.

PUBMED | CROSSREF

- Sakaguchi S, Miyara M, Costantino CM, Hafler DA. FOXP3⁺ regulatory T cells in the human immune system. *Nat Rev Immunol* 2010;10:490-500.
- Hsieh CS, Zheng Y, Liang Y, Fontenot JD, Rudensky AY. An intersection between the self-reactive regulatory and nonregulatory T cell receptor repertoires. *Nat Immunol* 2006;7:401-410.
 PUBMED | CROSSREF
- Wong J, Obst R, Correia-Neves M, Losyev G, Mathis D, Benoist C. Adaptation of TCR repertoires to selfpeptides in regulatory and nonregulatory CD4⁺ T cells. *J Immunol* 2007;178:7032-7041.
 PUBMED | CROSSREF
- Yadav M, Stephan S, Bluestone JA. Peripherally induced Tregs role in immune homeostasis and autoimmunity. *Front Immunol* 2013;4:232.
 PUBMED | CROSSREF
- Ziegler SF. FOXP3: of mice and men. Annu Rev Immunol 2006;24:209-226.
 PUBMED | CROSSREF
- 20. Roychoudhuri R, Hirahara K, Mousavi K, Clever D, Klebanoff CA, Bonelli M, Sciumè G, Zare H, Vahedi G, Dema B, et al. BACH2 represses effector programs to stabilize Treg-mediated immune homeostasis. *Nature* 2013;498:506-510.
 PUBMED | CROSSREF
- Igarashi K, Kurosaki T, Roychoudhuri R. BACH transcription factors in innate and adaptive immunity. Nat Rev Immunol 2017;17:437-450.
 PUBMED | CROSSREF

22. Kim EH, Gasper DJ, Lee SH, Plisch EH, Svaren J, Suresh M. Bach2 regulates homeostasis of Foxp3⁺ regulatory T cells and protects against fatal lung disease in mice. *J Immunol* 2014;192:985-995. PUBMED | CROSSREF

- 23. Overacre-Delgoffe AE, Chikina M, Dadey RE, Yano H, Brunazzi EA, Shayan G, Horne W, Moskovitz JM, Kolls JK, Sander C, et al. Interferon-γ drives Treg fragility to promote anti-tumor immunity. *Cell* 2017;169:1130-1141.e11.
 PUBMED | CROSSREF
- 24. Sarris M, Andersen KG, Randow F, Mayr L, Betz AG. Neuropilin-1 expression on regulatory T cells enhances their interactions with dendritic cells during antigen recognition. *Immunity* 2008;28:402-413. PUBMED | CROSSREF
- Getnet D, Grosso JF, Goldberg MV, Harris TJ, Yen HR, Bruno TC, Durham NM, Hipkiss EL, Pyle KJ, Wada S. A role for the transcription factor Helios in human CD4⁺CD25⁺ regulatory T cells. *Mol Immunol* 2010;47:1595-1600.
 PUBMED | CROSSREF

https://immunenetwork.org



- 26. Zheng Y, Josefowicz S, Chaudhry A, Peng XP, Forbush K, Rudensky AY. Role of conserved non-coding DNA elements in the Foxp3 gene in regulatory T-cell fate. *Nature* 2010;463:808-812. PUBMED | CROSSREF
- Lathrop SK, Santacruz NA, Pham D, Luo J, Hsieh CS. Antigen-specific peripheral shaping of the natural regulatory T cell population. *J Exp Med* 2008;205:3105-3117.
 PUBMED | CROSSREF
- Tran DQ, Ramsey H, Shevach EM. Induction of FOXP3 expression in naive human CD4⁺FOXP3 T cells by T-cell receptor stimulation is transforming growth factor-beta dependent but does not confer a regulatory phenotype. *Blood* 2007;110:2983-2990.
- 29. Liu W, Putnam AL, Xu-Yu Z, Szot GL, Lee MR, Zhu S, Gottlieb PA, Kapranov P, Gingeras TR, Fazekas de St Groth B, et al. CD127 expression inversely correlates with FoxP3 and suppressive function of human CD4⁺ T reg cells. J Exp Med 2006;203:1701-1711. PURMED L CROSSREF
- 30. Seddiki N, Santner-Nanan B, Martinson J, Zaunders J, Sasson S, Landay A, Solomon M, Selby W, Alexander SI, Nanan R, et al. Expression of interleukin (IL)-2 and IL-7 receptors discriminates between human regulatory and activated T cells. *J Exp Med* 2006;203:1693-1700. PUBMED | CROSSREF
- Miyara M, Yoshioka Y, Kitoh A, Shima T, Wing K, Niwa A, Parizot C, Taflin C, Heike T, Valeyre D, et al. Functional delineation and differentiation dynamics of human CD4⁺ T cells expressing the FoxP3 transcription factor. *Immunity* 2009;30:899-911.
 PUBMED | CROSSREF
- 32. Thornton AM, Shevach EM. CD4*CD25* immunoregulatory T cells suppress polyclonal T cell activation *in vitro* by inhibiting interleukin 2 production. *J Exp Med* 1998;188:287-296.
 PUBMED | CROSSREF
- 33. Takahashi T, Kuniyasu Y, Toda M, Sakaguchi N, Itoh M, Iwata M, Shimizu J, Sakaguchi S. Immunologic self-tolerance maintained by CD25*CD4* naturally anergic and suppressive T cells: induction of autoimmune disease by breaking their anergic/suppressive state. *Int Immunol* 1998;10:1969-1980. PUBMED | CROSSREF
- 34. Steinbrink K, Wölfl M, Jonuleit H, Knop J, Enk AH. Induction of tolerance by IL-10-treated dendritic cells. *J Immunol* 1997;159:4772-4780.
 PLIBMED
- 35. Collison LW, Workman CJ, Kuo TT, Boyd K, Wang Y, Vignali KM, Cross R, Sehy D, Blumberg RS, Vignali DA. The inhibitory cytokine IL-35 contributes to regulatory T-cell function. *Nature* 2007;450:566-569. PUBMED | CROSSREF
- 36. Turnis ME, Sawant DV, Szymczak-Workman AL, Andrews LP, Delgoffe GM, Yano H, Beres AJ, Vogel P, Workman CJ, Vignali DA. Interleukin-35 limits anti-tumor immunity. *Immunity* 2016;44:316-329. PUBMED | CROSSREF
- 37. Jarnicki AG, Lysaght J, Todryk S, Mills KH. Suppression of antitumor immunity by IL-10 and TGF-betaproducing T cells infiltrating the growing tumor: influence of tumor environment on the induction of CD4⁺ and CD8⁺ regulatory T cells. *J Immunol* 2006;177:896-904.
 PUBMED I CROSSREF
- Deaglio S, Dwyer KM, Gao W, Friedman D, Usheva A, Erat A, Chen JF, Enjyoji K, Linden J, Oukka M, et al. Adenosine generation catalyzed by CD39 and CD73 expressed on regulatory T cells mediates immune suppression. J Exp Med 2007;204:1257-1265.
 PUBMED | CROSSREF
- Wilson JM, Ross WG, Agbai ON, Frazier R, Figler RA, Rieger J, Linden J, Ernst PB. The A2B adenosine receptor impairs the maturation and immunogenicity of dendritic cells. *J Immunol* 2009;182:4616-4623.
 PUBMED | CROSSREF
- 40. Uyttenhove C, Pilotte L, Théate I, Stroobant V, Colau D, Parmentier N, Boon T, Van den Eynde BJ. Evidence for a tumoral immune resistance mechanism based on tryptophan degradation by indoleamine 2,3-dioxygenase. *Nat Med* 2003;9:1269-1274.
 PUBMED | CROSSREF
- 41. Saleh R, Elkord E. Treg-mediated acquired resistance to immune checkpoint inhibitors. *Cancer Lett* 2019;457:168-179.
 PUBMED | CROSSREF
- Burchell JT, Strickland DH, Stumbles PA. The role of dendritic cells and regulatory T cells in the regulation of allergic asthma. *Pharmacol Ther* 2010;125:1-10.
 PUBMED | CROSSREF



- 43. Wing K, Onishi Y, Prieto-Martin P, Yamaguchi T, Miyara M, Fehervari Z, Nomura T, Sakaguchi S. CTLA-4 control over Foxp3⁺ regulatory T cell function. *Science* 2008;322:271-275.
 PUBMED | CROSSREF
- 44. Schubert D, Bode C, Kenefeck R, Hou TZ, Wing JB, Kennedy A, Bulashevska A, Petersen BS, Schäffer AA, Grüning BA, et al. Autosomal dominant immune dysregulation syndrome in humans with CTLA4 mutations. *Nat Med* 2014;20:1410-1416.
 PUBMED | CROSSREF
- 45. Kuehn HS, Ouyang W, Lo B, Deenick EK, Niemela JE, Avery DT, Schickel JN, Tran DQ, Stoddard J, Zhang Y, et al. Immune dysregulation in human subjects with heterozygous germline mutations in CTLA4. *Science* 2014;345:1623-1627.
 PUBMED | CROSSREF
- 46. Perez VL, Van Parijs L, Biuckians A, Zheng XX, Strom TB, Abbas AK. Induction of peripheral T cell tolerance *in vivo* requires CTLA-4 engagement. *Immunity* 1997;6:411-417. PUBMED | CROSSREF
- Onizuka S, Tawara I, Shimizu J, Sakaguchi S, Fujita T, Nakayama E. Tumor rejection by *in vivo* administration of anti-CD25 (interleukin-2 receptor alpha) monoclonal antibody. *Cancer Res* 1999;59:3128-3133.
 PUBMED
- Shimizu J, Yamazaki S, Sakaguchi S. Induction of tumor immunity by removing CD25⁺CD4⁺ T cells: a common basis between tumor immunity and autoimmunity. *J Immunol* 1999;163:5211-5218.
 PUBMED
- 49. Tada Y, Togashi Y, Kotani D, Kuwata T, Sato E, Kawazoe A, Doi T, Wada H, Nishikawa H, Shitara K. Targeting VEGFR2 with Ramucirumab strongly impacts effector/activated regulatory T cells and CD8⁺ T cells in the tumor microenvironment. *J Immunother Cancer* 2018;6:106. PUBMED | CROSSREF
- 50. Saito T, Nishikawa H, Wada H, Nagano Y, Sugiyama D, Atarashi K, Maeda Y, Hamaguchi M, Ohkura N, Sato E, et al. Two FOXP3⁺CD4⁺ T cell subpopulations distinctly control the prognosis of colorectal cancers. *Nat Med* 2016;22:679-684.
 PUBMED I CROSSREF
- 51. Akimova T, Zhang T, Negorev D, Singhal S, Stadanlick J, Rao A, Annunziata M, Levine MH, Beier UH, Diamond JM, et al. Human lung tumor FOXP3⁺ Tregs upregulate four "Treg-locking" transcription factors. *JCI Insight* 2017;2:94075. PUBMED | CROSSREF
- 52. Curiel TJ, Coukos G, Zou L, Alvarez X, Cheng P, Mottram P, Evdemon-Hogan M, Conejo-Garcia JR, Zhang L, Burow M, et al. Specific recruitment of regulatory T cells in ovarian carcinoma fosters immune privilege and predicts reduced survival. *Nat Med* 2004;10:942-949. PUBMED | CROSSREF
- 53. Takeuchi Y, Nishikawa H. Roles of regulatory T cells in cancer immunity. *Int Immunol* 2016;28:401-409. PUBMED | CROSSREF
- 54. Gobert M, Treilleux I, Bendriss-Vermare N, Bachelot T, Goddard-Leon S, Arfi V, Biota C, Doffin AC, Durand I, Olive D, et al. Regulatory T cells recruited through CCL22/CCR4 are selectively activated in lymphoid infiltrates surrounding primary breast tumors and lead to an adverse clinical outcome. *Cancer Res* 2009;69:2000-2009. PUBMED | CROSSREF
- 55. Tan MC, Goedegebuure PS, Belt BA, Flaherty B, Sankpal N, Gillanders WE, Eberlein TJ, Hsieh CS, Linehan DC. Disruption of CCR5-dependent homing of regulatory T cells inhibits tumor growth in a murine model of pancreatic cancer. *J Immunol* 2009;182:1746-1755. PUBMED | CROSSREF
- 56. Kryczek I, Wei S, Zhu G, Myers L, Mottram P, Cheng P, Chen L, Coukos G, Zou W. Relationship between B7-H4, regulatory T cells, and patient outcome in human ovarian carcinoma. *Cancer Res* 2007;67:8900-8905. PUBMED | CROSSREF
- 57. Ladoire S, Martin F, Ghiringhelli F. Prognostic role of FOXP3⁺ regulatory T cells infiltrating human carcinomas: the paradox of colorectal cancer. *Cancer Immunol Immunother* 2011;60:909-918.
 PUBMED | CROSSREF
- 58. Salama P, Phillips M, Grieu F, Morris M, Zeps N, Joseph D, Platell C, Iacopetta B. Tumor-infiltrating FOXP3⁺ T regulatory cells show strong prognostic significance in colorectal cancer. *J Clin Oncol* 2009;27:186-192. PUBMED | CROSSREF
- 59. Badoual C, Hans S, Rodriguez J, Peyrard S, Klein C, Agueznay NH, Mosseri V, Laccourreye O, Bruneval P, Fridman WH, et al. Prognostic value of tumor-infiltrating CD4⁺ T-cell subpopulations in head and neck cancers. *Clin Cancer Res* 2006;12:465-472. PUBMED | CROSSREF

- 60. Petersen RP, Campa MJ, Sperlazza J, Conlon D, Joshi MB, Harpole DH Jr, Patz EF Jr. Tumor infiltrating Foxp3⁺ regulatory T-cells are associated with recurrence in pathologic stage I NSCLC patients. *Cancer* 2006;107:2866-2872. PUBMED | CROSSREF
- Marshall EA, Ng KW, Kung SH, Conway EM, Martinez VD, Halvorsen EC, Rowbotham DA, Vucic EA, Plumb AW, Becker-Santos DD, et al. Emerging roles of T helper 17 and regulatory T cells in lung cancer progression and metastasis. *Mol Cancer* 2016;15:67.
 PUBMED | CROSSREF
- 62. Fridman WH, Pagès F, Sautès-Fridman C, Galon J. The immune contexture in human tumours: impact on clinical outcome. *Nat Rev Cancer* 2012;12:298-306.
 PUBMED 1 CROSSREF
- 63. Sugiyama D, Nishikawa H, Maeda Y, Nishioka M, Tanemura A, Katayama I, Ezoe S, Kanakura Y, Sato E, Fukumori Y, et al. Anti-CCR4 mAb selectively depletes effector-type FoxP3*CD4* regulatory T cells, evoking antitumor immune responses in humans. *Proc Natl Acad Sci U S A* 2013;110:17945-17950. PUBMED | CROSSREF
- 64. De Simone M, Arrigoni A, Rossetti G, Gruarin P, Ranzani V, Politano C, Bonnal RJ, Provasi E, Sarnicola ML, Panzeri I, et al. Transcriptional landscape of human tissue lymphocytes unveils uniqueness of tumor-infiltrating T regulatory cells. *Immunity* 2016;45:1135-1147.
 PUBMED | CROSSREF
- 65. Nishikawa H, Kato T, Tawara I, Saito K, Ikeda H, Kuribayashi K, Allen PM, Schreiber RD, Sakaguchi S, Old LJ, et al. Definition of target antigens for naturally occurring CD4⁺ CD25⁺ regulatory T cells. *J Exp Med* 2005;201:681-686.
 PUBMED | CROSSREF
- 66. Nishikawa H, Kato T, Tawara I, Ikeda H, Kuribayashi K, Allen PM, Schreiber RD, Old LJ, Shiku H. IFN-gamma controls the generation/activation of CD4⁺ CD25⁺ regulatory T cells in antitumor immune response. J Immunol 2005;175:4433-4440. PUBMED | CROSSREF
- 67. Pace L, Tempez A, Arnold-Schrauf C, Lemaitre F, Bousso P, Fetler L, Sparwasser T, Amigorena S. Regulatory T cells increase the avidity of primary CD8⁺ T cell responses and promote memory. *Science* 2012;338:532-536. PUBMED | CROSSREF
- Maeda Y, Nishikawa H, Sugiyama D, Ha D, Hamaguchi M, Saito T, Nishioka M, Wing JB, Adeegbe D, Katayama I, et al. Detection of self-reactive CD8⁺ T cells with an anergic phenotype in healthy individuals. *Science* 2014;346:1536-1540.
 PUBMED | CROSSREF
- 69. Nishikawa H, Sakaguchi S. Regulatory T cells in cancer immunotherapy. *Curr Opin Immunol* 2014;27:1-7. PUBMED | CROSSREF
- 70. Kurose K, Ohue Y, Wada H, Iida S, Ishida T, Kojima T, Doi T, Suzuki S, Isobe M, Funakoshi T, et al. Phase Ia study of FoxP3⁺ CD4 Treg depletion by infusion of a humanized anti-CCR4 antibody, KW-0761, in cancer patients. *Clin Cancer Res* 2015;21:4327-4336.
 PUBMED I CROSSREF
- 71. Jia Z, Zhao R, Tian Y, Huang Z, Tian Z, Shen Z, Wang Q, Wang J, Fu X, Wu Y, et al. A novel splice variant of FR4 predominantly expressed in CD4⁺CD25⁺ regulatory T cells. *Immunol Invest* 2009;38:718-729. PUBMED | CROSSREF
- 72. Miyara M, Chader D, Sage E, Sugiyama D, Nishikawa H, Bouvry D, Claër L, Hingorani R, Balderas R, Rohrer J, et al. Sialyl Lewis x (CD15s) identifies highly differentiated and most suppressive FOXP3^{high} regulatory T cells in humans. *Proc Natl Acad Sci U S A* 2015;112:7225-7230. PUBMED | CROSSREF
- 73. Foss F. Clinical experience with denileukin diffitox (ONTAK). *Semin Oncol* 2006;33:S11-S16. PUBMED | CROSSREF
- 74. Steitz J, Brück J, Lenz J, Knop J, Tüting T. Depletion of CD25⁺ CD4⁺ T cells and treatment with tyrosinaserelated protein 2-transduced dendritic cells enhance the interferon alpha-induced, CD8⁺ T-cell-dependent immune defense of B16 melanoma. *Cancer Res* 2001;61:8643-8646.
 PUBMED
- 75. Rech AJ, Mick R, Martin S, Recio A, Aqui NA, Powell DJ Jr, Colligon TA, Trosko JA, Leinbach LI, Pletcher CH, et al. CD25 blockade depletes and selectively reprograms regulatory T cells in concert with immunotherapy in cancer patients. *Sci Transl Med* 2012;4:134ra62. PUBMED | CROSSREF
- 76. Jacobs JF, Punt CJ, Lesterhuis WJ, Sutmuller RP, Brouwer HM, Scharenborg NM, Klasen IS, Hilbrands LB, Figdor CG, de Vries IJ, et al. Dendritic cell vaccination in combination with anti-CD25 monoclonal antibody treatment: a phase I/II study in metastatic melanoma patients. *Clin Cancer Res* 2010;16:5067-5078. PUBMED | CROSSREF



- 77. Romano E, Kusio-Kobialka M, Foukas PG, Baumgaertner P, Meyer C, Ballabeni P, Michielin O, Weide B, Romero P, Speiser DE. Ipilimumab-dependent cell-mediated cytotoxicity of regulatory T cells *ex vivo* by nonclassical monocytes in melanoma patients. *Proc Natl Acad Sci U S A* 2015;112:6140-6145. PUBMED | CROSSREF
- 78. Ribas A. Tumor immunotherapy directed at PD-1. *N Engl J Med* 2012;366:2517-2519. PUBMED | CROSSREF
- 79. Arce Vargas F, Furness AJ, Litchfield K, Joshi K, Rosenthal R, Ghorani E, Solomon I, Lesko MH, Ruef N, Roddie C, et al. Fc effector function contributes to the activity of human anti-CTLA-4 antibodies. *Cancer Cell* 2018;33:649-663.e4.
 PUBMED | CROSSREF
- Bulliard Y, Jolicoeur R, Windman M, Rue SM, Ettenberg S, Knee DA, Wilson NS, Dranoff G, Brogdon JL. Activating Fc γ receptors contribute to the antitumor activities of immunoregulatory receptor-targeting antibodies. J Exp Med 2013;210:1685-1693.
- Selby MJ, Engelhardt JJ, Quigley M, Henning KA, Chen T, Srinivasan M, Korman AJ. Anti-CTLA-4 antibodies of IgG2a isotype enhance antitumor activity through reduction of intratumoral regulatory T cells. *Cancer Immunol Res* 2013;1:32-42.
 PUBMED I CROSSREF
- Simpson TR, Li F, Montalvo-Ortiz W, Sepulveda MA, Bergerhoff K, Arce F, Roddie C, Henry JY, Yagita H, Wolchok JD, et al. Fc-dependent depletion of tumor-infiltrating regulatory T cells co-defines the efficacy of anti-CTLA-4 therapy against melanoma. *J Exp Med* 2013;210:1695-1710.
 PUBMED | CROSSREF
- van Olffen RW, Koning N, van Gisbergen KP, Wensveen FM, Hoek RM, Boon L, Hamann J, van Lier RA, Nolte MA. GITR triggering induces expansion of both effector and regulatory CD4⁺ T cells *in vivo. J Immunol* 2009;182:7490-7500.
 PUBMED | CROSSREF
- 84. Nishikawa H, Kato T, Hirayama M, Orito Y, Sato E, Harada N, Gnjatic S, Old LJ, Shiku H. Regulatory T cell-resistant CD8⁺ T cells induced by glucocorticoid-induced tumor necrosis factor receptor signaling. *Cancer Res* 2008;68:5948-5954. PUBMED I CROSSREF
- Buchan SL, Rogel A, Al-Shamkhani A. The immunobiology of CD27 and OX40 and their potential as targets for cancer immunotherapy. *Blood* 2018;131:39-48.
 PUBMED | CROSSREF
- 86. Curti BD, Kovacsovics-Bankowski M, Morris N, Walker E, Chisholm L, Floyd K, Walker J, Gonzalez I, Meeuwsen T, Fox BA, et al. OX40 is a potent immune-stimulating target in late-stage cancer patients. *Cancer Res* 2013;73:7189-7198. PUBMED | CROSSREF
- Herman AE, Freeman GJ, Mathis D, Benoist C. CD4⁺CD25⁺ T regulatory cells dependent on ICOS promote regulation of effector cells in the prediabetic lesion. *J Exp Med* 2004;199:1479-1489.
 PUBMED | CROSSREF
- Burmeister Y, Lischke T, Dahler AC, Mages HW, Lam KP, Coyle AJ, Kroczek RA, Hutloff A. ICOS controls the pool size of effector-memory and regulatory T cells. *J Immunol* 2008;180:774-782.
 PUBMED | CROSSREF
- Nagase H, Takeoka T, Urakawa S, Morimoto-Okazawa A, Kawashima A, Iwahori K, Takiguchi S, Nishikawa H, Sato E, Sakaguchi S, et al. ICOS⁺ Foxp3⁺ TILs in gastric cancer are prognostic markers and effector regulatory T cells associated with Helicobacter pylori. *Int J Cancer* 2017;140:686-695.
 PUBMED | CROSSREF
- 90. Burris HA, Callahan MK, Tolcher AW, Kummar S, Falchook GS, Pachynski RK, Tykodi SS, Gibney GT, Seiwert TY, Gainor JF, et al. Phase 1 safety of ICOS agonist antibody JTX-2011 alone and with nivolumab (nivo) in advanced solid tumors; predicted vs observed pharmacokinetics (PK) in ICONIC. *J Clin Oncol* 2017;35:3033.
 - CROSSREF
- 91. Kurtulus S, Sakuishi K, Ngiow SF, Joller N, Tan DJ, Teng MW, Smyth MJ, Kuchroo VK, Anderson AC. TIGIT predominantly regulates the immune response via regulatory T cells. *J Clin Invest* 2015;125:4053-4062. PUBMED | CROSSREF
- 92. Joller N, Lozano E, Burkett PR, Patel B, Xiao S, Zhu C, Xia J, Tan TG, Sefik E, Yajnik V, et al. Treg cells expressing the coinhibitory molecule TIGIT selectively inhibit proinflammatory Th1 and Th17 cell responses. *Immunity* 2014;40:569-581. PUBMED | CROSSREF



93. Johnston RJ, Comps-Agrar L, Hackney J, Yu X, Huseni M, Yang Y, Park S, Javinal V, Chiu H, Irving B, et al. The immunoreceptor TIGIT regulates antitumor and antiviral CD8⁺ T cell effector function. *Cancer Cell* 2014;26:923-937. PUBMED | CROSSREF

94. Scurr M, Ladell K, Besneux M, Christian A, Hockey T, Smart K, Bridgeman H, Hargest R, Phillips S, Davies M, et al. Highly prevalent colorectal cancer-infiltrating LAP* Foxp3⁻ T cells exhibit more potent immunosuppressive activity than Foxp3⁺ regulatory T cells. *Mucosal Immunol* 2014;7:428-439. PUBMED | CROSSREF

95. Anderson AC, Joller N, Kuchroo VK. Lag-3, Tim-3, and TIGIT: co-inhibitory receptors with specialized functions in immune regulation. *Immunity* 2016;44:989-1004.
PUBMED | CROSSREF

- 96. Zhu C, Anderson AC, Schubart A, Xiong H, Imitola J, Khoury SJ, Zheng XX, Strom TB, Kuchroo VK. The Tim-3 ligand galectin-9 negatively regulates T helper type 1 immunity. *Nat Immunol* 2005;6:1245-1252. PUBMED | CROSSREF
- 97. Das M, Zhu C, Kuchroo VK. Tim-3 and its role in regulating anti-tumor immunity. *Immunol Rev* 2017;276:97-111.

PUBMED | CROSSREF

- Sakuishi K, Ngiow SF, Sullivan JM, Teng MW, Kuchroo VK, Smyth MJ, Anderson AC. TIM3*FOXP3* regulatory T cells are tissue-specific promoters of T-cell dysfunction in cancer. *Oncolmmunology* 2013;2:e23849.
 PUBMED | CROSSREF
- 99. Campbell DJ, Koch MA. Phenotypical and functional specialization of FOXP3⁺ regulatory T cells. *Nat Rev Immunol* 2011;11:119-130.

PUBMED | CROSSREF

- 100. Ishida T, Ueda R. CCR4 as a novel molecular target for immunotherapy of cancer. Cancer Sci 2006;97:1139-1146. PUBMED | CROSSREF
- 101. Nishikawa H, Sakaguchi S. Regulatory T cells in tumor immunity. *Int J Cancer* 2010;127:759-767. PUBMED | CROSSREF
- 102. Kurose K, Ohue Y, Oka M. Anti-CCR4 mAb and regulatory T cells. *Gan To Kagaku Ryoho* 2013;40:1150-1155. PUBMED
- 103. Zhang B, Chikuma S, Hori S, Fagarasan S, Honjo T. Nonoverlapping roles of PD-1 and FoxP3 in maintaining immune tolerance in a novel autoimmune pancreatitis mouse model. *Proc Natl Acad Sci U S A* 2016;113:8490-8495.
 PUBMED | CROSSREF
- 104. Gianchecchi E, Fierabracci A. Inhibitory receptors and pathways of lymphocytes: the role of PD-1 in Treg development and their involvement in autoimmunity onset and cancer progression. *Front Immunol* 2018;9:2374. PUBMED | CROSSREF
- 105. Kamada T, Togashi Y, Tay C, Ha D, Sasaki A, Nakamura Y, Sato E, Fukuoka S, Tada Y, Tanaka A, et al. PD-1⁺ regulatory T cells amplified by PD-1 blockade promote hyperprogression of cancer. *Proc Natl Acad Sci U S A* 2019;116:9999-10008.
 PUBMED | CROSSREF
- 106. Colak S, Ten Dijke P. Targeting TGF-β signaling in cancer. *Trends Cancer* 2017;3:56-71. PUBMED | CROSSREF
- 107. Holmgaard RB, Schaer DA, Li Y, Castaneda SP, Murphy MY, Xu X, Inigo I, Dobkin J, Manro JR, Iversen PW, et al. Targeting the TGFβ pathway with galunisertib, a TGFβRI small molecule inhibitor, promotes anti-tumor immunity leading to durable, complete responses, as monotherapy and in combination with checkpoint blockade. *J Immunother Cancer* 2018;6:47.
 PUBMED | CROSSREF
- 108. Strauss J, Heery CR, Schlom J, Madan RA, Cao L, Kang Z, Lamping E, Marté JL, Donahue RN, Grenga I, et al. Phase I trial of M7824 (MSB0011359C), a bifunctional fusion protein targeting PD-L1 and TGFβ, in advanced solid tumors. *Clin Cancer Res* 2018;24:1287-1295.
 PUBMED | CROSSREF
- 109. Ahmad S, Abu-Eid R, Shrimali R, Webb M, Verma V, Doroodchi A, Berrong Z, Samara R, Rodriguez PC, Mkrtichyan M, et al. Differential PI3Kδ signaling in CD4⁺ T-cell subsets enables selective targeting of T regulatory cells to enhance cancer immunotherapy. *Cancer Res* 2017;77:1892-1904. PUBMED | CROSSREF
- 110. Ali K, Soond DR, Pineiro R, Hagemann T, Pearce W, Lim EL, Bouabe H, Scudamore CL, Hancox T, Maecker H, et al. Inactivation of PI(3)K p110δ breaks regulatory T-cell-mediated immune tolerance to cancer. *Nature* 2014;510:407-411.
 PUBMED | CROSSREF



- 111. Huynh A, DuPage M, Priyadharshini B, Sage PT, Quiros J, Borges CM, Townamchai N, Gerriets VA, Rathmell JC, Sharpe AH, et al. Control of PI(3) kinase in Treg cells maintains homeostasis and lineage stability. *Nat Immunol* 2015;16:188-196.
 PUBMED | CROSSREF
- 112. Shrestha S, Yang K, Guy C, Vogel P, Neale G, Chi H. Treg cells require the phosphatase PTEN to restrain T_H1 and T_{FH} cell responses. *Nat Immunol* 2015;16:178-187.
 PUBMED | CROSSREF
- 113. Imagawa J, Tanaka H, Okada M, Nakamae H, Hino M, Murai K, Ishida Y, Kumagai T, Sato S, Ohashi K, et al. Discontinuation of dasatinib in patients with chronic myeloid leukaemia who have maintained deep molecular response for longer than 1 year (DADI trial): a multicentre phase 2 trial. *Lancet Haematol* 2015;2:e528-e535.
 PUBMED | CROSSREF
- 114. Vahl JC, Drees C, Heger K, Heink S, Fischer JC, Nedjic J, Ohkura N, Morikawa H, Poeck H, Schallenberg S, et al. Continuous T cell receptor signals maintain a functional regulatory T cell pool. *Immunity* 2014;41:722-736.

PUBMED | CROSSREF

115. Ohta A, Sitkovsky M. Extracellular adenosine-mediated modulation of regulatory T cells. *Front Immunol* 2014;5:304.

PUBMED | CROSSREF

- 116. Terme M, Pernot S, Marcheteau E, Sandoval F, Benhamouda N, Colussi O, Dubreuil O, Carpentier AF, Tartour E, Taieb J. VEGFA-VEGFR pathway blockade inhibits tumor-induced regulatory T-cell proliferation in colorectal cancer. *Cancer Res* 2013;73:539-549. PUBMED | CROSSREF
- 117. Zhu P, Hu C, Hui K, Jiang X. The role and significance of VEGFR2⁺ regulatory T cells in tumor immunity. Onco Targets Ther 2017;10:4315-4319.
 PUBMED | CROSSREF
- 118. Roland CL, Lynn KD, Toombs JE, Dineen SP, Udugamasooriya DG, Brekken RA. Cytokine levels correlate with immune cell infiltration after anti-VEGF therapy in preclinical mouse models of breast cancer. *PLoS One* 2009;4:e7669.
 PUBMED | CROSSREF
- 119. Voron T, Colussi O, Marcheteau E, Pernot S, Nizard M, Pointet AL, Latreche S, Bergaya S, Benhamouda N, Tanchot C, et al. VEGF-A modulates expression of inhibitory checkpoints on CD8⁺ T cells in tumors. *J Exp Med* 2015;212:139-148.
 PUBMED | CROSSREF