

Identification of Key Biomarkers in Systemic Lupus Erythematosus by a Multi-Cohort Analysis

Meilin Wei^{1*†}, Qiguan Dong^{2†}, Shaoqiu Chen^{3†}, Junlong Wang^{3†}, Hua Yang⁴ and Qin Xiong^{5*}

¹ Department of Endocrinology and Metabolism, The Second Affiliated Hospital of Nanchang University, Jiangxi, China, ² Department of Oncology, General Hospital of Fushun Mining Bureau of Liaoning Health Industry Group, Fushun, China, ³ Molecular Biosciences and Bioengineering Program, College of Tropical Agriculture and Human Resources, University of Hawaii at Manoa, Honolulu, HI, United States, ⁴ Chaminade University of Honolulu, Honolulu, HI, United States,

⁵ Department of Endocrinology and Metabolism, The First Affiliated Hospital of Nanchang University, Jiangxi, China

OPEN ACCESS

Edited by:

Xiaoyan Wang, Shanghai Jiao Tong University, China

Reviewed by:

Jian Yin Zou, Shanghai Jiao Tong University, China Kaifeng Guo, Fudan University, China

*Correspondence:

Meilin Wei weimeilin1007@163.com Qin Xiong elinor_20@foxmail.com [†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Autoimmune and Autoinflammatory Disorders, a section of the journal Frontiers in Immunology

> **Received:** 26 April 2022 **Accepted:** 09 May 2022 **Published:** 04 July 2022

Citation:

Wei M, Dong Q, Chen S, Wang J, Yang H and Xiong Q (2022) Identification of Key Biomarkers in Systemic Lupus Erythematosus by a Multi-Cohort Analysis. Front. Immunol. 13:928623. doi: 10.3389/fimmu.2022.928623 Systemic lupus erythematosus (SLE) is an autoimmune disease that affects multiple body systems with heterogeneous clinical manifestations. Since gene expression analyses have been accomplished on diverse types of samples to specify SLE-related genes, single-cohort transcriptomics have not produced reliable results. Using an integrated multi-cohort analysis framework, we analyzed whole blood cells from SLE patients from three transcriptomics cohorts (n=1222) and identified a five-gene signature that distinguished SLE patients from controls. We validated the diagnostic performance of this five-gene signature in six independent validation cohorts (n= 469), with an area under the receiver operating characteristic curve of 0.88 [95% CI 0.7 – 0.96]. This five-gene signature may be associated with the proportion of SLE immune cells, and generalizable across ages and sample types with real diagnostic value for clinical application.

Keywords: SLE, transcriptome, diagnosis, immune cells, multi-cohort

INTRODUCTION

Systemic lupus erythematosus (SLE) is a classic systemic autoimmune disease characterized by aberrant activity of the immune system and formation of nuclear autoantigens and immune complexes resulting in inflammation of multiple organs (1).

The incidence and prevalence of SLE varies among different ethnicities and minority groups and global prevalence is about 0-241/100000 (2). The Lupus nephritis is the most common target-organ manifestation in about 50% patients with SLE and is the most common cause of morbidity and mortality (3). Moreover, other organs involvement such as CNS involvement, cardiovascular disease, lung and skin damage contribute to mortality of SLE.

Without timely diagnosis and treatment or poor control, it will lead to irreversible damage to organs, even cause death (4). With more autoantibody tests available over the past decades, more than 180 autoantibodies can be detected in patients with SLE (5). A systematic literature search and meta-regression of diagnostic data found that the vast majority of SLE patients (97.8% [95% CI 96.8, 98.5]) have positive anti-nuclear antibodies (ANAs), suggesting ANAs are closely associated with SLE (6). Therefore, ANAs are important for diagnosis of SLE and are used as a screening parameter in routine

therapy. In recent years, many novel diagnostic markers for SLE have been reported, such as serum Galectin-9, exosomal miRNAs and non-coding RNAs, further studies are needed to confirm their value in diagnosis of SLE (7, 8). There are no universally accepted diagnostic criteria for SLE because of genetic heterogeneity. However, optimized classification criteria for SLE have been widely used based on clinical manifestations and autoimmune serology.

An interplay between genetic predisposition, epigenetic and environmental factors is involved in SLE development and activity. A fraction of genetic loci with SLE susceptibility has been demonstrated (9). The diagnostic accuracy and therapy for SLE have been dramatically improved over time. However, SLE is still a disease with poor prognosis and cause serious harm to the health of patients. Therefore, diagnosing, treating, and identifying novel therapies for SLE is still challenging and more accurate diagnostic markers are needed to assess the SLE. In essence, SLE is an autoimmune disease and we speculated dysregulated gene expression from whole blood or immune cells may provide new insights in SLE diagnosis.

In the study, we analyzed whole blood or immune cells from SLE patients from transcriptomics cohorts and identified novel biomarkers for SLE diagnosis. Additional biomarkers revealed by new technology may help us improve understanding of pathogenesis of SLE.

METHODS

Study Design and Sample Collection

We conducted a systematic search for expression profiling by array and whole-genome expression datasets that diagnosed SLE patients. We identified 9 datasets and divided them into 3 discovery cohorts (GSE65391, GSE49454, GSE50635) and 6 validation cohorts (GSE10325, GSE30153, GSE37356, GSE27427, GSE39088, GSE11909) from Gene Expression Omnibus (GEO) repository (**Table 1**).

In discovery cohorts, three microarray was performed to identify differentially expressed genes in SLE and control group (Control VS SLE). Expression profiling of whole blood samples from two validation cohorts (GSE39088, GSE11909). Additional four validation cohorts (GSE10325, GSE30153, GSE37356, GSE27427), patients samples obtained from CD4 T cells/CD19 B cells, B cells, Monocytes and Neutrophils, respectively.

 TABLE 1
 Demographic of the study population in discovery and validation dataset.

Dataset	Sample type	Sample size	Annotation
GSE49454	Whole Blood	177	Discovery
GSE10325	CD4 T cells, CD19 B cells	67	Validation
GSE50635	Whole Blood	49	Discovery
GSE65391	Whole Blood	996	Discovery
GSE30153	B cells	26	Validation
GSE37356	Monocytes	72	Validation
GSE27427	Neutrophils	47	Validation
GSE39088	Whole Blood	142	Validation
GSE11909	Whole Blood	115	Validation

Statistics and Bioinformatics Analyses

Affymetrix chips were normalized using GC Robust Multi-Array Average (GCRMA) and all other chips were normalized using normal-exponential correction and quantile normalization. Log2 normalization was performed in all arrays before analysis. DerSimonian-Laird random effects were used to combine gene expression effect sizes *via* Hedges'g (10). For differential expression, we set significance thresholds of FDR less than 1% and an effect size greater than 0.8 fold (in non-log space). All analyses were executed with R software (version 4.1.3).

Establishment of SLE Score

Using the MetaIntegrator R package, we ran a forward search to identify the most miserly gene signature that maximizes diagnostic performance (11). Each sample's SLE score is derived by subtracting the mean expression of the downregulated genes from the mean expression of the upregulated genes.

Deconvolution Analysis

Through the ImmuneDeconv package, we conducted cellmixture deconvolution analysis (12). Briefly, each microarray dataset was converted into a gene-expression matrix. The methodology uses a list of genes, which are Characteristics for each cell type, as markers. Based on the expression values of signature genes, each cell type is quantified separately.

RESULTS

The Five-Gene Signature Diagnostic of SLE in Three Discovery Cohorts

We conducted a systematic search for data on whole-genome expression in whole blood from patients with SLE. Our multicohort analysis found 93 significantly differentially expressed genes (FDR < 0.01, effect size > 0.8-fold) between patients who with SLE vs Control in the three discovery cohorts (N = 1222). Using forward search, we specified a signature of five significantly differentially expressed genes in SLE and Control that was optimized for diagnostic performance (Figure 1A). The forest plots showed that the 5 selected genes were differentially expressed (4 up-regulated, 1 down-regulated) according to the forward search (Figures 1B-F). In each sample, we calculated a SLE score by subtracting the mean expression of the one downregulated gene from the mean expression of the up-regulated genes (Figure 1G and Figure S1). There was a significant difference between patients with SLE and Control in the discovery cohorts with a summary area under the curve (AUC) equal to 0.93 (95% confidence interval 0.74-0.99, Figure 1H).

Validation of the Five-Gene Signature in Six Independent Validation Cohorts of SLE Patients

We then validated the five-gene set in the six validation cohorts (N=469). We validated the up-regulated and down-regulated genes



(Figures 2A–E). Also, we calculated the SLE score of each sample in the validation cohorts and meta score of each dataset (Figure 2F and Figure S1). Although the considerable heterogeneity between datasets, including sample type, race, platform, the SLE scores accurately identified SLE patients in all six datasets (summary AUC=0.88 [95% CI 0.7-0.96]) (Figure 2G). Notably, this signature could effectively discriminate the SLE patients for the four-validation dataset (GSE11909, GSE27427, GSE39088, GSE30153). However, the AUC of SLE scores in the GSE37356 (Monocytes) and GSE10325 (CD4 T cells, CD19 B cells) dataset were 0.67 and 0.73, respectively.

Deconvolution of Immune Cell Types

We also examined whether the five-gene signature is enriched in some immune cell types by using ImmuneDeconv analysis, which is based on publicly available whole-genome expression profiles from 10 molecular immune cells. We found that the percentages of NK Cell, B cells and neutrophils were significantly changed in the SLE group. Interestingly, no statistically consistent and reproducible differences in the proportions of T cells and monocytes (Figure 3 and Figure S2).

DISCUSSION

SLE is primarily caused by autoantibodies-induced tissue injury. An irregular immune response and inflammation are also significant pathological processes in SLE. Although numerous studies have been conducted, early diagnosis of SLE remains a challenging diagnostic problem, and many studies have suffered from a reproducibility crisis. The reproducibility crisis has been exacerbated by the fact that results from homogeneous, singlecenter studies do not generally generalize to heterogeneous populations in the real world. By combining data from various populations into one study, multi-cohort gene expression analysis has improved reproducibility. Here, through the use of a large, multicenter dataset of various types of samples, we identified five diagnostic biomarkers for patients with SLE



FIGURE 2 | Validation of the 5-Gene signature diagnosis of SLE. (A–E) The up-regulated genes and down-regulated genes in independent validation cohorts. (F) A representative violin plot illustrating the performance of the five-gene signature to differentiate SLE from Control in one of the discovery cohorts (GSE39088). The Wilcoxon p value and error bars are shown. (G) ROC curves of patients with SLE versus controls in six independent validation cohorts.



(IFI27, IFI44L, ANXA3, LY96, ABCB1). Furthermore, we utilized six independent validation sets to verify the diagnostic power of the five-gene signature.

There is strong evidence from previous studies that four of the above five genes are involved in SLE pathogenesis and therapeutics. In patients with SLE, the expression of IFI27, IFI44L, and ANXA3 were up-regulated as compared to controls (13, 14). A previous study demonstrated that IFI44L promoter methylation as a blood biomarker for systemic lupus erythematosus (15). Moreover, IFI27, IFI44L, and ANXA3 are associated with highly expressed interferon-stimulated genes. Up-regulation of IFI27 (the interferon alpha-inducible protein27) expression may be associated with inflammatory response (16), and is involved in the progression of an HIV infection by regulating immune response (17). P-glycoprotein encoded by ABCB1 gene is an ATP-dependent drug efflux pump, which has been detected in some autoimmune disease, such as SLE (18). And ABCB1 gene polymorphisms may be associated with clinical features of SLE (19). Additionally, synovial tissue from SLE patients up-regulated IFI27 and IFI44L (20). Notably, we first demonstrate that LY96 genes are novel markers of SLE in this study. Molecular analysis suggests that the LY96 coding protein binds to toll-like receptor 4 on the cell surface and grants responsiveness to lipopolysaccharide (LPS), thus connecting the receptor and LPS signaling (21). The effectiveness of LY96 to another type of auto-immune disease, namely Rheumatic, has also been reported (22).

Cell-type enrichment analysis suggest that reduced abundances of NK Cell, and B cells in patients with SLE. Interestingly, there was no significant difference in monocyte proportions between SLE and the control group, and diagnostic power was also limited in the independent validation set (AUC=0.67, [95% CI 0.61-0.73]).

In summary, we identified five genes that are significantly associated with SLE diagnosis. In addition, this result was validated across nine datasets and multiple types of human blood-born samples. This suggests that these genes could serve

REFERENCES

- Durcan L, O'Dwyer T, Petri MJTL. Management Strategies and Future Directions for Systemic Lupus Erythematosus in Adults. *Lancet* (2019) 393:2332–43. doi: 10.1016/S0140-6736(19)30237-5
- Rees F, Doherty M, Grainge MJ, Lanyon P, Zhang WJR. The Worldwide Incidence and Prevalence of Systemic Lupus Erythematosus: A Systematic Review of Epidemiological Studies. *Rheumatology* (2017) 56:1945–61. doi: 10.1093/rheumatology/kex260
- Fava A, Petri M. Systemic Lupus Erythematosus: Diagnosis and Clinical Management. J Autoimmun (2019) 96:1–13. doi: 10.1016/j.jaut.2018.11.001
- Liang R, Landry I, Saliaj MJC. Systemic Lupus Erythematosus vs. Mixed Connect Tissue Dis Disguised As Scabies (2022) 14(3):e23621. doi: 10.7759/ cureus.23621
- Yaniv G, Twig G, Shor DB-A, Furer A, Sherer Y, Mozes O, et al. A Volcanic Explosion of Autoantibodies in Systemic Lupus Erythematosus: A Diversity of 180 Different Antibodies Found in SLE Patients. *Autoimmun Rev* (2015) 14:75–9. doi: 10.1016/j.autrev.2014.10.003
- Leuchten N, Hoyer A, Brinks R, Schoels M, Schneider M, Smolen J, et al. And Research, Performance of Antinuclear Antibodies for Classifying Systemic Lupus

as biomarkers for the diagnosis of SLE in real clinical application. Understanding the mechanism of SLE pathogenesis through the five-gene signature can be beneficial. In a follow-up study, we hope to gain a deeper understanding of their possible functions.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/**Supplementary Material**.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Baylor Institute for Immunology Research, UT Southwestern Medical Center, and Mayo Clinic. The patients/ participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

MW, QX, and SC established the concept and the investigation. MW, QD, SC, JW collected and processed data. MW, SC drafted the manuscript, and all authors reviewed the manuscript and approved the version for publication. All authors contributed to the article and approved the submitted version.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2022. 928623/full#supplementary-material

Erythematosus: A Systematic Literature Review and Meta-Regression of Diagnostic Data. Arthritis Care Res (2018) 70:428–38. doi: 2010.1002/acr.23292

- Matsuoka N, Fujita Y, Temmoku J, Furuya MY, Asano T, Sato S, et al. Galectin-9 as a Biomarker for Disease Activity in Systemic Lupus Erythematosus. *PloS one* (2020) 15. doi: 10.1371/journal.pone.0227069
- Li W, Liu S, Chen Y, Weng R, Zhang K, He X, et al. Circulating Exosomal microRNAs as Biomarkers of Systemic Lupus Erythematosus. *Clinics* (2020) 75:e1528. doi: 10.6061/clinics/2020/e1528
- Kwon Y-C, Chun S, Kim K, Mak AJC. Update on the Genetics of Systemic Lupus Erythematosus: Genome-Wide Association Studies and Beyond. *Cells* (2019) 8:1180. doi: 10.3390/cells8101180
- Sweeney TE, Haynes WA, Vallania F, Ioannidis JP, Khatri P. Methods to Increase Reproducibility in Differential Gene Expression *via* Meta-Analysis. *Nucleic Acids Res* (2017) 45:e1–1. doi: 10.1093/nar/gkw797
- Haynes WA, Vallania F, Liu C, Bongen E, Tomczak A, Andres-Terrè M, et al. Empowering multi-Cohort Gene Expression Analysis to Increase Reproducibility. *Pacific Symposium On Biocomputing 2017 World Sci* (2017), 144–53. doi: 10.1142/9789813207813_0015
- 12. Sturm G, Finotello F, Petitprez F, Zhang JD, Baumbach J, Fridman WH, et al. Comprehensive Evaluation of Transcriptome-Based Cell-Type Quantification

Methods for Immuno-Oncology. *Bioinformatics* (2019) 35:i436-45. doi: 10.1093/bioinformatics/btz363

- Bing P-F, Xia W, Wang L, Zhang Y-H, Lei S-F, Deng F.-Y.J.P.O. Common Marker Genes Identified From Various Sample Types for Systemic Lupus Erythematosus. *PLoS One* (2016) 11 doi: 10.1371/journal.pone.0156234
- Okada Y, Terao C, Ikari K, Kochi Y, Ohmura K, Suzuki A, et al. Meta-Analysis Identifies Nine New Loci Associated With Rheumatoid Arthritis in the Japanese Population. *Meta Anal* (2012) 44:511–6. doi: 10.1038/ng. 2231
- Zhao M, Zhou Y, Zhu B, Wan M, Jiang T, Tan Q, et al. IFI44L Promoter Methylation as a Blood Biomarker for Systemic Lupus Erythematosus. Ann Rheumatol Dis (2016) 75:1998–2006. doi: 10.1136/annrheumdis-2015-208410
- Nagasawa Y, Okuzaki D, Muso E, Yamamoto R, Shinzawa M, Iwasaki Y, et al. IFI27 Is a Useful Genetic Marker for Diagnosis of Immunoglobulin A Nephropathy and Membranous Nephropathy Using Peripheral Blood. *PloS* One (2016) 11:e0153252. doi: 10.1371/journal.pone.0153252
- Huang H, Lv J, Huang Y, Mo Z, Xu H, Huang Y, et al. IFI27 Is a Potential Therapeutic Target for HIV Infection. *Ann Med* (2022) 54:314–25. doi: 10.1080/07853890.2021.1995624
- Tsujimura S, Saito K, Nakayamada S, Tanaka Y. Relevance of Multidrug Resistance 1 and P-Glycoprotein to Drug Resistance in Patients With Systemic Lupus Erythematosus. *Histol Histopathol* (2007) 22:465–8. doi: 10.14670/HH-22.465
- Gonzalez TP, Mucenic T, Brenol JC, Xavier RM, Schiengold M, Chies JA. ABCB1 C1236T, G2677T/A and C3435T Polymorphisms in Systemic Lupus Erythematosus Patients. *Braz J Med Biol Res* (2008) 41:769–72. doi: 10.1590/ S0100-879X2008000900005

- Toukap AN, Galant C, Theate I, Maudoux A, Lories R, Houssiau F, et al. And Rheumatism, Identification of Distinct Gene Expression Profiles in the Synovium of Patients With Systemic Lupus Erythematosus. *Arthritis Rheumatol* (2007) 56:1579–88. doi: 10.1002/art.22578
- Dou T, Fu M, Wang Y, Zhao Y, Wang Z, Bian Z, et al. Signatures of Positive Selection in LY96 Gene in Vertebrates. J Biosci (2013) 38:899–904. doi: 10.1007/s12038-013-9377-9
- 22. Wang L, Wu L-F, Lu X, Mo X-B, Tang Z-X, Lei S-F, et al. Integrated Analyses of Gene Expression Profiles Digs Out Common Markers for Rheumatic Diseases. *PLoS One* (2015) 10:. doi: 10.1371/journal.pone.0137522

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Wei, Dong, Chen, Wang, Yang and Xiong. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.