



Draft Genome Sequence of *Pseudomonas* sp. Strain FEN, Isolated from the Fe- and Organic Matter-Rich Schlöppnerbrunnen Fen

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ABSTRACT Here, we report the draft genome sequence of *Pseudomonas* sp. strain FEN, a nonfluorescent siderophore producer that was isolated from the Schlöppnerbrunnen fen, which is characterized by high concentrations of Fe, dissolved organic matter (DOM), and Fe-DOM complexes. This draft genome sequence provides insight into the mechanisms of siderophore biosynthesis and siderophore-mediated iron uptake by this bacterium.

Pseudomonas spp. are Gram-negative bacteria belonging to the *Pseudomonadaceae* family within the class *Gammaproteobacteria* and are characterized by metabolic versatility and colonization of a wide range of diverse habitats (1–3). A single nonfluorescent, siderophore-producing *Pseudomonas* sp. strain was isolated from soil samples collected from the moderately acidic (pH 4.5 to 6.0), Fe-rich minerotrophic Schlöppnerbrunnen fen located in the Fichtelgebirge (northern Bavaria, Germany; 50°7′54″N, 11°52′51″E). Enrichments were grown by inoculating Fe gradient tubes, prepared as described previously (4, 5), with 25 μ l soil slurry. Briefly, the gradient tubes contained an FeS bottom layer (1:1 FeS mixed with modified Wolfe’s mineral medium [MWMM] [ATCC medium 2672] and 3% [wt/vol] agarose) overlaid with 0.2% (wt/vol) agarose-stabilized MWMM supplemented with Wolfe’s vitamin solution, 1 mM ferulic acid, and 1 mM syringic acid. Active enrichment cultures were transferred to new gradient tubes every 4 to 6 weeks, and the dilution-to-extinction method was used to obtain the *Pseudomonas* sp. strain FEN isolate.

Pseudomonas sp. FEN stock cultures were cultivated at room temperature on solid agar plates containing PS medium (ATCC medium 3). Overnight shaking cultures grown in 100 ml PS medium at room temperature were inoculated from a single colony picked from the PS plate. Overnight cultures were used for genomic DNA extraction with the GenElute bacterial genomic DNA kit (Sigma-Aldrich, Taufkirchen, Germany). Whole-genome sequencing was performed on the RS II platform (Pacific Biosciences [PacBio], Menlo Park, CA) according to the standard manufacturer’s protocol. Briefly, a 10- to 20-kb library was prepared and sequenced on a PacBio RS II sequencer using C4-P6 chemistry on single-molecule real-time (SMRT) cells, with a 180-min collection protocol. Sequencing and subsequent filtering with Hierarchical Genome Assembly Process 4 (HGAP4) resulted in 153,019 reads with an average read length of 5,607 bp. The maximum read length was 74,202 bp. The genome was assembled *de novo* with HGAP4 using default parameters except for the estimated genome size (6.0 Mbp) (6). Genome sequence annotation and gene identification were performed with RASTtk v2.0 using default parameters (7–9).

The draft genome of *Pseudomonas* sp. FEN was assembled into 36 contigs totaling 6.79 Mbp, with a G+C content of 61.1%, an N_{50} value of 576,490 bp, and an L_{50} value of 4. Mapping of filtered raw reads onto the assembly revealed 90-fold coverage. The draft genome contains 6,294 coding sequences and 98 non-protein-coding genes. Initial cloning and 16S rRNA gene-based sequencing approaches indicated that the

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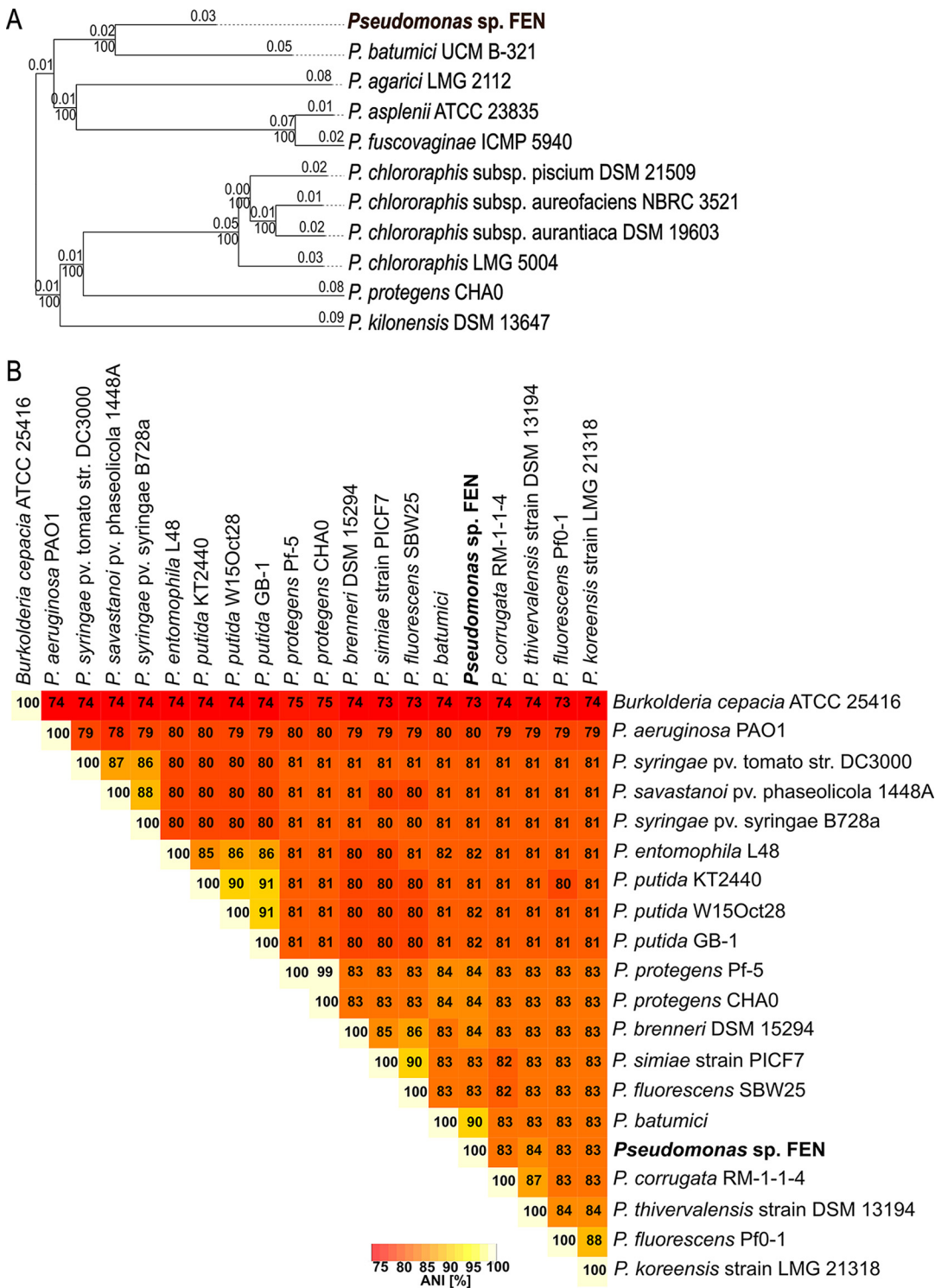


Fig 1 Phylogenetic analysis based on pairwise comparisons of *Pseudomonas* sp. FEN versus type strain genomes and ANI-based analysis of publicly available genome sequences of siderophore-producing *Pseudomonas* sp. strains. (A) Phylogenetic tree based on whole-genome comparisons using the TYGS. The phylogenetic tree was inferred using FASTME v2.1.6.1 from genome-based distance phylogeny (GBDP) distances calculated from closely related genome sequences. The branch lengths are scaled based on GBDP distances. The numbers above each branch represent GBDP pseudo-bootstrap values of >60% from 100 replications. The tree was rooted at the midpoint. (B) ANI genome-based distance matrix calculator output, based on pairwise ANI between different siderophore-producing *Pseudomonas* sp. strains within the phylum *Proteobacteria* with publicly available genomes, including the *Pseudomonas* sp. FEN isolate described here. All ANI values are shown in the matrix. *Pseudomonas cepacia* was renamed *Burkholderia cepacia* ATCC 25416 and was used in the ANI-based analysis due to its siderophore-producing phenotype.

isolate was most closely related to *Pseudomonas fluorescens*. However, recent advances in genomics and taxonomic classifications, specifically implementation of average nucleotide identity (ANI)-based comparisons, standardized taxonomic classifications (Genome Taxonomy Database [GTDB] taxonomy) (10, 11), and the automated genome-based taxonomic analysis tool Type Strain Genome Server (TYGS) (12), revealed that the isolate is more closely related to the proposed type strain *Pseudomonas batumici* UCM B-321 (3, 13), rather than to any of the publicly available *P. fluorescens* genome sequences (Fig. 1). According to GTDB-Tk v0.3.2 analysis (10, 11, 14–19), the ANI between *P. batumici* and *Pseudomonas* sp. FEN is 90.56%, based on an aligned fraction of 0.73 (proportion of regions displaying significant similarity). This taxonomic classification clarifies why fluorescent siderophores, such as pyoverdine (20), were not detected (21).

Analysis of the *Pseudomonas* sp. FEN genome revealed putative genes encoding pathways for siderophore biosynthesis, Fe-transport systems, Fe-siderophore sensor proteins, siderophore receptor proteins, and ferrichrome (hydroxamate siderophore) uptake systems and receptors. Several genes encoding nonspecific siderophore uptake systems were identified, suggesting that these systems play a role in the uptake of exogenous siderophores, including pyoverdines. Homologs of the ferric Fe-ABC transport system (*pitADC*) and a ferrous Fe-transport system (*efuUOB*) were also identified. The draft genome of *Pseudomonas* sp. FEN provides supporting evidence that this microorganism is capable of siderophore production, import, and export, as well as detection of exogenous siderophores. Further investigations may reveal additional genes or regulatory mechanisms involved in siderophore production and uptake in environments characterized by high concentrations of dissolved organic matter (DOM) and Fe-DOM complexes, such as the Schlöppnerbrunnen fen.

Data availability. The sequencing reads and assemblies for this whole-genome shotgun project are available in the European Nucleotide Archive (ENA) repository under the BioProject accession number [PRJEB40039](https://www.ebi.ac.uk/bioproject/14039). The version described in this paper is the first version. The BioSample number is [SAMEA7280114](https://www.ncbi.nlm.nih.gov/biosample/SAMEA7280114), and the individual genome assembly is available under the accession number [CAJFDC01000000](https://www.ncbi.nlm.nih.gov/genbank/CAJFDC01000000).

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REFERENCES

- Gomila M, Peña A, Mulet M, Lalucat J, García-Valdés E. 2015. Phylogenomics and systematics in *Pseudomonas*. *Front Microbiol* 6:214. <https://doi.org/10.3389/fmicb.2015.00214>.
- Lalucat J, Mulet M, Gomila M, García-Valdés E. 2020. Genomics in bacterial taxonomy: impact on the genus *Pseudomonas*. *Genes* 11:139. <https://doi.org/10.3390/genes11020139>.
- Peix A, Ramírez-Bahena M-H, Velázquez E. 2018. The current status on the taxonomy of *Pseudomonas* revisited: an update. *Infect Genet Evol* 57:106–116. <https://doi.org/10.1016/j.meegid.2017.10.026>.
- Emerson D, Floyd MM. 2005. Enrichment and isolation of iron-oxidizing bacteria at neutral pH. *Methods Enzymol* 397:112–123. [https://doi.org/10.1016/S0076-6879\(05\)97006-7](https://doi.org/10.1016/S0076-6879(05)97006-7).
- Lüdecke C, Reiche M, Eusterhues K, Nietzsche S, Küsel K. 2010. Acid-tolerant microaerophilic Fe(II)-oxidizing bacteria promote Fe(III)-accumulation in a fen. *Environ Microbiol* 12:2814–2825. <https://doi.org/10.1111/j.1462-2920.2010.02251.x>.
- Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <https://doi.org/10.1038/nmeth.2474>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formisano K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. *Sci Rep* 5:8365. <https://doi.org/10.1038/srep08365>.
- Chaumeil P-A, Mussig AJ, Hugenholtz P, Parks DH. 2019. GTDB-Tk: a toolkit to classify genomes with the Genome Taxonomy Database. *Bioinformatics* 36:1925–1927. <https://doi.org/10.1093/bioinformatics/btz848>.
- Jain C, Rodriguez-R LM, Phillippy AM, Konstantinidis KT, Aluru S. 2018. High throughput ANI analysis of 90K prokaryotic genomes reveals clear

- species boundaries. *Nat Commun* 9:5114. <https://doi.org/10.1038/s41467-018-07641-9>.
12. Meier-Kolthoff JP, Göker M. 2019. TYGS is an automated high-throughput platform for state-of-the-art genome-based taxonomy. *Nat Commun* 10:2182. <https://doi.org/10.1038/s41467-019-10210-3>.
 13. Kiprianova EA, Klochko VV, Zelena LB, Churkina LN, Avdeeva LV. 2011. *Pseudomonas batumici* sp. nov., the antibiotic-producing bacteria isolated from soil of the Caucasus Black Sea coast. *Mikrobiol Z.* 73:3–8.
 14. Parks DH, Chuvochina M, Chaumeil P-A, Rinke C, Mussig AJ, Hugenholtz P. 2020. A complete domain-to-species taxonomy for bacteria and archaea. *Nat Biotechnol* 38:1079–1086. <https://doi.org/10.1038/s41587-020-0501-8>.
 15. Matsen FA, Kodner RB, Armbrust EV. 2010. pplacer: linear time maximum-likelihood and Bayesian phylogenetic placement of sequences onto a fixed reference tree. *BMC Bioinformatics* 11:538. <https://doi.org/10.1186/1471-2105-11-538>.
 16. Hyatt D, Chen G-L, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. <https://doi.org/10.1186/1471-2105-11-119>.
 17. Price MN, Dehal PS, Arkin AP. 2010. FastTree 2: approximately maximum-likelihood trees for large alignments. *PLoS One* 5:e9490. <https://doi.org/10.1371/journal.pone.0009490>.
 18. Eddy SR. 2011. Accelerated profile HMM searches. *PLoS Comput Biol* 7:e1002195. <https://doi.org/10.1371/journal.pcbi.1002195>.
 19. Ondov BD, Treangen TJ, Melsted P, Mallonee AB, Bergman NH, Koren S, Phillippy AM. 2016. Mash: fast genome and metagenome distance estimation using MinHash. *Genome Biol* 17:132. <https://doi.org/10.1186/s13059-016-0997-x>.
 20. Cornelis P. 2010. Iron uptake and metabolism in pseudomonads. *Appl Microbiol Biotechnol* 86:1637–1645. <https://doi.org/10.1007/s00253-010-2550-2>.
 21. Kügler S, Cooper RE, Boessneck J, Küsel K, Wichard T. 2020. Rhizobactin B is the preferred siderophore by a novel *Pseudomonas* isolate to obtain iron from dissolved organic matter in peatland. *Biometals* 33:415–433. <https://doi.org/10.1007/s10534-020-00258-w>.