

# Clinical and immunologic features among COVID-19-affected mother–infant pairs: antibodies to SARS-CoV-2 detected in breast milk

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## Abstract

The coronavirus disease 2019 (COVID-19) pandemic remains threatening to women and children, but clinical evidence regarding women during pregnancy, puerperium and lactation is limited. We assessed clinical and immunologic features of and breastfeeding advice provided to mother–infant pairs. This observational analysis was conducted in a tertiary-care centre in Wuhan, China. Pregnant patients with laboratory-confirmed COVID-19 who delivered during hospitalization were enrolled. Clinical characteristics and serial specimens of the mother–infant pairs were examined, supplemented with follow-ups regarding recovery and breastfeeding. Fourteen pregnant patients had live births and recovered well; four patients continued breastfeeding while taking precautions. No neonatal infections were observed. No infants developed COVID-19 during breastfeeding. Common maternal symptoms were fever (11/14, 78.1%) and cough (6/14, 42.9%). A pregnancy-specific symptom was abnormal foetal movement, which was noticed by three patients (21.4%). The mean virus shedding time was 9 days (standard deviation, 6 days; range, 1–22 days). The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) genome was not detected in breast milk or maternal vaginal secretions. Immunologic assay revealed seroconversion of IgM on day 8 after onset and IgG on day 28. Both IgM and IgG antibodies to SARS-CoV-2 were detected in breast milk, cord blood and neonatal serum. The study results suggest that passive acquisition of antibodies against SARS-CoV-2 is available by ingesting breast milk. Breastfeeding has a low risk of transmitting SARS-CoV-2 or escalating maternal disease, so continuing breastfeeding with prudent precautions is encouraged. © 2020 The Authors. Published by Elsevier Ltd.

**Keywords:** Breastfeeding, COVID-19, infectious disease, lactation period, passive immunity, pregnancy

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## Introduction

Severe acute respiratory virus 2 (SARS-CoV-2), first identified in December 2019 in Wuhan, China, has continued to spread

rapidly, causing the ongoing pandemic [1]. As coronavirus disease 2019 (COVID-19) unfolded, various investigations have been launched to explore the pathophysiology of SARS-CoV-2. To date, the virus's genome has been detected in many different kinds of body fluids, including upper respiratory droplets, bronchoalveolar lavage, saliva, tears, conjunctival secretions, faeces, urine, blood and cerebrospinal fluid [2–4]. Breast milk, as a well-studied body fluid that delivers immunologic, nutritional and cognitive benefits and that provides maternal–foetal emotional effects [5], has not yet been fully characterized in terms of the novel coronavirus. Clinical data are also limited. Chen et al. [6] described a failure to detect the SARS-CoV-2 genome in the breast milk of six affected women in the third trimester of pregnancy. However, with more women in the early pregnancy, puerperium and lactation periods affected by and recovering from COVID-19, it is important to learn whether breastfeeding is safe, whether it plays a role in

transmitting SARS-CoV-2, what precautions ought to be taken and whether breastfeeding will affect the mother's disease course.

We assessed the clinical and immunologic features of COVID-19-affected mother-infant pairs; specifically tested breast milk for pathogens, SARS-CoV-2 neutralizing antibodies and immunologic components; and explored the feasibility of breastfeeding and related transmission possibilities.

## Patients and methods

### Study design and participants

The ambispective observational clinical analysis was conducted in a single tertiary-care centre, Tongji Hospital, affiliated with the Huazhong University of Science and Technology, located in Wuhan, China. The study retrospectively collected data of seven pregnant patients with laboratory-confirmed COVID-19 treated between 19 January and 7 February 2020, while 13 patients in various stages of pregnancy who were later diagnosed consented to the study from its baseline. Follow-up investigation focused on mothers' and infants' outcomes, breastfeeding and recovery. The last follow-up date was 5 April 2020. Patients who were enrolled met the following criteria [7]: positive real-time reverse transcription PCR (RT-PCR) test results of SARS-CoV-2 in oropharyngeal or nasopharyngeal swabs; tested positive by the IgM-IgG combined antibody test for SARS-CoV-2 1 week after disease onset, combined with epidemiology exposure and suspected symptoms; and agreed to and were compliant with testing and treatment protocols.

### Data and specimen collection

The collection and analysis of human clinical specimens was approved by the health and ethics boards of Tongji Hospital (approval TJ-IRB20200222). Written informed consent was provided by patients and the neonates' adult proxy.

Demographic information and clinical characteristics of the enrolled patients were extracted from the electronic medical system, supplemented with telephone follow-up. Within 7 days after giving birth, breast milk was self-pumped after hand sanitization, then placed in containers. Mother-child isolation was performed immediately after cord clamping. Neonates were either observed in neonatal intensive care units (NICU) or kept in a quarantine center as close contacts with family member's accompany. Neonates in the NICU were sampled by nasal or oropharyngeal swabbing for SARS-CoV-2 two times in a row, 24 hours apart. Neonatal faeces were sampled after meconium passed. All clinical specimens underwent RT-PCR assays for SARS-CoV-2 ORF1ab/N genes via detection kits [8], as authorized by the Chinese centre for disease control and

prevention. SARS-CoV-2 IgM-IgG combined antibody tests [9] and fully automated chemiluminescence immunoassay were performed of breast milk and of maternal and neonatal sera after the test's introduction on 26 February 2020. IgM or IgG titres of more than 10 arbitrary unit (AU)/mL were considered positive; other results were considered negative.

### Statistical analysis

Statistical analysis was performed by SPSS 20.0 software (IBM, Armonk, NY, USA). Continuous variables were presented as mean and standard deviations (SDs), or medians and interquartile ranges. Categorical variables were expressed as counts and percentages.

## Results

### Clinical characteristics and perinatal outcomes

Demographic information and clinical features of 14 mothers with confirmed COVID-19 are presented in Table 1, where data of four patients who were cotested with immunologic assays of SARS-CoV-2 are detailed.

In this cohort, mean maternal age was 31 years (SD, 2.4 years; range, 27–35 years). All had a singleton pregnancy; four (28.5%) were the mothers' first babies and ten were not. All patients were Wuhan residents, two (14.2%) were healthcare workers, three (21.4%) had contact with confirmed or suspected cases and two (14.2%) had family aggregation occurrence. The most common symptoms were fever (11/14, 78.1%) and cough (6/14, 42.9%). A pregnancy-specific symptom, abnormal foetal movement, occurred in three patients (21.4%), with one feeling evidently increased foetal movement and two decreased movement. All patients had abnormalities found by chest computed tomography (CT). Typical findings were ground-glass opacities, multiple patches in lung fields and subpleural adhesions. Laboratory results showed lymphopenia (lymphocytes  $<1.1 \times 10^{12}$ ) in seven patients (50%); five patients had abnormal liver functions, two of whom had coexisting intrahepatic cholestasis of pregnancy. We also performed immunoserologic testing of other common respiratory pathogens, including respiratory syncytial virus, adenovirus, influenza A, influenza B, parainfluenza virus, *Mycoplasma pneumoniae* and *Legionella pneumophila*. Four patients (26.1%) tested positive for influenza A IgM and one (4.3%) tested positive for *Mycoplasma pneumoniae* IgM.

Respiratory support was applied for 4 hours after surgery for those who underwent caesarean sections and for those whose blood oxygen saturation dropped below 93%. Eight patients (57.1%) received oxygen via nasal catheter; no respirator or mechanical ventilation was indicated.

**TABLE 1. Baseline information of patients with COVID-19 during pregnancy**

Characteristic	Patients with COVID-19 during pregnancy (n = 14)	Patients who developed SARS-CoV-2-neutralized IgM/IgG antibodies after delivery			
		Patient 1	Patient 2	Patient 3	Patient 4
Age (years), mean (SD, range)	31 (2.4, 27–35)	30	33	27	30
Mean GA at disease onset (weeks)	36 <sup>+3</sup>	32	37	41	32 <sup>+3</sup>
Mean GA at admission (weeks)	38	33 <sup>+3</sup>	39 <sup>+1</sup>	41 <sup>+2</sup>	39 <sup>+3</sup>
Epidemiology exposure					
Wuhan or Hubei travel history	14 (100)	Yes	Yes	Yes	Yes
Healthcare worker	2 (14.2)	Yes	No	No	Yes
Contact with confirmed or suspected case	3 (21.4)	Yes	No	No	Yes
Family aggregation occurrence	2 (14.2)	No	No	No	Yes
Gestational comorbidity					
ICP	2 (14.2)	No	No	No	No
Gestational diabetes	1 (7.1)	No	No	No	No
Thyroid dysfunction	3 (21.4)	No	No	Yes	No
Congenital heart disease	1 (7.1)	No	No	Yes	No
Chronic HBV infection	1 (7.1)	Yes	No	No	No
Tuberculosis	1 (7.1)	No	No	No	No
Signs and symptoms					
Fever	11 (78.6)	Yes	Yes	No	No
Fever during pregnancy	8 (57.1)	Yes	Yes	No	No
Postpartum fever	8 (57.1)	No	No	No	No
Malaise	2 (14.2)	No	No	No	No
Myalgia	2 (14.2)	Yes	No	No	No
Headache	1 (7.1)	No	No	No	No
Sore throat	2 (14.2)	No	No	No	Yes
Cough	6 (42.9)	Yes	No	No	Yes
Chest fullness	2 (14.2)	No	No	No	No
Chest pain	2 (14.2)	No	No	No	No
Diarrhoea	1 (7.1)	No	No	No	No
Vomiting	1 (7.1)	No	No	No	No
Anorexia	3 (21.4)	Yes	No	No	No
Abnormal foetal movement	3 (21.4)	No	No	No	No
Respiratory support	8 (57.1)	Yes	No	No	No
Laboratory results, reference value, mean (SD, range)					
Chest CT abnormalities	13 (92.9)	Yes	Yes	Yes	Yes
WBC, 3.50–9.50 × 10 <sup>9</sup> /L	8.6 (2.9, 5.0–16.4)	5.7	9.2	8.5	NA
Neutrophils, 1.80–6.30 × 10 <sup>9</sup> /L	6.9 (2.5, 3.0–13.59)	4.8	7.9	6.3	NA
Lymphocytes, 1.10–3.20 × 10 <sup>9</sup> /L	3.0 (0.4, 0.6–1.9)	0.7	0.8	1.7	NA
Platelets, 125–350 × 10 <sup>9</sup> /L	199 (64, 121–318)	214	313	236	NA
Haemoglobin, 115–50 g/L	123 (15.2, 100–55)	113	109	100	NA
CRP, <1 mg/L (n = 7)	27.5 (22.1, 1.1–0.5)	70.5	1.1	NA	NA
ESR, 0–20 mm/h (n = 3)	50 (24.6, 3–5)	95	NA	NA	NA
PCT, 0.02–0.05 ng/L (n = 6)	0.4 (0.7, 0.04–0.05)	0.32	0.06	NA	NA
ALT, <33 U/L	105 (226, 5–82)	18	7	5	NA
AST, <32 U/L	126 (220, 13–83)	38	13	14	NA
LDH, 135–14 U/L (n = 10)	283.9 (159.4, 141–86)	304	167	NA	NA
D-D, <0.5 µg/mL (n = 11)	2 (0.8, 0.6–3.2)	1.7	0.65	NA	NA
Coinfection	5 (35.7)	Yes <sup>a</sup>	No	No	No

Data are presented as n (%) unless otherwise indicated. AST, aspartate aminotransferase; COVID-19, coronavirus disease 2019; CRP, C-reactive protein; D-D, D-dimer; ESR, erythrocyte sedimentation rate; GA, gestational age; HBV, hepatitis B virus; ICP, intrahepatic cholestasis of pregnancy; LDH, lactate dehydrogenase; LT, alanine transaminase; NA, not applicable; PCT, procalcitonin; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2; SD, standard deviation; WBC, white blood cell.  
<sup>a</sup>Coinfected with influenza A.

All patients underwent successful term delivery with no severe complications or admission to the intensive care unit. Perinatal outcomes and neonate baseline information are shown in Table 2. The mean interval from onset of disease to delivery was 5.4 days (SD, 6.3 days; range, 1–46 days). Two patients (14.2%) had foetal distress, which was marked by variable decelerations observed on foetal heart tracing and

third-degree amniotic foetal meconium pollution. Twelve women (85.8%) chose to deliver via caesarean section; two patients (14.2%) gave birth vaginally without mechanical assistance. The surgeries were performed in an isolated surgical suite with continuous lumbar epidural analgesia. The mean birth weight of neonates was 3224 g (SD, 421 g; range, 2700–4120 g). One neonate, born to a mother with complications related

**TABLE 2.** Perinatal outcomes of patients with COVID-19 during pregnancy

Outcome	Patients with COVID-19 during pregnancy (n = 14)	Patients who developed SARS-CoV-2 neutralized IgM/IgG antibodies after delivery			
		Patient 1	Patient 2	Patient 3	Patient 4
GA of delivery (weeks), mean (range)	38 + 3	39	39 + 1	41 + 2	39 + 5
Time from disease onset to delivery (days), mean (SD, range)	5.4 (6.3, 1–21)	46	15	2	43
PROM	2 (14.2)	No	No	No	No
Foetal distress	2 (14.2)	No	No	No	No
Caesarean section	12 (85.7)	Yes	Yes	Yes	No
Amniotic fluid pollution	6 (42.8)	No	No	No	No
Live birth	14 (100)	Yes	Yes	Yes	Yes
Hospital stay (days), mean (SD, range)	19 (7.9, 9–9)	39	16	10	9
Neonate baseline characters					
Birth weight (g), mean (SD, range)	3224 (421, 2700–120)	2700	3930	4120	2900
NICU admission	7 (50)	Yes	Yes	Yes	No
Neonatal asphyxia	1 (7.1)	No	No	No	No
Neonatal jaundice	5 (35.7)	Yes	Yes	No	No
Fever	1 (7.1)	Yes	No	No	No
Anaemia	1 (7.1)	No	No	No	No

Data are presented as n (%) unless otherwise indicated. COVID-19, coronavirus disease 2019; GA, gestational age; NICU, neonatal intensive care unit; PROM, premature rupture of membrane; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2; SD, standard deviation.

to intrahepatic cholestasis of pregnancy, had mild asphyxia at birth. One neonate had transient fever (anal temperature 37.9°C) at 30 hours after birth.

Maternal and neonatal RT-PCR SARS-CoV-2 detection results are shown in Table 3. Samples were taken by oropharyngeal or nasopharyngeal swabbing every 3 days during hospitalization. SARS-CoV-2 virus shedding days were described as the first positive RT-PCR result to the first continuously negative RT-PCR result. The mean maternal virus shedding time was 9 days (SD, 6 days; range, 1–22 days). SARS-CoV-2 nucleic acids were not detected in maternal breast milk (n = 12), vaginal secretions (n = 10), neonatal oropharyngeal swabs (n = 12) or meconium specimens (n = 6).

#### SARS-CoV-2 immunoassay in breast milk and in maternal and neonatal sera

Immunologic features and the disease courses of four mother–infant pairs are presented in Fig. 1. Maternal

seroconversion of IgM was observed on day 8 after disease onset, and IgG on day 28. Three breast milk samples tested positive for SARS-CoV-2 IgM or IgG. Three neonates tested positive for SARS-CoV-2 IgG. One neonate tested positive for IgM within 24 hours of birth. IgG was detected in one cord blood sample.

Patient 1, a healthcare worker, had fever and cough on 24 January at 32 weeks of pregnancy. She tested positive from a oropharyngeal swab sample on 1 February. Chest CT revealed bilateral pneumonia (Fig. 2). She was admitted and received oxygen and other supportive treatment. She had two consecutive negative results on 12 and 14 February, but she still complained of persistent dry cough and malaise. On 3 March, at 37 + 5 weeks of pregnancy, her SARS-CoV-2 antibody assay showed evidently increased antibodies, with IgG 280.96 AU/mL and IgM 2581.57 AU/mL, while the nasopharyngeal swab test results were negative. An elective caesarean section was planned to avoid maternal exhaustion. A boy was born, with an Apgar score of 1 minute after birth is 8 and 9 for 5 minutes; birth weight was 2700 g. At testing within 24 hours of birth, the neonatal SARS-CoV-2 immunologic assays showed positive titres (IgG 147.21 AU/mL, IgM 184.3 AU/mL). At 72 hours after birth, IgG titre was 95.48 AU/mL and IgM 188.33 AU/mL. SARS-CoV-2 nucleic acids were not detected in neonatal oropharyngeal swab or meconium samples. Breast milk was examined on postpartum day 6; IgG titre was 145.31 AU/mL and IgM 92.01 AU/mL. The immunologic components (IgA, IgM, IgG, C3, C4) of breast milk were within normal limit. During isolation, the neonate was fed formula only. The mother resumed breastfeeding when discharged with bottle feeding of expressed breast milk.

Patient 2 was asymptomatic when admitted on 29 February at 39 weeks of pregnancy. Her nasopharyngeal swab results at admission were negative. She reported fever and cough during 36 weeks of pregnancy and was advised to quarantine at home, where the symptoms self-resolved. She underwent an elective caesarean section as a result of a scarred uterus. The surgery was uncomplicated; a 3930 g boy was born. Antibody testing of serum and breast milk was performed on postpartum day 3; results indicated markedly increased IgG titres and was negative for IgM. Breast milk level of IgG was 103.15 AU/mL (an immunologic component in the normal range). The neonate was tested within 24 hours of birth; increased IgG (78.41 AU/mL) and negative IgM were noted. Patient 2 continued breastfeeding with expressed breast milk. The neonate was well appearing, with normal weight gain according to the father during follow-up.

Patient 3 was admitted on 20 February with low-grade fever (temperature 37.3°C) at 41 weeks of pregnancy. Outpatient chest CT scan obtained 2 days before revealed bilateral ground-

**TABLE 3. Detection of SARS-CoV-2 nucleic acids in confirmed mother-infant pairs in early 2020**

Characteristic	Pair no.													
	1	2	3	4	5	6	7	8	9	10	11	12	13	14
Delivery date	19 Jan	22 Jan	24 Jan	30 Jan	31 Jan	31 Jan	3 Feb	5 Feb	9 Feb	9 Feb	20 Feb	29 Feb	5 Mar	16 Mar
Gestational age at delivery (weeks)	36 <sup>+4</sup>	38 <sup>+4</sup>	38 <sup>+1</sup>	36 <sup>+5</sup>	37	38 <sup>+2</sup>	40 <sup>+5</sup>	38 <sup>+4</sup>	39 <sup>+1</sup>	39	41 <sup>+1</sup>	39	38	39 <sup>+5</sup>
Caesarean section	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	No	Yes	Yes	Yes	Yes	No
Maternal SARS-CoV-2 nucleic acid detection														
Positive throat swab date	31 Jan	5 Feb	28 Jan	8 Feb	31 Jan	31 Jan	6 Feb	5 Feb	10 Feb	9 Feb	NA	NA	2 Feb	4 Feb
Negative throat swab date 1	18 Feb	17 Feb	31 Jan	9 Feb	8 Feb	22 Feb	9 Feb	13 Feb	21 Feb	12 Feb	17 Feb	3 Feb	12 Feb	12 Feb
Negative throat swab date 2	22 Feb	19 Feb	NA	11 Feb	13 Feb	24 Feb	NA	16 Feb	24 Feb	14 Feb	19 Feb	1 Mar	4 Mar	14 Feb
SARS-CoV-2 shedding time (days)	18	12	3	1	8	22	3	8	11	3	NA	NA	10	8
Breast milk (n = 12)	NA	NA	—	—	—	—	—	—	—	—	—	—	—	—
Vaginal secretion (n = 10)	NA	NA	NA	—	NA	—	—	—	—	—	—	—	—	—
Anal swab (n = 12)	NA	—	NA	—	—	—	—	—	—	—	—	—	—	—
Neonatal SARS-CoV-2 nucleic acids detection														
Throat swab (n = 12)	—	—	NA	—	NA	—	—	—	—	—	—	—	—	—
Meconium (n = 6)	NA	NA	NA	NA	NA	NA	NA	—	—	—	—	—	—	NA

Maternal negative throat swab dates were presented as two continuous dates separated by at least 48 hours. SARS-CoV-2 shedding time was counted as duration from first positive date to first consecutive negative date. Disease of mothers 11 and 12 was confirmed by epidemiologic exposure, characteristic symptoms and positive SARS-CoV-2 combined IgM/IgG tests; their throat swab results remained negative two separate times. Dash indicates negative results. COVID-19, coronavirus disease 2019; NA, not available; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

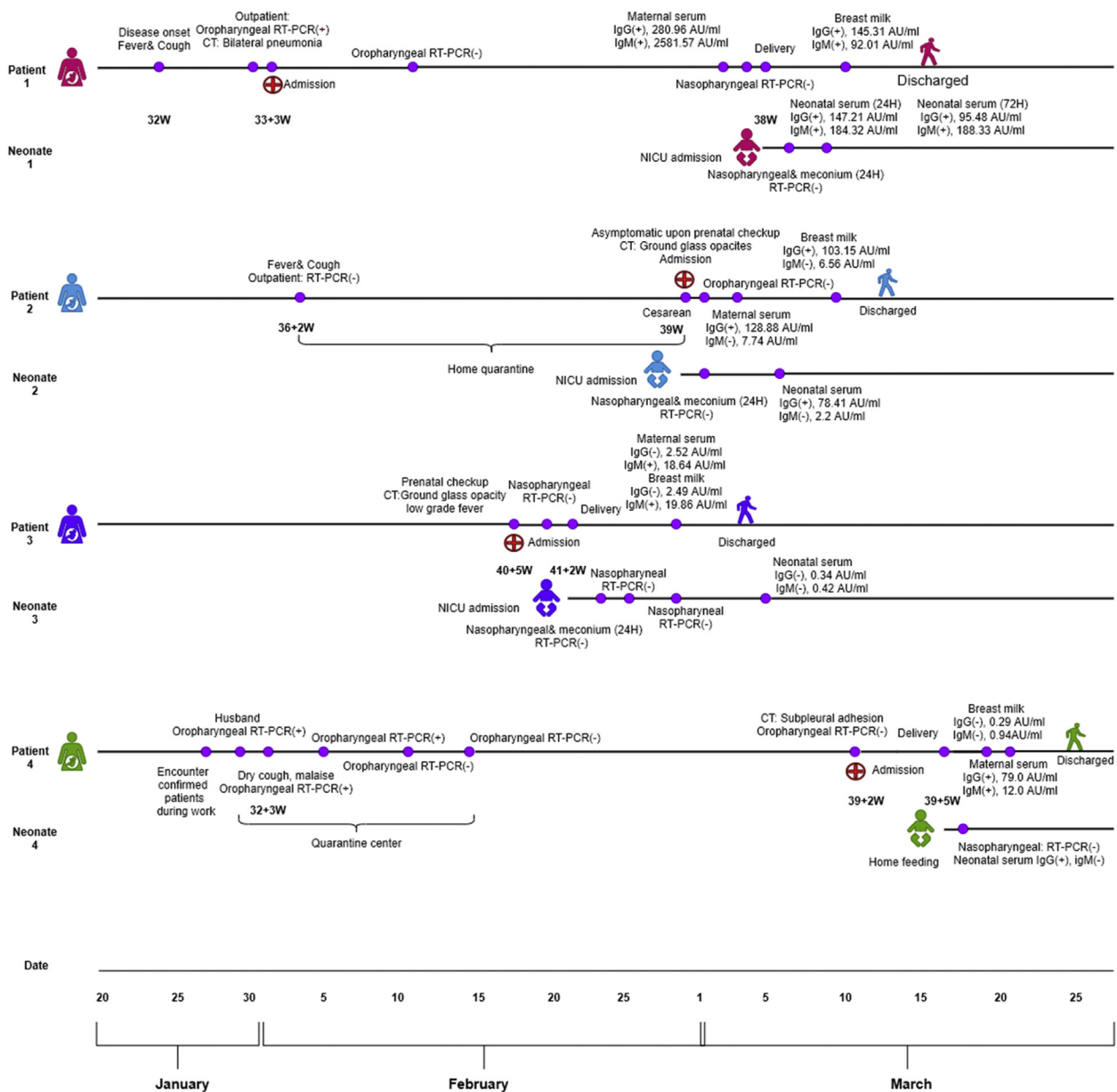
glass opacities. Elective caesarean section was performed, and the 4120 g female neonate was transferred to the NICU for observation. Breast milk and maternal serum were tested for antibodies on postpartum day 6, showing increased IgM antibody titres and negative IgG. Breast milk IgM level was 19.86 AU/mL, and maternal serum IgM was 18.64 AU/mL. Neonatal serum antibody test at 14-day checkup was negative for both IgM and IgG. The neonate was fed formula in the NICU and breastfed after isolation finished. During follow-up, the mother reported bottle feeding her baby with breast milk on most occasions. She was advised to wear a mask during direct breastfeeding.

Patient 4 was a nurse in outpatient triage centre. She encountered patients with confirmed COVID-19 on 27 January; her husband was also confirmed to have COVID-19. She had dry cough and malaise on 2 February, at 32 + 3 weeks of pregnancy, and subsequently tested positive. She was admitted to the quarantine centre for observation with no further treatment. Her symptoms spontaneously resolved 1 week later. Testing of oropharyngeal swabs on 12 and 14 February had negative results, and the patient was discharged. At a prenatal checkup on 11 March, her nasopharyngeal swab test results

remained negative; CT revealed clear lung fields with subpleural adhesions. Testing results for SARS-CoV-2 antibodies were positive for IgG and negative for IgM. She was considered cured and admitted to the regular obstetrics ward. On 16 March, at 39 + 5 weeks of pregnancy, she vaginally delivered a baby girl (Apgar score 8–9, weight 2900 g). The cord blood tested positive for SARS-CoV-2 IgG and negative for IgM. Breast milk was tested on postpartum day 3, with negative SARS-CoV-2 antibody components; maternal serum samples taken the same day had IgG 79.0 AU/mL and IgM 12.0 AU/mL. She did not isolate with her baby and breastfed directly without a mask. At birth, the neonate tested positive for SARS-CoV-2 IgG and negative for IgM, and negative nasopharyngeal swab test results were found on the day 14 checkup.

### Discussion

The study enrolled 14 patients who delivered during their hospitalization with COVID-19 from 32 weeks of pregnancy to postpartum day 2 in Wuhan, China. All of them experienced good recovery, with no reported COVID-19-related sequelae.



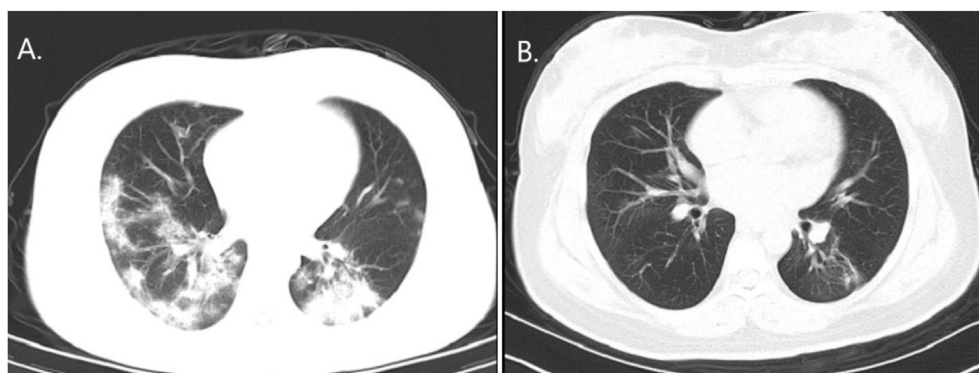
**FIG. 1.** Timeline and immunologic findings in breast milk and sera among four mother–infant pairs.

No infection of the neonate was noticed. SARS-CoV-2 nucleic acids were not detected in 12 breast milk samples of mothers with COVID-19 in various stages of disease. In contrast, neutralized SARS-CoV-2 antibodies were identified in three breast milk samples. Among the symptomatic breastfeeding mothers, extreme precautions were taken to avoid postnatal infection. For recovered or cured mothers, direct breastfeeding did not result in neonatal infection.

Breastfeeding has been proven to have many benefits to both mother and child [10]. With the ongoing COVID-19 pandemic, the general population is overall vulnerable to the novel coronavirus. Women in pregnant and puerperium states are

believed to be particularly susceptible to infection, as they are in other coronavirus diseases, such as SARS-CoV and Middle East respiratory syndrome coronavirus [11,12], in which poorer clinical outcomes and higher morbidity and mortality were reported. So far, COVID-19 during pregnancy has been reported to be mostly mild in presentation and has a good prognosis [13], but neonatal infection was reported during the Wuhan epidemic [14], thus focusing discussion on postnatal management and breastfeeding advice.

Known pathogens that can be transmitted during breastfeeding include HIV-1, human T-lymphotropic virus I and cytomegalovirus; the pathogen can be isolated from breast



**FIG. 2.** Chest computed tomographic (CT) image of pregnant patients diagnosed with coronavirus disease 2019 (COVID-19). (A) CT image of patient 1 on 6 February 2020 (day 13 after disease onset) showing bilateral lung multiple patches and ground-glass opacities with ragged edges. (B) CT image of patient 1 on 9 March 2020 (postpartum day 4, or 45 days after disease onset) showing evident absorption and improvement, with partial stranding in left lung subpleural area.

milk and causes corresponding illness in neonates. On other occasions, breast lesions can serve as an infection source while the neonate sucks; such lesions may be caused by tuberculosis or varicella zoster virus [15]. In our findings, SARS-CoV-2 was not detected in breast milk, and the mothers who continued breastfeeding had no breast lesions. Direct breastfeeding does carry certain risks; for example, maternal respiratory droplets are considered contagious, especially during the acute symptomatic stage, and they have an exceedingly high virus load [16]. SARS-CoV-2's viraemia provides an opportunity for breast lesions to transmit the virus to the neonate during sucking. Neonates also have weaker immune systems and inadequate protection against pathogens, which makes them vulnerable to the overall environment during breastfeeding. However, those risks are avoidable if certain measures are taken. Wearing a surgical mask during breastfeeding, handwashing before and after pumping milk and thoroughly sanitizing milk-expressing devices can help minimize the risk of virus transmission.

The effect of passive acquisition of antibody to SARS-CoV-2 from breast milk and maternal serum remains to be addressed. Zeng et al. [17] described neonatal acquisition of both IgM and IgG antibodies from a COVID-19-affected mother. In our findings, antibodies were also identified in cord blood and breast milk samples, indicating passive acquisition via the placenta and through breast milk. A prospective controlled study discussing the effect of anticholera antibodies in breast milk [18] showed that breast milk ingestion did not stop the colonization of *Vibrio cholerae*. However, colonized infants had a milder disease presentation and a shorter disease course. For *Enterovirus* infections, one study reported that breast milk antibodies had specific protective effects [19]. During combat with COVID-19, convalescent plasma (CP) therapy has shown

promising effects. Chen et al. [20] described five critically ill patients who experienced remission after CP therapy, concluding that CP contributed to disease remission and virus clearance. However, concerns regarding transfusion-related allergy and possible lethal hyperimmunity attacks of CP must be addressed. Breast milk from recovered mothers possesses similar neutralization antibody components, is nonallergenic, is easy to acquire and is delivered orally. In addition, breast milk contains a wide variety of cell-rich components, including macrophages, immunoglobulin, complement bodies and cytokines, which have wide-spectrum antiviral effects.

On the basis of current information, we hypothesize that breastfeeding's role in transmission is low, and passive acquisition of antibodies to SARS-CoV-2 is beneficial to the infants. However, because there are other possible infection routes during mother–infant contact, prudent precautions should be taken.

Further investigation should focus on creating regulations for COVID-19 mother–infant contact that both prevent neonatal infection and permit emotional bonding. The SARS-CoV-2 mother–infant immunologic trends require long-term follow-up regarding antibody classes, duration and titre changes. Moreover, prospective studies of infants with or without passive acquisition of antibodies to SARS-CoV-2 should be conducted to evaluate the immunity effects.

## Conclusions

Breastfeeding has a low risk of transmitting SARS-CoV-2 or escalating maternal disease; mothers should continue to breastfeed but should take prudent precautions. Infants can additionally benefit from direct acquisition of antibodies against SARS-CoV-2 through breast milk.

## Conflict of interest

None declared.

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## References

- [1] Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel coronavirus from patients with pneumonia in China, 2019. *N Engl J Med* 2020;382:727–33.
- [2] Zou L, Ruan F, Huang M, Liang L, Huang H, Hong Z, et al. SARS-CoV-2 viral load in upper respiratory specimens of infected patients. *N Engl J Med* 2020;382:1177–9.
- [3] Zhang JC, Wang SB, Xue YD. Fecal specimen diagnosis 2019 novel coronavirus–infected pneumonia. *J Med Virol* 2020;92:680–2.
- [4] Young BE, Ong SWX, Kalimuddin S, Low JG, Tan SY, Loh J, et al. Epidemiologic features and clinical course of patients infected with SARS-CoV-2 in Singapore. *JAMA* 2020;323:1488–94.
- [5] Lawrence RM, Lawrence RA. Breast milk and infection. *Clin Perinatol* 2004;31:501–28.
- [6] Chen H, Guo J, Wang C, Luo F, Yu X, Zhang W, et al. Clinical characteristics and intrauterine vertical transmission potential of COVID-19 infection in nine pregnant women: a retrospective review of medical records. *Lancet* 2020;395(10226):809–15.
- [7] Guan W, Ni Z, Hu Y, Liang W, Ou C, He J, et al. Clinical characteristics of coronavirus disease 2019 in China. *N Engl J Med* 2020;382:1708–20.
- [8] Chinese Center for Disease Control and Prevention; National Institute for Viral Disease Control and Prevention. Specific primers and probes for detection 2019 novel coronavirus. [http://ivdc.chinacdc.cn/kyjz/202001/t20200121\\_211337.html](http://ivdc.chinacdc.cn/kyjz/202001/t20200121_211337.html).
- [9] Xu WZ, Li J, He XY, Zhang CQ, Mei SQ, Li CR, et al. Application of chemiluminescence immunoassay in SARS-CoV-2 IgM-IgG tests. *Chin Lab Med* 2020. <https://doi.org/10.3760/cma.j.cn114452-20200223-00109>.
- [10] Brock EG, Long L. Breast feeding. *Obst Gynaecol Reprod Med*, Churchill Livingstone. 2019 May 1. <https://doi.org/10.1016/j.ogrm.2019.02.003>.
- [11] Wong SF, Chow KM, Leung TN, Ng WF, Ng TK, Shek CC, et al. Pregnancy and perinatal outcomes of women with severe acute respiratory syndrome. *Am J Obstet Gynecol* 2004;191:292–7.
- [12] Assiri A, Abedi GR, Al Masri M, Bin Saeed A, Gerber SI, Watson JT. Middle East respiratory syndrome coronavirus infection during pregnancy: a report of 5 cases from Saudi Arabia. *Clin Infect Dis* 2016;63:951–3.
- [13] Zhang L, Jiang Y, Wei M, Cheng BH, Zhou XC, Li J, et al. Analysis of pregnancy outcomes in pregnant women with COVID-19 in Hubei province. *Zhonghua Fu Chan Ke Za Zhi* 2020;55:166–71.
- [14] Wang S, Guo L, Chen L, Liu W, Cao Y, Zhang J, et al. A case report of neonatal COVID-19 infection in China. *Clin Infect Dis* 2020;71:853–7.
- [15] Lanari M, Sogno Valin P, Natale F, Capretti MG, Serra L. Human milk, a concrete risk for infection. *J Matern Neonatal Med* 2012;25(Suppl. 4):67–9.
- [16] Woelfel R, Corman VM, Guggemos W, Seilmaier M, Zange S, Mueller MA, et al. Clinical presentation and virological assessment of hospitalized cases of coronavirus disease 2019 in a travel-associated transmission cluster. *medRxiv*. 8 March 2020. <https://doi.org/10.1101/2020.03.05.20030502>.
- [17] Zeng H, Xu C, Fan J, Tang Y, Deng Q, Zhang W, et al. Antibodies in infants born to mothers with COVID-19 pneumonia. *JAMA* 2020;323:1848–9.
- [18] Glass RI, Svennerholm AM, Stoll BJ, Khan MR, Hossain KMB, Hug MI, et al. Protection against cholera in breast-fed children by antibodies in breast milk. *N Engl J Med* 1983;308:1389–92.
- [19] Sadeharju K, Knip M, Virtanen SM, Savilahti E, Tauriainen S, Koskela P, et al. Maternal antibodies in breast milk protect the child from enterovirus infections. *Pediatrics* 2007;119:941–6.
- [20] Chen L, Xiong J, Bao L, Shi Y. Convalescent plasma as a potential therapy for COVID-19. *Lancet Infect Dis* 2020;20:398–400.