# A Patient with Asymptomatic Severe Acute Respiratory Syndrome (SARS) and Antigenemia from the 2003–2004 Community Outbreak of SARS in Guangzhou, China

## Xiao-yan Che,<sup>1</sup> Biao Di,<sup>2</sup> Guo-ping Zhao,<sup>3</sup> Ya-di Wang,<sup>1</sup> Li-wen Qiu,<sup>1</sup> Wei Hao,<sup>1</sup> Ming Wang,<sup>2</sup> Peng-zhe Qin,<sup>2</sup> Yu-fei Liu,<sup>2</sup> Kwok-hong Chan,<sup>4</sup> Vincent C. C. Cheng,<sup>4</sup> and Kwok-yung Yuen<sup>4</sup>

<sup>1</sup>Center of Laboratory, Zhujiang Hospital, Southern Medical University, and <sup>2</sup>Center for Disease Control and Prevention of Guangzhou, Guangzhou, <sup>3</sup>Laboratory of Disease and Health Genomics, Chinese National Human Genome Center at Shanghai, Shanghai, and <sup>4</sup>Department of Microbiology, The University of Hong Kong, Hong Kong Special Administrative Region, People's Republic of China

An asymptomatic case of severe acute respiratory syndrome (SARS) occurred early in 2004, during a community outbreak of SARS in Guangzhou, China. This was the first time that a case of asymptomatic SARS was noted in an individual with antigenemia and seroconversion. The asymptomatic case patient and the second index case patient with SARS in the 2003–2004 outbreak both worked in the same restaurant, where they served palm civets, which were found to carry SARS-associated coronaviruses. Epidemiological information and laboratory findings suggested that the findings for the patient with asymptomatic infection, together with the findings from previously reported serological analyses of handlers of wild animals and the 4 index case patients from the 2004 community outbreak, reflected a likely intermediate phase of animal-to-human transmission of infection, rather than a case of human-to-human transmission. This intermediate phase may be a critical stage for virus evolution and disease prevention.

After the announcement of the end of the severe acute respiratory syndrome (SARS) epidemic on 5 July 2003, 4 new community-acquired cases of SARS were reported between 16 December 2003 and 8 January 2004, in Guangzhou City in the province of Guangdong, China [1]. The genomic sequences of SARS-associated coronavirus (SARS-CoV) isolates recovered from these 4 index case patients with SARS and from palm civets that were associated with this SARS outbreak were nearly identical. The results of phylogenetic analysis, combined with epidemiological lineage information,

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suggested that SARS-CoV was reintroduced to humans from animals [2, 3]. Although it was reported that none of these patients were involved in the secondary spread of symptomatic SARS-CoV infection [1], we describe a patient with asymptomatic SARS-CoV infection who was identified from 115 contacts of the index case patients in this community outbreak. This asymptomatic case of SARS-CoV infection was confirmed by a sensitive method of viral antigen detection and by serological tests. The asymptomatic case also highlights some issues associated with disease prevention.

### PATIENTS AND METHODS

The present study was performed retrospectively. Seventeen serum specimens were collected from 4 index case patients who exhibited recurrence of SARS with laboratory-confirmed SARS-CoV infection in Guangzhou City, China, from 22 December 2003 through 30 January 2004. An additional 118 serum specimens were collected from 115 contacts of the 4 index case patients with SARS.

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Reprints or correspondence: Dr. Xiao-yan Che, Center of Laboratory, Zhujiang Hospital, Southern Medical University, Guangzhou 510282, People's Republic of China (linche@pub.guangzhou.gd.cn); or Dr. Guo-ping Zhao, Laboratory of Disease and Health Genomics, Chinese National Human Genome Center at Shanghai, Shanghai 201203, People's Republic of China (gpzhao@sibs.ac.cn).

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To identify the nucleocapsid (N) protein of SARS-CoV in serum specimens, 2 antigen-capture assays were performed: an N antigen-capture ELISA and an N antigen-capture chemiluminescent immunoassay (CIA). The development of these 2 assays was based on the detection of 3 monoclonal antibodies directed against the N protein of SARS-CoV, as described elsewhere [4-6]. Serum IgG and IgM antibodies to SARS-CoV were detected using commercially available indirect ELISA kits (BGI-GBI; Biotech) according to the manufacturer's instructions [6]. The sensitivity and specificity of ELISA for the detection of IgG antibodies to SARS-CoV were 99%-100% and 89%-97.2%, respectively, and the sensitivity and specificity of ELISA for the detection of IgM antibodies to SARS-CoV were 89.8% and 97.6%, respectively [6-8]. An indirect immunofluorescent assay (IFA) for the detection of IgG antibodies to SARS-CoV was performed using a commercially available IFA kit (Euroimmun), according to the manufacturer's instructions [9]. This IFA has a specificity of 100% (with detection of IgG antibodies to SARS-CoV in 200 healthy blood donors) and a sensitivity of 97%-100% (with detection of IgG antibodies to SARS-CoV in 150 serum specimens obtained, at least 10 days after the onset of symptoms, from patients with SARS). In-house IFAs for the detection of IgG specific to human coronaviruses 229E and OC43 were performed as described by us elsewhere [10]. A microneutralization assay was performed according to procedures described elsewhere, with modifications [1]. In brief, a 96-well microtiter plate that contained confluent fetal rhesus kidney-4 cells in 100 µL of maintenance medium was prepared. Two-fold dilutions of patient serum samples (50  $\mu$ L), which started at 1:10 and increased to 1:320, were premixed with 50  $\mu$ L TCID<sub>50</sub> of SARS-CoV (the HKU-39849 isolate) and were incubated at 37°C for 1.5 h. Then, 100  $\mu$ L of the virus-serum mixture was inoculated in duplicate wells with fetal rhesus kidney-4 cells and was further incubated at 37°C. A cytopathic effect was observed at 72 and 96 h. The neutralization titer was determined to be the reciprocal of the highest serum dilution that produced 50% cytopathic effect on cells. A neutralizing antibody titer of ≥10 was considered to be positive for SARS-CoV.

## **RESULTS AND DISCUSSION**

All serum specimens were tested for the N protein of SARS-CoV by use of both N antigen–capture ELISA and N antigen– capture CIA. The findings of these 2 assays had been validated previously with the use of serum specimens obtained from patients with serologically confirmed SARS, and the sensitivity and specificity of the N antigen–capture ELISA were documented [4–6]. Although the characteristic of the N antigen– capture CIA was shown to be equivalent to that of the N antigen–capture ELISA (data not shown), the N antigen–capture CIA seemed to be more sensitive than N antigen–capture ELISA in the present study. Both N antigen–capture assays detected N protein in serially obtained serum specimens that were collected from 3 patients 6–9 days after the onset of symptoms; CIA was also able to detect N protein in serum samples collected from the second index case patient on the ninth and 10th days after the onset of symptoms, although these samples had negative results according to the N antigen–capture ELISA (table 1). This result was not surprising, because enzyme-amplified chemiluminescent chemical analysis is generally more sensitive than conventional ELISA [11]. For index case patient 4, the first serum sample was obtained on day 17 after the onset of symptoms, and N protein was not detectable in serially obtained serum specimens during this patient's illness.

Of the 118 serum specimens obtained from the 115 contacts of the index case patients, 1 specimen, which was collected on 6 January 2004, was from a female contact of the second index case patient, who experienced onset of symptoms on 26 December 2003. This female contact was found to be positive for the N protein by means of N antigen-capture CIA, and the presence of SARS-CoV infection was further confirmed by serological tests (table 2). IFA detected a 16-fold increase in the SARS-CoV antibody titer in paired serum samples collected over a 1-week period. IgM and IgG antibodies to SARS-CoV were also detected by ELISA. Antibodies to SARS-CoV were also detected in the convalescent-phase serum sample obtained from this female contact case patient 12 weeks later, and the results of the SARS-CoV microneutralization assay were also found to be positive, with a titer of 20 determined for the convalescent-phase serum sample obtained from this female contact case patient. The serological results were verified by the Center for Disease Control and Prevention of Guangzhou, which is one of the SARS reference laboratories in China.

To exclude the possibility of a false-positive result, serum specimens from the contact case patient were tested for antibodies against the human coronaviruses 229E and OC43, by use of IFA. Paired serum samples showed no increase in the titer of antibodies to either 229E or OC43. These tests were also performed for serially obtained serum specimens from the 4 index case patients (table 1). Although the paired serum samples obtained from the first and second index case patients did show an increase in the titers of antibodies to 229E and OC43, no neutralizing antibodies against 229E and OC43 were detected in these index case patients during a previous study [1].

Although none of the 4 index case patients showed evidence of secondary spread of the infection [1], the direct detection of SARS-CoV N protein by the highly sensitive CIA, complemented by the results of serologic tests, clearly demonstrated that a fifth case of SARS-CoV infection occurred, although the 4 index cases occurred during the 2003–2004 community outbreak of SARS in China. On the other hand, because the contact case patient did not show any signs or symptoms of respiratory

			Antibodies to SARS-CoV detected						
Patient (SARS onset date), day after onset of symptoms	SARS-CoV N protein detected		By IFA,	By ELISA, OD <sub>450</sub> (ratio) <sup>b</sup>		IgG antibody titer <sup>a</sup> determined by IFA			
	By CIA	By ELISA	lgG titer <sup>a</sup>	lgM	lgG	For 229E	For OC43		
1 (16 Dec 2003)									
6	+	+	160	0.724 (1)	0.355 (1)	40	320		
8	-	-	ND	1.416 (1.9)	0.843 (2.4)	80	ND		
10	-	-	ND	1.193 (1.6)	0.924 (2.6)	ND	ND		
12	_	-	640	1.193 (1.6)	0.873 (2.5)	160	2560		
2 (26 Dec 2003)									
6	+	+	<10	0.142 (1)	0.129 (1)	40	320		
7	+	+	ND	0.050 (0.4)	0.031 (0.2)	ND	ND		
8	+	+	320	0.513 (3.6)	0.584 (4.5)	ND	ND		
9	+	-	ND	0.675 (4.8)	0.727 (5.6)	80	640		
10	+	-	ND	0.282 (2.0)	0.526 (4.1)	ND	ND		
11	_	-	ND	0.721 (5.1)	0.568 (4.4)	ND	ND		
12	_	-	1280	0.462 (3.3)	0.785 (6.1)	320	1280		
3 (30 Dec 2003)									
9	+	+	640	0.387 (1)	0.665 (1)	160	320		
10	_	-	ND	0.162 (0.4)	ND	160	320		
11	_	-	640	0.355 (0.9)	0.795 (1.2)	160	320		
4 (8 Jan 2004)									
16	-	-	640	1.433 (1)	0.593 (1)	80	320		
17	-	-	ND	1.135 (0.8)	0.539 (0.9)	ND	ND		
22	_	_	640	1.219 (0.9)	0.618 (1.0)	80	320		

Table 1. Results of detection of severe acute respiratory syndrome (SARS)–associated coronavirus (SARS-CoV) nucleocapsid (N) antigen, antibodies to SARS-CoV, and antibodies to the human coronaviruses 229E and OC43 in the 4 index case patients with SARS from the 2003–2004 outbreak in Guangzhou City, China.

**NOTE.** CIA, chemiluminescent immunoassay; IFA, indirect immunofluorescent assay; ND, not done; OD, optical density measured at 450 nm; +, positive result; -, negative result.

<sup>a</sup> Serum samples in a serial 2-fold dilution (from 10-fold to 5120-fold).

<sup>b</sup> The highest ratio of acute-phase to convalescent-phase absorbance values at the same serum dilution tested.

illness, her case is the first confirmed and documented asymptomatic case of SARS in this outbreak.

This asymptomatic case of infection occurred in a 31-year-old woman who was working as a service chief in the local restaurant TDLR [2], where the second index case patient worked as a waitress [2, 3]. Although the asymptomatic case patient had only indirect contact with the second index case patient, she had a history of clear and close animal (i.e., palm civet) contact. She often showed the wild animals to customers who were ordering exotic food. Subsequently, SARS-CoV N protein was detected, by use of N antigen-capture ELISA, in 5 of a total of 6 palm civet serum samples obtained during the same period from the restaurant TDLR; results revealed a high level of N protein, with a mean  $OD_{450}$  value of 1.556, compared with the cutoff value of 0.21. Viral RNA was also detected in all throat and rectal swab specimens obtained from 6 palm civets, by use of RT-PCR for the detection of polyprotein and N genes of SARS-CoV, and 3 full genome sequences and 2 complete S gene sequences of SARS-CoV were identified in the specimens obtained from 5 of 6 palm civets [3]. When this information is considered together with the

viral genomic information from the palm civets that was collected from the same restaurant during the same period [2–3], the clear conclusion is that the palm civets used for exotic food in the restaurant must have been heavily contaminated with SARS-CoV.

With the limited samples available, the kinematics of the antigenemia and antibody responses of the second index case patient were compared with those of the asymptomatic case patient. The asymptomatic case patient had antigenemia (i.e., detection of N antigen in serum; antigenemia was detected in the asymptomatic case patient on 6 January 2004) disappear at almost the same time as it disappeared in the second index case patient (who had detectable antigenemia up to 5 January 2004). Meanwhile, the second index case patient had seroconversion occur on 3 January 2004, whereas the asymptomatic case patient had a positive antibody result from 6 January 2004; however, because serum samples were unavailable before this date, the exact date of seroconversion cannot be determined. Nevertheless, because the mean incubation period for SARS is 6 days [12], it seems that the asymptomatic case patient was unlikely to have acquired infection directly from the second

Table 2. Results of detection of severe acute respiratory syndrome (SARS)-associated coronavirus (SARS-CoV) nucleocapsid (N) antigen, antibodies to SARS-CoV, and antibodies to human coronaviruses 229E and OC43 in a patient with an asymptomatic case of SARS in the 2003–2004 community outbreak of SARS in Guangzhou City, China.

			Anti	bodies to				
Date of serum sample collection	SARS-CoV N protein detected		By IFA,	By NA,	By ELISA, OD <sub>450</sub> (ratio) <sup>b</sup>		lgG antibody titer <sup>a</sup> detected by IFA	
	By CIA	By ELISA	lgG titer <sup>a</sup>	titer	lgM	lgG	For 229E	For OC43
6 Jan 2004	+	_	80	ND	0.031 (1)	0.74 (1)	160	320
9 Jan 2004	_	_	640	ND	0.38 (12)	0.773 (1.0)	160	320
13 Jan 2004	_	-	1280	ND	1.451 (47)	0.801 (1.1)	160	320
24 Mar 2004	-	-	640	20	0.044 (1.3)	1.08 (1.5)	160	320

**NOTE.** CIA, chemiluminescent immunoassay; IFA, indirect immunofluorescent assay; NA, neutralization assay; ND, not done; OD<sub>450</sub>, optical density measured at 450 nm; +, positive result; -, negative result.

<sup>a</sup> Serum samples in a serial 2-fold dilution (from 10-fold to 5120-fold).

<sup>b</sup> The highest ratio of acute-phase to convalescent-phase absorbance values at the same serum dilution tested.

index case patient. Combining the epidemiological information regarding the contact history with the molecular diagnostic profiling of either the animals (the palm civets) or the human (the second index case patient), we infer that this asymptomatic case of SARS was more likely to have resulted from an animalto-human infection (likely due to SARS-CoV carried by the palm civets from the restaurant) than from the second index case patient.

SARS is a zoonotic disease. SARS-CoV evolved consistently and rapidly within its animal and human hosts, while both the infectivity of the virus and the severity of the disease varied, along with the variation/adaptation of the virus to its hosts [2, 13]. It is particularly significant that even in the "early phase" of the 2002-2003 SARS epidemic [13], human-to-human transmission of infection was observed, and the symptoms were so severe that the disease was eventually named after its primary symptomatic characteristics. However, in the 2003-2004 community outbreak of SARS, none of the 4 index case patients with confirmed SARS had severe illness, and they all seemed to have acquired infection with SARS-CoV directly from animals. Although SARS-CoV-specific antibodies previously had been detected at a relatively high rate among the population handling wild animals or had been observed in health care workers [14-16], it was impossible to correlate serological results with the corresponding onset of clinical symptoms in these retrospective screenings. Therefore, the asymptomatic case patient described in the present article is the first patient with asymptomatic SARS-CoV infection with detectable antigenemia and seroconversion caused by animal-to-human infection.

This finding, together with the previously reported findings of serological analysis of the handlers of wild animals and the 4 index case patients from the community outbreak of SARS in 2004, reveals that there likely is an intermediate epidemiological phase, which might be critical for genetic adaptation of the virus to its new hosts before critical mutation enables it to eventually cause severe symptomatic SARS, as was seen in early 2003. Although it is usually difficult to observe the virus in the intermediate phase, SARS-CoV is one of the rare viruses for which both genotypes [2, 13] and the antigen/antibody corresponding to the intermediate phase were detected.

This finding may also be significant for the prevention of SARS epidemics. Although the most contagious human epidemic strain of SARS-CoV from the middle and late phases of the 2002-2003 pandemic has not been seen in nature since June 2003 [13], the risk of asymptomatic or mildly symptomatic infection with SARS-CoV should not be ignored. To date, molecular epidemiological data have indicated that, although it might take decades for SARS-CoV to evolve into a contagious strain for human infection, it would not be too difficult for SARS-CoV to reach the stage in which it might cause infection either without symptoms or with mild symptoms in humans. This process could be accelerated if appropriate susceptible animal hosts, such as palm civets, were largely present together with the human population. On the other hand, if the highrisk population with infection can be properly monitored at the asymptomatic stage and the disease can be promptly controlled at the mildly symptomatic stage (both objectives are technically feasible now), future threats of a reemerging SARS epidemic could be much lower than once was surmised.

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## References

- 1. Liang GD, Chen QX, Xu JG, et al. Laboratory diagnosis of four recent, sporadic cases of community-acquired SARS, Guangdong Province, China. Emerg Infect Dis **2004**; 10:1774–81.
- Song HD, Tu CC, Zhang GW, et al. Cross-host evolution of severe acute respiratory syndrome coronavirus in palm civet and human. Proc Natl Acad Sci USA 2005; 102:2430–5.
- 3. Wang M, Yan MY, Xu HF, et al. SARS-CoV infection in a restaurant from palm civet. Emerg Infect Dis **2005**;11:1860–5.
- 4. Che XY, Hao W, Wang YD, et al. Nucleocapsid protein as early diagnostic marker for SARS. Emerg Infect Dis **2004**; 10:1947–9.
- Che XY, Qiu LW, Pan YX, et al. Sensitive and specific monoclonal antibody–based capture enzyme immunoassay for detection of nucleocapsid antigen in sera from patients with severe acute respiratory syndrome. J Clin Microbiol 2004; 42:2629–35.
- Li YH, Li J, Liu XE, et al. Detection of the nucleocapsid protein of severe acute respiratory syndrome coronavirus in serum: comparison with results of other viral markers. J Virol Methods 2005; 130:45–50.
- 7. Mo HY, Xu J, Ren XL, et al. Evaluation by indirect immunofluorescent assay and enzyme linked immunosorbent assay of the dynamic changes of serum antibody responses against severe acute respiratory syndrome coronavirus. Chin Med J (Engl) **2005**; 118:446–50.
- Chen WQ, Lu CY, Wong TW, et al. Anti–SARS-CoV immunoglobulin G in healthcare workers, Guangzhou, China. Emerg Infect Dis 2005; 11:89–94.
- 9. He Q, Manopo I, Lu L, et al. Novel immunofluorescence assay using

recombinant nucleocapsid-spike fusion protein as antigen to detect antibodies against severe acute respiratory syndrome coronavirus. Clin Diagn Lab Immunol **2005**; 12:321–8.

- Che XY, Qiu LW, Liao ZY, et al. Antigenic cross-reactivity between severe acute respiratory syndrome–associated coronavirus and human coronaviruses 229E and OC43. J Infect Dis 2005; 191:2033–7.
- Lewkowich IP, Campbell JD, HayGlass KT. Comparison of chemiluminescent assays and colorimetric ELISAs for quantification of murine IL-12, human IL-4 and murine IL-4: chemiluminescent substrates provide markedly enhanced sensitivity. J Immunol Methods 2001;247: 111–8.
- 12. Peiris JS, Yuen KY, Osterhaus AD, Stohr K. The severe acute respiratory syndrome. N Engl J Med **2003**; 349:2431–41.
- The Chinese SARS Molecular Epidemiology Consortium. Molecular evolution of the SARS-coronavirus during the course of the SARS epidemic in China. Science 2004; 303:1666–9.
- 14. Guan Y, Zheng BJ, He YQ, et al. Isolation and characterization of viruses related to the SARS coronavirus from animals in southern China. Science **2003**; 302:276–8.
- Wilder-Smith A, Teleman MD, Heng BH, et al. Asymptomatic SARS coronavirus infection among healthcare workers, Singapore. Emerg Infect Dis 2005; 11:1142–5.
- 16. Ho KY, Singh KS, Habib AG, et al. Mild illness associated with severe acute respiratory syndrome coronavirus infection: lessons from a prospective seroepidemiologic study of health-care workers in a teaching hospital in Singapore. J Infect Dis 2004; 189:642–7.