

Diffuse Myocardial Fibrosis Reduces Electrocardiographic Voltage Measures of Left Ventricular Hypertrophy Independent of Left Ventricular Mass

Maren Maanja; Björn Wieslander, MD, PhD; Todd T. Schlegel, MD; Ljuba Bacharova, MD, DSc, MBA; Hussein Abu Daya, MD; Yaron Fridman, MD; Timothy C. Wong, MD, MS; Erik B. Schelbert, MD, MS; Martin Ugander, MD, PhD

Background—Myocardial fibrosis quantified by myocardial extracellular volume fraction (ECV) and left ventricular mass (LVM) index (LVMI) measured by cardiovascular magnetic resonance might represent independent and opposing contributors to ECG voltage measures of left ventricular hypertrophy (LVH). Diffuse myocardial fibrosis can occur in LVH and interfere with ECG voltage measures. This phenomenon could explain the decreased sensitivity of LVH detectable by ECG, a fundamental diagnostic tool in cardiology.

Methods and Results—We identified 77 patients (median age, 53 [interquartile range, 26–60] years; 49% female) referred for contrast-enhanced cardiovascular magnetic resonance with ECV measures and 12-lead ECG. Exclusion criteria included clinical confounders that might influence ECG measures of LVH. We evaluated ECG voltage-based LVH measures, including Sokolow-Lyon index, Cornell voltage, 12-lead voltage, and the vectorcardiogram spatial QRS voltage, with respect to LVMI and ECV. ECV and LVMI were not correlated (R^2 =0.02; P=0.25). For all voltage-related parameters, higher LVMI resulted in greater voltage (r=0.33–0.49; P<0.05 for all), whereas increased ECV resulted in lower voltage (r=-0.32 to -0.57; P<0.05 for all). When accounting for body fat, LV end-diastolic volume, and mass-to-volume ratio, both LVMI (β =0.58, P=0.03) and ECV (β =-0.46, P<0.001) were independent predictors of QRS voltage (multivariate adjusted R^2 =0.39; P<0.001).

Conclusions—Myocardial mass and diffuse myocardial fibrosis have independent and opposing effects upon ECG voltage measures of LVH. Diffuse myocardial fibrosis quantified by ECV can obscure the ECG manifestations of increased LVM. This provides mechanistic insight, which can explain the limited sensitivity of the ECG for detecting increased LVM. (*J Am Heart Assoc.* 2017;6:e003795. DOI: 10.1161/JAHA.116.003795.)

Key Words: electrocardiography • extracellular matrix • hypertrophy • left ventricular • magnetic resonance imaging

D iffuse myocardial fibrosis and increased left ventricular (LV) mass may represent independent and opposing contributors to ECG voltage measures of left ventricular hypertrophy (LVH). Decreased ECG voltage from myocardial fibrosis could explain the limited sensitivity of LVH detectable by ECG, a fundamental diagnostic parameter in cardiology. LVH is broadly defined as an increased left ventricular mass (LVM)¹ and remains relevant because LVH usually indicates

prognostically adverse myocardial remodeling. In addition to enlargement of cardiomyocytes, pathologically hypertrophied myocardium undergoes complex changes in the interstitium, including diffuse myocardial fibrosis, defined as a global abnormal accumulation of collagen in the extracellular matrix of the myocardium.² The cardiomyocyte and interstitial compartments are thought to be regulated independently of one another.³ Previously, myocardial fibrosis was measured

Received August 1, 2016; accepted December 2, 2016.

© 2017 The Authors. Published on behalf of the American Heart Association, Inc., by Wiley Blackwell. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

From the Department of Clinical Physiology, Karolinska Institutet, and Karolinska University Hospital, Stockholm, Sweden (M.M., B.W., T.T.S., M.U.); Department of Medicine, University of Pittsburgh Medical Center, Pittsburgh, PA (M.M., H.A.D., Y.F., T.C.W., E.B.S.); Nicollier-Schlegel SARL, Trélex, Switzerland (T.T.S.); International Laser Center, Bratislava, Slovak Republic (L.B.); Institute of Pathophysiology, Medical School, Comenius University, Bratislava, Slovak Republic (L.B.).

This article was handled by N.A. Mark Estes III, MD, as a guest editor. The editors had no role in the evaluation of the manuscript, or in the decision about its acceptance.

Data from the current article have been presented, in part, at the Swedish Heart Spring Meeting, April 27 to 29, 2016, in Gothenburg, Sweden, the MALT meeting, June 15 to 18, 2016, in Helsingborg, Sweden, and at the European Society of Cardiology Congress, August 27 to 31, 2016, in Rome, Italy.

Correspondence to: Martin Ugander, MD, PhD, Department of Clinical Physiology, C8:27 Karolinska University Hospital, SE-17176 Stockholm, Sweden. E-mail: martin@ugander.com

only by invasive myocardial biopsy. Novel extracellular volume fraction (ECV) measures with cardiac magnetic resonance (CMR) have now been repeatedly histologically validated for noninvasive quantification of diffuse myocardial fibrosis.^{4–9} In summary, quantitative histomorphometry studies showed a strong correlation between myocardial collagen fraction and myocardial ECV fraction by CMR in patients with aortic stenosis (R^2 =0.86; P<0.001)⁴ and (R^2 =0.69; P<0.01).⁶ Another study showed a strong correlation (r=0.85; P=0.01) between endomyocardial biopsies and myocardial ECV fraction by CMR in patients.⁷

No data exist on the contributions of diffuse myocardial fibrosis to ECG changes in LVH despite the central diagnostic role that the ECG occupies in cardiology. Several criteria for LVH from both the standard 12-lead ECG and the vectorcardiogram (VCG) have been used, including the Sokolow–Lyon index, Cornell voltage criteria, and others.^{10,11} However, diagnostic performance of LVH by ECG varies greatly, with sensitivity typically close to 50% and specificity approaching 90%,¹⁰ presumably attributable to confounding by myocardial fibrosis or other conditions.

To evaluate the relationship between diffuse myocardial fibrosis and ECG voltage, we evaluated LVH-associated changes in the ECG with respect to LVM indexed to body surface area (LVMI) and ECV, as well as body fat, end-diastolic volume (EDV), and LVM/EDV^{2/3}, a surrogate for mass-to-volume ratio.¹² We studied carefully selected patients who had varying levels of LVM and ECV but who lacked any clinical or CMR-related findings that might otherwise obviously influence ECG measures of LVH. The hypothesis of the study was that both ECV and LVMI are independent and opposing contributors to ECG voltage criteria for LVH.

Methods

Study Patients

In this cross-sectional observational study, patients were identified from a prospectively acquired database of 1707 enrolled patients who had undergone a clinically indicated CMR scan at University of Pittsburgh Medical Center (UPMC; Pittsburgh, PA) between 2010 and 2014. Inclusion criteria were ECG with sinus rhythm with a heart rate <100 beats/ min and an ECG acquired within 30 days of CMR. The study was approved by the UPMC Institutional Review Board, and all participants provided written informed consent. Exclusion criteria were ECG confounders such as previous myocardial infarction (MI) or nonischemic myocardial scar as determined by late gadolinium enhancement (LGE) CMR, treatment with digitalis, atrial fibrillation or flutter, bundle branch- or fascicular block, abundant premature ventricular contractions (bigeminy/trigeminy), abundant premature atrial contractions,

QRS duration >110 ms, paced rhythm, LV end-diastolic volume (EDV) index (EDVI) >120 g/m², body mass index (BMI) <18 or >30 kg/m², previous cardiac surgery, pathologic cardiac stress perfusion CMR, cardiac amyloidosis, severe valvular disease, and significant coronary artery stenosis by invasive angiography.

Based on an alpha of 0.05, power of 0.9, and a minimum detectable correlation coefficient (*r*) of 0.4, our power calculation¹³ resulted in a required sample size of n=62. In order to provide a margin of error, we set our target sample size at n=80 patients. Patients were divided into 4 groups according to the CMR parameters of LVMI and ECV, as shown in Figure 1, with a target sample size of n=20 in each group, with as equal contributions of both sexes as possible. Selection of subjects into groups was performed before ECG analysis in order to achieve a study population with a balanced and broad range of ECV and LVMI findings. A total of 77 patients were identified; 20 in 3 groups and 17 in the group with increased ECV and high LVMI, because of a shortage of eligible patients in that group who did not have ECG confounders.

Baseline data included age at CMR, sex, LVM, LVMI, ECV, EDV, EDVI, left ventricular ejection fraction (LVEF), BMI, body surface area (BSA), tobacco use, hypertension, diabetes mellitus, treatment with beta-blockers, angiotensin-converter enzyme inhibitors (ACEi), angiotensin receptor blockers (ARB), and diuretics as reflected in the medical record. Body fat was derived from BMI, as described by Deurenberg et al, ¹⁴ according to the formula: adult body fat (%)=(1.20×BMI)+ (0.23×Age)-(10.8×sex)-5.4, where sex is 1 for males and 0 for females.

Subgroups

From a database of 1707 patients, 1170 met the exclusion criteria and 537 had data for both LVMI and myocardial ECV measurements, as shown in Table 1. We aimed to achieve a study population with both sexes equally represented, as well as with wide ranges of LVMI and ECV. Therefore, the target sample size was 10 subjects of each sex in the following subgroups: (1) normal ECV (defined as <28.5% based on normal values from volunteers at UPMC) and low LVMI (defined as \leq 55 g/m² based on the distribution of LVMI in the study population; see below); (2) normal ECV and high LVMI (>55 g/m²); (3) increased ECV (\geq 28.5%) and low LVMI; and (4) both increased ECV and high LMVI.

There are several sources for LVMI reference ranges in the literature with varying limits.^{15,16} We adopted a subgroup division strategy based on quartiles of LVMI. Specifically, we first calculated the quartile ranges of the 537 patients who met inclusion criteria and that had data on LVMI and ECV available. Next, we enrolled subjects for the high-LVMI



Figure 1. Flow chart of patient selection. The cutoff separating low from high left ventricular mass index (LVMI) was 55 g/m², and the cutoff separating normal from increased myocardial extracellular volume fraction (ECV) was 28.5%. See Methods for details on cut off criteria and selection of patients into the 4 subgroups. CMR indicates cardiovascular magnetic resonance; ECV, extracellular volume; LMVI, left ventricular mass index; MI, myocardial infarction; UPMC, University of Pittsburgh Medical Center.

subgroups from the fourth quartile and downward. Because this approach did not identify 20 patients for each subgroup, we further included patients with the highest LVMI from the third quartile. In a corresponding fashion, we enrolled subjects for the low-LVMI subgroups starting from the bottom of the second quartile and upward. To meet the sample size, we further included patients with the highest LVMI from the first quartile as well as patients with the lowest LVMI from the third quartile. Finally, this approach identified 55 g/m² as the cutoff between high and low LVMI in the population.

CMR Acquisition and Analysis

Quantification of volumes and dimensions

CMR images were acquired with a 1.5 Tesla scanner (Magnetom Espree; Siemens Medical Solutions, Erlangen, Germany) and a 32-channel phased array cardiovascular coil by dedicated CMR technologists. The exam included standard breath held segmented cine imaging with a steady-state free precession (SSFP) sequence. LVM and LVEF were measured from short-axis stacks of end-diastolic and end-systolic cine frames without geometric assumptions. Ten minutes after a 0.2-mmol/kg intravenous gadoteridol bolus (Prohance; Bracco Diagnostics, Princeton, NJ) LGE imaging was performed. To optimize LGE, a phase-sensitive inversion recovery (PSIR) pulse sequence was used, rendering signal intensity proportional to T1 recovery and corrected for surface coil intensity variation. When patients could not breath hold, single-shot SSFP, averaged PSIR, motion-corrected images were acquired.^{17,18} Typical acquisition parameters were field of view 360×270 mm, matrix 256×128 mm, and slice thickness 6 mm.

Quantification of the myocardial ECV fraction

ECV measures in myocardium without infarction or nonischemic scar were acquired after the gadolinium bolus, with minimal variation related to time elapsed following the bolus or to heart rate. T1 measures were recorded using an ECGgated single-shot-modified Look Locker inversion recovery

Characteristics	All	ECV Normal, LVMI Low	ECV Normal, LVMI High	ECV Increased, LVMI Low	ECV Increased, LVMI High	P Value
No., n	77	20	20	20	17	0.95
Age, y	53 (26–60)	48 (25–59)	47 (24–60)	54 (48–63)	54 (31–60)	0.65
Female sex	38 (49)	10 (50)	10 (50)	9 (45)	9 (53)	0.99
LVM, g	102 (85–126)	83 (80–90)	139 (124–156)	86 (75–95)	117 (102–138)	<0.001
LVMI, g/m ²	55 (45–68)	45 (43–46)	75 (68–78)	46 (42–50)	64 (59–74)	<0.001
ECV, %	28.5±3.8	26.3±1.8	24.9±2.4	30.9±2.6	32.3±2.6	<0.001
EDV, mL	149.0±37.0	136.2±28.6	164.6±34.4	137.2±39.3	159.6±38.9	0.02
EDVI, mL/m ²	79.3±17.1	72.6±14.3	86.2±12.7	71.7±18.0	87.9±17.5	0.001
LVEF, %	63 (57–67)	64 (59–68)	62 (57–67)	63 (58–69)	60 (52–66)	0.52
LVM/EDV ^{2/3} , g/mL ^{2/3}	3.9±0.9	3.3±0.5	4.8±0.8	3.4±0.5	4.2±0.7	<0.001
BMI, kg/m ²	25.2 (22.8–28.1)	24.6 (22.7–27.7)	25.3 (23.6–28.1)	26.7 (24.8–28.2)	24.1 (21.6–27.9)	0.43
Body fat, %	30.1±7.2	28.9±8.4	29.8±9.2	31.7±4.9	29.8±4.9	0.63
BSA, m ²	1.9±0.2	1.9±0.2	1.9±0.2	1.9±0.2	1.8±0.2	0.54
Hypertension	26 (34)	8 (40)	4 (20)	6 (30)	8 (47)	0.64
Diabetes mellitus	7 (9)	0 (0)	2 (10)	1 (5)	4 (24)	0.17
Smoking status						
Current smoker	8 (10)	0 (0)	3 (15)	2 (10)	3 (18)	0.39
Ex-smoker	28 (36)	10 (50)	6 (30)	6 (30)	6 (35)	0.63
Medication						
Beta-blockers	19 (25)	6 (30)	4 (20)	3 (15)	6 (35)	0.70
ACEi/ARB	18 (23)	3 (15)	7 (35)	3 (15)	5 (29)	0.49
Diuretics	29 (38)	6 (30)	9 (45)	7 (35)	7 (41)	0.88

Table 1. Baseline Variables for the Different Subgroups

Continuous data are given as mean±SD, median (interquartile range), or number (%). ACEi indicates angiotensin-converting enzyme inhibitors; ARB, angiotensin II receptor blocker; BMI, body mass index; BSA, body surface area; CMR, cardiovascular magnetic resonance; ECV, extracellular volume fraction; EDV, end-diastolic volume; EDVI, end-diastolic volume index; LVEF, left ventricular ejection fraction; LVM, left ventricular mass; LVMI, left ventricular mass index.

sequence. To avoid partial ECV effects, the middle third of the myocardium was the traced region of interest (ROI). A circular ROI was traced in the center of the blood pool as large as possible while avoiding the papillary muscles.

MI was identified when LGE involved the subendocardium in a coronary distribution. Nonischemic scar was defined as not having a distribution of LGE corresponding to MI. We quantified ECV from the myocardium without any apparent scar with the following formula:

$$ECV = \lambda \cdot (1 - hematocrit)$$

where $\lambda = \Delta R 1_{myocardium} / \Delta R 1_{bloodpool}$ and $\Delta R 1 = 1/T 1_{precontrast-1/T1postcontrast}$. ECV was measured in 2 short-axis slices in the basal and midventricular level in precontrast and postcontrast T1 images. Postcontrast T1 mapping images were acquired after acquisition of LGE images. These images were usually acquired 20 minutes after the contrast bolus. Hematocrit was acquired on the day of CMR and analyzed in the clinical laboratory. The final ECV measures were averaged

from the basal and midventricular short-axis slices. Because of obliquity between myocardium and the image plane, which could introduce partial volume error, ECV was not measured in apical slices.¹⁹ CMR data were analyzed using existing clinical software. Furthermore, myocardial extracellular mass and myocardial cellular mass were calculated as LVM×ECV, and LVM×(1–ECV), respectively, and both were indexed to BSA.

ECG Acquisition and Analysis

ECG data for each subject were collected from the local UPMCbased ECG system (MUSE Cardiology Information System, Version 8.0 SP2; GE Healthcare, Little Chalfont, UK) and exported into anonymized .xml files with coded subject identification. Data were analyzed digitally using semiautomatic software developed in-house. The following variables of the ECG were analyzed: Sokolow–Lyon index, defined as the sum of S wave voltage in lead V₁ (S_{V1}) plus the larger of the R wave voltage in lead V5 or V6 (R_{V5} or R_{V6}), where LVH is defined as >3.5 mV; Cornell voltage, defined as $S_{V3}+R_{aVL}$, where LVH is defined as >2.8 mV for men and >2.0 mV for women; Cornell voltage product, defined as Cornell voltage times QRS duration, where LVH is defined as >244 mV×ms; the 12-lead QRS voltage sum as the sum of the peak of R to nadir of S in each of the 12 leads; the QRS maximum spatial magnitude, defined as the maximum instantaneous amplitude, in millivolts, of the vectorcardiographic QRS loop²⁰ as derived from the 12-lead ECG²¹; and the QRS duration, defined as the time from the beginning of the Q-wave to the end of the S-wave.

Statistical Analysis

Statistical analysis was performed using SPSS software (version 23; IBM Corp, Armonk, NY). The Kolmogorov-Smirnov test was used to determine whether continuous variables were normally distributed. Differences between the subgroups' baseline data were tested using ANOVA, the Kruskal–Wallis test, or the chi-square test, as appropriate. Normally distributed measures were described using mean and SD and compared using the Student t test. Nonnormally distributed measures were described using median and interguartile range and compared using the Mann-Whitney U test. Linear correlations were evaluated using Pearson's correlation coefficient (r) and expressed as its square (R^2) . Multivariable linear regression was used to evaluate the relative contributions of ECV, LVMI, EDV, body fat, and $LVM/EDV^{2/3}$ to the investigated ECG variables. Nonsignificant univariable measures were not included in the multivariable analysis. A P<0.05 was considered statistically significant.

Results

Baseline characteristics of the population are presented in Table 1. A total of 77 patients were included in the study: patients with normal ECV and low LVMI (n=20), normal ECV and high LVMI (n=20), increased ECV and low LMVI (n=20), and increased ECV and high LVMI (n=17). Six patients met LVH by Sokolow–Lyon criteria only, 5 by Cornell voltage criteria only, 2 by Cornell product only, and 1 patient by all 3 sets of criteria.

Subgroup Comparisons

ECG measures for low/high LVMI and normal/increased ECV were compared, respectively. All ECG voltage variables, including Sokolow–Lyon, Cornell, 12-lead, and QRS maximum spatial magnitude voltages, were higher with greater LVMI and lower with increased ECV (*P*<0.05 for all). However, QRS duration did not differ by either ECV or LVMI.



Figure 2. Differences in voltage/mass ratio with regards to extracellular volume fraction (ECV). The voltage mass ratio was measured as the 12-lead voltage sum divided by left ventricular mass index

Impact of ECV on Voltage/Mass Ratio

(LMVI). Data are shown as mean ± 1 SD.

Figure 2 shows that the voltage/mass ratio, measured as the 12-lead voltage per LVMI, decreased with increased ECV (P<0.05). Thus, each unit of myocardial mass contributed less ECG voltage in the setting of increased ECV.

Uni- and Multivariable Linear Correlations

As shown in Figure 3, LVMI was positively correlated with all ECG voltage variables whereas ECV was negatively correlated with all ECG voltage variables. QRS duration did not correlate with ECV or LVMI. All correlations remained significant when comparing against LVM rather than LVMI.

Table 2 summarizes the results for uni- and multivariable linear regression. The results show that ECV and LVMI contributed approximately equally, but in opposing directions, to the ECG voltage. Both ECV and LVMI were independent predictors of Sokolow-Lyon voltage and QRS maximum spatial magnitude, and together with body fat, EDV, and LVM/EDV^{2/3} they could explain \approx 40% of voltage. For the other ECG voltage parameters, ECV was the only independent predictor of voltage. Univariable correlations between Cornell voltage and Cornell product and LVMI and ECV, respectively, were similar in magnitude and significance, as well as the multivariable adjusted R^2 . Figure 4 shows an illustrative case of a subject with high LVMI and ECV, yet absence of increased voltage amplitudes on the ECG. Furthermore, ECG voltage correlations with myocardial extracellular mass index and myocardial cellular mass index were also explored, but did not yield independent incremental information (data not shown). ECV and LVMI were not correlated with each other (R^2 =0.02; P=0.25). There was a weak correlation between LVEF and LVMI (R^2 =0.05; P=0.04), but there was no correlation between LVEF and ECV (R^2 =0.01; P=0.52).



Figure 3. Linear correlations between ECG measures and left ventricular mass index (LVMI) and extracellular volume fraction (ECV). Note that LVMI is positively correlated with ECG voltage, whereas ECV is negatively correlated with ECG voltage. QRS duration is not correlated with LVMI or ECV. Linear correlations between: A, Sokolow-Lyon Index and LVMI; B, Sokolow-Lyon Index and ECV; C, Cornell voltage and LVMI; D, Cornell voltage and ECV; E, 12-lead voltage sum and LVMI; F, 12-lead voltage sum and ECV; G, QRS maximum spatial magnitude and LVMI; H, QRS maximum spatial magnitude and ECV; I, QRS duration and LVMI and; J, QRS duration and ECV.

Table 2. Uni- a	nd Multivaria	ıble Linear Reg	gression Betwee	en ECG Varial	bles and LVI	MI, ECV, Bod)	y Fat, EDV, an	d LVM/EDV ^{2/3}	, Respectively	,	
ECG Variable	Univariable <i>r</i> , LVMI (<i>P</i> Value)	Univariable <i>r</i> , ECV (<i>P</i> Value)	Univariable <i>r</i> , Body Fat (<i>P</i> Value)	Univariable <i>r</i> , EDV (<i>P</i> Value)	Univariable <i>r</i> , LVM/EDV ^{2/3} (<i>P</i> Value)	Multivariable β, LVMI (<i>P</i> Value)	Multivariable β, ECV (<i>P</i> Value)	Multivariable β, Body Fat (<i>P</i> Value)	Multivariable β, EDV (<i>P</i> Value)	Multivariable β , LVM/EDV ^{2/3} (<i>P</i> Value)	Multivariable Adjusted R ² (P Value)
Sokolow-Lyon index, mV	0.49 (<0.001)	-0.43 (<0.001)	-0.27 (0.010)	0.37 (<0.001)	0.37 (0.001)	0.51 (0.045)	-0.34 (0.001)	0.16 (0.10)	0.05 (0.70)	-0.11 (0.62)	0.42 (<0.001)
Cornell voltage, mV	0.34 (0.001)	-0.32 (0.002)	-0.006 (0.48)	0.18 (0.06)	0.31 (0.003)	0.25 (0.21)	-0.28 (0.011)	N/A	N/A	0.06 (0.76)	0.19 (0.001)
Cornell product, mV×ms	0.28 (0.006)	-0.29 (0.006)	-0.034 (0.38)	0.23 (0.023)	0.27 (0.008)	-0.16 (0.60)	-0.22 (0.055)	N/A	0.25 (0.13)	0.36 (0.18)	0.17 (0.008)
12-lead voltage, mV	0.33 (0.002)	-0.57 (<0.001)	-0.24 (0.019)	0.21 (0.034)	0.27 (0.009)	0.46 (0.07)	-0.53 (<0.001)	-0.13 (0.16)	-0.10 (0.47)	0.19 (0.40)	0.42 (<0.001)
QRS maximum spatial magnitude, mV	0.38 (0.001)	-0.51 (<0.001)	-0.26 (0.013)	-0.27 (0.008)	0.27 (0.010)	0.58 (0.03)	-0.46 (<0.001)	-0.15 (0.12)	-0.077 (0.60)	-0.27 (0.23)	0.39 (<0.001)
QRS duration, ms	-0.10 (0.20)	0.00 (0.50)	-0.13 (0.12)	0.24 (0.017)	-0.07 (0.28)	N/A	N/A	N/A	N/A	N/A	N/A
Nonsignificant univaria	ble measures were	not included in the	e multivariable analysis	s. ECV indicates ex	xtracellular volum	e; EDV, end-diastol	lic volume; LVM, left	t ventricular mass; LV	/MI, left ventricular	mass index; N/A,	not applicable.

Discussion

The major finding of this study is that there is a positive relationship between ECG voltage amplitude and LVMI as well as an inverse relationship between ECG voltage amplitude and ECV. ECV and LVMI were not related to each other. After excluding confounding conditions, and when accounting for body fat, EDV, and LVM/EDV^{2/3}, a surrogate for mass-to-volume ratio, we found that both LVMI and ECV were sole independent predictors of QRS voltage.

To our knowledge, this is the first investigation to show the influence of diffuse myocardial fibrosis upon ECG criteria for LVH. A plausible physiological explanation for these findings is that increased LVM may give rise to larger and longer living activation boundaries, which thereby increase QRS amplitude. However, the complex process beyond the increased LVM, such as myocyte hypertrophy and apoptosis, changes in contractile and electrical phenotype, and alterations in the quantity and composition of the extracellular matrix, are additional factors that have been shown to affect QRS voltage.²² Conversely, increased ECV, consisting mostly of collagen, might serve as an electrical insulator, thus decreasing the voltage recorded by ECG, a phenomenon also observed in cardiac amyloidosis whereby extensive interstitial protein deposition occurs.²³ Our results indicate that increased ECV may disguise the ECG manifestations of increased LVM, in line with the previously described concept of "relative voltage deficiency."24

ECV and LVMI did not correlate with each other, and this has previously been postulated by others.³ There is some evidence indirectly supporting the notion that increased ECV is associated with decreased ECG voltages. For instance, the increase in both LVM and ECV associated with cardiac amyloidosis has been shown to correlate with decreased ECG voltages.^{25,26} Furthermore, both focal and diffuse myocardial fibrosis have been suggested to affect the impulse propagation recorded by ECG through at least 3 distinct mechanisms: decreases in the proportion of electrically active tissue; slowing of impulse generation; and fractionation of the electrical front.^{27,28}

We found no relationship between LVMI (or LVM) and QRS duration, nor a relationship between QRS duration and ECV. Previous studies have found a relationship between QRS duration and LVM measured by echocardiography or CMR.^{29–33} It may be that relationships between LVM and QRS duration are more pronounced in the high ranges of both variables. Thus, the reason for the discrepancy may lie in our strict selection of subjects to avoid confounders, given that all patients in our study had a QRS duration <110 ms. Notably, however, in the current study, both LVMI and ECV correlated with voltage measures regardless of the absence of correlation with QRS duration. ECV, but not



Figure 4. Example of a 24-year-old woman illustrating the coexistence of normal QRS amplitudes on the ECG in patients with high left ventricular mass index (LVMI) and increased myocardial extracellular volume fraction (ECV). A, The 12-lead ECG. The Sokolow-Lyon index was 2.8 mV, Cornell voltage 0.3 mV, and the 12-lead voltage was 14.7 mV. B, An end-diastolic cine image from a midventricular short-axis slice, which was part of the short-axis stack from which the LVMI (60 g/m²) was measured (left), a corresponding late gadolinium enhancement (LGE) showing absence of focal abnormalities (middle), and a corresponding ECV image showing diffusely increased ECV (30%) consistent with diffuse myocardial fibrosis (right).

LVMI, was an independent predictor for Cornell voltage and 12-lead voltage. LVM/EDV^{2/3}, was correlated with all ECG voltage variables; however, it was not an independent predictor for ECG voltage. However, mean LVM was lower in the current study compared to a previous study,¹² resulting in relatively low LVM/EDV^{2/3} values. It may be that LVM/EDV^{2/3} may predict ECG voltage in the higher ranges of LVM/EDV^{2/3}.

The ECG has repeatedly been shown to lack both sensitivity and specificity in diagnosing LVH.^{34,35} The Journal of Electrocardiology LVH Working Group has proposed that instead of the currently widely accepted model, that is, that heart disease leads to increased myocardial mass that causes ECG abnormalities, it is likely that heart and vascular disease lead to structural, bioelectrical, and biochemical changes that cause ECG abnormalities.³⁶ The results of the current study add knowledge to this ongoing debate, including the discussion related to the use of the term left ventricular electrical remodeling (LVER) as opposed to "ECG-LVH."

Study Limitations

Because of the characteristics of the subjects available for the study, the cohort was grouped according to high/low LVMI based on values above or below the median of the population as opposed to above or below the upper limit of normal values, which may vary by technique. However, we judged this grouping strategy suitable for studying the basic relationships between ECG, ECV, and LVMI, respectively. A further limitation was that neither age nor sex was taken into account when exploring these relationships. Stratifying the population according to age and sex would require increased sample size to maintain adequate power, and such studies are justified. Notably, age and sex distribution did not differ across the 4 subgroups of high/low LVMI and normal/increased ECV. The current study did not address the effects of focal myocardial scarring attributable to infarction or nonischemic scar upon the ECG, and thus the current findings may not be applicable to such a population. However, the current study was specifically designed to enroll a carefully selected population without myocardial scar in order to specifically address the isolated and respective effects of diffuse myocardial fibrosis and LVM on ECG. We believe that such a carefully selected study population was an important and unique strength of the study necessary to address the hypothesis.

Clinical Applications

The present study provides the first in vivo data supporting a novel mechanistic explanation behind the lack of sensitivity of voltage-based ECG criteria for detecting increased LVM. Also,

the results illustrate that some patients may have pathologically increased ECV that can normalize voltages and thereby disguise ECG detection of LVH.

Conclusions

Myocardial mass and diffuse fibrosis have independent and opposing effects upon ECG voltage measures of LVH. Thus, diffuse myocardial fibrosis may disguise the ECG manifestations of increased LVM. These competing effects reflect LV structural and electrical remodeling of the hypertrophied myocardium and provide mechanistic insight into the limited sensitivity of the ECG for detecting increased LVM.

Sources of Funding

The study was funded, in part, by the Swedish Research Council, Swedish Heart and Lung Foundation, Stockholm County Council, and Karolinska Institutet. This work was supported by a grant from The Pittsburgh Foundation (PA), Grant M2009-0068; and an American Heart Association Scientist Development grant (09SDG2 180083) including a T. Franklin Williams Scholarship Award; funding provided by: Atlantic Philanthropies, Inc., the John A. Hartford Foundation, the Association of Specialty Professors, and the American Heart Association (Dallas, TX). This work was also supported by Grant No. UL1 RR024153 from the National Center for Research Resources (NCRR), a component of the National Institutes of Health (NIH), and NIH Roadmap for Medical Research.

Disclosures

Dr Schlegel reports ownership interest; Modest; Nicollier-Schlegel Sarl. Dr Schelbert has accepted contrast material from Bracco Diagnostics for research purposes beyond the scope of this work. Dr Wong is supported by a grant from the American Heart Association and the Children's Cardiomyopathy Foundation. Dr Ugander is principal investigator for the Karolinska Cardiovascular Magnetic Resonance Group, and Karolinska University Hospital has a collaborative research and development agreement with Siemens regarding cardiovascular magnetic resonance imaging.

References

- Bacharova L, Ugander M. Left ventricular hypertrophy: the relationship between the electrocardiogram and cardiovascular magnetic resonance imaging. Ann Noninvasive Electrocardiol. 2014;19:524–533.
- Broberg CS, Chugh SS, Conklin C, Sahn DJ, Jerosch-Herold M. Quantification of diffuse myocardial fibrosis and its association with myocardial dysfunction in congenital heart disease. *Circ Cardiovasc Imaging.* 2010;3:727–734.
- Brilla CG, Janicki JS, Weber KT. Cardioreparative effects of lisinopril in rats with genetic hypertension and left ventricular hypertrophy. *Circulation*. 1991;83: 1771–1779.

- Flett AS, Hayward MP, Ashworth MT, Hansen MS, Taylor AM, Elliott PM, McGregor C, Moon JC. Equilibrium contrast cardiovascular magnetic resonance for the measurement of diffuse myocardial fibrosis: preliminary validation in humans. *Circulation*. 2010;122:138–144.
- Miller CA, Naish JH, Bishop P, Coutts G, Clark D, Zhao S, Ray SG, Yonan N, Williams SG, Flett AS, Moon JC, Greiser A, Parker GJ, Schmitt M. Comprehensive validation of cardiovascular magnetic resonance techniques for the assessment of myocardial extracellular volume. *Circ Cardiovasc Imaging*. 2013;6:373–383.
- White SK, Sado DM, Fontana M, Banypersad SM, Maestrini V, Flett AS, Piechnik SK, Robson MD, Hausenloy DJ, Sheikh AM, Hawkins PN, Moon JC. T1 mapping for myocardial extracellular volume measurement by CMR: bolus only versus primed infusion technique. *JACC Cardiovasc Imaging*. 2013;6:955–962.
- aus dem Siepen F, Buss SJ, Messroghli D, Andre F, Lossnitzer D, Seitz S, Keller M, Schnabel PA, Giannitsis E, Korosoglou G, Katus HA, Steen H. T1 mapping in dilated cardiomyopathy with cardiac magnetic resonance: quantification of diffuse myocardial fibrosis and comparison with endomyocardial biopsy. *Eur Heart J Cardiovasc Imaging*. 2015;16:210–216.
- Fontana M, White SK, Banypersad SM, Sado DM, Maestrini V, Flett AS, Piechnik SK, Neubauer S, Roberts N, Moon JC. Comparison of T1 mapping techniques for ECV quantification. Histological validation and reproducibility of ShMOLLI versus multibreath-hold T1 quantification equilibrium contrast CMR. J Cardiovasc Magn Reson. 2012;14:88.
- de Meester de Ravenstein C, Bouzin C, Lazam S, Boulif J, Amzulescu M, Melchior J, Pasquet A, Vancraeynest D, Pouleur AC, Vanoverschelde JL, Gerber BL. Histological validation of measurement of diffuse interstitial myocardial fibrosis by myocardial extravascular volume fraction from Modified Look-Locker imaging (MOLLI) T1 mapping at 3 T. J Cardiovasc Magn Reson. 2015;17:48.
- 10. Hancock EW, Deal BJ, Mirvis DM, Okin P, Kligfield P, Gettes LS, Bailey JJ, Childers R, Gorgels A, Josephson M, Kors JA, Macfarlane P, Mason JW, Pahlm O, Rautaharju PM, Surawicz B, van Herpen G, Wagner GS, Wellens H. AHA/ACCF/HRS recommendations for the standardization and interpretation of the electrocardiogram: part V: electrocardiogram changes associated with cardiac chamber hypertrophy: a scientific statement from the American Heart Association Electrocardiography and Arrhythmias Committee, Council on Clinical Cardiology; the American College of Cardiology Foundation; and the Heart Rhythm Society: endorsed by the International Society for Computerized Electrocardiology. *Circulation*. 2009;119:e251–e261.
- Bacharova L, Chen H, Estes EH, Mateasik A, Bluemke DA, Lima JA, Burke GL, Soliman EZ. Determinants of discrepancies in detection and comparison of the prognostic significance of left ventricular hypertrophy by electrocardiogram and cardiac magnetic resonance imaging. *Am J Cardiol.* 2015;115:515–522.
- Khouri MG, Peshock RM, Ayers CR, de Lemos JA, Drazner MH. A 4-tiered classification of left ventricular hypertrophy based on left ventricular geometry: the Dallas Heart Study. *Circ Cardiovasc Imaging*. 2010;3:164–171.
- Hulley SB, Cummings SR, Browner WS, Grady D, Newman TB. *Designing Clinical Research: An Epidemiologic Approach*. 4th ed. Philadelphia, PA: Lippincott Williams & Wilkins; 2013.
- Deurenberg P, Weststrate JA, Seidell JC. Body mass index as a measure of body fatness: age- and sex-specific prediction formulas. *Br J Nutr.* 1991;65:105–114.
- Kawel-Boehm N, Maceira A, Valsangiacomo-Buechel ER, Vogel-Claussen J, Turkbey EB, Williams R, Plein S, Tee M, Eng J, Bluemke DA. Normal values for cardiovascular magnetic resonance in adults and children. *J Cardiovasc Magn Reson.* 2015;17:29.
- 16. Le Ven F, Bibeau K, De Larochellière É, Tizón-Marcos H, Deneault-Bissonnette S, Pibarot P, Deschepper CF, Larose É. Cardiac morphology and function reference values derived from a large subset of healthy young Caucasian adults by magnetic resonance imaging. *Eur Heart J Cardiovasc Imaging*. 2016;17:981–990.
- Kellman P, Larson AC, Hsu LY, Chung YC, Simonetti OP, McVeigh ER, Arai AE. Motion-corrected free-breathing delayed enhancement imaging of myocardial infarction. *Magn Reson Med.* 2005;53:194–200.
- Piehler KM, Wong TC, Puntil KS, Zareba KM, Lin K, Harris DM, Lacomis JM, Czeyda-Pommersheim F, Cook SC, Kellman P, Schelbert EB. Free-breathing,

motion-corrected late gadolinium enhancement is robust and extends risk stratification to vulnerable patients. *Circ Cardiovasc Imaging*. 2013;6:423–432.

- Moon JC, Messroghli DR, Kellman P, Piechnik SK, Robson MD, Ugander M, Gatehouse PD, Arai AE, Friedrich MG, Neubauer S, Schulz-Menger J, Schelbert EB. Myocardial T1 mapping and extracellular volume quantification: a Society for Cardiovascular Magnetic Resonance (SCMR) and CMR Working Group of the European Society of Cardiology consensus statement. *J Cardiovasc Magn Reson.* 2013;15:92.
- Pipberger HV, Goldman MJ, Littmann D, Murphy GP, Cosma J, Snyder JR. Correlations of the orthogonal electrocardiogram and vectorcardiogram with consitutional variables in 518 normal men. *Circulation*. 1967;35:536–551.
- Kors JA, van Herpen G, Sittig AC, van Bemmel JH. Reconstruction of the Frank vectorcardiogram from standard electrocardiographic leads: diagnostic comparison of different methods. *Eur Heart J.* 1990;11:1083–1092.
- Bacharova L. Electrical and structural remodeling in left ventricular hypertrophy-a substrate for a decrease in QRS voltage? *Ann Noninvasive Electrocardiol.* 2007;12:260–273.
- Banypersad SM, Moon JC, Whelan C, Hawkins PN, Wechalekar AD. Updates in cardiac amyloidosis: a review. J Am Heart Assoc. 2012;1:e000364. DOI: 10. 1161/JAHA.111.000364.
- Bacharova L, Kyselovic J. Electrocardiographic diagnosis of left ventricular hypertrophy: is the method obsolete or should the hypothesis be reconsidered? *Med Hypotheses*. 2001;57:487–490.
- Madias JE. Low QRS voltage and its causes. J Electrocardiol. 2008;41:498– 500.
- Banypersad SM, Sado DM, Flett AS, Gibbs SD, Pinney JH, Maestrini V, Cox AT, Fontana M, Whelan CJ, Wechalekar AD, Hawkins PN, Moon JC. Quantification of myocardial extracellular volume fraction in systemic AL amyloidosis: an equilibrium contrast cardiovascular magnetic resonance study. *Circ Cardio*vasc Imaging. 2013;6:34–39.
- Gaudesius G, Miragoli M, Thomas SP, Rohr S. Coupling of cardiac electrical activity over extended distances by fibroblasts of cardiac origin. *Circ Res.* 2003;93:421–428.
- Nguyen TP, Xie Y, Garfinkel A, Qu Z, Weiss JN. Arrhythmogenic consequences of myofibroblast-myocyte coupling. *Cardiovasc Res.* 2012;93:242–251.
- Kansal S, Roitman DI, Sheffield LT. A quantitative relationship of electrocardiographic criteria of left ventricular hypertrophy with echocardiographic left ventricular mass: a multivariate approach. *Clin Cardiol.* 1983;6:456–463.
- Fragola PV, De Nardo D, Calo L, Cannata D. Use of the signal-averaged QRS duration for diagnosing left ventricular hypertrophy in hypertensive patients. *Int J Cardiol.* 1994;44:261–270.
- Carlsson MB, Tragardh E, Engblom H, Hedstrom E, Wagner G, Pahlm O, Arheden H. Left ventricular mass by 12-lead electrocardiogram in healthy subjects: comparison to cardiac magnetic resonance imaging. *J Electrocardiol.* 2006;39:67–72.
- Stewart RA, Young AA, Anderson C, Teo KK, Jennings G, Cowan BR. Relationship between QRS duration and left ventricular mass and volume in patients at high cardiovascular risk. *Heart.* 2011;97:1766–1770.
- Sjoberg S, Sundh F, Schlegel T, Maynard C, Ruck A, Wagner G, Ugander M. The relationship between electrocardiographic left ventricular hypertrophy criteria and echocardiographic mass in patients undergoing transcatheter aortic valve replacement. J Electrocardiol. 2015;48:630–636.
- Sundström J, Lind L, Arnlöv J, Zethelius B, Andrén B, Lithell HO. Echocardiographic and electrocardiographic diagnoses of left ventricular hypertrophy predict mortality independently of each other in a population of elderly men. *Circulation*. 2001;103:2346–2351.
- Pewsner D, Jüni P, Egger M, Battaglia M, Sundström J, Bachmann LM. Accuracy of electrocardiography in diagnosis of left ventricular hypertrophy in arterial hypertension: systematic review. *BMJ*. 2007;335:711.
- Bacharova L, Estes EH, Bang LE, Hill JA, Macfarlane PW, Rowlandson I, Schillaci G. Second statement of the working group on electrocardiographic diagnosis of left ventricular hypertrophy. J Electrocardiol. 2011;44:568–570.