



# Article **Prognostic Value of Salivary Biochemical Indicators in Primary Resectable Breast Cancer**

Lyudmila V. Bel'skaya \* D and Elena A. Sarf

Biochemistry Research Laboratory, Omsk State Pedagogical University, 14 Tukhachevsky str, 644043 Omsk, Russia; nemcha@mail.ru

\* Correspondence: belskaya@omgpu.ru

Abstract: Despite the fact that breast cancer was detected in the early stages, the prognosis was not always favorable. In this paper, we examined the impact of clinical and pathological characteristics of patients and the composition of saliva before treatment on overall survival and the risk of recurrence of primary resectable breast cancer. The study included 355 patients of the Omsk Clinical Oncology Center with a diagnosis of primary resectable breast cancer  $(T_{1-3}N_{0-1}M_0)$ . Saliva was analyzed for 42 biochemical indicators before the start of treatment. We have identified two biochemical indicators of saliva that can act as prognostic markers: alkaline phosphatase (ALP) and diene conjugates (DC). Favorable prognostic factors were ALP activity above 71.7 U/L and DC level above 3.93 c.u. Additional accounting for aspartate aminotransferase (AST) activity allows for forming a group with a favorable prognosis, for which the relative risk is reduced by more than 11 times (HR = 11.49, 95%CI 1.43–88.99, p = 0.01591). Salivary AST activity has no independent prognostic value. Multivariate analysis showed that tumor size, lymph nodes metastasis status, malignancy grade, tumor HER2 status, and salivary ALP activity were independent predictors. It was shown that the risk of recurrence decreased with menopause and increased with an increase in the size of the primary tumor and lymph node involvement. Significant risk factors for recurrence were salivary ALP activity below 71.7 U/L and DC levels below 3.93 c.u. before treatment. Thus, the assessment of biochemical indicators of saliva before treatment can provide prognostic information comparable in importance to the clinicopathological characteristics of the tumor and can be used to identify a risk group for recurrence in primary resectable breast cancer.

**Keywords:** saliva; biochemistry; primary resectable breast cancer; prognosis; overall survival; recurrence

### 1. Introduction

Breast cancer is the second most common cancer after lung cancer [1,2]. The high incidence of breast cancer is due to both genetic and environmental factors [3]. Despite the fact that the methods of treatment of breast cancer are constantly being improved and the mortality rate is decreasing, it is impossible to completely avoid regional and distant recurrences [3,4]. Early detection of primary and recurrent breast cancer is of great clinical importance for making breast cancer treatment decisions to improve survival rates [5]. Early (primarily operable) breast cancer includes stages 0, I, IIA, IIB, IIIA ( $T_{1-3}N_{0-1}M_0$ ) [6]. However, even if breast cancer is detected in the early stages, the prognosis is not always favorable.

Traditional prognostic factors for breast cancer are tumor size, axillary lymph node status, lymphatic and vascular invasion, hormone receptor status, and expression of human epidermal growth factor receptor 2 (HER2), but they do not fully reflect the prognosis of breast cancer [3,7,8]. Various serum tumor markers have been identified as prognostic, predictive factors in breast cancer patients, such as CEA, CA15-3, CA19-9, and CA125 [9,10]. Elevated levels of CA 15-3 or CEA have been shown to be associated



Citation: Bel'skaya, L.V.; Sarf, E.A. Prognostic Value of Salivary Biochemical Indicators in Primary Resectable Breast Cancer. *Metabolites* **2022**, *12*, 552. https://doi.org/ 10.3390/metabo12060552

Academic Editor: Silvia Ravera

Received: 20 May 2022 Accepted: 14 June 2022 Published: 16 June 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). with poor disease-free and overall survival [3,10,11]. The prognostic role of angiotensinconverting enzyme 2 (ACE2) has been described, elevated levels of which correlated with a decrease in disease-free survival [12]. Prognostic indicators based on inflammation are mentioned in the literature, including the ratio of neutrophils to lymphocytes, the ratio of lymphocytes to monocytes, the ratio of platelets to lymphocytes, and the ratio of C-reactive protein/albumin [10,13,14]. High levels of Ki-67 have also been significantly associated with poor survival [15]. Several studies have described the use of molecular markers characterizing the cellular pathways involved in tumor growth and spread (p53, RB, PI3K/Akt/mTOR, and Ras/MAPK) [16,17], as well as the tumor immune microenvironment, including tumor lymphoid infiltration (TILs) to determine the prognosis of breast cancer [18–20]. However, at present, none of the listed prognostic factors in blood serum are generally accepted in clinical practice.

A large number of studies are devoted to the use of saliva for the diagnosis of breast cancer [21–31]; however, the prognostic value of biochemical indicators of saliva has not yet been described. Previously, we have shown the potential applicability of biochemical indicators of saliva to determine the prognosis of lung cancer [32–34]. In particular, unfavorable signs are the level of imidazole compounds in the saliva of more than 0.478 mmol/L and lactate dehydrogenase activity of less than 545 U/L (HR = 4.17; 95% CI 1.36–12.51; p = 0.00000) [32]. We found that for different histological subtypes of lung cancer, different biochemical indicators of saliva are prognostically significant [33]. Due to the complete lack of literature data on the prognostic significance of biochemical indicators of saliva in breast cancer, we conducted a new selection of prognostically important signs. In this work, we tested the effect of 34 biochemical indicators of saliva on the overall survival of patients with primary resectable breast cancer. We also examined the impact of clinicopathological characteristics of patients and the composition of saliva before treatment on the risk of recurrence of primary resectable breast cancer.

#### 2. Results

# 2.1. Overall Survival Rates Depending on the Clinicopathological Characteristics of Patients and Type of Treatment

During the follow-up, 63 (17.7%) patients died; 59 (16.6%) patients had a disease relapse. Overall survival and relative risk are shown in Table 1. A statistically significant factor of poor prognosis was the age of patients older than 70, disease stages  $pT_2$  and  $pT_3$ , lymph node involvement  $pN_1$ , high grade of G3 malignancy, luminal B-like (HER2–) molecular biological tumor subtype (Figure 1). Favorable prognostic factors include the HER2-positive status of breast cancer (Table 1). It was found that the presence of menopause was not a predictive factor of the disease nor the histological type of breast cancer (Table 1). There was no statistically confirmed improvement in overall survival with a positive ER and PR status.

It was found that the amount of surgical treatment did not significantly affect overall survival (Table 2); however, the relative risk was significantly higher in groups of patients who underwent radiation and/or chemotherapy. A risk reduction was observed for patients who received hormone therapy after treatment (Table 2).



**Figure 1.** Overall survival of patients with primary resectable breast cancer depending on age (**A**), tumor size (**B**), lymph node disease status (**C**), tumor differentiation (**D**), molecular biological sub-type (**E**), and HER2 status (**F**).

Category		OS, Months	HR (95% CI)	<i>p</i> -Value
Age, years	30-39, n = 34 40-49, n = 68 50-59, n = 117 60-69, n = 105 70+, n = 27	56.7 59.3 65.4 61.0 48.1	1 1.24 (0.40–3.83) 1.12 (0.39–3.25) 1.04 (0.35–3.07) 3.41 (1.00–11.47) *	0.06851
Menopause	No, <i>n</i> = 106 Yes, <i>n</i> = 248	57.2 61.8	1 1.32 (0.71–2.43)	0.95402
рТ	1, <i>n</i> = 133 2, <i>n</i> = 172 3, <i>n</i> = 47	62.3 60.9 56.3	1 2.73 (1.32–5.56) * 6.88 (2.91–15.84) *	0.00000
pN	0, <i>n</i> = 245 1, <i>n</i> = 110	60.7 60.5	1 3.93 (2.22–6.82) *	0.00000
Grade	1, <i>n</i> = 28 2, <i>n</i> = 58 3, <i>n</i> = 88	64.3 59.5 60.8	1 3.71 (0.44–30.98) 9.00 (1.15–68.17) *	0.06269
Histological type	Ductal, $n = 171$ Lobular, $n = 58$	65.2 59.6	1 1.18 (0.56–2.47)	0.61737
Subtype	Luminal A-like, $n = 50$ Luminal B-like (HER2–), $n = 41$ Luminal B-like (HER2+), $n = 181$ Non-Luminal (HER2+), $n = 22$ Basal-like, $n = 20$	68.0 55.1 60.5 68.6 58.8	1 2.72 (1.01–7.26) * 0.80 (0.34–1.91) 0.83 (0.20–3.45) 1.75 (0.50–6.11)	0.00652
HER2-status	(-), n = 112 (+), n = 98 (++), n = 62 (+++), n = 48	60.1 61.5 60.0 60.1	1 0.63 (0.32–1.24) 0.38 (0.16–0.94) * 0.35 (0.13–0.97) *	0.03198
ER-status	(-), n = 49 (+), n = 41 (++), n = 57 (+++), n = 173	61.7 60.4 64.9 60.3	1 1.27 (0.49–3.29) 0.83 (0.33–2.10) 0.56 (0.25–1.23)	0.09137
PR-status	(-), n = 85 (+), n = 44 (++), n = 60 (+++), n = 131	60.4 60.1 62.6 60.2	1 0.89 (0.35–2.25) 0.71 (0.29–1.71) 0.85 (0.43–1.70)	0.79103

 Table 1. Overall survival rates depending on the clinicopathological characteristics of patients.

Note: \*—Differences are statistically significant, p < 0.05; G1—highly, G2—moderately, and G3—poorly differentiated breast cancer; OS—overall survival; HR—hazard ratio; CI—confidence interval.

Table 2. Overall survival	rates depending or	n type of treatment.

Category		OS, Months	HR (95% CI)	<i>p</i> -Value
Operation status	BCS, <i>n</i> = 61 TM, <i>n</i> = 286	59.0 61.3	1 1.50 (0.64–3.47)	0.37724
Radiation therapy	Done, <i>n</i> = 191 Not done, <i>n</i> = 163	60.5 60.9	1 0.34 (0.19–0.64) *	0.00452
Chemotherapy	Done, <i>n</i> = 181 Not done, <i>n</i> = 173	59.5 61.8	1 0.33 (0.18–0.60) *	0.00005
Endocrine therapy	Done, <i>n</i> = 241 Not done, <i>n</i> = 113	61.3 58.3	1 2.18 (1.24–3.77) *	0.00413

Note: BCS—breast-conserving surgery, TM—total mastectomy; \*—differences are statistically significant, p < 0.05; OS—overall survival; HR—hazard ratio; CI—confidence interval.

#### 2.2. Overall Survival Rates Depending on the Biochemical Composition of Saliva

We have identified three biochemical indicators of saliva that can act as prognostic markers, namely: alkaline phosphatase (ALP), aspartate aminotransferase (AST), and diene conjugates (DC) (Supplementary Table S3). As a threshold value, the medians of the corresponding indicators were used: for ALP—71.7 U/L, for DC—3.93 c.u., for AST—6.33 U/L (Supplementary Table S1). The results of the calculation of overall survival in groups, taking into account the threshold values of indicators, are shown in Table 3.

OS, Months HR (95% CI) p-Value Category >71.7, n = 175 61.4 1 ALP, U/L 0.00243 <71.7, n = 179 58.5 2.60 (1.44-4.62) \* >6.33, *n* = 174 61.5 1 AST, U/L 0.36144<6.33, *n* = 163 58.5 1.13 (0.64-1.97) >3.93, *n* = 176 58.7 1 DC, c.u. 0.08518 1.78 (1.02-3.08) \* <3.93, *n* = 178 60.2 >71.7, >6.33, n = 64 59.6 1 >71.7, <6.33, *n* = 98 63.3 1.80 (0.61-5.27) 0.02068 ALP + AST <71.7, >6.33, n = 76 60.5 3.15 (1.08-9.01) \* <71.7, <6.33, *n* = 97 56.7 4.10 (1.47-11.18) \* >71.7, >3.93, n = 87 61.7 1 3.15 (1.08-9.02) \* >71.7, <3.93, *n* = 87 61.0 ALP + DC0.00580 4.22 (1.49-11.74) \* <71.7, >3.93, n = 8856.5<71.7, <3.93, *n* = 91 6.21 (2.24-16.79) \* 58.9 Favorable, n = 5562.8 1 ALP + AST + DC0.01591 Unfavorable, n = 2911.49 (1.43-88.99) \* 58.9

Table 3. Biochemical composition of the saliva of patients with breast cancer depends on the stage.

Note: ALP—alkali phosphatase, AST—aspartate aminotransferase, DC—diene conjugates, \*—differences are statistically significant, p < 0.05; OS—overall survival; HR—hazard ratio; CI—confidence interval.

It was found that a favorable prognosis of breast cancer was associated with ALP activity above 71.7 U/L (Figure 2A), while the values of three-year survival were 90.4 and 95.3%, five-year survival—76.3 and 91.1%, seven-year survival 59.0 and 83.0% with an unfavorable and favorable prognosis, respectively. The content of diene conjugates is more than 3.93 c.u. was also a prognostically favorable sign (Figure 2C); the corresponding three-, five- and seven-year survival rates were 93.6, 87.0, and 75.7% in case of a favorable prognosis, respectively, and 91.3, 79.6 and 67.8% in case of unfavorable prognosis. AST activity had no independent prognostic value (Figure 2B); however, in combination with ALP activity, it made it possible to refine the prognosis of breast cancer (Figure 2E). The combination of ALP activity above 71.7 U/L and AST above 6.33 U/L was a prognostically favorable sign, while values below the threshold were unfavorable. All other combinations were intermediate. Differences in three-, five- and seven-year survival between groups with favorable and unfavorable prognosis, in this case, were more significant; the corresponding values were 96.8, 91.1, and 91.1% for a favorable prognosis and 89.4, 72.9, and 48.2% for poor prognosis, respectively. The simultaneous recording of ALP and AST activity made it possible to single out a group with the least favorable prognosis (overall survival—48.2%). Multivariate analysis of all three biochemical salivary indicators showed that AST activity was not an independent prognostic factor (Supplementary Table S4). Similarly, groups with a favorable prognosis were obtained with a combination of ALP activity above 71.7 U/L and the level of diene conjugates above 3.93 c.u. and an unfavorable prognosis with a combination of ALP activity below 71.7 U/L and the level of diene conjugates below 3.93 c.u. (Figure 2E). In this case, three-, five-, and seven-year survival rates were 96.4, 94.6, and 92.4% with a favorable prognosis, and 88.8, 72.3, and 61.6%, respectively, with an unfavorable prognosis. With a combination of all three favorable factors, the relative risk decreased by more than 11 times (Table 3). In this case, the survival rate was 96.6%, with a



favorable prognosis throughout the entire follow-up period, with an unfavorable prognosis three-, five-, and seven-year survival rates were 90.5, 69.5, and 56.6%, respectively.

**Figure 2.** Overall survival of patients with primary resectable breast cancer depending on the biochemical composition of saliva before treatment: (**A**) unfavorable prognosis (curve 1, ALP < 71.7 U/L), favorable prognosis (curve 2, ALP > 71.7 U/L); (**B**) unfavorable prognosis (curve 1, AST > 6.33 U/L), favorable prognosis (curve 2, AST < 6.33 U/L); (**C**) unfavorable prognosis (curve 1, DC > 3.93 c.u.), favorable prognosis (curve 2, DC < 3.93 c.u.); (**D**) unfavorable prognosis (curve 1, ALP < 71.7 and

AST < 6.33 U/L), favorable prognosis (curve 4, ALP > 71.7 and AST > 6.33 U/L), intermediate options (curve 2, ALP > 71.7 and AST < 6.33 U/L, curve 3—ALP < 71.7 and AST > 6.33 U/L); (E) unfavorable prognosis (curve 1, ALP < 71.7 U/L and DC < 3.93 c.u.), favorable prognosis (curve 4, ALP > 71.7 U/L and DC > 3.93 c.u.), intermediate options (curve 2, ALP > 71.7 U/L and DC < 3.93 c.u., curve 3, ALP < 71.7 U/L and DC > 3.93 c.u.); (F) unfavorable prognosis (curve 1, ALP < 71.7 U/L, AST < 6.33 U/L); AST < 6.33 U/L and DC < 3.93 c.u.), favorable prognosis (curve 2, ALP > 71.7 U/L and DC < 3.93 c.u.) and DC < 3.93 c.u.); (F) unfavorable prognosis (curve 1, ALP < 71.7 U/L, AST < 6.33 U/L); AST < 6.33 U/L and DC < 3.93 c.u.), favorable prognosis (curve 2, ALP > 71.7 U/L, AST > 6.33 U/L); AST < 6.33 U/L and DC < 3.93 c.u.), favorable prognosis (curve 2, ALP > 71.7 U/L, AST > 6.33 U/L); AST < 6.33 U/L and DC < 3.93 c.u.), favorable prognosis (curve 2, ALP > 71.7 U/L, AST > 6.33 U/L); AST < 6.33 U/L and DC < 3.93 c.u.), favorable prognosis (curve 2, ALP > 71.7 U/L); AST > 6.33 U/L and DC < 3.93 c.u.), favorable prognosis (curve 2, ALP > 71.7 U/L); AST > 6.33 U/L and DC < 3.93 c.u.).

At the next stage, we conducted a multivariate analysis, which included parameters whose contribution to the one-way analysis was statistically significant (Table 4). Tumor size, lymph nodes metastasis status, malignancy grade, tumor HER2 status, and salivary ALP activity were shown to be independent prognostic features.

<b>Prognostic Factors</b>	β	Standard Error	t-Value	<i>p</i> -Value
Age group	0.2421	0.1363	1.7761	0.0757
рТ	0.8087	0.2213	3.6550	0.0003
pN	0.9110	0.2749	3.3138	0.0009
Grade	0.7704	0.3017	2.5538	0.0107

0.1184

0.3460

0.3821

0.3829

0.3822

1.0577

-3.3477

-2.6446

0.4709

-1.2997

0.2902

0.0008

0.0082

0.6377

0.1937

**Table 4.** Results of multivariate survival analysis using the Cox regression model ( $\chi^2 = 69.67$ , p < 0.00001).

Note: ALP—alkali phosphatase, AST—aspartate aminotransferase, DC—diene conjugates.

Molecular biological subtype

HER2-status

ALP, U/L

AST, U/L

DC, c.u.

#### 2.3. Analysis of the Risk of Relapse in Patients with Primary Operable Breast Cancer

0.1252

-1.1583

-1.0105

0.1803

-0.4967

An unfavorable prognosis for primary resectable breast cancer is associated primarily with the recurrence of the disease. Thus, 59 patients had a relapse during the observation period. We analyzed the main factors that could be associated with an increased risk of disease recurrence (Table 5). The risk of recurrence has been shown to decrease with menopause and increase with primary tumor size and lymph node involvement. The absence of radiation and chemotherapy reduced the risk of recurrence. A significant risk factor for recurrence was the activity of salivary alkaline phosphatase below 71.7 U/L before treatment and the level of DC below 3.93 c.u. We determined that the ALP activity in the saliva of patients with breast cancer recurrence was 60.8 [47.8; 78.2] U/L, while in the group without recurrence, alkaline phosphatase activity was higher—76.1 [48.9; 108.7] U/L. Differences between groups were statistically significant (p = 0.0274). The DC level in the groups with and without relapse was 3.89 [3.70; 4.10] and 3.93 [3.72; 4.17] c.u., respectively (p = 0.2529), so we used this indicator as an auxiliary one when calculating the prognosis in combination with ALP activity.

Category		Relapse, <i>n</i> = 59	No Relapse, <i>n</i> = 292	HR (95% CI)	<i>p</i> -Value
	Clinicopat	hological ch	aracteristics of	patients	
	30–39	9	28	1	0.69523
Age group	40–49	16	51	0.98 (0.38-2.48)	
	50–59	19	98	0.60 (0.25–1.48)	
-	60–69	13	91	0.44 (0.17–1.15)	
-	70+	2	25	0.25 (0.05–1.26)	
Mananauaa	No	40	84	1	0.0050(
Menopause	Yes	19	208	0.19 (0.11–0.35) *	- 0.00526
	1	10	123	1	
рТ	2	30	141	2.62 (1.23–5.51) *	0.00032
	3	19	28	8.35 (3.47–19.50) *	-
nN	0	26	216	1	0.00070
pin -	1	33	76	3.61 (2.02–6.34) *	- 0.00079
	1	4	24	1	
Grade	2	8	50	0.96 (0.27–3.48)	0.46751
	3	16	71	1.35 (0.41-4.40)	-
Histological	Ductal	32	137	1	- 0.98523
type	Lobular	14	44	1.36 (0.67–2.76)	
	Luminal A-like	11	39	1	0.72153
	Luminal B-like (HER2–)	8	33	0.86 (0.31–2.37)	
biological subtype	Luminal B-like (HER2+)	30	149	0.71 (0.33–1.55)	
71 -	Non-Luminal (HER2+)	2	20	0.35 (0.07–1.75)	
	Basal-like	4	15	0.95 (0.26–3.41)	
	(—)	24	87	1	- 0.12697
-	(+)	16	81	0.72 (0.36–1.44)	
HER2-status	(++)	10	52	0.70 (0.31–1.57)	
	(+++)	7	39	0.65 (0.26–1.63)	
ER-status	(-)	8	39	1	- 0.56214
	(+)	11	30	1.79 (0.64–4.94)	
	(++)	9	48	0.91 (0.32–2.57)	
	(+++)	29	142	1.00 (0.42–2.34)	
	(-)	16	69	1	- - 0.64852 -
	(+)	6	36	0.72 (0.26–1.99)	
PR-status	(++)	10	50	0.86 (0.36–2.05)	
	(+++)	25	105	1.03 (0.51–2.05)	

 Table 5. The relative risk of relapse in patients with primary operable breast cancer.

Category		Relapse, <i>n</i> = 59	No Relapse, <i>n</i> = 292	HR (95% CI)	<i>p</i> -Value		
Type of treatment							
Operation _ status	BCS	5	56	1	- 0.16325		
	TM	51	235	2.43 (0.93-6.29)			
Radiation	Done	41	150	1			
therapy	Not done	18	145	0.45 (0.25–0.83) *	- 0.00289		
Charry ath arrange	Done	42	139	1			
Chemotherapy -	Not done	17	156	0.36 (0.20–0.66) *	- 0.00117		
Endocrine	Done	38	203	1	- 0.62547		
therapy	Not done	21	92	1.22 (0.68–2.18)			
Biochemical indicators of saliva							
	>71.7	22	163	1	- 0.00524		
ALP, U/L -	<71.7	36	129	2.07 (1.16-3.66) *			
	>6.33	29	142	1	- 0.69441		
AS1, U/L -	<6.33	28	133	1.03 (0.58–1.82)			
	>3.93	26	148	1	- 0.45597		
DC, c.u	<3.93	33	144	1.30 (0.74–2.28)			
ALP + AST -	>71.7, >6.33	16	86	1	- 0.14965		
	<71.7, <6.33	23	67	1.85 (0.90–3.73)			
ALP + DC	>71,7, >3.93	10	81	1	- 0.00124		
	<71.7, <3.93	21	61	2.79 (1.22-6.27) *			
ALP + AST +	Favorable	7	45	1	- 0.08963		
DC	Unfavorable	14	37	2.43 (0.89–6.57)			

Table 5. Cont.

Note: \*—The differences are statistically significant, p < 0.05. ALP—alkali phosphatase, AST—aspartate amino-transferase, DC—diene conjugates.

## 3. Discussion

We have shown that the factors of unfavorable prognosis are, first, the size of the tumor, the defeat of the lymph nodes and the high degree of malignancy, which is consistent with the literature data. It is known that despite a generally favorable prognosis for primary operable breast cancer, in cases of high malignancy or HER2 overexpression, survival rates decrease to 70–75% [35]. The invasion of the lymphatic vessels around the tumor significantly correlates with the size of the primary lesion, the histological malignancy of breast cancer, the involvement of the axillary lymph nodes, and the expression of hormone receptors [36]. The type of treatment was determined by the tumor's corresponding clinical and pathological characteristics; therefore, it naturally affected the prognosis of breast cancer. Thus, patients with a large tumor size and lymph node involvement are subject to chemotherapy and radiation therapy and have worse overall survival rates (Table 2).

It has been shown for the first time that a number of biochemical indicators of saliva can be of prognostic value in breast cancer (Table 3). Thus, an independent prognostic factor is the activity of salivary alkaline phosphatase before treatment (Table 4). It is known that ALP is formed in the liver, skeletal tissue, intestines, kidneys, placenta, and various tumors and is usually considered a serum marker of hepatobiliary pathology and fractures [37]. Previous studies have shown that ALP is associated with systemic inflammation and tumor development [38,39]. ALP is a hydrolytic enzyme that dephosphorylates various types of molecules, including nucleotides, proteins, and alkaloids [40]. It plays an anti-inflammatory

and tissue-protective role by enhancing the conversion of ATP to adenosine and increasing the level of adenosine [41]. A previous study showed that ALP activity is associated with cancer cell death, migration, and transition from mesenchyme to epithelium [39]. The albumin-to-alkaline phosphatase ratio (AAPR) is a combined measure associated with systemic inflammation, which is calculated by dividing the albumin level by the ALP level [42]. Higher serum ALP levels have been shown to be associated with a worse prognosis in nasopharyngeal carcinoma, prostate cancer, and colorectal cancer [43–45]. Chen et al. reported that pretreatment serum ALP was an independent adverse predictor of disease-free and overall survival in patients with basal-like breast cancer [46]. Women with breast cancer have ALP activities generally higher than in normal, healthy women. The progressive increase in the serum ALP activities with breast cancer indicates metastasis [47]. However, according to our data, increased ALP activity in saliva is, on the contrary, a favorable prognostic sign. We have previously shown that the activities of a number of enzymes in blood serum and saliva do not always coincide; therefore, the corresponding indicators in saliva should be considered independently and have their own reference values [48].

Salivary AST activity and the level of diene conjugates were additional prognostic indicators of saliva. According to current concepts, the effect of AST on cancer recurrence and survival is unclear [49]. Researchers' understanding of the potential role of AST in human carcinogenesis remains speculative. Unlike normal cells, most cancer cells rely on aerobic glycolysis to generate the energy needed for cellular processes. Warburg suggested that mitochondrial dysfunction exists in cancer cells, as he observed that cancer cells could convert most glucose into lactate, regardless of the presence of oxygen [50]. Some studies have shown that cancer cell proliferation can also be powered by the metabolism of glutamine, which is required by tumor cells to support the biosynthesis of nucleotides and non-essential amino acids catalyzed by AST and ALT [51]. Thornburg et al. reported that oxamate could inhibit the proliferation of transformed mammary adenocarcinoma cells in vitro, and AST acts as an important metabolic target [52]. A number of studies have shown the predictive value of the De Ritis coefficient (AST/ALT-ratio) [53]. However, according to our data, it makes sense to consider only AST activity.

The pathogenetic role of oxygen free radicals and the processes of lipid peroxidation initiated by them in the development of diseases, including cancer, is widely known [34,54,55]. Oxidative stress manifests itself in the accumulation of damaged DNA bases, products of protein oxidation and lipid peroxidation, as well as in a decrease in the level of antioxidants and, as a result, an increase in the susceptibility of membrane lipids and lipoproteins to the action of prooxidants [56]. As the results of the observations showed, in the group of patients with primary operable breast cancer (stages I-IIA), the content of lipid peroxidation products was increased in the zone of breast neoplasia compared to the neighboring tissues of the gland—in the zone of the visible absence of malignant cells [57]. However, the prognostic role of DC has not yet been discussed. Prognostically favorable, according to our data, was an increase in the level of DC in saliva, which is due to a shift in the equilibrium of lipid peroxidation processes towards primary products (DC); against this background, the content of toxic Schiff bases and MDA decreases. Simultaneous assessment of ALP activity, AST, and DC level made it possible to form a group of patients with a favorable prognosis (all indicators were above the threshold values), for which the relative risk was 11.5 times lower compared to the group in which all of the listed indicators were below the threshold values (Table 3).

The picture of breast cancer recurrence has not changed significantly in recent years; however, there is a statistically significant increase in relapse-free survival. There is an improvement in the results of treatment of all types of breast cancer, especially HER2-positive and triple-negative, with a significant decrease in the frequency of early recurrences [58]. The likelihood of local recurrence is higher in patients with high-risk molecular subtypes [59,60]. However, a number of studies have shown that local recurrences after organ-preserving operations are not associated with a specific molecular subtype of the

tumor [61,62]. In general, the incidence of local recurrence of breast cancer is from 10 to 30% and remains high after both breast-conserving operations and after mastectomy, despite the use of adjuvant radiation therapy [63]. We analyzed a group of patients with a local or distant breast cancer recurrence during the observation period (Table 5). It was shown that in the group with recurrence, there are more patients with preserved menstrual function (67.8% vs. 28.8%), a higher proportion of larger tumors (38.7% vs. 9.6% for pT2), and more often affected lymph nodes (67.3% vs. 26.0%). The remaining factors are not statistically significant factors in the occurrence of relapse. Additionally, it was shown that in the group with relapse of the disease, the activity of salivary alkaline phosphatase is lower than in the group without relapse. Moreover, the combination of ALP activity above 71.7 U/l and DC level above 3.93 c.u. is a favorable factor and reduces the risk of breast cancer recurrence by 2.5 times (Table 5).

The study's limitations include the lack of information on anti-HER2 therapy performed in the study groups, as well as on Ki-67, which could provide additional information.

Thus, the assessment of biochemical indicators of saliva before treatment can provide prognostic information comparable in importance to the clinical and pathological characteristics of the tumor and can be used to identify a risk group for recurrence in primary resectable breast cancer.

#### 4. Materials and Methods

#### 4.1. Study Design and Group Description

The study included 355 patients of the Clinical Oncological Dispensary in Omsk, hospitalized with a diagnosis of primary resectable breast cancer in the period 2014–2017. Patients were enrolled after informed consent, and the study was performed following the approval from the ethical committee of the Omsk Regional Clinical Oncological Dispensary (21 July 2016, Protocol No. 15) and in accordance with Helsinki principles. All patients were divided into age groups with a step of ten years: 30–39 years old—34 (9.6%), 40–49 years old—68 (19.2%), 50–59 years old—117 (33.0%), 60–69 years old—105 (29.6%), over 70 years old—31 people (8.6%); 248 (69.9%) patients were in the postmenopausal state. For breast cancer staging, the AJCC TNM classification (8th edition, 2017) was used. Thus, pT<sub>1</sub> stage was verified in 133 (37.5%) patients, pT<sub>2</sub>—in 172 (48.5%), pT<sub>3</sub>—in 50 (14.0%) patients. The status of lymph node involvement  $pN_0$  was confirmed in 245 (69.0%) patients and  $pN_1$ in the remaining 110 patients. According to the histological type, 171 (74.7%) patients had ductal breast cancer, 58 (25.3%) had lobular cancer. According to the histological degree of malignancy, G1 status was established for 28 (16.1%) patients, G2-for 58 (33.3%), G3—for 88 (50.6%) patients. According to the molecular biological subtype, 50 (15.9%) patients were assigned to luminal A-like, 41 (13.1%)-to luminal B-like (HER2-negative), 181 (57.6%)—to luminal B-like (HER2-positive), 22 (7.0%)—to Non-Luminal and 20 (6.4%) to basal-like subtypes, respectively. According to the receptor status, HER2-negative status was 112 (35.0%), HER2-positive status—208 (65.0%), ER-negative—49 (15.3%), ER-positive-271 (84.7%), PR-negative—85 (26.6%), PR-positive—235 (73.4%) patients, respectively. Organ-preserving surgical treatment (sectoral resection) was performed in 61 (17.2%) patients, mastectomy in 286 (80.6%) patients, and in 8 (2.2%) patients; surgical treatment was not performed due to contraindications. Adjuvant radiation therapy was performed in 191 (53.8%) patients; adjuvant chemotherapy was performed in 181 (51.0%) patients and hormone therapy in 241 (67.9%) patients.

#### 4.2. Determination of the Expression of the Receptors for Estrogen, Progesterone, and HER2

The Allred Scoring Guideline was used to assess the level of expression of estrogen receptors (ER), progesterone receptors (PR), and HER2 [64]. The level of expression of estrogen, progesterone, and HER2 receptors was assigned to one of four categories (-, +, ++, +++) in accordance with the ASCO/CAP recommendations [65]. According to the obtained values, breast cancer was classified into five groups: basal-like, luminal A-like, luminal B-like (HER2-negative), luminal B-like (HER2-positive), and Non-Luminal.

#### 4.3. Collection and Analysis of Saliva

Saliva samples were collected at baseline, right before the start of treatment. Collection of saliva samples was carried out on an empty stomach after rinsing the mouth with water in the interval of 8–10 am by spitting into sterile polypropylene tubes. Saliva samples were centrifuged ( $10,000 \times g$  for 10 min) (CLb-16, Moscow, Russia), and biochemical analysis was performed without storage and freezing on a Stat Fax 3300 semi-automatic biochemical analyzer (Awareness Technology, Palm City, FL, USA) for 34 biochemical indicators, as described previously [28,66]. The full results of biochemical analysis of saliva for 34 indicators for a group of patients with primary resectable breast cancer compared with healthy controls are given in Supplementary Table S1. Table S2 shows the results of the biochemical analysis of saliva, depending on the presence/absence of relapse.

#### 4.4. Statistical Analysis

The total follow-up time was 7 years; the median follow-up time was 59.8 months. The patient's overall survival (OS) was assessed from the date of hospitalization to the date of the last observation (censored) or the date of death of the patient (complete). OS was assessed using the Kaplan–Meier method (Statistica 13.0, StatSoft, Tulsa, OK, USA). A univariate Cox proportional hazards regression analysis was initially carried out to investigate the relationships between salivary parameters and survival data (Supplementary Table S3). Finally, variables with p < 0.10 were chosen to formulate multivariate Cox proportional hazards regression models and determine the independent prognostic factors for OS (Supplementary Table S4).

**Supplementary Materials:** The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/metabo12060552/s1, Table S1: Biochemical composition of the saliva of patients with primary resectable breast cancer and the control group; Table S2: Biochemical composition of the saliva of patients with primary resectable breast cancer depending on the presence/absence of relapse; Table S3: Results of univariate Cox proportional hazards regression analysis; Table S4: Results of multivariate survival analysis using the Cox regression model for biochemical indicators of saliva only.

**Author Contributions:** Conceptualization, L.V.B.; methodology, L.V.B.; validation, L.V.B. and E.A.S.; formal analysis, L.V.B.; investigation, E.A.S.; resources, L.V.B.; data curation, L.V.B.; writing—original draft preparation, E.A.S.; writing—review and editing, L.V.B.; visualization, E.A.S.; supervision, L.V.B.; project administration, L.V.B. All authors have read and agreed to the published version of the manuscript.

Funding: This research received no external funding.

**Institutional Review Board Statement:** The study was conducted according to the guidelines of the Declaration of Helsinki and approved by the Ethics Committee of Omsk Regional Clinical Oncological Dispensary (21 July 2016, Protocol No. 15).

Informed Consent Statement: Informed consent was obtained from all subjects involved in the study.

**Data Availability Statement:** The data presented in this study are available on request from the corresponding author. The data are not publicly available because they are required for the preparation of a Ph.D. thesis.

Conflicts of Interest: The authors declare no conflict of interest.

#### References

- Kabel, A.M.; Baali, F.H. Breast cancer: Insights into risk factors, pathogenesis, diagnosis and management. J. Cancer Res. Treat. 2015, 3, 28–33.
- Sung, H.; Ferlay, J.; Siegel, R.L.; Laversanne, M.; Soerjomataram, I.; Jemal, A.; Bray, F. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J. Clin. 2021, 71, 209–249. [CrossRef]

- 3. Uygur, M.M.; Gümüş, M. The utility of serum tumor markers CEA and CA 15–3 for breast cancer prognosis and their association with clinicopathological parameters. *Cancer Treat. Res. Commun.* **2021**, *28*, 100402. [CrossRef]
- 4. Kabel, A.M.; Elkhoely, A.A. Ameliorative potential of fluoxetine/raloxifene combination on experimentally induced breast cancer. *Tissue Cell* **2016**, *48*, 89–95. [CrossRef] [PubMed]
- Shah, R.; Rosso, K.; Nathanson, S.D. Pathogenesis, prevention, diagnosis and treatment of breast cancer. World J. Clin. Oncol. 2014, 5, 283–298. [CrossRef]
- 6. Tyulyandin, S.A.; Zhukova, L.G.; Koroleva, I.A.; Parokonnaya, A.A.; Semiglazova, T.Y.; Stenina, M.B.; Frolova, M.A. Practical recommendations for the drug treatment of breast cancer. *Malig. Tumors Russco. Pract. Guidel.* **2021**, *11*, 119–157.
- Soerjomataram, I.; Louwman, M.W.; Ribot, J.G.; Roukema, J.A.; Coebergh, J.W. An overview of prognostic factors for long-term survivors of breast cancer. *Breast Cancer Res. Treat.* 2008, 107, 309–330. [CrossRef]
- 8. Mohanty, S.S.; Sahoo, C.R.; Padhy, R.N. Role of hormone receptors and HER2 as prospective molecular markers for breast cancer: An update. *Genes Dis.* **2022**, *9*, 648–658. [CrossRef]
- 9. Wang, W.; Xu, X.; Tian, B.; Wang, Y.; Du, L.; Sun, T.; Shi, Y.; Zhao, X.; Jing, J. The Diagnostic Value of Serum Tumor Markers CEA, CA19-9, CA125, CA15-3, and TPS in Metastatic Breast Cancer. *Clin. Chim. Acta* **2017**, *470*, 51–55. [CrossRef]
- Greten, F.; Grivennikov, S. Inflammation and Cancer: Triggers, Mechanisms, and Consequences. *Immunity* 2019, 51, 27–41. [CrossRef]
- Hing, J.X.; Mok, C.W.; Tan, P.T.; Sudhakar, S.S.; Seah, C.M.; Lee, W.P.; Tan, S.M. Clinical utility of tumour marker velocity of cancer antigen 15-3 (CA15-3) and carcinoembryonic antigen (CEA) in breast cancer surveillance. *Breast* 2020, 52, 95–101. [CrossRef] [PubMed]
- 12. Nair, M.G.; Prabhu, J.S.; Ts, S. High expression of *ACE2* in HER2 subtype of breast cancer is a marker of poor prognosis. *Cancer Treat. Res. Commun.* **2021**, 27, 100321. [CrossRef]
- Zhang, F.; Huang, M.; Zhou, H.; Chen, K.; Jin, J.; Wu, Y.; Ying, L.; Ding, X.; Su, D.; Zou, D. A Nomogram to Predict the Pathologic Complete Response of Neoadjuvant Chemotherapy in Triple-Negative Breast Cancer Based on Simple Laboratory Indicators. *Ann. Surg. Oncol.* 2019, 26, 3912–3919. [CrossRef] [PubMed]
- 14. Tiainen, S.; Rilla, K.; Hämäläinen, K.; Oikari, S.; Auvinen, P. The prognostic and predictive role of the neutrophil-to-lymphocyte ratio and the monocyte-to-lymphocyte ratio in early breast cancer, especially in the HER2+ subtype. *Breast Cancer Res. Treat.* **2021**, *185*, 63–72. [CrossRef] [PubMed]
- Choi, S.B.; Park, J.M.; Ahn, J.H.; Go, J.; Kim, J.; Park, H.S.; Kim, S.I.; Park, B.W.; Park, S. Ki-67 and breast cancer prognosis: Does it matter if Ki-67 level is examined using preoperative biopsy or postoperative specimen? *Breast Cancer Res. Treat.* 2022, 192, 343–352. [CrossRef]
- 16. Kourea, H.P.; Zolota, V.; Scopa, C.D. Targeted pathways in breast cancer: Molecular and protein markers guiding therapeutic decisions. *Curr. Mol. Pharmacol.* **2014**, *7*, 4–21. [CrossRef]
- 17. Bertheau, P.; Lehmann-Che, J.; Varna, M.; Dumay, A.; Poirot, B.; Porcher, R.; Turpin, E.; Plassa, L.F.; de Roquancourt, A.; Bourstyn, E.; et al. P53 in breast cancer subtypes and new insights into response to chemotherapy. *Breast* **2013**, *22*, S27–S29. [CrossRef]
- Denkert, C.; von Minckwitz, G.; Brase, J.C.; Sinn, B.V.; Gade, S.; Kronenwett, R.; Pfitzner, B.M.; Salat, C.; Loi, S.; Schmitt, W.D.; et al. Tumor-infiltrating lymphocytes and response to neoadjuvant chemotherapy with or without carboplatin in human epidermal growth factor receptor 2-positive and triple-negative primary breast cancers. J. Clin. Oncol. 2015, 33, 983–991. [CrossRef]
- 19. Krüger, J.M.; Wemmert, C.; Sternberger, L.; Bonnas, C.; Dietmann, G.; Gançarski, P.; Feuerhake, F. Combat or surveillance? Evaluation of the heterogeneous inflamatory breast cancer microenvironment. *J. Pathol.* **2013**, 229, 569–578. [CrossRef]
- 20. Savas, P.; Salgado, R.; Denkert, C.; Sotiriou, C.; Darcy, P.K.; Smyth, M.J.; Loi, S. Clinical relevance of host immunity in breast cancer: From TILs to the clinic. *Nat. Rev. Clin. Oncol.* **2016**, *13*, 228–241. [CrossRef]
- Cavaco, C.; Pereira, J.A.M.; Taunk, K.; Taware, R.; Rapole, S.; Nagarajaram, H.; Câmara, J.S. Screening of salivary volatiles for putative breast cancer discrimination: An exploratory study involving geographically distant populations. *Anal. Bioanal. Chem.* 2018, 410, 4459–4468. [CrossRef] [PubMed]
- 22. Al-Muhtaseb, S.I. Serum and saliva protein levels in females with breast cancer. *Oncol. Lett.* **2014**, *8*, 2752–2756. [CrossRef] [PubMed]
- 23. Liu, X.; Yu, H.; Qiao, Y.; Yang, J.; Shu, J.; Zhang, J.; Zhang, Z.; He, J.; Li, Z. Salivary glycopatterns as potential biomarkers for screening of early-stage breast cancer. *EBioMedicine* **2018**, *28*, 70–79. [CrossRef]
- 24. Takayama, T.; Tsutsui, H.; Shimizu, I.; Toyama, T.; Yoshimoto, N.; Endo, Y.; Inoue, K.; Todoroki, K.; Min, J.Z.; Mizuno, H.; et al. Diagnostic approach to breast cancer patients based on target metabolomics in saliva by liquid chromatography with tandem mass spectrometry. *Clin. Chim. Acta* **2016**, 452, 18–26. [CrossRef]
- Wang, X.; Zhao, X.; Chou, J.; Yu, J.; Yang, T.; Liu, L.; Zhang, F. Taurine, glutamic acid and ethylmalonic acid as important metabolites for detecting human breast cancer based on the targeted metabolomics. *Cancer Bio.* 2018, 23, 255–268. [CrossRef] [PubMed]
- Murata, T.; Yanagisawa, T.; Kurihara, T.; Kaneko, M.; Ota, S.; Enomoto, A.; Tomita, M.; Sugimoto, M.; Sunamura, M.; Hayashida, T.; et al. Salivary metabolomics with alternative decision tree-based machine learning methods for breast cancer discrimination. Breast Cancer Res. Treat. 2019, 177, 591–601. [CrossRef]
- Rapado-González, Ó.; Martínez-Reglero, C.; Salgado-Barreira, Á.; Takkouche, B.; López-López, R.; Suárez-Cunqueiro, M.M.; Muinelo-Romay, L. Salivary biomarkers for cancer diagnosis: A meta-analysis. *Ann. Med.* 2020, 52, 131–144. [CrossRef]

- Bel'skaya, L.V.; Sarf, E.A.; Solomatin, D.V.; Kosenok, V.K. Metabolic Features of Saliva in Breast Cancer Patients. *Metabolites* 2022, 12, 166. [CrossRef]
- López-Jornet, P.; Aznar, C.; Ceron, J.; Asta, T. Salivary biomarkers in breast cancer: A cross-sectional study. *Support. Care Cancer* 2021, 29, 889–896. [CrossRef]
- 30. Porto-Mascarenhas, E.C.; Assad, D.X.; Chardin, H.; Gozal, D.; De Luca Canto, G.; Acevedo, A.C.; Guerra, E.N. Salivary biomarkers in the diagnosis of breast cancer: A review. *Crit. Rev. Oncol./Hematol.* **2017**, *110*, 62–73. [CrossRef]
- Koopaie, M.; Kolahdooz, S.; Fatahzadeh, M.; Manifar, S. Salivary biomarkers in breast cancer diagnosis: A systematic review and diagnostic meta-analysis. *Cancer Med.* 2022, 1–18. [CrossRef] [PubMed]
- 32. Bel'skaya, L.V.; Sarf, E.A.; Kosenok, V.K. Survival Rates of Patients with Non-Small Cell Lung Cancer Depending on Lymph Node Metastasis: A Focus on Saliva. *Diagnostics* **2021**, *11*, 912. [CrossRef] [PubMed]
- 33. Bel'skaya, L.V.; Kosenok, V.K. A new field of application of saliva tests for prognostic purpose: Focus on lung cancer. *Biomedical Chemistry: Res. Methods* **2020**, *3*, e00133. [CrossRef]
- Bel'skaya, L.V.; Sarf, E.A.; Kosenok, V.K.; Gundyrev, I.A. Biochemical Markers of Saliva in Lung Cancer: Diagnostic and Prognostic Perspectives. *Diagnostics* 2020, 10, 186. [CrossRef] [PubMed]
- Ruibal, Á.; Aguiar, P.; Del Rio, M.C.; Arias, J.I.; Menéndez-Rodríguez, P.; Gude, F.; Herranz, M. Histological grade (HG) in invasive ductal carcinomas of the breast of less than 1 cm: Clinical and biological assosiations during progression from HG1 to HG3. *Anticancer Res.* 2015, 35, 569–573.
- Gurleyik, G.; Gurleyik, E.; Aker, F.; Aktekin, A.; Emir, S.; Gungor, O.; Saglam, A. Lymphovascular invasion, as a prognostic marker in patients with invasive breast cancer. *Acta Chir. Belg.* 2007, 107, 284–287. [CrossRef]
- Haarhaus, M.; Brandenburg, V.; Kalantar-Zadeh, K.; Stenvinkel, P.; Magnusson, P. Alkaline Phosphatase: A Novel Treatment Target for Cardiovascular Disease in CKD. *Nat. Rev. Nephrol.* 2017, 13, 429–442. [CrossRef]
- 38. Damera, S.; Raphael, K.; Baird, B.; Cheung, A.; Greene, T.; Beddhu, S. Serum Alkaline Phosphatase Levels Associate with Elevated Serum C-Reactive Protein in Chronic Kidney Disease. *Kidney Int.* **2011**, *79*, 228–233. [CrossRef]
- Rao, S.R.; Snaith, A.E.; Marino, D.; Cheng, X.; Lwin, S.T.; Orriss, I.R.; Hamdy, F.C.; Edwards, C.M. Tumour-Derived Alkaline Phosphatase Regulates Tumour Growth, Epithelial Plasticity and Disease-Free Survival in Metastatic Prostate Cancer. *Br. J. Cancer* 2017, 116, 227–236.
- 40. Cauwels, A.; Rogge, E.; Vandendriessche, B.; Shiva, S.; Brouckaert, P. Extracellular ATP Drives Systemic Inflammation, Tissue Damage and Mortality. *Cell Death Dis.* **2014**, *5*, e1102. [CrossRef]
- Peters, E.; Geraci, S.; Heemskerk, S.; Wilmer, M.J.; Bilos, A.; Kraenzlin, B.; Gretz, N.; Pickkers, P.; Masereeuw, R. Alkaline Phosphatase Protects Against Renal Inflammation Through Dephosphorylation of Lipopolysaccharide and Adenosine Triphosphate. *Br. J. Pharm.* 2015, 172, 4932–4945. [CrossRef] [PubMed]
- Qu, F.; Li, Z.; Lai, S.; Zhong, X.; Fu, X.; Huang, X.; Li, Q.; Liu, S.; Li, H. Construction and Validation of a Serum Albumin-to-Alkaline Phosphatase Ratio-Based Nomogram for Predicting Pathological Complete Response in Breast Cancer. *Front. Oncol.* 2021, 11, 681905. [CrossRef] [PubMed]
- 43. Li, G.; Gao, J.; Tao, Y.L.; Xu, B.Q.; Tu, Z.W.; Liu, Z.G.; Zeng, M.S.; Xia, Y.F. Increased Pretreatment Levels of Serum LDH and ALP as Poor Prognostic Factors for Nasopharyngeal Carcinoma. *Chin. J. Cancer* **2012**, *31*, 197–206. [CrossRef] [PubMed]
- Sonpavde, G.; Pond, G.R.; Berry, W.R.; de Wit, R.; Armstrong, A.J.; Eisenberger, M.A.; Tannock, I.F. Serum Alkaline Phosphatase Changes Predict Survival Independent of PSA Changes in Men with Castration-Resistant Prostate Cancer and Bone Metastasis Receiving Chemotherapy. Urol. Oncol. 2012, 30, 607–613. [CrossRef]
- 45. Maisano, R.; Azzarello, D.; Del Medico, P.; Maisano, M.; Bottari, M.; Egitto, G.; Nardi, M. Alkaline Phosphatase Levels as a Prognostic Factor in Metastatic Colorectal Cancer Treated with the FOLFOX 4 Regimen: A Monoinstitutional Retrospective Study. *Tumori* **2011**, *97*, 39–42. [CrossRef] [PubMed]
- 46. Chen, B.; Dai, D.; Tang, H.; Chen, X.; Ai, X.; Huang, X.; Wei, W.; Xie, X. Pre-Treatment Serum Alkaline Phosphatase and Lactate Dehydrogenase as Prognostic Factors in Triple Negative Breast Cancer. *J. Cancer* **2016**, *7*, 2309–2316. [CrossRef] [PubMed]
- 47. Singh, A.K.; Pandey, A.; Tewari, M.; Kumar, R.; Sharma, A.; Singh, K.A.; Pandey, H.P.; Shukla, H.S. Advanced stage of breast cancer hoist alkaline phosphatase activity: Risk factor for females in India. *3 Biotech* **2013**, *3*, 517–520. [CrossRef]
- 48. Bel'skaya, L.V.; Sarf, E.A.; Kosenok, V.K. Age and gender characteristics of the biochemical composition of saliva: Correlations with the composition of blood plasma. *J. Oral Biol. Craniofacial Res.* **2020**, *10*, 59–65. [CrossRef]
- Chen, S.-L.; Xue, N.; Wu, M.-T.; Chen, H.; He, X.; Li, J.-P.; Liu, W.-L.; Dai, S.-Q. Influence of Preoperative Serum Aspartate Aminotransferase (AST) Level on the Prognosis of Patients with Non-Small Cell Lung Cancer. *Int. J. Mol. Sci.* 2016, 17, 1474. [CrossRef]
- 50. Hsu, P.P.; Sabatini, D.M. Cancer cell metabolism: Warburg and beyond. Cell 2008, 134, 703–707. [CrossRef]
- 51. Elf, S.E.; Chen, J. Targeting glucose metabolism in patients with cancer. Cancer 2014, 120, 774–780. [CrossRef] [PubMed]
- 52. Thornburg, J.M.; Nelson, K.K.; Clem, B.F.; Lane, A.N.; Arumugam, S.; Simmons, A.; Eaton, J.W.; Telang, S.; Chesney, J. Targeting aspartate aminotransferase in breast cancer. *Breast Cancer Res.* **2008**, *10*, R84. [CrossRef] [PubMed]
- Lee, H.; Choi, Y.H.; Sung, H.H.; Han, D.H.; Jeon, H.G.; Chang Jeong, B.; Seo, S.I.; Jeon, S.S.; Lee, H.M.; Choi, H.Y. De Ritis Ratio (AST/ALT) as a Significant Prognostic Factor in Patients with Upper Tract Urothelial Cancer Treated with Surgery. *Clin. Genitourin. Cancer* 2017, 15, e379–e385. [CrossRef]

- 54. Bel'skaya, L.V.; Kosenok, V.K.; Massard, G.; Zav'yalov, A.A. Status Indicators of Lipid Peroxidation and Endogenous Intoxication in Lung Cancer Patients. *Ann. Russ. Acad. Med. Sci.* 2016, *71*, 313–322.
- Barrera, G. Oxidative Stress and Lipid Peroxidation Products in Cancer Progression and Therapy. ISRN Oncol. 2012, 2012, 137289.
   [CrossRef]
- Gęgotek, A.; Nikliński, J.; Žarković, N.; Žarković, K.; Waeg, G.; Łuczaj, W.; Charkiewicz, R.; Skrzydlewska, E. Lipid mediators involved in the oxidative stress and antioxidant defense of human lung cancer cells. *Redox Biol.* 2016, 9, 210–219. [CrossRef]
- 57. Barsukov, V.Y.; Plokhov, V.N.; Tchesnokova, N.P. Activation of lipid peroxidation as typical cellular disintegration process in neoplastic area in breast cancer. *Vestn. VolGMU* **2007**, *3*, 3–6.
- 58. Cossetti, R.J.; Tyldesley, S.K.; Speers, C.H.; Zheng, Y.; Gelmon, K.A. Comparison of breast cancer recurrence and outcome patterns between patients treated from 1986 to 1992 and from 2004 to 2008. *J. Clin. Oncol.* **2015**, *33*, 65–73. [CrossRef]
- 59. Morrow, M. Personalizing extent of breast cancer surgery according to molecular subtypes. Breast 2013, 22, S106–S109. [CrossRef]
- 60. Pilewskie, M.; King, T.A. Age and molecular subtypes: Impact on surgical decisions. J. Surg. Oncol. 2014, 110, 8–14. [CrossRef]
- 61. Mersin, H.; Gülben, K.; Berberoğlu, U.; Yazi, M.; Acun, G.; Kinaş, V.; Erdoğan, S. Prognostic factors affecting postmastectomy locoregional recurrence in patients with early breast cancer: Are intrinsic subtypes effective? *World J. Surg.* **2011**, *35*, 2196–2202. [CrossRef] [PubMed]
- 62. Sabatier, R.; Finetti, P.; Cervera, N.; Tallet, A.; Benchalal, M.; Houvenaeghel, G.; Jacquemier, J.; Birnbaum, D.; Bertucci, F. Gene expression profiling and its utility in prediction of local relapse after breast-conserving therapy in early breast cancer. *Cancer Genom. Proteom.* **2011**, *8*, 199–209.
- Roka, S.; Rudas, M.; Taucher, S.; Dubsky, P.; Bachleitner-Hofmann, T.; Kandioler, D.; Gnant, M.; Jakesz, R. High nuclear grade and negative estrogen receptor are significant risk factors for recurrence in DCIS. *Eur. J. Surg. Oncol.* 2004, 30, 243–247. [CrossRef] [PubMed]
- 64. Ilić, I.R.; Stojanović, N.M.; Radulović, N.S.; Živković, V.V.; Randjelović, P.J.; Petrović, A.S.; Božić, M.; Ilić, R.S. The Quantitative ER Immunohistochemical Analysis in Breast Cancer: Detecting the 3 + 0, 4 + 0, and 5 + 0 Allred Score Cases. *Medicina* 2019, 55, 461. [CrossRef] [PubMed]
- 65. Human Epidermal Growth Factor Receptor 2 Testing in Breast Cancer: American Society of Clinical Oncology/College of American Pathologists Clinical Practice Guideline Focused Update. J. Clin. Oncol. 2018, 36, 2105–2122. [CrossRef]
- 66. Bel'skaya, L.V.; Kosenok, V.K.; Sarf, E.A. Chronophysiological features of the normal mineral composition of human saliva. *Arch. Oral Biol.* **2017**, *82*, 286–292. [CrossRef]