



Barley Melanoidins: Key Dietary Compounds With Potential Health Benefits

Jitendra Kumar Sharma^{1†}, Monika Sihmar^{1†}, Anita Rani Santal^{2†}, Louis Prager^{3†}, Franck Carbonero^{4†} and Nater Pal Singh^{1*†}

¹ Centre for Biotechnology, Maharshi Dayanand University, Rohtak, India, ² Department of Microbiology, Maharshi Dayanand University, Rohtak, India, ³ Department of Crop and Soil Science, College of Agricultural, Human, and Natural Resource Sciences, Washington State University, Pullman, WA, United States, ⁴ Department of Nutrition and Exercise Physiology, Elson Floyd College of Medicine, Washington State University, Spokane, WA, United States

OPEN ACCESS

Edited by:

Amélia M. Silva, University of Trás-os-Montes and Alto Douro, Portugal

Reviewed by:

José Ángel Rufián Henares, University of Granada, Spain Salvatore Parisi, Lourdes Matha Institute of Hotel Management and Catering Technology, India Juana Ines Mosele, University of Buenos Aires, Argentina

*Correspondence:

Nater Pal Singh npsinghcbt@gmail.com; npsingh.cbt@mdurohtak.ac.in

[†]These authors have contributed equally to this work

Specialty section:

This article was submitted to Food Chemistry, a section of the journal Frontiers in Nutrition

Received: 11 May 2021 Accepted: 30 August 2021 Published: 28 September 2021

Citation:

Sharma JK, Sihmar M, Santal AR, Prager L, Carbonero F and Singh NP (2021) Barley Melanoidins: Key Dietary Compounds With Potential Health Benefits. Front. Nutr. 8:708194. doi: 10.3389/fnut.2021.708194 This paper is a review of the potential health benefits of barley melanoidins. Food melanoidins are still rather understudied, despite their potential antioxidant, antimicrobial, and prebiotic properties. Free radicals are villainous substances in humans produced as metabolic byproducts and causing cancers and cardiovascular diseases, and the melanoidins alleviate the effects of these free radicals. Malt is produced from cereal grains such as barley, wheat, and maize, and barley is predominantly used in beer production. Beer (alcoholic and non-alcoholic) is a widely consumed beverage worldwide and a good source of dietary melanoidins, which enhance the beers' flavor, texture, and sensorial properties. Melanoidins, the final products of the Maillard reaction, are produced at different stages during the brewing process. Beer melanoidins protect the cells from oxidative damage of DNA. The high reducing capacity of melanoidins can induce hydroxyl radicals from H_2O_2 in the presence of ferric ion (Fe³⁺). Melanoidins inhibit lipid peroxidation during digestion due to their chelating metal property. However, lower digestibility of melanoidins leads to less availability to the organisms but is considered to function as dietary fiber that can be metabolized by the lower gut microbiota and possibly incur prebiotic properties. Melanoidins promote the growth of Lactobacilli and Bifidobacteria in the gastrointestinal tract, preventing the colonization of potential pathogens. Barley is already popular through beer production and increasingly as a functional food. Considering this economic and industrial importance, more research to explore the chemical properties of barley melanoidins and corresponding health benefits as barley is warranted.

Keywords: barley, melanoidins, health benefits, gut microbiome, antioxidant

INTRODUCTION

Barley is the fourth most important cereal worldwide, used as human food and animal feed, and the most common raw material in the brewing industry. It is rich in soluble dietary fiber, specifically beta-glucan-like oats. Potential health benefits of barley have been reported, including against diabetes, hypertension, obesity, and colon inflammation (1-3). The health benefits of barley grains can be enhanced by heat treatment of barley grains during their processing. Heat treatment facilitates the formation of melanoidins (4, 5). Melanoidins are brown-colored, polymeric high

1

Melanoidins structure	References
Melanoidins formed by polycondensation reaction of units of furan and pyrroles.	(13)
Skeleton of melanoidins formed at an early stage of MR by sugar degradation, then polymerization and linked by amino compounds.	(14)
Crosslink between free amino groups of lysine or arginine in proteins.	(15)
Skeleton of melanoidins formed by crosslinking of MR products and proteins.	(16)

molecular weight (HMW) compounds found in thermally processed foods and are the end product of the Maillard reaction (MR) (6). The MR, a non-enzymatic browning reaction, occurs between reducing sugars and free amino acids and peptides at high temperatures and produces melanoidins (7, 8). Usually, food or food products are processed at high temperatures for cooking, baking, roasting, frying, or even sterilization. The temperature ranged from 90 to 220°C. At this high temperature, MR, a Non-enzymatic browning reaction, occurs (9). In the formation of melanoidins, a brown nitrogenous polymer and copolymer, an array of reactions include cyclizations, dehydrations, retro aldolization, rearrangements, isomerizations, and condensations take place (9). The chemical structure of barley melanoidins is very complex and largely unknown because polymerization of complex melanoidins is influenced by various factors such as starting materials and their concentration, reaction conditions, such as pH, water activity, temperature, and reaction time, and solvent used (10-12). In order to predict the chemical structure of melanoidins, some recommendations are listed in Table 1.

Melanoidins are classified based on their skeleton composition. The skeleton of melanosaccharides is composed of polysaccharides, whereas melanoproteins are composed of proteins (17, 18). Melanosaccharides compounds are negatively charged, readily soluble in water developed from amino acids and polysaccharides (19). Melanoproteins have resulted when proteins and sugars of protein-rich food are cross-linking with each other. Melanoproteins are usually water-insoluble and contain HMW molecules (18).

In beverages like coffee and beer, melanosaccharides are the predominant melanoidins, whereas melanoproteins are predominant in bakery products like bread and biscuits (20). Since melanoidins are complex and diverse molecules, they are poorly soluble in water and organic solvents, making it hard to define their specific properties (21). However, the properties of melanoidins are greatly influenced by the low molecular weight (LMW) compounds (<10 kDa), which polymerize during MR and form HMW melanoidins (>300 kDa) (22). Also, the properties of melanoidins depend on the time and temperature applied during the brewing process (23).

Several studies were conducted to investigate the health benefits, daily dietary intake of melanoidins in different populations. For example, Papetti et al. (5) investigated the antioxidant activity of melanoidins isolated from roasted barley solution/barley coffee and reported that HMW melanoidins are associated with antioxidant activity. Daily dietary intake of melanoidins in different populations varies (**Figure 1**). For example, in the Spanish population, the daily dietary intake of melanoidins is 12.2 g/person/day. Therefore, it contributes 20% of the total antioxidant capacity intake, while in the Brazilian population, the daily dietary intake of melanoidins is 10.7 g/person/day and contributes around 21% of dietary antioxidant capacity intake (18, 19).

Several studies reported that melanoidins prevent lipid peroxidation, oxidative damage of DNA, and have antimicrobial, antihypertensive, antiallergenic, and prebiotic properties (**Figure 2**) (9, 24). The present review will focus on the health benefits of melanoidins, especially from barley sources. It is very important to conduct more research to explore the chemical properties of barley melanoidins and their corresponding health properties as barley has economic and industrial importance.

BARLEY SOURCES OF MELANOIDINS

Barley Malt

Based on color, barley malts can be categorized into pale and dark malt. In the production of beer, pale malt is commonly used as the main ingredient. Malting of barley grains is achieved in three steps, steeping, germination, and kilning. The malting process begins with steeping raw grain underwater to bring moisture content up from 12 to (42-46%). The first steeping lasts between 6 and 16 h and brings the grain up to 33-37% moisture content. Next, the water is removed from the grain bed, removing any moisture film and carbon dioxide produced during respiration. The grain then goes through an air rest for 12-24 h, exposing the embryo to oxygen. Next, the grain is immersed in the water again for 10-20 h to bring the final moisture content of 42-46% (25, 26). After steeping, the hydrated grain or green malt is stored in a well-ventilated humid area kept between 14 and 20°C. Enzymes are activated and begin degrading cell walls, proteins, and starches. The green malt is kept in this germination stage for 4-6 days, during which some moisture is lost and replaced by spraying the green malt with water. Germination is measured by the rootlets that begin to grow, and once the rootlets are between 1.5 and 2 times, the germination period is complete (25, 26).

Kilning or roasting is the final step in the malting process. Here the green malt is thermally processed, drying the malt to a moisture content of \sim 5%. This step inactivates and preserves the enzymes and ensures stability for the storage and transportation of the finished malt. The first phase is referred to as the "whitering" or "free drying" phase. Next, the grain is air-dried at 25°C, decreasing the moisture content from 44 to 12%. The next phase is a much longer process taking the malt from 12% moisture content to 4%. This is referred to as the "falling rate phase." After the grain is dried, the final step is an increase in temperature, initiating the "curing stage" followed by a cooling period and finalized with the packaging of the final product (25, 26). It is important to note that the kilning and roasting





step contributes the most to the color and flavor profiles of the malted barley (27). Dark malt is further classified into color brew malts, caramel malts, and roasted malts, and their production is dependent on the temperature on which they are produced. Malt produced from roasted barley can be classified into Pale malt (products include amber, chocolate, and black malt), green malt (products include cara, crystal, dark crystal, caramel malt), and colored kilned malts (includes Munich malt and Vienna malt) (26). Temperature up to 105° C is used to produce color brew malt, while caramel malts and roasted malts are up to 160 and 220–250°C for 2–2.5 h, respectively (28).

Beer Melanoidins

Various types of beers are produced and consumed worldwide. They differ from each other in color, flavor, and alcohol content, including ale, lager, porter, stout, blonde ale, brown ale, pale ale, India pale ale pilsner, etc. Melanoidins influenced beer characteristics, such as color, flavor, and body of the beer despite having positive effects on health (23). The type of beers melanoidins influence content in the beer and ranges between 0.06 and 10.3 g/100 ml (29, 30).

The quantification of melanoidins, phenolic compounds and sulfur dioxide, as well as the assessment of antioxidant activity of

40 lager beer was performed by Zhao et al. (30). They reported 8-fold variation in the melanoidins content of beer which ranged from 1.64 to 13.71 g/L. The variation in melanoidins content of different beers occurred due to differences in the raw material used and the brewing process. Rivero et al. (29) compared the melanoidins content of dark, blond, and alcohol-free beer and reported the highest level of melanoidins in dark beer with values of 1.49 g/L than blond beer (0.61 g/L) and alcohol-free beer (0.58 g/L). Tagliazucchi and Verzelloni (31) analyzed dark beer and found melanoidins content of 12.3 g/L.

Beer melanoidins intake is depending on the consumption of beer in the populations and data varying between countries. Beer consumption per capita in different countries is different. In 2011, Ireland was on top in per capita beer consumption per year that was 138 L, and this consumption was 1.7 times greater than consumed by a person per year in the United States, while beer consumed by the Czech Republican population was almost equal to the Ireland population (32). As discussed above, beer melanoidins have positive effects on human health, melanoidins from the dark beer demonstrated antioxidant activity and may be used to prevent oxidative damage. In addition, dark beer melanoidins might inhibit lipid peroxidation during gastric digestion and prevent the formation of secondary lipoxidation products, which have adverse health effects, like atherosclerosis (33).

HEALTH BENEFITS OF MELANOIDINS

Melanoidins provide a health benefit to human health in various ways. Melanoidins can trap positively charged ion species (electrophiles), scavenge free radicals, and also have reducing power; chelating metal ions lessen oxidative damage (34). Antioxidative capacity of melanoidins can be assayed using DPPH (2,2-diphenyl-1-picryl-hydrazyl-hydrate) assay for free radical scavenging activity ABTS+ (2,2-azinobis-3-ethylbenzothiazoline-6-sulfonate) assay for antioxidant activity, and Ferric Reducing Antioxidant Power (FRAP) assay for ferric reducing ability (21). The occurrence of free ions of iron (Fe²⁺)/ free heme (HmFeIII) groups/hemecontaining peptides like metmyoglobin in gastric fluid catalyzes lipid peroxidation (33). Reactive oxygen species (ROS) induces oxidative damage to the DNA, and the melanoidins can counter this harmful effect of ROS (23). Barley melanoidins acting as dietary fiber and increases the growth of beneficial bacteria. Prebiotic properties of melanoidins can be assessed by quantifying Fecal Short-Chain Fatty Acids (SCFAs) (35). Barley sources of melanoidins and their biological activities and assaying methods are listed in Table 2.

Melanoidins as Antioxidants

Maillard reaction products (MRPs), melanoidins, contribute antioxidant capacity to the food products. For example, wheat bread exhibits increased antioxidant capacity by adding barley fiber with wheat flour, and baking enhances the antioxidant property (41). Heat treatment of the malt produces melanoidins and has antioxidant properties, resulting from reducing sugars, and amino acids or proteins (42). The strong reducing capacity of melanoidins proves substantial health-promoting effects (9, 22). Zhao et al. (30) studied the antioxidant activity of lager beer and reported a positive correlation between antioxidant property and melanoidins content (p < 0.05), also phenolic content correlated with the antioxidant activity. Morales (43) evaluated the hydroxyl radical scavenging potential of three different European beers: dry-stout beer, abbeys style beer, and pilsener beer. The abbeys-style beer and pilsener beer.

In India, barley is commonly consumed as "Sattu" prepared by roasting barley grains in sand heated to 250-300°C for a short time, removing the husk, and then ground to flour. Roasting is attributed to the increased antioxidant potential of barley grains and their products. In roasted barley, there are high indicators of antioxidant activity such as metal chelating activity, radical scavenging capacity, and reducing power (44, 45). Carvalho et al. (22) studied the antioxidant potential of three different barley malt produced at different kilning temperatures, 80-85°C for pale malt, 130°C for melano, 80 230°C for black malt. They assayed the antiradical potential and reducing the power of these three barley malt using metmyoglobin assay, deoxyribose assay, and FRAP assay, respectively. High reducing the power of melanoidins can induce conversion of H₂O₂ into •OH (hydroxyl radical) in the presence of Fe³⁺ ion. Black malt showed high reducing properties than melano 80 and pale malt. Also, black malt's radical scavenging capacity is higher than melano 80, and pale malt in metmyoglobin assay and deoxyribose assay result is reciprocal of metmyoglobin assay (22). Pastoriza and Rufián-Henares (19) have been investigated the contribution of melanoidins to the antioxidant capacity of the Spanish diet. They reported 8.7 and 15.0 g/100 mL in Pilsen and black beers, respectively (19). Alves et al. (18) studied the various heatprocessed food consumed in the Brazilian diet. On average, 10.7 g of melanoidins are consumed daily by this population and contribute up to 21% of the dietary antioxidant capacity of the Brazilian population (18). Antioxidant properties of food items consumed were evaluated using FRAP and TEAC (Trolox Equivalent Antioxidant Capacity) assay. In dark and blonde beer, monosaccharides contents were 4.2 \pm 0.6 and 2.0 \pm 1.3 g/100 mL, respectively (18). Brewers spent grains (barley) or distilled spent grains (sorghum) are high in moisture and can easily get rotten and cause an environmental problem. However, these byproducts are used in animal feed production or used for biogas production (46). Nonetheless, brewers spent grains or distilled spent grains are rich in melanoidins and established for their potential of antioxidant activities. Spent grains of barley exhibit three times more antioxidant capacity than sorghum spent grain (46, 47). Piggott and coworkers evaluated brewers' antioxidant activity and found greater antioxidant potential in black spent grain than pale spent grain. Ultra filtrate fraction of black spent grain, HMW fraction (>100 kDa) dominate over LMW (<kDa) fraction in antioxidant activity and metal chelating ability, and these properties are attributed by the melanoidins present in HMW fraction (47).

TABLE 2 | Methodology and source of melanoidins used in the assessment of biological activities.

Assessment of activity	Barley melanoidins source	Methodology	References
Lipid peroxidation	Barley coffee and dark beer	Radical-Scavenging Activity by ABTS assay, Lipid Hydroperoxides (LHP) Measurements by FOX assay.	(33)
Antioxidant activity	Beer melanoidins	ABTS radical cation scavenging activity, DPPH radical scavenging activity, Oxygen radical absorbance capacity assay	(30)
Antioxidant activity	Dark and blonde beer	FRAP and TEAC assays	(18)
DNA damage, Antioxidant activity	Lager beers	Folin–Ciocalteus reaction, catechin content by vanillin reaction, DMPD (N, N-dimethyl-p-phenylenediamine) assay	(29)
DNA damage, Antioxidative capacity	Barley flour	DPPH radical scavenging activity.	(36)
Antioxidant activity	Dark beer, barley coffee,	Radical scavenging activity- ABTS assay	(31)
Antioxidant activity, antiradical activity	Black beer and pilsner beer	ABTS assay, DPPH assay (antiradical activity), FRAP assay (ferric-reducing ability), Oxygen Radical Absorbance Capacity (ORAC) assay, HOSC assay (hydroxyl radical-scavenging capacity)	(19)
Antioxidant activity	Roasted barley	DPPH Assay, Linoleic Acid-β-Carotene Assay	(5)
Prebiotic potential	Melanoidins-rich barley malts	Fecal SCFAs Quantification	(35)
Antimicrobial activity	Barley coffee	Microtiter plate assay for <i>Streptococcus mutans</i> Biofilm	(37)
Antimicrobial activity	Barley coffee	Minimum inhibitory and bactericidal concentration assay for <i>Streptococcus mutans</i> Biofilm	(38)
Antimicrobial activity	Pilsner-style beer, Abbeys-style beer, and dry-stout beer	Antimicrobial assay for E. coli and S. aureus	(39)
Antioxidant activity	Brewers' spent grains	DPPH and FR assay	(40)

Melanoidins as an Inhibitor of Lipid Peroxidation

The elevated production of ROS leads to tissue dysfunction and is associated with various pathophysiologies. Lipid peroxidation by ROS produces peroxides and aldehydes. Tissues that are far distant from the site of aldehyde generation are also damaged because these aldehydes are highly stable than parent ROS and can quickly diffuse. The lipid-derived products by ROS are highly reactive and cause noticeable biological effects. Various ailments in humans are caused by these lipid peroxidation products, such as atherosclerosis, reperfusion injury, coronary artery disease, Alzheimer's, Rheumatoid arthritis, and tumors, neurodegenerative diseases, and other disorders. Lipid peroxidation also causes damage to protein and DNA, changes in cell signaling (48-50). Modern-day foods are rich in oxidized and oxidizable lipids, especially fast foods, which elevated lipid hydroperoxides in plasma. Increased plasma lipid hydroperoxides can be integrated into lipoproteins and resulted in lipoprotein oxidation (51, 52). Absorption of malondialdehyde (MDA) and 4-hydroxynonenal in the gastrointestinal tract can cause the pathogenesis of cardiovascular diseases (53).

Various studies and reviews on melanoidins and their antilipid peroxidation role are reported. Dietary melanoidins can mitigate the adverse effects of lipid peroxidation products mainly by two strategies (i) melanoidins possibly hinder the lipid peroxidation during digestion of lipid-rich food, and (ii) inhibit the production of secondary lipoxidation products (33). Metal chelating ability of barley coffee, dark beer, and coffee melanoidins proves positive health benefits, chelation of metals during gastric digestion inhibits lipid peroxidation. On the other hand, consumption of red meat renders the heme in circulation, which catalyzes oxidative damage, although melanoidins bind with heme contained in meat and check its absorption in the gastrointestinal tract (33).

Melanoidins Prevent Oxidative Damage of DNA

Cellular structures and molecules are vulnerable to reactive oxygen species, damaging lipid, protein, or DNA. ROS can cause oxidative damage to the DNA in many ways, such as a base lesion, single-strand breaks, double-strand breaks, cross-linking DNA and protein, and chromosomal breakage rearrangement (29, 54–56). Hydroxyl radical (•OH), generated during the reaction between hydro-peroxides and transition metal ions, is very reactive can assail DNA, proteins, and fatty acids (polyunsaturated) (57). An increased risk of the onset of cancer is highly associated with alcohol consumption. Processing alcohol inside the body generated acetaldehyde, which belongs to group1 carcinogen, which interferes with the DNA repair (58). ROS can attack and modify the guanine base of the DNA into 8-hydroxydeoxyguanosine (8-OHdG). During replication, 8-OHdG is paired with thymine or adenine base instead of cytosine, and eventually, accumulation of this transverse mutation in DNA has a detrimental effect on cells or tissue (59). Rivero et al. (29) signifying the beer melanoidins, which can effectively combat the oxidative stress to the cells. The level of 8-OHdG and DNA damage are negatively correlated with the concentration of melanoidins in beer, especially dark beer (29). However, melanoidins consist of high molecular weight compounds and cannot cross the cellular barrier. Morales et al. (60) suggest that intestinal digestion cleaved melanoidins into an absorbable compound with antioxidant activity.

Barley Melanoidins and the Gut Microbiome

Dietary Maillard Reaction Products (MRP) has been studied sparsely when it comes to their impact on the gut microbiome and their fate in the lower digestive tract (61), while there is slightly more information published on their precursors, such as the Advanced glycation end products (AGEs) (62-64). Even if their complex and diverse structure make it difficult to draw clear predictions, it is widely thought that a significant portion of dietary melanoidins pass through the upper digestive tract and become available for fermentative metabolism by the gut microbiome (65). This model mostly derives from animal studies. For example, Helou et al. (66) used different bread models and confirmed that a significant amount of dietary melanoidins were excreted in rat's feces. Unlike many other fermentable dietary compounds, very little is known about the microbially derived metabolites from dietary melanoidins. N ɛcarboxymethyl-lysine (CML) has been used as a biomarker of dietary and endogenous MRP intake (67). The impact of CML and other MRPs on the growth of different bacterial strains has earlier been studied (68). However, it is not clear if CML can be considered a biomarker of microbiome metabolic activities. There has been no published report on metabolomics data from dietary melanoidins, while there have been recent reports of metabolites associated with other MRPs (69-71). As expected, protein and carbohydrates derivatives were identified as likely by-products of gut microbiome metabolism. Therefore, it can be hypothesized that melanoidins would be degraded into smaller MRPs than similar metabolites, but exploratory metabolomics studies are needed to identify microbially-derived metabolites.

In the absence of baseline knowledge about the microbial metabolism of melanoidins, most studies have focused on their effect on the microbiota. One of the earliest studies relied on *in vitro* small intestine and gut microbiota batch fermentation (72). Peptic and pancreatic digestion did not affect the model melanoidin, supporting the hypothesis that they are mostly non-digestible. On the other hand, melanoidins stimulated the overall growth of anaerobic gut bacteria, suggesting that gut microbiota members can metabolize them to gain energy (72). It was further demonstrated that native dietary melanoidins could be excreted in feces but very low amounts in urine (66, 73). These observations strengthen the hypothesis that microbiota can utilize melanoidins. Based on melanoidins physical similarity with dietary fibers (74) and their apparent fermentability by the gut microbiota, melanoidins have subsequently been proposed

analogous to dietary fiber and, thereby, potential prebiotics (75). In this study, bread melanoidins were submitted to an anaerobic batch culture, and stimulation of Bifidobacteria was observed.

It should be noted that the ability of a food extract to stimulate the growth of so-called prebiotic species cannot be used as conclusive evidence of a prebiotic or even prebiotic-like effect. Indeed, when similar *in vitro* fermentation experiments were conducted with other bread melanoidins products, bifidogenic effects or the *Lactobacillus* stimulation were subject-specific and not across the board (76). The inter individuality in gut microbiota response has been commonly described (77, 78) and should not be considered as contradicting the potential dietary fiber or prebiotic effect, but it also emphasizes the need for more evidence.

In the context of barley melanoidins, it should be stated that barley dietary fiber and specifically beta-glucan, can be expected to exert beneficial prebiotic-like effects (79-81). Intriguingly, some of the initial studies on barley melanoidins have focused on their antimicrobial properties. For example, barley coffee and the melanoidin fraction specifically inhibited Streptococcus mutans biofilm formation (37, 38). Furthermore, potential antioxidant effects from barley and other dietary melanoidins were ascribed to the gastrointestinal tract in general and possibly mediated by the gut microbiota (33, 82, 83). More recently, we conducted an animal study to test the hypothesis of barley melanoidins acting as dietary fiber (35). We used an increasing proportion of melanoidin-rich malted barley combined with simple malted barley (0, 25, 50, 75, and 100%) fed ad libitum to mice over 3 weeks. Mice gut microbiota was significantly clustered by dietary treatment, and many bacterial taxa were differentially affected.

Interestingly, *Lactobacillus* responded slower to the treatment with melanoidins, while Bacteroidetes were the most responsive after 7 days. *Bifidobacterium* and *Akkermansia*, two genera considered beneficial, were stimulated by barley melanoidins, especially toward the end of the experiment. The almost immediate increase in Bacteroidetes supports the model of barley melanoidins acting like dietary fiber. However, it appears that any prebiotic-like can only be expected after a relatively long adaptation period from the gut microbiota. It can be hypothesized that this delayed effect is due to the melanoidins structure being even more complex than dietary fiber and the need for cellulose and other fermentative consortia (84, 85) to such a new energy source.

NEGATIVE HEALTH IMPACT OF MELANOIDINS

Melanoidins have been extensively investigated for their health beneficial effects, and reports are signifying their other health beneficial properties. The beneficial health properties of food melanoidins are associated with various activities such as antioxidant, antimicrobial, anti-inflammatory, antihypertensive, and prebiotic activity (35, 86, 87). However, very few studies on the negative impacts of melanoidins on health are reported, an extensive review has been done on the impact of MRPs on nutrition and health by ALjahdali and Carbonero (61). MRPs affects the bioavailability of trace elements in rat magnesium digestion is reduced in the presence of MRPs. Nutritional quality of foods and protein digestibility is also reported to be reduced under the influence of MRPs (61, 65, 88). Diabetes, cardiovascular diseases, and even carcinogenetic effects are also associated with MRPs. High dietary carboxymethyl lysine (CML) promotes diabetes and cardiovascular diseases, whereas acrylamide inciting tumors (9, 89). Melanoidins are recalcitrant molecules and hard to biodegrade by microorganisms; industrial wastes of breweries and distilleries are rich in melanoidins, act as pollutants to the environment, and ultimately affect human health (90–92).

CONCLUSION

People's food habits and living habits lead them to obesity, diabetes, hypertension, and gastrointestinal diseases, cancer, and others in the current situation. Today's need is to improve our food habits and incorporate such food materials in our daily diet to reduce the conditions mentioned above. Various

REFERENCES

- Farag MA, Xiao J, Abdallah HM. Nutritional value of barley cereal and better opportunities for its processing as a value-added food: a comprehensive review. *Crit Rev Food Sci Nutr.* (2020) 1–13. doi: 10.1080/10408398.2020.1835817
- 2. Goudar G, Sharma P, Janghu S, Longvah T. Effect of processing on barley β -glucan content, its molecular weight and extractability. *Int J Biol Macromol.* (2020) 162:1204–16. doi: 10.1016/j.ijbiomac.2020.06.208
- Lin S, Guo H, Gong JDB, Lu M, Lu M-Y, Wang LU, et al. Phenolic profiles, βglucan contents, and antioxidant capacities of colored Qingke (Tibetan hulless barley) cultivars. J Cereal Sci. (2018) 81:69–75. doi: 10.1016/j.jcs.2018.04.001
- Milic BL, Grujic-Injac B, Piletic MV, Lajsic S, Kolarov LA. Melanoidins and carbohydrates in roasted barley. J Agric Food Chem. (1975) 23:960– 3. doi: 10.1021/jf60201a031
- Papetti A, Daglia M, Aceti C, Quaglia M, Gregotti C, Gazzani G. Isolation of an *in vitro* and *ex vivo* antiradical melanoidin from roasted barley. *J Agric Food Chem.* (2006) 54:1209–16. doi: 10.1021/jf058133x
- Fogliano V, Morales FJ. Estimation of dietary intake of melanoidins from coffee and bread. *Food Funct*. (2011) 2:117–23. doi: 10.1039/c0fo00156b
- Moreira AS, Nunes FM, Simões C, Maciel E, Domingues P, Domingues MRM, et al. Transglycosylation reactions, a main mechanism of phenolics incorporation in coffee melanoidins: inhibition by Maillard reaction. *Food Chem.* (2017) 227:422–31. doi: 10.1016/j.foodchem.2017. 01.107
- Bertrand E, El Boustany P, Faulds C, Berdagué J-L. The Maillard reaction in food: an introduction. *Reference Module Food Science Elsevier*. (2018) 2018:9780081005965. doi: 10.1016/B978-0-08-100596-5.21459-5
- Echavarría AP, Pagán J, Ibarz A. Melanoidins formed by Maillard reaction in food and their biological activity. *Food Eng Rev.* (2012) 4:203– 23. doi: 10.1007/s12393-012-9057-9
- Fay LB, Brevard H. Contribution of mass spectrometry to the study of the Maillard reaction in food. *Mass Spectrom Rev.* (2005) 24:487– 507. doi: 10.1002/mas.20028
- Benzing-Purdie LM, Ripmeester JA, Ratcliffe CI. Effects of temperature on Maillard reaction products. J Agric Food Chem. (1985) 33:31– 3. doi: 10.1021/jf00061a009
- Ekielski A, Mishra PK, Zelaziński T. Assessing the influence of roasting process parameters on mepiquat and chlormequat formation in dark barley malts. *Food Bioprocess Technol.* (2018) 11:1177–87. doi: 10.1007/s11947-018-2087-4

studies suggested that the intake of melanoidins associated with health benefitting properties. However, melanoidins structure's chemical characteristics are sparsely known. Barley as a source of melanoidins needs more attention because barley can be grown in marginal areas with low input, used mainly in malt and brewing industries, which produces many by-products containing melanoidins. Therefore, comprehensive studies are required to understand better the chemical structure and specific health-benefiting properties of food melanoidins.

AUTHOR CONTRIBUTIONS

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

ACKNOWLEDGMENTS

The Science and Engineering Research Board (SERB), New Delhi (SB/EMEQ-085/2014; SB/YS/LS-334/2013) and DST-FIST are highly acknowledged for lab support.

- Cämmerer B, Kroh LW. Investigation of the influence of reaction conditions on the elementary composition of melanoidins. *Food Chem.* (1995) 53:55– 9. doi: 10.1016/0308-8146(95)95786-6
- Tressl R, Wondrak GT, Garbe L-A, Krüger R-P, Rewicki D. Pentoses and hexoses as sources of new melanoidin-like Maillard polymers. J Agric Food Chem. (1998) 46:1765–76. doi: 10.1021/jf970973r
- Hofmann T. Studies on the relationship between molecular weight and the color potency of fractions obtained by thermal treatment of glucose/amino acid and glucose/protein solutions by using ultracentrifugation and color dilution techniques. J Agric Food Chem. (1998) 46:3891– 5. doi: 10.1021/jf980397e
- Hofmann T, Bors W, Stettmaier K. Radical-assisted melanoidin formation during thermal processing of foods as well as under physiological conditions. *J Agric Food Chem.* (1999) 47:391–6. doi: 10.1021/jf980627p
- Cämmerer B, Jalyschko W, Kroh LW. Intact carbohydrate structures as part of the melanoidin skeleton. J Agric Food Chem. (2002) 50:2083– 7. doi: 10.1021/jf011106w
- Alves G, Xavier P, Limoeiro R, Perrone D. Contribution of melanoidins from heat-processed foods to the phenolic compound intake and antioxidant capacity of the Brazilian diet. J Food Sci Technol. (2020) 57:3119– 31. doi: 10.1007/s13197-020-04346-0
- Pastoriza S, Rufián-Henares JA. Contribution of melanoidins to the antioxidant capacity of the Spanish diet. *Food Chem.* (2014) 164:438– 45. doi: 10.1016/j.foodchem.2014.04.118
- Alves G, Lobo LA, Domingues RMCP, Monteiro M, Perrone D. Bioaccessibility and gut metabolism of free and melanoidin-bound phenolic compounds from coffee and bread. *Front Nutr.* (2021) 8:708928. doi: 10.3389/fnut.2021.708928
- Langner E, Rzeski W. Biological properties of melanoidins: a review. Int J Food Prop. (2014) 17:344–53. doi: 10.1080/10942912.2011.631253
- Carvalho DO, Correia E, Lopes L, Guido LF. Further insights into the role of melanoidins on the antioxidant potential of barley malt. *Food Chem.* (2014) 160:127–33. doi: 10.1016/j.foodchem.2014.03.074
- 23. Martinez-Gomez A, Caballero I, Blanco CA. Phenols and melanoidins as natural antioxidants in beer. Structure, reactivity and antioxidant activity. *Biomolecules*. (2020) 10:400. doi: 10.3390/biom10030400
- 24. Olas B, Bryś M. Beer components and their beneficial effect on the hemostasis and cardiovascular diseases-truth or falsehood. *Food Chem Toxicol.* (2020) 146:111782. doi: 10.1016/j.fct.2020.111782
- Briggs DE, Hough JS, Stevens R, Young TW. Malting and Brewing Science: Malt and Sweet Wort. Springer Science & Business Media (1981).

- Carvalho DO, Gonçalves LM, Guido LF. Overall antioxidant properties of malt and how they are influenced by the individual constituents of barley and the malting process. *Compr Rev Food Sci Food Saf.* (2016) 15:927– 43. doi: 10.1111/1541-4337.12218
- Yahya H, Linforth RS, Cook DJ. Flavour generation during commercial barley and malt roasting operations: a time course study. *Food Chem.* (2014) 145:378–87. doi: 10.1016/j.foodchem.2013.08.046
- Carvalho DO, Øgendal LH, Andersen ML, Guido LF. High molecular weight compounds generated by roasting barley malt are pro-oxidants in metal-catalyzed oxidations. *Eur Food Res Technol.* (2016) 242:1545– 53. doi: 10.1007/s00217-016-2655-7
- Rivero D, Pérez-Magariño S, González-Sanjosé ML, Valls-Belles V, Codoñer P, Muñiz P. Inhibition of induced DNA oxidative damage by beers: correlation with the content of polyphenols and melanoidins. *J Agric Food Chem.* (2005) 53:3637–42. doi: 10.1021/jf048146v
- Zhao H, Li H, Sun G, Yang B, Zhao M. Assessment of endogenous antioxidative compounds and antioxidant activities of lager beers. J Sci Food Agric. (2013) 93:910–7. doi: 10.1002/jsfa.5824
- Tagliazucchi D, Verzelloni E. Relationship between the chemical composition and the biological activities of food melanoidins. *Food Sci Biotechnol.* (2014) 23:561–8. doi: 10.1007/s10068-014-0077-5
- Xie W, Xiong W, Pan J, Ali T, Cui Q, Guan D, et al. Decreases in global beer supply due to extreme drought and heat. *Nat Plants*. (2018) 4:964– 73. doi: 10.1038/s41477-018-0263-1
- Tagliazucchi D, Verzelloni E, Conte A. Effect of dietary melanoidins on lipid peroxidation during simulated gastric digestion: their possible role in the prevention of oxidative damage. J Agric Food Chem. (2010) 58:2513– 9. doi: 10.1021/jf903701h
- Mesías M, Delgado-Andrade C. Melanoidins as a potential functional food ingredient. *Curr Opin Food Sci.* (2017) 14:37– 42. doi: 10.1016/j.cofs.2017.01.007
- Aljahdali N, Gadonna-Widehem P, Anton PM, Carbonero F. Gut microbiota modulation by dietary barley malt melanoidins. *Nutrients*. (2020) 12:241. doi: 10.3390/nu12010241
- Baba WN, Rashid I, Shah A, Ahmad M, Gani A, Masoodi FA, et al. Effect of microwave roasting on antioxidant and anticancerous activities of barley flour. J Saudi Soc Agric Sci. (2016) 15:12–9. doi: 10.1016/j.jssas.2014. 06.003
- Stauder M, Papetti A, Daglia M, Vezzulli L, Gazzani G, Varaldo PE, et al. Inhibitory activity by barley coffee components towards Streptococcus mutans biofilm. *Curr Microbiol.* (2010) 61:417–21. doi: 10.1007/s00284-010-9630-5
- Papetti A, Pruzzo C, Daglia M, Grisoli P, Bacciaglia A, Repetto B, et al. Effect of barley coffee on the adhesive properties of oral streptococci. J Agric Food Chem. (2007) 55:278–84. doi: 10.1021/jf062090i
- Rufián-Henares JA, Morales FJ. Microtiter plate-based assay for screening antimicrobial activity of melanoidins against *E. coli* and *S. aureus. Food Chem.* (2008) 111:1069–74. doi: 10.1016/j.foodchem.2008.05.027
- Patrignani M, González-Forte L del S. Characterisation of melanoidins derived from Brewers' spent grain: new insights into their structure and antioxidant activity. *Int J Food Sci Technol.* (2021) 56:384–91. doi: 10.1111/ijfs.14653
- Ragaee S, Guzar I, Dhull N, Seetharaman K. Effects of fiber addition on antioxidant capacity and nutritional quality of wheat bread. *LWT-Food Sci Technol.* (2011) 44:2147–53. doi: 10.1016/j.lwt.2011.06.016
- Gasior J, Kawa-Rygielska J, Kucharska AZ. Carbohydrates profile, polyphenols content and antioxidative properties of beer worts produced with different dark malts varieties or roasted barley grains. *Molecules*. (2020) 25:3882. doi: 10.3390/molecules25173882
- Morales FJ. Non-specific hydroxyl radical scavenging properties of melanoidins from beer. In: Preedy VR, editors. *Beer in Health and Disease Prevention*. Elsevier. p. 765–74. doi: 10.1016/B978-0-12-373891-2.00078-X
- Sharma P, Gujral HS. Effect of sand roasting and microwave cooking on antioxidant activity of barley. *Food Res Int.* (2011) 44:235–40. doi: 10.1016/j.foodres.2010.10.030
- Omwamba M, Li F, Sun G, Hu Q. Antioxidant effect of roasted barley (Hordeum vulgare L.) grain extract towards oxidative stress in vitro and in vivo. Food Nutr Sci. (2013) 4:139–146. doi: 10.4236/fns.2013.48A017

- 46. Yang S, Fan W, Xu Y. Melanoidins from Chinese distilled spent grain: content, preliminary structure, antioxidant, and ACE-inhibitory activities *in vitro*. *Foods*. (2019) 8:516. doi: 10.3390/foods8100516
- Piggott CO, Connolly A, FitzGerald RJ. Application of ultrafiltration in the study of phenolic isolates and melanoidins from pale and black brewers' spent grain. *Int J Food Sci Technol.* (2014) 49:2252–9. doi: 10.1111/jifs.12540
- Mylonas C, Kouretas D. Lipid peroxidation and tissue damage. In Vivo. (1999) 13:295–309.
- Ramana KV, Srivastava S, Singhal SS. Lipid peroxidation products in human health and disease. Oxid Med Cell Longev. (2013) 2013:e583438. doi: 10.1155/2013/583438
- Petrovic S, Arsic A, Ristic-Medic D, Cvetkovic Z, Vucic V. Lipid peroxidation and antioxidant supplementation in neurodegenerative diseases: a review of human studies. *Antioxidants*. (2020) 9:1128. doi: 10.3390/antiox9111128
- Staprans I, Rapp JH, Pan X-M, Kim KY, Feingold KR. Oxidized lipids in the diet are a source of oxidized lipid in chylomicrons of human serum. *Arterioscler Thromb J Vasc Biol.* (1994) 14:1900– 5. doi: 10.1161/01.ATV.14.12.1900
- 52. Ursini F, Sevanian A. Postprandial oxidative stress. *Biol Chem.* (2002) 383:599–605. doi: 10.1515/BC.2002.062
- Uchida K. Role of reactive aldehyde in cardiovascular diseases. Free Radic Biol Med. (2000) 28:1685–96. doi: 10.1016/S0891-5849(00)00226-4
- 54. Simic MG, Jovanovic SV. Free radical mechanisms of DNA base damage. In: Simic MG, Grossman L, Upton AC, Bergtold DS, editors. *Mechanisms of DNA Damage and Repair*. Boston: Springer, 39–49. doi: 10.1007/978-1-4615-9462-8_5
- 55. Kaneko T, Tahara S, Matsuo M. Retarding effect of dietary restriction on the accumulation of 8-hydroxy-2'-deoxyguanosine in organs of Fischer 344 rats during aging. *Free Radic Biol Med.* (1997) 23:76–81. doi: 10.1016/S0891-5849(96)00622-3
- Hemnani T, Parihar M. Reactive oxygen species and oxidative DNA damage. Indian J Physiol Pharmacol. (1998) 42:440–52.
- Auroma O. Free radicals, antioxidants and international nutrition review. Asia Pac J Clin Nutr. (1999) 8:53–63. doi: 10.1046/j.1440-6047.1999.00036.x
- Brooks PJ, Zakhari S. Acetaldehyde and the genome: beyond nuclear DNA adducts and carcinogenesis. *Environ Mol Mutagen*. (2014) 55:77– 91. doi: 10.1002/em.21824
- Emam AN, Girgis E, Khalil WK, Mohamed MB. Toxicity of plasmonic nanomaterials and their hybrid nanocomposites. In: Fishbein JC, Heilman JM, editors. Advances in Molecular Toxicology. Elsevier. p. 173–202. doi: 10.1016/B978-0-444-63406-1.00005-2
- Morales FJ, Somoza V, Fogliano V. Physiological relevance of dietary melanoidins. *Amino Acids*. (2012) 42:1097– 109. doi: 10.1007/s00726-010-0774-1
- ALjahdali N, Carbonero F. Impact of Maillard reaction products on nutrition and health: Current knowledge and need to understand their fate in the human digestive system. *Crit Rev Food Sci Nutr.* (2019) 59:474– 87. doi: 10.1080/10408398.2017.1378865
- 62. Hellwig M, Auerbach C, Müller N, Samuel P, Kammann S, Beer F, et al. Metabolization of the advanced glycation end product N-εcarboxymethyllysine (CML) by different probiotic *E. coli* strains. *J Agric Food Chem.* (2019) 67:1963–72. doi: 10.1021/acs.jafc.8b06748
- Snelson M, Coughlan MT. Dietary advanced glycation end products: digestion, metabolism and modulation of gut microbial ecology. *Nutrients*. (2019) 11:215. doi: 10.3390/nu11020215
- Nie C, Li Y, Qian H, Ying H, Wang L. Advanced glycation end products in food and their effects on intestinal tract. *Crit Rev Food Sci Nutr.* (2020) 1–13. doi: 10.1080/10408398.2020.1863904
- Pérez-Burillo S, Rajakaruna S, Pastoriza S, Paliy O, Rufián-Henares JÁ. Bioactivity of food melanoidins is mediated by gut microbiota. *Food Chem.* (2020) 316:126309. doi: 10.1016/j.foodchem.2020.126309
- Helou C, Anton PM, Niquet-Leridon C, Spatz M, Tessier FJ, Gadonna-Widehem P. Fecal excretion of Maillard reaction products and the gut microbiota composition of rats fed with bread crust or bread crumb. *Food Funct*. (2017) 8:2722–30. doi: 10.1039/C7FO00430C
- Tessier FJ, Birlouez-Aragon I. Health effects of dietary Maillard reaction products: the results of ICARE and other studies. *Amino Acids*. (2012) 42:1119–31. doi: 10.1007/s00726-010-0776-z

- Helou C, Marier D, Jacolot P, Abdennebi-Najar L, Niquet-Léridon C, Tessier FJ, et al. Microorganisms and Maillard reaction products: a review of the literature and recent findings. *Amino Acids*. (2014) 46:267– 77. doi: 10.1007/s00726-013-1496-y
- 69. Qu W, Nie C, Zhao J, Ou X, Zhang Y, Yang S, et al. Microbiome–metabolomics analysis of the impacts of long-term dietary advanced-Glycation-end-product consumption on C57BL/6 mouse fecal microbiota and metabolites. J Agric Food Chem. (2018) 66:8864–75. doi: 10.1021/acs.jafc.8b01466
- Zhou X, Ulaszewska MM, Cuparencu C, De Gobba C, Vázquez-Manjarrez N, Gürdeniz G, et al. Urine metabolome profiling reveals imprints of food heating processes after dietary intervention with differently cooked potatoes. *J Agric Food Chem.* (2020) 68:6122–31. doi: 10.1021/acs.jafc.0c01136
- 71. Quan W, Jiao Y, Li Y, Xue C, Liu G, Wang Z, et al. Metabolic changes from exposure to harmful Maillard reaction products and high-fat diet on Sprague-Dawley rats. *Food Res Int.* (2021) 141:110129. doi: 10.1016/j.foodres.2021.110129
- Ames JM, Wynne A, Hofmann A, Plos S, Gibson GR. The effect of a model melanoidin mixture on faecal bacterial populations *in vitro*. Br J Nutr. (1999) 82:489–95. doi: 10.1017/S0007114599001749
- Faist V, Erbersdobler HF. Metabolic transit and *in vivo* effects of melanoidins and precursor compounds deriving from the Maillard reaction. *Ann Nutr Metab.* (2001) 45:1–12. doi: 10.1159/000046699
- 74. Pérez-Jiménez J, Díaz-Rubio ME, Mesías M, Morales FJ, Saura-Calixto F. Evidence for the formation of maillardized insoluble dietary fiber in bread: a specific kind of dietary fiber in thermally processed food. *Food Res Int.* (2014) 55:391–6. doi: 10.1016/j.foodres.2013.11.031
- Borrelli RC, Fogliano V. Bread crust melanoidins as potential prebiotic ingredients. Mol Nutr Food Res. (2005) 49:673– 8. doi: 10.1002/mnfr.200500011
- Helou C, Denis S, Spatz M, Marier D, Rame V, Alric M, et al. Insights into bread melanoidins: fate in the upper digestive tract and impact on the gut microbiota using *in vitro* systems. *Food Funct.* (2015) 6:3737– 45. doi: 10.1039/C5FO00836K
- Mayta-Apaza AC, Pottgen E, De Bodt J, Papp N, Marasini D, Howard L, et al. Impact of tart cherries polyphenols on the human gut microbiota and phenolic metabolites *in vitro* and *in vivo*. J Nutr Biochem. (2018) 59:160– 72. doi: 10.1016/j.jnutbio.2018.04.001
- Cortés-Martín A, Selma MV, Tomás-Barberán FA, González-Sarrías A, Espín JC. Where to look into the puzzle of polyphenols and health? The postbiotics and gut microbiota associated with human metabotypes. *Mol Nutr Food Res.* (2020) 64:1900952. doi: 10.1002/mnfr.201900952
- Maccaferri S, Klinder A, Cacciatore S, Chitarrari R, Honda H, Luchinat C, et al. *In vitro* fermentation of potential prebiotic flours from natural sources: impact on the human colonic microbiota and metabolome. *Mol Nutr Food Res.* (2012) 56:1342–52. doi: 10.1002/mnfr.201200046
- Sandberg J, Kovatcheva-Datchary P, Björck I, Bäckhed F, Nilsson A. Abundance of gut Prevotella at baseline and metabolic response to barley prebiotics. *Eur J Nutr.* (2019) 58:2365–76. doi: 10.1007/s00394-018-1788-9
- Mio K, Otake N, Nakashima S, Matsuoka T, Aoe S. Ingestion of high β-glucan barley flour enhances the intestinal immune system of diet-induced obese mice by prebiotic effects. *Nutrients*. (2021) 13:907. doi: 10.3390/nu13030907
- Verzelloni E, Tagliazucchi D, Conte A. From balsamic to healthy: traditional balsamic vinegar melanoidins inhibit lipid peroxidation during simulated gastric digestion of meat. *Food Chem Toxicol.* (2010) 48:2097– 102. doi: 10.1016/j.fct.2010.05.010

- Tagliazucchi D, Bellesia A. The gastro-intestinal tract as the major site of biological action of dietary melanoidins. *Amino Acids*. (2015) 47:1077– 89. doi: 10.1007/s00726-015-1951-z
- Cann I, Bernardi RC, Mackie RI. Cellulose degradation in the human gut: *Ruminococcus champanellensis* expands the cellulosome paradigm. *Environ Microbiol.* (2016) 18:307–10. doi: 10.1111/1462-2920. 13152
- Moraïs S, David YB, Bensoussan L, Duncan SH, Koropatkin NM, Martens EC, et al. Enzymatic profiling of cellulosomal enzymes from the human gut bacterium, *Ruminococcus champanellensis*, reveals a fine-tuned system for cohesin-dockerin recognition. *Environ Microbiol.* (2016) 18:542–56. doi: 10.1111/1462-2920. 13047
- Wang H-Y, Qian H, Yao W-R. Melanoidins produced by the Maillard reaction: structure and biological activity. *Food Chem.* (2011) 128:573– 84. doi: 10.1016/j.foodchem.2011.03.075
- Kandylis P, Bekatorou A, Dimitrellou D, Plioni I, Giannopoulou K. Health promoting properties of cereal vinegars. *Foods.* (2021) 10:344. doi: 10.3390/foods10020344
- Sá AGA, Moreno YMF, Carciofi BAM. Food processing for the improvement of plant proteins digestibility. *Crit Rev Food Sci Nutr.* (2020) 60:3367– 86. doi: 10.1080/10408398.2019.1688249
- Tamanna N, Mahmood N. Food processing and maillard reaction products: effect on human health and nutrition. *Int J Food Sci.* (2015) 2015:e526762. doi: 10.1155/2015/526762
- Santal AR, Singh N. Biodegradation of melanoidin from distillery effluent: role of microbes and their potential enzymes. *Biodegrad Hazard Spec Prod.* (2013) 5:71–100. doi: 10.5772/56252
- Santal AR, Singh NP, Saharan BS. Biodegradation and detoxification of melanoidin from distillery effluent using an aerobic bacterial strain SAG5 of Alcaligenes faecalis. J Hazard Mater. (2011) 193:319–24. doi: 10.1016/j.jhazmat.2011.07.068
- 92. Santal AR, Singh NP, Saharan BS. A novel application of *Paracoccus* pantotrophus for the decolorization of melanoidins from distillery effluent under static conditions. J Environ Manage. (2016) 169:78–83. doi: 10.1016/j.jenvman.2015.12.016

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 Sharma, Sihmar, Santal, Prager, Carbonero and Singh. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.