



# Review Amyloid Beta Dynamics in Biological Fluids—Therapeutic Impact

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**Abstract:** Despite the significant impact of Alzheimer's disease (AD) at individual and socioeconomic levels and the numerous research studies carried out on this topic over the last decades, the treatments available in daily clinical practice remain less than satisfactory. Among the accepted etiopathogenic hypotheses, the amyloidogenic pathway theory, although intensively studied and even sometimes controversial, is still providing relevant theoretical elements for understanding the etiology of AD and for the further development of possible therapeutic tools. In this sense, this review aims to offer new insights related to beta amyloid (A $\beta$ ), an essential biomarker in AD. First the structure and function of A $\beta$  in normal and pathological conditions are presented in detail, followed by a discussion on the dynamics of A $\beta$  at the level of different biological compartments. There is focus on A $\beta$  elimination modalities at central nervous system (CNS) level, and clearance via the blood–brain barrier seems to play a crucial/dominant role. Finally, different theoretical and already-applied therapeutic strategy" and "cerebrospinal fluid sinks therapeutic strategy". These data outline the need for a multidisciplinary approach designed to deliver a solution to stimulate A $\beta$  clearance in more direct ways, including from the cerebrospinal fluid level.

**Keywords:** Alzheimer's disease; amyloid beta; amyloidogenic pathway; peripheral sink therapeutic strategy; cerebrospinal fluid sink therapeutic strategy

## 1. Introduction

Alzheimer's disease (AD), a neurodegenerative disease with a huge impact at the public health level, remains a challenge for neurologists in terms of finding an effective therapy [1]. Epidemiologically, it is estimated that over 5 million Americans currently suffer from AD, and predictions for the next few decades show an increase in the prevalence of the disease [2]. In addition, AD is increasingly correlated not only with mortality [3], but also with the financial burden incurred both at the individual level and at the global socioeconomic level [4].

Despite certain already demonstrated risk factors such as age, head injuries, coexisting vascular diseases, infections, and genetic predisposition [5], the cause remains unknown, with no curative treatment available. Over the past 30 years, various studies have tried to explain as completely as possible AD's etiology and evolution and to use this knowledge for the purpose of developing an effective treatment, leading to several theories [6]. There are two main hypotheses: the cholinergic hypothesis [7] and the amyloidogenic theory [8], although other theories related to neuroinflammation [9], abnormal Tau protein metabolism [10], or free radical damage [11] must also be considered.



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Regarding the cholinergic hypothesis, it is based on research initiated in the 1970s related to acetylcholine (ACh), its synthesis and transport at synapse level, and its role in the cognitive process [12]. The synthesis of ACh takes place in cholinergic neurons, starting from choline and acetyl-coenzyme A, the process being performed under the action of the choline acetyltransferase (ChAT) enzyme [13]. Subsequently, ACh is transported by the vesicular acetylcholine transporter to the postsynaptic neuron [14]. At the CNS level, ACh is involved in many higher cognitive processes, such as memory, attention, and learning [15]. Studies on AD have also shown a degeneration of cholinergic neurons [16], explaining the symptoms of the disease, which mainly involve memory loss and impaired cognitive function [17]. Based on these considerations, currently used drug therapies seek to compensate for reduced ACh biosynthesis by inhibiting the ACh degradation enzyme, acetylcholinesterase [18]. Drugs such as donepezil, rivastigmine, or galantamine are currently in use for the symptomatic treatment of AD [19]. In addition, related to ACh, we mention a new class of drugs whose role is to target choline transporter (CHT1) by competitive inhibition, the studies being still in their infancy [20]. CHT1 is a high-affinity choline transporter present on the presynaptic terminal of cholinergic neurons which takes up choline, one of the precursors of acetylcholine [21]. The cholinergic hypothesis is closely related to the amyloid hypothesis (detailed below), as amyloid beta (A $\beta$ ) is thought to modulate cholinergic neurotransmission by reducing the choline uptake and subsequently the release of ACh [22]. Studies have demonstrated that as the cholinergic synaptic loss and the amyloid fibril formation are related to  $A\beta$  and interaction with ACh enzymes, more attention should be directed toward amyloid precursor protein (APP) metabolism or Wnt signaling [23].

The amyloid hypothesis, one of the oldest and most studied of the AD pathophysiological hypotheses, is currently undergoing a renaissance, with still many clues to offer which might be helpful for the development of new therapies. The link between A $\beta$ accumulation in the brain and the onset of dementia has been observed for decades [24]. However, the fact that abnormal deposition of amyloid beta-sheets was also found in the brains of healthy elderly people poses a question about the causal relationship between A $\beta$ deposits and AD onset [25]. Moreover, the AD classification into two subtypes, depending on the time of onset, although first described in 1997 [26], was supported by subsequent research and is still relevant, representing a starting point for the development of effective therapies. On the one hand, we are talking about early-onset AD (EOAD) which represents only a small percentage [27], while on the other hand the late-onset AD (LOAD) is the sporadic form, and the most common one (95% of cases) [28]. Even though both forms are associated with an imbalance between A $\beta$  production and clearance, resulting in the excessive accumulation of toxic forms of A $\beta$ , the hypotheses claim that the clinical and pathophysiological aspects are different for each form [29].

While the main cause for EOAD dementia would be the increased A $\beta$  production, LOAD is the result of a defective clearance of A $\beta$  [30]. In this context, researchers' attention has focused in recent years on the dynamics of A $\beta$  between the brain compartment and other biological compartments such as cerebrospinal fluid (CSF) and blood, with respect to the A $\beta$  clearance pathways from the brain. This review aims to present an overview of this very idea, according to which improvement of the deficient A $\beta$  clearance in AD could represent a viable method of treatment. Accordingly, after presenting in detail the structure and function of A $\beta$  and its clinical correlations, the authors provide an overview of the A $\beta$  dynamics between different biological compartments, focusing on the elimination via blood–brain barrier (BBB), which is one of the most important ways of A $\beta$  elimination at the brain level. In the last part, the therapeutic possibilities and the models/hypotheses that can lead to new therapies to increase the A $\beta$  clearance at CNS level are highlighted. The originality of this study, comprising many intensely debated elements about the pathophysiology and therapeutic options of AD, lies in fresh approach to a topic that, despite numerous available studies, still presents unknown facts to researchers.

### 2. Aβ—Production, Structure, Functions, and Therapeutic Correlations

Aβ protein is the result of non-physiological proteolysis of the amyloid precursor protein (APP) [31]. APP, encoded by APP genes located on Chromosome 21, is a Type I transmembrane protein, having a long N-terminal domain and a short cytoplasmic tail [32]. The APP protein family also contains APP Type 1 and 2 proteins (APLP1 and APLP2), which, although very similar in sequence to APP, lack the Aβ domain. In addition, both APLP1 and APLP2 are processed similarly, undergoing ectodomain shedding [33]. In terms of expression and localization, APP and APLP2 are expressed ubiquitously, with particularly high expression in neurons, while APLP1 is found primarily in the nervous system [34].

APP can follow two degradation pathways, one non-amyloidogenic and one amyloidogenic. Under normal conditions, APP is processed via the non-amyloidogenic pathway, first under the action of alpha-secretase (a zinc metalloproteinase), resulting in the sAPP $\alpha$  fragment, a broadly soluble ectodomain of APP [35]. Currently, three members of the ADAM family (a disintegrin and metalloproteinase) are suspected to play a role in this degradation pathway, having  $\alpha$ -secretase-like activity: ADAM9, ADAM10, and ADAM17 [36]. Alteration of ADAM17 can alter  $\alpha$  cleavage of APP and A $\beta$  generation,  $\alpha$  cleavage being abolished in ADAM17-deficient cells [37]. In humans, inhibiting ADAM17 prevented regulated  $\alpha$ -secretase activity in human neurons [38]. Regarding ADAM10, its overexpression increases  $\alpha$ -cleavage [39], while sAPP $\alpha$  generation was nearly abolished in the neurons of mice with neural ADAM10 conditionally knocked out [40]. sAPP $\alpha$  has multiple roles, demonstrating its action in neuronal plasticity/survival and protection against excitotoxicity in the mature brain [41]. In the early stages of development, sAPP $\alpha$  regulates neural stem cell proliferation [42].

Regarding the amyloidogenic pathway that will end with the production of A $\beta$ , cleavage of APP will be performed under the action of  $\beta$ -secretase. The main  $\beta$ -secretase is BACE1, a membrane-bound aspartyl protease, and considered the rate-limiting factor in A $\beta$  generation from APP [43]. BACE1 was also studied as a potential therapeutic target, with the studies on AD mouse models being worth mentioning, where the deficiency of BACE1 was correlated with an important reduction in A $\beta$ 40/42 levels, reduced neuronal loss, and memory deficits [44]. BACE2, a homologue of BACE1, also processes APP at the  $\beta$  site, contributing to AD pathogenesis [45]. Although expressed in neurons substantially lower than BACE1, BACE2 could become a therapeutic target for AD, without the side effects of BACE1 inhibition [46]. Besides BACE1 and BACE2, a lysosomal cysteine protease, cathepsin B, was also proposed as an additional  $\beta$ -secretase, as cathepsin B inhibitors demonstrated mitigation of memory deficit and reduction in beta-amyloid concentration related to AD [47].

After the cleavage by either alpha-secretase or beta-secretase, the carboxyl terminal fragments (CTFs) of APP are further altered by gamma-secretase, generating p83 and Aβ, respectively. The gamma-secretase complex comprises at least four components: presenilin (PS, PS1 or PS2), Nicastrin, anterior pharynx-defective-1 (APH-1), and presenilin enhancer-2 (PEN-2) [48]. Several other factors such as CD147 or TMP21/p23 have been proposed as gamma-secretase components, although playing only modulatory roles [49]. Moreover, other enzymes such as caspases (especially caspase-3) can cleave APP in specific positions (position Asp664), releasing a 31 amino acid fragment of APP called C31, with cytotoxic characteristics [50]. For a detailed overview, see Figure 1.



Figure 1. Non-amyloidogenic vs. amyloidogenic pathway.

Knowing in detail the amyloidogenic metabolic pathway of A $\beta$  provides essential knowledge for developing new therapies. B-secretase cleavage, a fundamental step in A $\beta$  production, was already the subject of research when referring to BACE inhibitors (see below), and although clinical trials conducted so far were discontinued for safety reasons, BACE1 remains a well-validated therapeutic target for AD [51].

When referring to the structure of  $A\beta$  and its influence on  $A\beta$  toxicity, quaternary structures play the major role. Moreover, it has been shown that oligomers are the most toxic/pathogenic structures, compared to monomers or even stable senile plaques [52–54]. The formation of oligomers occurs through the nucleation process, which can be a primary process—when two or more monomers are assembled—or a secondary one, when there are already oligomers that facilitate additional nucleation [55]. These two nucleation processes are of therapeutic importance, with conformation-specific antibodies being used experimentally to prevent the formation of toxic oligomers and being a possible treatment modality [56]. However, an important element when talking about the study of oligomers is the differentiation of their influence in vivo compared to in vitro studies. Only Type 2 oligomers (formed by secondary nucleation) share the basic structural features of amyloid fibrils, being found in the vicinity of amyloid plaques. However, regarding the toxic effects at the CNS level, both types of oligomers alter neuronal signaling pathways, leading from varying degrees of synaptic and axonal transport dysfunction to synaptic loss, and neuron death [57]. In order to develop efficient therapies, more research on A $\beta$  oligomers must be conducted in humans, as there are still questions regarding the correlation between temporal and spatial oligomer patterns and neurological dysfunction.

# 3. A B Dynamics between ISF, CSF, and Blood

A $\beta$  dynamics, with a focus on the pathways of A $\beta$  elimination from the CNS level, have provoked great interest among researchers in recent years, given that studies increasingly show the link between defective A $\beta$  elimination and LOAD [58]. In addition, if in the past it was thought that most A $\beta$  elimination is performed via BBB, the glymphatic pathway is becoming increasingly important nowadays, with studies showing that astroglial aquaporin-4 (AQP4) channels eliminate significant amounts of extracellular A $\beta$  [59].

Currently, several A $\beta$  purification systems are known at the CNS level (see Table 1), with soluble extracellular A $\beta$  forming the bulk of the total amount of A $\beta$  that has a high mobility between the different biological compartments. The elimination of proteins, and by implication of A $\beta$ , is achieved both at the intracellular level, under the action of proteolytic enzymes, and at the extracellular level, being eliminated in the blood or recirculated in the CSF. The actual percentage contribution of each protein degradation/elimination method remains to be unraveled.

Clearance Pathway	Direction/Biological Compartment	Key Players	Alterations in Pathological Conditions (AD)
Blood-brain barrier	ISF to peripheral circulation	LRP1, LRP2, ABCB1, ABCA1, α2-macroglobulin, IDE, ApoE, RAGE	Reduced efflux Increased RAGE-influx
Intracellular degradation	Microglia, Astrocyte	ubiquitin–proteasome pathway, autophagy–lysosome pathway, endosome–lysosome pathway	Reduced
Extracellular degradation	ISF	Proteases, phagocytosis (microglia/astrocyte uptake) Diffusion	Reduced
Perivascular drainage	ISF to CSF	Transporter-mediated active mechanism Bulk flow	Reduced diffusion
Glymphatic system	ISF to CSF	AQP4 Sleep	Unknown (probably reduced)
CSF absorption	CSF to peripheral circulation CSF to peripheral lymphatic system	Brain–CSF barrier Arachnoid villi Lymphatic absorption	Reduced flow via brain-CSF barrier and arachnoid villi
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### Table 1. A $\beta$ clearance pathways from CNS.

AD—Alzheimer's disease; ISF—interstitial fluid; CSF—cerebrospinal fluid.

Degradation clearance occurs at the intra- and extracellular level, being supported by different enzyme systems. At the intracellular level, along with protease, A $\beta$  degradation can occur as follows: via ubiquitin–proteasome pathway, via autophagy–lysosome pathway, and via endosome–lysosome pathway [60]. However, these systems are affected in aging or in neurodegenerative diseases, the evidence suggesting that proteasomal activity may be inhibited by A $\beta$ . Dysfunction in the ubiquitin–proteasome pathway appears to promote APP processing, with the formation of increased amounts of A $\beta$  [61].

At the extracellular level, the degradative processes are much simpler, being performed under the action of proteases and phagocytes produced by the astrocyte. However, extracellular proteins can be embedded at the intracellular level, where they will be degraded under the phagocytic action of the microglia or astrocyte [62].

A $\beta$  from the ISF level is cleared mostly via BBB directly into the bloodstream, under the action of specialized transporter systems [63]. It is already known that BBB is one of the three highly selective interfaces between the circulatory system and the CNS, with the role of maintaining cerebral homeostasis, along with the blood–cerebrospinal fluid (CSF) barrier and the arachnoid barrier [64].

Located at the interface between the vascular system and the cerebral substance, more specifically at the level of the endothelium of micro vessels, BBB is not a simple barrier but a complex structure, unique in the human body, its integrity being essential to fulfill its functions correctly [65]. The presence of inflow and outflow transporters is important for A $\beta$  dynamics through BBB. Responsible for the soluble A $\beta$  efflux at the ISF level are LDL receptor (LDLR) family members such as LRP1, and ATP-binding cassette transporters (ABC transporters) [66].

ABCB1 (also known as P-glycoprotein 1 or MDR1) is the main transporter in the ABC transporters family and mediates  $A\beta$  efflux directly into the circulation [67]. Studies have further shown a possible association between oxidative stress and downregulation of

ABCB1 and other BBB efflux transporters in AD, as AD is considered a chronic inflammatory state with sustained systemic oxidative stress [68].

Another transporter, ABCA1, appears to mediate A $\beta$  efflux in an ApoE-dependent manner. The data are also supported by a meta-analysis [69], which showed that ABCA1 rs2422493 polymorphism was a risk factor for AD, while other mutations may play a role in AD pathogenesis when interacting with ApoE- $\epsilon$ 4. Another study showed that ABCA1 controls ApoE lipidation, as ApoE4 promotes ABCA1 trapping in late-endosomes and impairs its recycling to the cell membrane. That is conducive to lower ABCA1-mediated cholesterol efflux activity, a greater percentage of lipid-free ApoE particles, and consequently lower A $\beta$  degradation capacity [70].

LRP1 has been intensively studied in relation to the Aß accumulation found in AD, its involvement being demonstrated in several stages of the A $\beta$  formation and elimination process. First, there are studies that have shown that LRP1 has the ability to modulate APP processing, influencing A $\beta$  generation. On the one hand, Ref. [71] showed that blocking LRP1 activity will lead to an increased cell surface level of APP and a significant reduction in A $\beta$  synthesis. Other studies demonstrate the opposite, considering that overexpression of the LRP1 C-terminal transmembrane domain can play the role of a competitor for APP when referring to  $\beta$ -secretase and  $\gamma$ -secretase processing, leading to reduced A $\beta$ production [72]. LRP1 is directly involved in the endocytic receptor mediation and trafficking, and in lysosomal degradation of several substrates, with recent works demonstrating receptor-mediated clearance of A $\beta$  through LRP1 in neurons and astrocytes [73]. Besides neural cells, LRP1 acts also at vascular cell level, as recent research confirms LRP1-mediated internalization of A $\beta$  at BBB level in animal models [74]. The same mechanism of active elimination of A $\beta$  from CSF through LRP1 is found at choroid plexus level, forming another important pathway to regulate CSF A $\beta$  concentration [75]. With regard to BBB, LRP1 seems to play an important role in vascular smooth muscle cells and pericytes also. The group of (Kanekiyo et al., 2012) demonstrated the influence of LRP1 suppression in the human brain's vascular smooth muscle cells, which significantly reduces uptake and degradation of both endogenous and exogenous A $\beta$  [76]. Normal LRP1 activity is also a condition for pericytes' normal functioning, as these cells regulate BBB integrity and cerebral blood flow by also clearing A $\beta$  through LRP1 mediated pathway [77].

When thinking of A $\beta$  clearance and A $\beta$  equilibrium between different biological fluids or compartments, LRP1 is also important in the peripheral clearance of A $\beta$ . As the peripheral sink therapeutic strategy states, there is a dynamic equilibrium between A $\beta$  pools in the brain and at the peripheries, where A $\beta$  is cleared through peripheral organs such as the liver [78]. However, many studies found no evidence of a therapeutic relevance for AD evolution when A $\beta$  peripheral elimination was increased through different methods, showing that other solutions such as CSF clearance need to be found in order to delay AD evolution [79].

Finally, LRP1 is of therapeutic importance because it can regulate neuronal transmission by direct or indirect interaction with synaptic proteins such as NMDA receptor unit or GluA1 [80]. Studies have shown the LRP1-mediated insulin signaling and glucose uptake regulation in animal models, with reduction in GLUT3 and GLUT4 levels in neurons [81]. In addition, LRP1 regulates the leptin/leptin receptor complex in the hypothalamus, with depletion in neuronal LRP1 being associated with increased food intake, and the subsequent increased risk of obesity and diabetes [82].

Another member of the LDL receptors is LDLR-related protein 2 (LRP2), also known as megalin, which seems to mediate the clearance of  $A\beta$  through the BBB when forming a complex with clusterin (also known as ApoJ). Megalin also has a protective function at choroid plexus level, forming a complex with gelsolin, a protein that is produced in the epithelial cells of the choroid plexus [83].

Clearance of A $\beta$  through the BBB is also mediated by  $\alpha$ 2-macroglobulin ( $\alpha$ 2M). The interest in the influence of  $\alpha$ 2M in AD is not new, as earlier reports suggested an association between  $\alpha$ 2M polymorphisms and an increased risk of neurodegenerative diseases [84,85].

Although newer data does not sustain this correlation [86], elevated levels of  $\alpha$ 2M were found in the CSF of AD patients (especially males), with clinical significance still to be determined. Besides the interactions with A $\beta$  aggregation and clearance,  $\alpha$ 2M is involved in other relevant biological processes, such as the interaction with different apoE isoforms [87]. Thus, so far, there is strong evidence suggesting the role of  $\alpha$ 2M in the inhibition of A $\beta$ peptide aggregation and toxicity, mainly by facilitating its clearance [88]; however, direct individual implication of  $\alpha$ 2M in the prevention or promotion of neurodegeneration independent of AD remains to be determined.

Insulin-degrading enzyme (IDE) has been proposed to have a role in A $\beta$  clearance through the BBB, which might explain why BBB clearance is sensitive to insulin. IDE along with neprilysin is involved in the intracellular and extracellular degradation of A $\beta$  [89]. Moreover, IDE seems to act specifically on beta-structure forming substrates, its specificity being a potential advantage in therapeutic approaches for AD [90]. Even in a proteolytically inactive form, according to recent research, IDE appears to inhibit A $\beta$ fibrillogenesis through a chaperone-like role [91]. However, the sensitive cerebral A $\beta$ clearance mediated by insulin and IDE cannot be fully explained based only on present known molecules; the involvement of other, still to be detected, transport systems or proteases has also to be taken into consideration.

Aβ transport is bidirectional, including influx into the ISF/CNS, via RAGE (advanced glycosylation end product-specific receptor). RAGE, a 35kD transmembrane receptor of the immunoglobulin super family, was initially identified and characterized for its ability to bind advanced glycosylation end products (AGEs), but also other ligands such as A $\beta$ , with implication in AD pathogenesis [92]. It seems that RAGE is involved through several mechanisms in neurodegeneration. Firstly, Aβ–RAGE interactions are important for the influx of A $\beta$  from blood to the cerebral sector, but also regarding the BBB integrity. RAGE-mediated AB cytotoxicity is directed towards the brain's microvascular endothelial cells, resulting in structural damage of the neurovascular unit [93]. Breakage of the BBB integrity is also induced by A $\beta$ -RAGE interactions that disrupt tight junction proteins via the Ca<sup>2+</sup>-calcineurin (CaN) pathway [94]. Moreover, RAGE is supposed to be also an important contributor to AB generation, as increased expression of RAGE in AD enhances the activities of beta- and/or gamma-secretases, stimulating A $\beta$  production [95]. Finally, the role of oxidative stress and sustained chronic neuroinflammation are two other potential mechanisms related to AD pathogenesis. RAGE, acting as a critical player in both initiation and progression of oxidative stress and inflammation, has intricate pathways with AB production and clearance at CNS level [96].

Intensive research was conducted during the last decade on alternative ways of ISF clearance to CSF sink. Two main hypotheses have been circulating, namely the perivascular drainage pathway and the glymphatic pathway. As it is still unclear if these two are distinct pathways, or they simply reflect transport along the same pathway captured under differing physiological or experimental conditions, both mechanisms are detailed below. However, recent data are in favor of the non-mutual exclusiveness of the two clearance pathways, as both could be active depending on specific conditions, while anatomical data reveals that they could use different vessel systems [97].

Previous research has demonstrated that perivascular drainage of ISF was directed to the subarachnoid CSF, taking place ubiquitously throughout the brain [98]. Several mechanisms were proposed to explain the directional movement of solutes from ISF to CSF: passive mechanisms such as diffusion, and active mechanisms such as advection and channel- or transporter-mediated facilitative mechanism. More recent studies, where high-performance fluorescence microscopy was used, confirmed that special structures are involved in the bidirectional ISF–CSF fluid exchange. AQP4 channels play a major role, with significantly impaired flow being observed in mice models lacking AQP4; however, other still to be discovered structures also contribute to this pseudo-lymphatic function of this pathway [99]. Several factors influence glymphatic flow, with maneuvers that lead to arterial pulsatility reduction being associated to impaired solute drainage [100]. On the

other hand, gravitational factors and circadian rhythm, especially deep stages of sleep, seem to enhance solutes drainage through the glymphatic system [101]. As research is still emerging in this subfield relating to the glymphatic system, its functions and roles in physiological and pathological conditions is not completely known, and we can only assume that the glymphatic system contributes to  $A\beta$  clearance from ISF, although its exact share is still to be determined.

From CSF, solutes (including  $A\beta$ ) enter the final part of the clearance pathway in order to be eliminated in the peripheral circulatory system. Two main modalities were highlighted by physiologic and imagistic studies: clearance through arachnoid villi and blood–CSF barrier (BCSFB) in blood systemic circulation, concomitantly with perivascular and perineural flow into the lymphatic system [102]. Several factors influence CSF absorption by circular and lymphatic peripheral systems, including CSF production, structural integrity of BCSFB and arachnoid villi, and lymphatic flow at perivascular level, with all these factors being affected by pathological conditions. AD alters CSF production in different ways, by impairing the function of the choroid plexus which suffers calcification and fibrosis [64]. The main limitation in the AD context remains the impaired clearance of CSF, based on multiple dysfunctions in CNS solutes elimination mechanisms. Firstly, BCSFB suffers structural changes, with the activity of influx and efflux transporters such as LRP1 and ABCB1 also being significantly altered [103]. Impairment in the circulatory absorption pathway is found also at arachnoid villi level, where outflow resistance is increased in AD patients. Similar to normal pressure hydrocephalus, the proposed mechanism is based on decreased CSF bulk outflow resulting from structural degradation (amyloid deposition and fibrosis) at arachnoid villi level. Regarding lymphatic clearance, it is known that age is a non-modifiable factor that contributes to decrease in absorption activity of the peripheral lymphatic system [104], with the same suppositions being valid also for CSF lymphatic absorption.

Finally, it is observed that during the last decade the paradigm has shifted, as  $A\beta$  clearance via BBB pathway has lost some of its importance, with other ways of elimination coming into the spotlight. Moreover, the hype related to the discovery and better understanding of the glymphatic pathway has offered this alternative way of clearance increasing importance in physiological, as also pathological, conditions.

#### 4. Current and Future Therapeutic Directions for Aβ Reduction at CNS Level

Starting with the abovementioned protein clearance mechanisms at CNS level, all being to a varying extent affected in AD, several ways to compensate for their impairment have been examined, including increasing A $\beta$  clearance through alternative mechanisms (see Table 2).

A first way to compensate for the decreased  $A\beta$  clearance at CNS level is to inhibit its production. Based on theoretical knowledge about the amyloid cascade, one of the first suggested treatments, no longer valid today, consisted in inhibiting BACE. Studies on murine models were first performed, with several working groups succeeding in synthesizing molecules that had the ability to penetrate the BBB and selectively inhibit BACE1 and BACE2. The results showed a decrease in A $\beta$  brain accumulation [105], but without a real improvement in memory and behavioral deficits [106]. Despite extensive research performed in several mouse models such as double transgenic mice (APP23  $\times$  PS45)–[107] or Tg2576 transgenic mice—[108], none of the major BACE1 inhibitors tested were documented to show behavioral effects in AD animal models. Clinical trials in humans followed, involving the administration of different compounds (Verubecestat, Atabecestat, Umibecestat) among various groups of patients in different stages of evolution (preclinical/early AD—[109]; mild AD and mild-to-moderate AD—[110]), with most of them being stopped in the early stages considering the lack of clinical benefit, compounded also, however, by significant side effects such as drug worsened cognition, brain atrophy, and weight loss [111]. Given the results, expert opinion considers the failure of this therapy a sufficient

reason to forget this approach and focus on other directions/points of interest on the amyloidogenic pathway.

Type of Treatment	Pathophysiological Mechanism	Use/Efficiency
BACE inhibitors	Inhibit BACE1 and BACE2 in order to	Inefficient
	minimize Aß production	Several important adverse effects (brain
		atrophy, weight loss)
		Currently not recommended
Aβ monoclonal antibodies	Immunotherapy (Antigen-antibody	Inconsistent results
	complex)—favors Aβ elimination	Recently approved Adacunumab for clinical use
Aβ vaccine	DNA vaccination for anti-Aβ immunotherapy	Phase III clinical trials ongoing
RAGE inhibitors	Inhibition of RAGE	Azeliragon tested in phase 2/3
	Inhibition of Aß influx to CNS	trials—missed endpoints
	Reduction in oxidative stress and neuroinflammation	Research in progress
Plasmapheresis	Reduction in Aß peripheral level	Positive preliminary results
*	The "peripheral sink therapeutic strategy"	1
Peritoneal dialysis	Reduction in A $\beta$ peripheral level	Positive preliminary results
ý	The "peripheral sink therapeutic strategy"	1 2
Implantable intrathecal pumps	Reduction in A $\beta$ CSF level	Near future approach
	The "CSF sink therapeutic strategy"	
Aβ cleavage	Degradation of A $\beta$ at both CNS and peripheral	Intracerebral delivery of
C C	level	neprilysin—positive preliminary results
		Peripheral delivery of neprilysin—no
		impact on $A\beta$ at brain level

Table 2. Alzheimer's disease—therapies with focus on the amyloidogenic pathway.

A completely opposite approach to the A $\beta$  production inhibition is to stimulate the elimination of A $\beta$  from the CNS and/or A $\beta$  enzymatic degradation. Therapeutic clearance of A $\beta$  involves several strategies, but the most researched remains immunotherapy. The principle of immunotherapy consists in activating the immune system, the method's final role being the degradation of A $\beta$ . There are two ways to achieve this goal: active immunotherapy, in which the production of anti-A $\beta$  antibodies is stimulated, and passive immunotherapy, in which antibodies are administered directly [112]. There are currently several antibodies available, including for clinical use, that have been shown to reduce cerebral A $\beta$  load in both basic research and human clinical trials. However, there is a lack of correlation between the neuropathological benefits and the expected clinical improvement, due to several suspected reasons: the efficiency of antibodies is maximum on A $\beta$  oligomers that have not yet aggregated in the form of senile plaques, thus requiring administration in the prodromal/early stages; there are also other factors such as age or ApoE which influence the behavior of A $\beta$  oligomers and the interaction with possible antibodies [113].

Even if the research carried out so far has not brought the desired results, the mechanisms underlying the process by which anti-A $\beta$  antibodies remove A $\beta$  from the brain are key in the design of new therapeutic approaches. Two separate mechanisms have been proposed, which are not mutually exclusive. On the one hand, microglia with their phagocytic capabilities would capture the antigen-antibody complex, once it is formed at the ISF level. On the other hand, clearance also occurs in the periphery. There are data which support "the peripheral sink therapeutic strategy" and show that the elimination/decrease in the peripheral level of A $\beta$  leads to a reduction in the cerebral A $\beta$ level [114]. The validity of this method has been questioned by subsequent studies [115]; however, many variables remain under discussion, given that results on different mouse models have delivered different, sometimes opposite, conclusions. For example, mice with the Dutch and Iowa mutations tend to accumulate more A $\beta$  in the brain, as A $\beta$  is in this case a poor substrate for LRP transporter. Consequently, peripheral sink therapeutic strategy remains inefficient, with low blood A $\beta$  levels being correlated to high, unmodifiable brain/ISF A $\beta$  concentration [116]. Even in human studies, the results were not as desired, a recent meta-analysis showing that, despite being well tolerated, peripherally administered anti-A $\beta$  immunotherapy does not significantly improve the primary outcome measures. In addition, it must be considered that the peripheral A $\beta$  pool is not only the result of the passage of A $\beta$  from the brain to the peripheral circulation, but also of A $\beta$  being produced in the periphery from cells other than neurons (hepatocytes, platelets) [117].

Based on these assumptions, alternative strategies have been tried, which consist in reducing the A $\beta$  blood level. Among them, we mention plasmapheresis, with positive preliminary results in a phase 2b/3 trial in a group of mild-to-moderate AD patients, where a decrease in cognitive and functional decline was observed [118]. In addition, based on the "peripheral sink therapeutic strategy", A $\beta$  peripheral clearance was tried by peritoneal dialysis [119] and hemodialysis [120], with satisfactory preliminary results.

However, a much more direct way to eliminate  $A\beta$  remains to be explored, more precisely at the CSF level. Knowing the equilibrium between ISF and CSF, as well as the shortcomings of peripheral A $\beta$  clearance methods, the approach based on the "CSF sink therapeutic strategy" seems much more natural and having better therapeutic potential. The decrease in A $\beta$  concentration at the CSF level will determine the balancing of the A $\beta$ concentrations between the communicating fluidic compartments CSF-ISF, as also the decrease of the soluble  $A\beta$  concentration at the ISF level. Hypothetically, the elimination of soluble A $\beta$  will also mobilize the insoluble, initially difficult to mobilize, A $\beta$  from the stocks, possibly even influencing the degradation of senile plaques. However, this hypothesis must be validated by studies on human and murine models in the near future. There is nowadays the possibility of accessing the CSF compartment by using implantable biocompatible devices that are based on nanotechnologies. Several technologies such as unidirectional and bidirectional ventriculoperitoneal derivation and lumboperitoneal shunting are already being employed in other pathological conditions, e.g., hydrocephalus [121]. Implantable intrathecal pumps are more advanced biomedical devices that have seen an increase in interest during the last year, as they help in delivering medications directly into the lumbar subarachnoid space [122]. With an inspirational starting point linked to current existing technologies, a very successful future direction could be the development of similar medical devices tailored for A $\beta$  clearance as at least an adjuvant therapeutic approach.

Finally, we present another potentially effective method, although it has not yet been adopted in current clinical practice, whose mechanism of action is based on amyloid cleavage. Among the enzymes capable of degrading physiologically relevant peptides, neprilysin has attracted interest because of its ability to degrade both amyloid beta peptides 1-40 and 1-42 rapidly and efficiently [123]. Neprilysin, also known as neutral endopeptidase (NEP), is an integral Type II membrane-bound zinc-dependent peptidase normally expressed by a variety of tissues [124]. NEP has a quite broad substrate specificity, having however a stronger preference for peptides such as enkephalins, tachykinins, and natriuretic peptides [125]. By cleaving peptides at the N-terminal side of hydrophobic amino acid residues, NEP is responsible for the degradation of a variety of physiological substrates, limiting their activity. As regards AD, NEP is considered a potential therapeutic strategy for the prevention and treatment of the disease, being able to degrade A $\beta$  at brain level. Research in animal models has revealed that manipulation of the levels of brain NEP has a significant effect on A $\beta$  levels. Numerous study groups have shown that NEP gene transfer reduces human amyloid pathology in transgenic mice [126–130]. These promising preliminary results have led to extensive research on this topic, with two directions being relevant here. Firstly, in order to create a potent therapeutic tool for AD, the activity and specificity of NEP towards A ß need to be better understood and improved. In this regard, the works of [131,132] bring new insights at the molecular level of the A $\beta$  degradation by NEP, paving the way to creating a NEP mutant enzyme with higher efficiency in degrading A $\beta$  in vivo. Secondly, Ref. [133] assessed the therapeutic potential of the intracerebral delivery of neprilysin as a dynamically controllable A $\beta$ (40)-degrading therapeutic strategy for AD. Recent studies [134,135] concentrate on the improvement of NEP delivery across

the BBB using novel transport systems, as peripheral administration of NEP does not affect central levels of  $A\beta$ . Moreover, immunogenicity may be a side effect in some cases [115].

## 5. Conclusions

Although incomplete and lacking full explanation of the etiopathogenesis of AD, the amyloidogenic hypothesis remains one of the most important theories for understanding the disease and developing potential therapies [136]. In addition, continuous discoveries in recent years regarding A $\beta$  dynamics, such as the growing importance of the role of the glymphatic system in the clearance of molecules involved in AD pathogenesis [137], are challenging the classical paradigms. Moreover, acceptance of the existence of two forms of AD, EOAD and LOAD, with the latter being predominant and based on impaired Aß clearance, suggests that in order to be successful in the treatment of AD, future approaches must focus on ways that increase  $A\beta$  elimination from the CNS; or, as the direct elimination from ISF is currently unfeasible, new theories such as the "peripheral sink therapeutic strategy" or the "CSF sink therapeutic strategy" become the theoretical basis for the development of new technologies that will be able to eliminate  $A\beta$  directly from circulation and CSF level, ultimately reducing the cerebral A $\beta$  concentration. This therapeutic path remains open and of great interest, requiring interdisciplinary research on biocompatible nanostructures in order to come up with a feasible solution first on murine models, and later with satisfactory results in daily clinical practice.

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#### References

- 1. Briggs, R.; Kennelly, S.P.; O'Neill, D. Drug treatments in Alzheimer's disease. Clin. Med. 2016, 16, 247–253. [CrossRef]
- 2. Alzheimer's Association 2016 Alzheimer's disease facts and figures. Alzheimer's Dement. 2016, 12, 459–509. [CrossRef]
- 3. Sandoval, J.D.J.; Turra, C.M.; Loschi, R.H. Adjusted Mortality Rates Attributable to Alzheimer's Disease Dementia, Brazil, 2009–2013. *Cad. Saude Publica* 2019, 35, e00091918. [CrossRef]
- 4. Gresenz, C.R.; Mitchell, J.M.; Marrone, J.; Federoff, H.J. Effect of early-stage Alzheimer's disease on household financial outcomes. *Health Econ.* **2020**, 29, 18–29. [CrossRef]
- 5. Armstrong, R.A. Risk factors for Alzheimer's disease. Folia Neuropathol. 2019, 57, 87–105. [CrossRef]
- 6. Abeysinghe, A.A.D.T.; Deshapriya, R.D.U.S.; Udawatte, C. Alzheimer's disease; a review of the pathophysiological basis and therapeutic interventions. *Life Sci.* **2020**, *256*, 117996. [CrossRef]
- Hampel, H.; Mesulam, M.-M.; Cuello, A.C.; Farlow, M.R.; Giacobini, E.; Grossberg, G.T.; Khachaturian, A.S.; Vergallo, A.; Cavedo, E.; Snyder, P.J.; et al. The cholinergic system in the pathophysiology and treatment of Alzheimer's disease. *Brain* 2018, 141, 1917–1933. [CrossRef] [PubMed]
- 8. Hardy, J.A.; Higgins, G.A. Alzheimer's disease: The amyloid cascade hypothesis. *Science* 1992, 256, 184–185. [CrossRef]
- Calsolaro, V.; Edison, P. Neuroinflammation in Alzheimer's disease: Current evidence and future directions. *Alzheimer's Dement*. 2016, 12, 719–732. [CrossRef] [PubMed]
- Bakota, L.; Brandt, R. Tau Biology and Tau-Directed Therapies for Alzheimer's Disease. Drugs 2016, 76, 301–313. [CrossRef] [PubMed]
- 11. Sonnen, J.A.; Larson, E.B.; Gray, S.L.; Wilson, A.; Kohama, S.G.; Crane, P.K.; Breitner, J.C.S.; Montine, T.J. Free radical damage to cerebral cortex in Alzheimer's disease, microvascular brain injury, and smoking. *Ann. Neurol.* 2009, 65, 226–229. [CrossRef]
- 12. Beers, W.H.; Reich, E. Structure and Activity of Acetylcholine. *Nature* **1970**, 228, 917–922. [CrossRef] [PubMed]
- 13. Mautner, H.G.; Nachmansohn, D. Choline Acetyltransferas. CRC Crit. Rev. Biochem. 1977, 4, 341–370. [CrossRef] [PubMed]
- 14. Prado, V.F.; Roy, A.; Kolisnyk, B.; Gros, R.; Prado, M.A.M. Regulation of cholinergic activity by the vesicular acetylcholine transporter. *Biochem. J.* **2013**, 450, 265–274. [CrossRef]

- 15. Brown, D.A. Acetylcholine and cholinergic receptors. Brain Neurosci. Adv. 2019, 3, 2398212818820506. [CrossRef]
- 16. Schliebs, R.; Arendt, T. The cholinergic system in aging and neuronal degeneration. *Behav. Brain Res.* 2011, 221, 555–563. [CrossRef]
- 17. Lester, D.; Rogers, T.D.; Blaha, C.D. Acetylcholine-Dopamine Interactions in the Pathophysiology and Treatment of CNS Disorders. *CNS Neurosci. Ther.* **2010**, *16*, 137–162. [CrossRef] [PubMed]
- Bagri, K.; Kumar, A.; Manisha; Kumar, P. Computational Studies on Acetylcholinesterase Inhibitors: From Biochemistry to Chemistry. *Mini-Rev. Med. Chem.* 2020, 20, 1403–1435. [CrossRef]
- 19. Tiwari, S.; Atluri, V.; Kaushik, A.; Yndart, A.; Nair, M. Alzheimer's disease: Pathogenesis, diagnostics, and therapeutics. *Int. J. Nanomed.* **2019**, *14*, 5541–5554. [CrossRef]
- 20. Okuda, T.; Nomura, Y.; Konishi, A.; Misawa, H. Competitive inhibition of the high-affinity choline transporter by tetrahydropyrimidine anthelmintics. *Eur. J. Pharmacol.* **2021**, *898*, 173986. [CrossRef]
- 21. Haga, T. Molecular properties of the high-affinity choline transporter CHT1. J. Biochem. 2014, 156, 181–194. [CrossRef] [PubMed]
- 22. Fang, L.; Duan, J.; Ran, D.; Fan, Z.; Yan, Y.; Huang, N.; Gu, H.; Zhu, Y. Amyloid-β depresses excitatory cholinergic synaptic transmission in *Drosophila*. *Neurosci. Bull.* **2012**, *28*, 585–594. [CrossRef]
- 23. Majdi, A.; Sadigh-Eteghad, S.; Aghsan, S.R.; Farajdokht, F.; Vatandoust, S.M.; Namvaran, A.; Mahmoudi, J. Amyloid-β, tau, and the cholinergic system in Alzheimer's disease: Seeking direction in a tangle of clues. *Rev. Neurosci.* **2020**, *31*, 391–413. [CrossRef]
- Murphy, M.P.; LeVine, H., 3rd. Alzheimer's Disease and the Amyloid-β Peptide. J. Alzheimer's Dis. 2010, 19, 311–323. [CrossRef] [PubMed]
- Fjell, A.M.; McEvoy, L.; Holland, D.; Dale, A.M.; Walhovd, K.B.; Alzheimer's Disease Neuroimaging Initiative. What is normal in normal aging? Effects of aging, amyloid and Alzheimer's disease on the cerebral cortex and the hippocampus. *Prog. Neurobiol.* 2014, 117, 20–40. [CrossRef] [PubMed]
- Hampel, H.; Kötter, H.U.; Möller, H.-J. Blood-Cerebrospinal Fluid Barrier Dysfunction for High Molecular Weight Proteins in Alzheimer Disease and Major Depression: Indication for Disease Subsets. *Alzheimer Dis. Assoc. Disord.* 1997, 11, 78–87. [CrossRef]
- Cacace, R.; Sleegers, K.; Van Broeckhoven, C. Molecular genetics of early-onset Alzheimer's disease revisited. *Alzheimer's Dement*. 2016, 12, 733–748. [CrossRef]
- 28. Pierce, A.L.; Bullain, S.S.; Kawas, C.H. Late-Onset Alzheimer Disease. Neurol. Clin. 2017, 35, 283–293. [CrossRef]
- Tellechea, P.; Pujol, N.; Esteve-Belloch, P.; Echeveste, B.; García-Eulate, M.; Arbizu, J.; Riverol, M. Early- and Late-Onset Alzheimer Disease: Are They the Same Entity? *Neurologia* 2018, 33, 244–253. [CrossRef]
- 30. Mawuenyega, K.G.; Sigurdson, W.; Ovod, V.; Munsell, L.; Kasten, T.; Morris, J.C.; Yarasheski, K.E.; Bateman, R.J. Decreased Clearance of CNS β-Amyloid in Alzheimer's Disease. *Science* **2010**, *330*, 1774. [CrossRef]
- 31. Zhang, Y.-W.; Thompson, R.; Zhang, H.; Xu, H. APP processing in Alzheimer's disease. Mol. Brain 2011, 4, 3. [CrossRef]
- 32. Van Giau, V.; Bagyinszky, E.; Youn, Y.C.; An, S.S.A.; Kim, S. APP, PSEN1, and PSEN2 Mutations in Asian Patients with Early-Onset Alzheimer Disease. *Int. J. Mol. Sci.* 2019, 20, 4757. [CrossRef]
- Müller, U.C.; Zheng, H. Physiological Functions of APP Family Proteins. Cold Spring Harb. Perspect. Med. 2012, 2, a006288. [CrossRef]
- 34. Lorent, K.; Overbergh, L.; Moechars, D.; de Strooper, B.; van Leuven, F.; van den Berghe, H. Expression in mouse embryos and in adult mouse brain of three members of the amyloid precursor protein family, of the alpha-2-macroglobulin receptor/low density lipoprotein receptor-related protein and of its ligands apolipoprotein E, lipoprotein lipase, alpha-2-macroglobulin and the 40,000 molecular weight receptor-associated protein. *Neuroscience* 1995, *65*, 1009–1025. [CrossRef]
- 35. Kojro, E.; Fahrenholz, F. The Non-Amyloidogenic Pathway: Structure and Function of α-Secretases. *Subcell. Biochem.* **2005**, *38*, 105–127. [CrossRef] [PubMed]
- 36. Sammel, M.; Peters, F.; Lokau, J.; Scharfenberg, F.; Werny, L.; Linder, S.; Garbers, C.; Rose-John, S.; Becker-Pauly, C.; John, R.; et al. Differences in Shedding of the Interleukin-11 Receptor by the Proteases ADAM9, ADAM10, ADAM17, Meprin α, Meprin β and MT1-MMP. *Int. J. Mol. Sci.* **2019**, *20*, 3677. [CrossRef] [PubMed]
- 37. Buxbaum, J.D.; Liu, K.-N.; Luo, Y.; Slack, J.L.; Stocking, K.L.; Peschon, J.J.; Johnson, R.S.; Castner, B.J.; Cerretti, D.P.; Black, R.A. Evidence That Tumor Necrosis Factor α Converting Enzyme Is Involved in Regulated α-Secretase Cleavage of the Alzheimer Amyloid Protein Precursor. *J. Biol. Chem.* **1998**, 273, 27765–27767. [CrossRef] [PubMed]
- Tachida, Y.; Nakagawa, K.; Saito, T.; Saido, T.C.; Honda, T.; Saito, Y.; Murayama, S.; Endo, T.; Sakaguchi, G.; Kato, A.; et al. Interleukin-1β up-regulates TACE to enhance α-cleavage of APP in neurons: Resulting decrease in Aβ production. *J. Neurochem.* 2007, 104, 1387–1393. [CrossRef] [PubMed]
- Kuhn, P.-H.; Wang, H.; Dislich, B.; Colombo, A.; Zeitschel, U.; Ellwart, J.W.; Kremmer, E.; Roßner, S.; Lichtenthaler, S.F. ADAM10 is the physiologically relevant, constitutive α-secretase of the amyloid precursor protein in primary neurons. *EMBO J.* 2010, 29, 3020–3032. [CrossRef]
- Jorissen, E.; Prox, J.; Bernreuther, C.; Weber, S.; Schwanbeck, R.; Serneels, L.; Snellinx, A.; Craessaerts, K.; Thathiah, A.; Tesseur, I.; et al. The Disintegrin/Metalloproteinase ADAM10 Is Essential for the Establishment of the Brain Cortex. *J. Neurosci.* 2010, 30, 4833–4844. [CrossRef]
- Livingstone, R.W.; Elder, M.K.; Barrett, M.C.; Westlake, C.M.; Peppercorn, K.; Tate, W.P.; Abraham, W.C.; Williams, J.M. Secreted Amyloid Precursor Protein-Alpha Promotes Arc Protein Synthesis in Hippocampal Neurons. *Front. Mol. Neurosci.* 2019, 12, 198. [CrossRef] [PubMed]

- Coronel, R.; Bernabeu-Zornoza, A.; Palmer, C.; Muñiz-Moreno, M.; Zambrano, A.; Cano, E.; Liste, I. Role of Amyloid Precursor Protein (APP) and Its Derivatives in the Biology and Cell Fate Specification of Neural Stem Cells. *Mol. Neurobiol.* 2018, 55, 7107–7117. [CrossRef]
- 43. Koelsch, G. BACE1 Function and Inhibition: Implications of Intervention in the Amyloid Pathway of Alzheimer's Disease Pathology. *Molecules* 2017, 22, 1723. [CrossRef] [PubMed]
- 44. Hu, X.; Das, B.; Hou, H.; He, W.; Yan, R. BACE1 deletion in the adult mouse reverses preformed amyloid deposition and improves cognitive functions. *J. Exp. Med.* **2018**, *215*, 927–940. [CrossRef] [PubMed]
- 45. Wang, Z.; Xu, Q.; Cai, F.; Liu, X.; Wu, Y.; Song, W. BACE2, a conditional β-secretase, contributes to Alzheimer's disease pathogenesis. *JCI Insight* **2019**, *4*, e123431. [CrossRef]
- 46. Voytyuk, I.; Mueller, S.A.; Herber, J.; Snellinx, A.; Moechars, D.; Van Loo, G.; Lichtenthaler, S.F.; De Strooper, B. BACE2 distribution in major brain cell types and identification of novel substrates. *Life Sci. Alliance* **2018**, *1*, e201800026. [CrossRef]
- 47. Hook, V.; Hook, G.; Kindy, M. Pharmacogenetic features of cathepsin B inhibitors that improve memory deficit and reduce β-amyloid related to Alzheimer's disease. *Biol. Chem.* **2010**, *391*, 861–872. [CrossRef]
- 48. Wolfe, M.S. Structure and Function of the γ-Secretase Complex. *Biochemistry* **2019**, *58*, 2953–2966. [CrossRef] [PubMed]
- 49. Kanyenda, L.J.; Verdile, G.; Boulos, S.; Krishnaswamy, S.; Taddei, K.; Meloni, B.P.; Mastaglia, F.L.; Martins, R.N. The Dynamics of CD147 in Alzheimer's Disease Development and Pathology. *J. Alzheimer's Dis.* **2011**, *26*, 593–605. [CrossRef] [PubMed]
- 50. Park, S.A.; Shaked, G.M.; Bredesen, D.E.; Koo, E.H. Mechanism of cytotoxicity mediated by the C31 fragment of the amyloid precursor protein. *Biochem. Biophys. Res. Commun.* 2009, *388*, 450–455. [CrossRef] [PubMed]
- 51. Hampel, H.; Vassar, R.; De Strooper, B.; Hardy, J.; Willem, M.; Singh, N.; Zhou, J.; Yan, R.; Vanmechelen, E.; De Vos, A.; et al. The β-Secretase BACE1 in Alzheimer's Disease. *Biol. Psychiatry* **2021**, *89*, 745–756. [CrossRef] [PubMed]
- Walsh, D.M.; Klyubin, I.; Fadeeva, J.V.; Rowan, M.J.; Selkoe, D.J. Amyloid-β oligomers: Their production, toxicity and therapeutic inhibition. *Biochem. Soc. Trans.* 2002, 30, 552–557. [CrossRef]
- Glabe, C.C. Amyloid Accumulation and Pathogensis of Alzheimer's Disease: Significance of Monomeric, Oligomeric and Fibrillar Abeta. Subcell. Biochem. 2005, 38, 167–177. [CrossRef]
- Katzmarski, N.; Ziegler-Waldkirch, S.; Scheffler, N.; Witt, C.; Abou-Ajram, C.; Nuscher, B.; Prinz, M.; Haass, C.; Meyer-Luehmann, M. Aβ oligomers trigger and accelerate Aβ seeding. *Brain Pathol.* 2019, 30, 36–45. [CrossRef]
- Ayala, S.; Genevaux, P.; Hureau, C.; Faller, P. (Bio)chemical Strategies To Modulate Amyloid-β Self-Assembly. ACS Chem. Neurosci. 2019, 10, 3366–3374. [CrossRef] [PubMed]
- 56. Kayed, R.; Canto, I.; Breydo, L.; Rasool, S.; Lukacsovich, T.; Wu, J.; Albay, R., 3rd; Pensalfini, A.; Yeung, S.; Head, E.; et al. Conformation dependent monoclonal antibodies distinguish different replicating strains or conformers of prefibrillar Aβ oligomers. *Mol. Neurodegener.* **2010**, *5*, 57. [CrossRef]
- Sherman, M.A.; Lacroix, M.; Amar, F.; Larson, M.E.; Forster, C.; Aguzzi, A.; Bennett, D.A.; Ramsden, M.; Lesné, S.E. Soluble Conformers of A and Tau Alter Selective Proteins Governing Axonal Transport. *J. Neurosci.* 2016, 36, 9647–9658. [CrossRef] [PubMed]
- 58. Wildsmith, K.R.; Holley, M.; Savage, J.C.; Skerrett, R.; Landreth, G.E. Evidence for impaired amyloid β clearance in Alzheimer's disease. *Alzheimer's Res. Ther.* **2013**, *5*, 33–36. [CrossRef]
- 59. Mader, S.; Brimberg, L. Aquaporin-4 Water Channel in the Brain and Its Implication for Health and Disease. *Cells* **2019**, *8*, 90. [CrossRef]
- Tarasoff-Conway, J.M.; Carare, R.O.; Osorio, R.S.; Glodzik, L.; Butler, T.; Fieremans, E.; Axel, L.; Rusinek, H.; Nicholson, C.; Zlokovic, B.V.; et al. Clearance systems in the brain-implications for Alzheimer disease. *Nat. Rev. Neurol.* 2015, 11, 457–470. [CrossRef]
- 61. Harris, L.D.; Jasem, S.; Licchesi, J.D.F. The Ubiquitin System in Alzheimer's Disease. *Adv. Exp. Med. Biol.* **2020**, *1233*, 195–221. [CrossRef] [PubMed]
- 62. Agrawal, I.; Jha, S. Mitochondrial Dysfunction and Alzheimer's Disease: Role of Microglia. *Front. Aging Neurosci.* **2020**, *12*, 252. [CrossRef] [PubMed]
- 63. Gosselet, F.; Saint-Pol, J.; Candela, P.; Fenart, L. Amyloid-β peptides, Alzheimer's disease and the blood-brain barrier. *Curr. Alzheimer Res.* **2013**, *10*, 1015–1033. [CrossRef]
- 64. Solár, P.; Zamani, A.; Kubíčková, L.; Dubový, P.; Joukal, M. Choroid plexus and the blood–cerebrospinal fluid barrier in disease. *Fluids Barriers CNS* **2020**, *17*, 1–29. [CrossRef]
- 65. Abbott, N.J.; Patabendige, A.A.; Dolman, D.E.; Yusof, S.R.; Begley, D.J. Structure and function of the blood-brain barrier. *Neurobiol. Dis.* **2010**, *37*, 13–25. [CrossRef]
- Pereira, C.D.; Martins, F.; Wiltfang, J.; da Cruz E Silva, O.A.B.; Rebelo, S. ABC Transporters Are Key Players in Alzheimer's Disease. J. Alzheimer's Dis. 2018, 61, 463–485. [CrossRef] [PubMed]
- Mora Lagares, L.; Minovski, N.; Caballero Alfonso, A.Y.; Benfenati, E.; Wellens, S.; Culot, M.; Gosselet, F.; Novič, M. Homology Modeling of the Human P-glycoprotein (ABCB1) and Insights into Ligand Binding through Molecular Docking Studies. *Int. J. Mol. Sci.* 2020, 21, 4058. [CrossRef] [PubMed]
- 68. Erickson, M.A.; Hartvigson, P.E.; Morofuji, Y.; Owen, J.B.; Butterfield, D.A.; Banks, W.A. Lipopolysaccharide impairs amyloid beta efflux from brain: Altered vascular sequestration, cerebrospinal fluid reabsorption, peripheral clearance and transporter function at the blood–brain barrier. *J. Neuroinflammation* **2012**, *9*, 150. [CrossRef]

- 69. Chen, Q.; Liang, B.; Wang, Z.; Cheng, X.; Huang, Y.; Liu, Y.; Huang, Z. Influence of four polymorphisms in ABCA1 and PTGS2 genes on risk of Alzheimer's disease: A meta-analysis. *Neurol. Sci.* **2016**, *37*, 1209–1220. [CrossRef]
- 70. Rawat, V.; Wang, S.; Sima, J.; Bar, R.; Liraz, O.; Gundimeda, U.; Parekh, T.; Chan, J.; Johansson, J.O.; Tang, C.; et al. ApoE4 Alters ABCA1 Membrane Trafficking in Astrocytes. *J. Neurosci.* 2019, *39*, 9611–9622. [CrossRef] [PubMed]
- 71. Ulery, P.G.; Beers, J.; Mikhailenko, I.; Tanzi, R.E.; Rebeck, G.W.; Hyman, B.T.; Strickland, D.K. Modulation of β-Amyloid Precursor Protein Processing by the Low Density Lipoprotein Receptor-related Protein (LRP). Evidence that LRP contributes to the pathogenesis of Alzheimer's disease. *J. Biol. Chem.* 2000, 275, 7410–7415. [CrossRef] [PubMed]
- 72. von Einem, B.; Schwanzar, D.; Rehn, F.; Beyer, A.-S.; Weber, P.; Wagner, M.; Schneckenburger, H.; von Arnim, C.A. The role of low-density receptor-related protein 1 (LRP1) as a competitive substrate of the amyloid precursor protein (APP) for BACE1. *Exp. Neurol.* 2010, 225, 85–93. [CrossRef] [PubMed]
- 73. Liu, C.-C.; Hu, J.; Zhao, N.; Wang, J.; Wang, N.; Cirrito, J.R.; Kanekiyo, T.; Holtzman, D.M.; Bu, G. Astrocytic LRP1 Mediates Brain Aβ Clearance and Impacts Amyloid Deposition. *J. Neurosci.* **2017**, *37*, 4023–4031. [CrossRef]
- 74. Storck, S.; Meister, S.; Nahrath, J.; Meißner, J.N.; Schubert, N.; Di Spiezio, A.; Baches, S.; Vandenbroucke, R.; Bouter, Y.; Prikulis, I.; et al. Endothelial LRP1 transports amyloid-β1–42 across the blood-brain barrier. *J. Clin. Investig.* 2015, *126*, 123–136. [CrossRef] [PubMed]
- 75. Shen, X.; Xia, L.; Liu, L.; Jiang, H.; Shannahan, J.; Du, Y.; Zheng, W. Altered clearance of beta-amyloid from the cerebrospinal fluid following subchronic lead exposure in rats: Roles of RAGE and LRP1 in the choroid plexus. *J. Trace Elem. Med. Biol.* **2020**, *61*, 126520. [CrossRef]
- 76. Kanekiyo, T.; Liu, C.-C.; Shinohara, M.; Li, J.; Bu, G. LRP1 in Brain Vascular Smooth Muscle Cells Mediates Local Clearance of Alzheimer's Amyloid-β. *J. Neurosci.* **2012**, *32*, 16458–16465. [CrossRef]
- 77. Ma, Q.; Zhao, Z.; Sagare, A.P.; Wu, Y.; Wang, M.; Owens, N.C.; Verghese, P.B.; Herz, J.; Holtzman, D.M.; Zlokovic, B.V. Blood-brain barrier-associated pericytes internalize and clear aggregated amyloid-β42 by LRP1-dependent apolipoprotein E isoform-specific mechanism. *Mol. Neurodegener.* 2018, 13, 1–13. [CrossRef]
- 78. Georgievska, B.; Gustavsson, S.; Lundkvist, J.; Neelissen, J.; Eketjäll, S.; Ramberg, V.; Bueters, T.; Agerman, K.; Jureus, A.; Svensson, S.; et al. Revisiting the peripheral sink hypothesis: Inhibiting BACE1 activity in the periphery does not alter β-amyloid levels in the CNS. *J. Neurochem.* 2014, *132*, 477–486. [CrossRef]
- Kim, D.E.; Priefer, R. Therapeutic Potential of Direct Clearance of the Amyloid-β in Alzheimer's Disease. *Brain Sci.* 2020, 10, 93.
  [CrossRef]
- Nakajima, C.; Kulik, A.; Frotscher, M.; Herz, J.; Schäfer, M.K.; Bock, H.H.; May, P. Low Density Lipoprotein Receptor-related Protein 1 (LRP1) Modulates N-Methyl-d-aspartate (NMDA) Receptor-dependent Intracellular Signaling and NMDA-induced Regulation of Postsynaptic Protein Complexes. J. Biol. Chem. 2013, 288, 21909–21923. [CrossRef]
- 81. Dato, V.A.; Sánchez, M.C.; Chiabrando, G.A. LRP1 mediates the IGF-1-induced GLUT1 expression on the cell surface and glucose uptake in Müller glial cells. *Sci. Rep.* **2021**, *11*, 4742. [CrossRef]
- 82. Liu, Q.; Zhang, J.; Zerbinatti, C.; Zhan, Y.; Kolber, B.J.; Herz, J.; Muglia, L.J.; Bu, G. Lipoprotein Receptor LRP1 Regulates Leptin Signaling and Energy Homeostasis in the Adult Central Nervous System. *PLoS Biol.* **2011**, *11*, e1000575. [CrossRef]
- Marzolo, M.-P.; Farfán, P. New Insights into the Roles of Megalin/LRP2 and the Regulation of its Functional Expression. *Biol. Res.* 2011, 44, 89–105. [CrossRef] [PubMed]
- 84. Saunders, A.J.; Bertram, L.; Mullin, K.; Sampson, A.J.; Latifzai, K.; Basu, S.; Jones, J.; Kinney, D.; MacKenzie-Ingano, L.; Yu, S.; et al. Genetic association of Alzheimer's disease with multiple polymorphisms in alpha-2-macroglobulin. *Hum. Mol. Genet.* 2003, 12, 2765–2776. [CrossRef]
- 85. Xu, X.; Wang, Y.; Wang, L.; Liao, Q.; Chang, L.; Xu, L.; Huang, Y.; Ye, H.; Xu, L.; Chen, C.; et al. Meta-Analyses of 8 Polymorphisms Associated with the Risk of the Alzheimer's Disease. *PLoS ONE* **2013**, *8*, e73129. [CrossRef]
- Shen, L.; Jia, J. An Overview of Genome-Wide Association Studies in Alzheimer's Disease. *Neurosci. Bull.* 2016, 32, 183–190. [CrossRef] [PubMed]
- 87. Krimbou, L.; Tremblay, M.; Davignon, J.; Cohn, J.S. Association of apolipoprotein E with alpha2-macroglobulin in human plasma. *J. Lipid Res.* **1998**, *39*, 2373–2386. [CrossRef]
- 88. Whiten, D.R.; Cox, D.; Horrocks, M.H.; Taylor, C.G.; De, S.; Flagmeier, P.; Tosatto, L.; Kumita, J.R.; Ecroyd, H.; Dobson, C.M.; et al. Single-Molecule Characterization of the Interactions between Extracellular Chaperones and Toxic α-Synuclein Oligomers. *Cell Rep.* 2018, 23, 3492–3500. [CrossRef] [PubMed]
- Malito, E.; Hulse, R.E.; Tang, W.-J. Amyloid β-degrading cryptidases: Insulin degrading enzyme, presequence peptidase, and neprilysin. *Cell. Mol. Life Sci.* 2008, 65, 2574–2585. [CrossRef]
- 90. Shen, Y.; Joachimiak, A.; Rosner, M.R.; Tang, W.-J. Structures of human insulin-degrading enzyme reveal a new substrate recognition mechanism. *Nature* **2006**, *443*, 870–874. [CrossRef]
- De Tullio, M.B.; Castelletto, V.; Hamley, I.W.; Adami, P.V.M.; Morelli, L.; Castaño, E.M. Proteolytically Inactive Insulin-Degrading Enzyme Inhibits Amyloid Formation Yielding Non-Neurotoxic Aβ Peptide Aggregates. *PLoS ONE* 2013, *8*, e59113. [CrossRef] [PubMed]
- 92. Cai, Z.; Liu, N.; Wang, C.; Qin, B.; Zhou, Y.; Xiao, M.; Chang, L.; Yan, L.-J.; Zhao, B. Role of RAGE in Alzheimer's Disease. *Cell. Mol. Neurobiol.* **2015**, *36*, 483–495. [CrossRef] [PubMed]

- Liu, B.; Liu, G.; Wang, Y.; Yao, Y.; Wang, G.; Lei, X.; Zhang, N.; Dong, X. Protective Effect of Buyang Huanwu Decoction on Neurovascular Unit in Alzheimer's Disease Cell Model via Inflammation and RAGE/LRP1 Pathway. *Med. Sci. Monit.* 2019, 25, 7813–7825. [CrossRef]
- 94. Kook, S.-Y.; Hong, H.S.; Moon, M.; Ha, C.M.; Chang, S.; Mook-Jung, I. A 1-42-RAGE Interaction Disrupts Tight Junctions of the Blood-Brain Barrier Via Ca2+-Calcineurin Signaling. *J. Neurosci.* 2012, *32*, 8845–8854. [CrossRef] [PubMed]
- 95. Cho, H.J.; Son, S.M.; Jin, S.M.; Hong, H.S.; Shin, D.H.; Kim, S.J.; Huh, K.; Mook-Jung, I. RAGE regulates BACE1 and Aβ generationviaNFAT1 activation in Alzheimer's disease animal model. *FASEB J.* **2009**, *23*, 2639–2649. [CrossRef]
- 96. Tobon-Velasco, J.C.; Cuevas, E.; Torres-Ramos, M.A. Receptor for AGEs (RAGE) as Mediator of NF-kB Pathway Activation in Neuroinflammation and Oxidative Stress. *CNS Neurol. Disord.-Drug Targets* **2014**, *13*, 1615–1626. [CrossRef]
- 97. Hladky, S.B.; Barrand, M.A. Mechanisms of fluid movement into, through and out of the brain: Evaluation of the evidence. *Fluids Barriers CNS* **2014**, *11*, 1–32. [CrossRef] [PubMed]
- Szentistvanyi, I.; Patlak, C.S.; Ellis, R.A.; Cserr, H.F. Drainage of interstitial fluid from different regions of rat brain. Am. J. Physiol. Physiol. 1984, 246, F835–F844. [CrossRef]
- 99. Iliff, J.J.; Wang, M.; Liao, Y.; Plogg, B.A.; Peng, W.; Gundersen, G.A.; Benveniste, H.; Vates, G.E.; Deane, R.; Goldman, S.A.; et al. A Paravascular Pathway Facilitates CSF Flow through the Brain Parenchyma and the Clearance of Interstitial Solutes, Including Amyloid β. *Sci. Transl. Med.* 2012, *4*, 147ra111. [CrossRef] [PubMed]
- 100. Goodman, J.R.; Iliff, J.J. Vasomotor influences on glymphatic-lymphatic coupling and solute trafficking in the central nervous system. *J. Cereb. Blood Flow Metab.* **2019**, *40*, 1724–1734. [CrossRef]
- 101. Hablitz, L.M.; Plá, V.; Giannetto, M.; Vinitsky, H.S.; Stæger, F.F.; Metcalfe, T.; Nguyen, R.; Benrais, A.; Nedergaard, M. Circadian control of brain glymphatic and lymphatic fluid flow. *Nat. Commun.* **2020**, *11*, 4411. [CrossRef]
- 102. Proulx, S.T. Cerebrospinal fluid outflow: A review of the historical and contemporary evidence for arachnoid villi, perineural routes, and dural lymphatics. *Cell. Mol. Life Sci.* 2021, *78*, 2429–2457. [CrossRef]
- Reiber, H. Blood-cerebrospinal fluid (CSF) barrier dysfunction means reduced CSF flow not barrier leakage-conclusions from CSF protein data. Arg. Neuro-Psiquiatr. 2021, 79, 56–67. [CrossRef]
- 104. Pollay, M. The function and structure of the cerebrospinal fluid outflow system. Cereb. Fluid Res. 2010, 7, 9. [CrossRef] [PubMed]
- 105. Coimbra, J.; Marques, D.F.F.; Baptista, S.J.; Pereira, C.M.F.; Moreira, P.I.; Dinis, T.C.P.; Santos, A.E.; Salvador, J.A.R. Highlights in BACE1 Inhibitors for Alzheimer's Disease Treatment. *Front. Chem.* **2018**, *6*, 178. [CrossRef] [PubMed]
- 106. Imbimbo, B.P.; Watling, M. Investigational BACE inhibitors for the treatment of Alzheimer's disease. *Expert Opin. Investig. Drugs* **2019**, *28*, 967–975. [CrossRef] [PubMed]
- 107. Keskin, A.D.; Kekuš, M.; Adelsberger, H.; Neumann, U.; Shimshek, D.R.; Song, B.; Zott, B.; Peng, T.; Förstl, H.; Staufenbiel, M.; et al. BACE inhibition-dependent repair of Alzheimer's pathophysiology. *Proc. Natl. Acad. Sci. USA* 2017, 114, 8631–8636. [CrossRef]
- 108. Fukumoto, H.; Takahashi, H.; Tarui, N.; Matsui, J.; Tomita, T.; Hirode, M.; Sagayama, M.; Maeda, R.; Kawamoto, M.; Hirai, K.; et al. A Noncompetitive BACE1 Inhibitor TAK-070 Ameliorates Aβ Pathology and Behavioral Deficits in a Mouse Model of Alzheimer's Disease. J. Neurosci. 2010, 30, 11157–11166. [CrossRef]
- Sperling, R.; Henley, D.; Aisen, P.S.; Raman, R.; Donohue, M.C.; Ernstrom, K.; Rafii, M.S.; Streffer, J.; Shi, Y.; Karcher, K.; et al. Findings of Efficacy, Safety, and Biomarker Outcomes of Atabecestat in Preclinical Alzheimer Disease. *JAMA Neurol.* 2021, 78, 293–301. [CrossRef]
- 110. Egan, M.F.; Kost, J.; Tariot, P.N.; Aisen, P.S.; Cummings, J.L.; Vellas, B.; Sur, C.; Mukai, Y.; Voss, T.; Furtek, C.; et al. Randomized Trial of Verubecestat for Mild-to-Moderate Alzheimer's Disease. *N. Engl. J. Med.* **2018**, *378*, 1691–1703. [CrossRef] [PubMed]
- 111. Shugart, J.; Strobel, G. Cognitive decline trips up API trials of BACE inhibitor. *Alzheimer Research Forum*, 12 July 2019.
- 112. Menendez-Gonzalez, M.; Perez-Pinera, P.; Martinez-Rivera, M.; Muniz, A.L.; Vega, J.A. Immunotherapy for Alzheimer's Disease: Rational Basis in Ongoing Clinical Trials. *Curr. Pharm. Des.* **2011**, *17*, 508–520. [CrossRef]
- 113. Morrone, C.D.; Liu, M.; Black, S.E.; McLaurin, J. Interaction between therapeutic interventions for Alzheimer's disease and physiological Aβ clearance mechanisms. *Front. Aging Neurosci.* **2015**, *7*, 64. [CrossRef]
- 114. Xiang, Y.; Bu, X.-L.; Liu, Y.-H.; Zhu, C.; Shen, L.-L.; Jiao, S.-S.; Zhu, X.-Y.; Giunta, B.; Tan, J.; Song, W.; et al. Physiological amyloid-beta clearance in the periphery and its therapeutic potential for Alzheimer's disease. *Acta Neuropathol.* 2015, 130, 487–499. [CrossRef] [PubMed]
- 115. Henderson, S.J.; Andersson, C.; Narwal, R.; Janson, J.; Goldschmidt, T.J.; Appelkvist, P.; Bogstedt, A.; Steffen, A.-C.; Haupts, U.; Tebbe, J.; et al. Sustained peripheral depletion of amyloid-β with a novel form of neprilysin does not affect central levels of amyloid-β. *Brain* 2014, *137*, 553–564. [CrossRef] [PubMed]
- 116. Davis, J.; Xu, F.; Deane, R.; Romanov, G.; Previti, M.L.; Zeigler, K.; Zlokovic, B.V.; Van Nostrand, W.E. Early-onset and Robust Cerebral Microvascular Accumulation of Amyloid β-Protein in Transgenic Mice Expressing Low Levels of a Vasculotropic Dutch/Iowa Mutant Form of Amyloid β-Protein Precursor. J. Biol. Chem. 2004, 279, 20296–20306. [CrossRef]
- 117. Moreira, P.I.; Sayre, L.M.; Zhu, X.; Nunomura, A.; Smith, M.A.; Perry, G. Detection and Localization of Markers of Oxidative Stress by In Situ Methods: Application in the Study of Alzheimer Disease. *Methods Mol. Biol.* 2010, *610*, 419–434. [CrossRef]
- 118. Boada, M.; López, O.L.; Olazarán, J.; Núñez, L.; Pfeffer, M.; Paricio, M.; Lorites, J.; Piñol-Ripoll, G.; Gámez, J.E.; Anaya, F.; et al. A randomized, controlled clinical trial of plasma exchange with albumin replacement for Alzheimer's disease: Primary results of the AMBAR Study. *Alzheimer's Dement.* 2020, 16, 1412–1425. [CrossRef]

- 119. Jin, W.-S.; Shen, L.-L.; Bu, X.-L.; Zhang, W.-W.; Chen, S.-H.; Huang, Z.-L.; Xiong, J.-X.; Gao, C.-Y.; Dong, Z.; He, Y.-N.; et al. Peritoneal dialysis reduces amyloid-beta plasma levels in humans and attenuates Alzheimer-associated phenotypes in an APP/PS1 mouse model. Acta Neuropathol. 2017, 134, 207–220. [CrossRef]
- 120. Kitaguchi, N.; Tatebe, H.; Sakai, K.; Kawaguchi, K.; Matsunaga, S.; Kitajima, T.; Tomizawa, H.; Kato, M.; Sugiyama, S.; Suzuki, N.; et al. Influx of Tau and Amyloid-β Proteins into the Blood During Hemodialysis as a Therapeutic Extracorporeal Blood Amyloid-β Removal System for Alzheimer's Disease. J. Alzheimer's Dis. 2019, 69, 687–707. [CrossRef] [PubMed]
- 121. González, M.M. Implantable Systems for Continuous Liquorpheresis and CSF Replacement. Cureus 2017, 9, e1022. [CrossRef]
- 122. Bolash, R.; Mekhail, N. Intrathecal Pain Pumps: Indications, patient selection, techniques, and outcomes. *Neurosurg. Clin. N. Am.* **2014**, *25*, 735–742. [CrossRef]
- 123. Shirotani, K.; Tsubuki, S.; Iwata, N.; Takaki, Y.; Harigaya, W.; Maruyama, K.; Kiryu-Seo, S.; Kiyama, H.; Iwata, H.; Tomita, T.; et al. Neprilysin Degrades Both Amyloid β Peptides 1–40 and 1–42 Most Rapidly and Efficiently among Thiorphan- and Phosphoramidon-sensitive Endopeptidases. *J. Biol. Chem.* 2001, 276, 21895–21901. [CrossRef]
- 124. Nalivaeva, N.N.; Zhuravin, I.A.; Turner, A.J. Neprilysin expression and functions in development, ageing and disease. *Mech. Ageing Dev.* **2020**, *192*, 111363. [CrossRef]
- 125. Salazar, J.; Rojas-Quintero, J.; Cano, C.; Pérez, J.L.; Ramirez, P.P.; Carrasquero, R.; Torres, W.; Espinoza, C.; Chacín-González, M.; Bermudez, V. Neprilysin: A Potential Therapeutic Target of Arterial Hypertension? *Curr. Cardiol. Rev.* 2020, *16*, 25–35. [CrossRef]
- 126. Marr, R.A.; Rockenstein, E.; Mukherjee, A.; Kindy, M.S.; Hersh, L.B.; Gage, F.H.; Verma, I.M.; Masliah, E. Neprilysin Gene Transfer Reduces Human Amyloid Pathology in Transgenic Mice. J. Neurosci. 2003, 23, 1992–1996. [CrossRef]
- 127. Leissring, M.A.; Farris, W.; Chang, A.Y.; Walsh, D.M.; Wu, X.; Sun, X.; Frosch, M.P.; Selkoe, D.J. Enhanced proteolysis of beta-amyloid in APP transgenic mice prevents plaque formation, secondary pathology, and premature death. *Neuron* 2003, 40, 1087–1093. [CrossRef]
- 128. Iwata, N.; Tsubuki, S.; Takaki, Y.; Shirotani, K.; Lu, B.; Gerard, N.P.; Gerard, C.; Hama, E.; Lee, H.J.; Saido, T.C. Metabolic regulation of brain Abeta by neprilysin. *Science* **2001**, 292, 1550–1552. [CrossRef]
- 129. Hemming, M.L.; Patterson, M.; Reske-Nielsen, C.; Lin, L.; Isacson, O.; Selkoe, D.J. Reducing Amyloid Plaque Burden via Ex Vivo Gene Delivery of an Aβ-Degrading Protease: A Novel Therapeutic Approach to Alzheimer Disease. *PLoS Med.* 2007, 4, e262. [CrossRef]
- 130. El-Amouri, S.S.; Zhu, H.; Yu, J.; Marr, R.; Verma, I.M.; Kindy, M.S. Neprilysin: An Enzyme Candidate to Slow the Progression of Alzheimer's Disease. *Am. J. Pathol.* **2008**, *172*, 1342–1354. [CrossRef]
- Webster, C.I.; Burrell, M.; Olsson, L.-L.; Fowler, S.B.; Digby, S.; Sandercock, A.; Snijder, A.; Tebbe, J.; Haupts, U.; Grudzinska, J.; et al. Engineering Neprilysin Activity and Specificity to Create a Novel Therapeutic for Alzheimer's Disease. *PLoS ONE* 2014, 9, e104001. [CrossRef]
- Kamble, S.; Barale, S.; Dhanavade, M.; Sonawane, K. Structural significance of Neprylysin from *Streptococcus suis GZ1* in the degradation of Aβ peptides, a causative agent in Alzheimer's disease. *Comput. Biol. Med.* 2021, 136, 104691. [CrossRef] [PubMed]
- 133. Barua, N.U.; Miners, J.S.; Bienemann, A.S.; Wyatt, M.J.; Welser, K.; Tabor, A.B.; Hailes, H.C.; Love, S.; Gill, S.S. Convection-Enhanced Delivery of Neprilysin: A Novel Amyloid-β-Degrading Therapeutic Strategy. J. Alzheimer's Dis. 2012, 32, 43–56. [CrossRef] [PubMed]
- Campos, C.R.; Kemble, A.M.; Niewoehner, J.; Freskgård, P.-O.; Urich, E. Brain Shuttle Neprilysin reduces central Amyloid-β levels. *PLoS ONE* 2020, 15, e0229850. [CrossRef] [PubMed]
- 135. Rofo, F.; Yilmaz, C.U.; Metzendorf, N.; Gustavsson, T.; Beretta, C.; Erlandsson, A.; Sehlin, D.; Syvänen, S.; Nilsson, P.; Hultqvist, G. Enhanced neprilysin-mediated degradation of hippocampal Aβ42 with a somatostatin peptide that enters the brain. *Theranostics* 2021, 11, 789–804. [CrossRef] [PubMed]
- 136. Castro, M.A.; Hadziselimovic, A.; Sanders, C.R. The vexing complexity of the amyloidogenic pathway. *Protein Sci.* 2019, 28, 1177–1193. [CrossRef] [PubMed]
- 137. Nedergaard, M.; Goldman, S.A. Glymphatic failure as a final common pathway to dementia. *Science* **2020**, *370*, 50–56. [CrossRef] [PubMed]