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# Cyclooxygenase-2 expression correlates with development, progression, metastasis, and prognosis of osteosarcoma: a meta-analysis and trial sequential analysis

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#### Keywords

clinical features; cyclooxygenase-2; expression; osteosarcoma; prognosis

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Cyclooxygenase-2 (COX-2), a key enzyme in arachidonic acid metabolism, is involved in several cancers, including osteosarcoma. The prognostic significance of COX-2 in osteosarcoma remains controversial. This study was to analyze the potential clinical and prognostic effects of COX-2 protein expression in patients with osteosarcoma. Eligible articles were searched via online databases. The combined odds ratios (ORs) or hazard ratios (HRs) with their 95% confidence intervals (95% CIs) were calculated using the random-effects model. Trial sequential analysis (TSA) was applied to analyze the required information size and determine the reliability of the evidence. Twenty-three studies on COX-2 expression were identified, which included a total of 1084 patients with malignant osteosarcoma and 247 patients with benign osteochondroma. COX-2 protein expression in osteosarcoma was higher than in benign osteochondroma (OR = 7.66, P < 0.001). COX-2 expression was not correlated with age, gender, tumor location, cancer histology, or necrosis (P > 0.1), but was significantly associated with tumor grade (high grade vs. low grade: OR = 4.81, P < 0.001), clinical stage (stage 3-4 vs. stage 1-2: OR = 4.89, P < 0.001), and metastasis (yes vs. no: OR = 3.53, P < 0.001). Based on TSA results, we suggest that additional studies are not required to examine osteosarcoma vs. benign osteochondroma, tumor grade, clinical stage, or metastasis. No heterogeneity was observed in these analyses. COX-2 expression is linked to poor prognosis in metastasis-free survival, overall survival, and relapse-free survival, as indicated by multivariate analysis. Therefore, the expression of COX-2 may correlate with the development, progression, metastasis, and poor prognosis of osteosarcoma.

Osteosarcoma is the most frequent primary bone sarcoma that occurs mainly in children and adolescents [1]. Approximately 4.4 per million are diagnosed with osteosarcoma every year [1,2]. Osteogenic sarcoma consists of several main histotypes: osteoblastic, chondroblastic, and fibroblastic osteosarcoma [3]. Osteogenic osteosarcoma is the most common pathological subtype [3]. Although the biology of osteosarcoma is

#### Abbreviations

95% CI, 95% confidence interval; APIS, a priori anticipated information size; COX-2, cyclooxygenase-2; HR, hazard ratio; IHC, immunohistochemistry; OR, odds ratio; PRISMA, Preferred Reporting Items for Systematic Reviews and Meta-Analyses; PTGS, prostaglandin-endoperoxide synthase; RRR, relative risk reduction; TSA, trial sequential analysis.

complex, treatment options have not significantly changed over the past several decades [4]. Osteosarcoma can be caused by other types of cancer and some environmental factors such as viruses and radiation [5]. This rare disease is treated by advanced surgery and adjuvant/neoadjuvant chemotherapy [6,7]. Current treatment for osteosarcoma achieves a 5-year survival rate of 20% to 30% in metastatic or recurrent osteosarcoma patients [8]. When diagnosed, 40% of metastasis occurs in osteosarcoma patients at an advanced stage [9].

Increasing evidence shows that molecular mechanisms are correlated with the development, progresand prognosis of osteosarcoma [10–13]. sion. Cyclooxygenase (COX), also called prostaglandinendoperoxide synthase (PTGS), is a key enzyme catalyzing the conversion of arachidonic acid to prostaglandin. COX has two isoforms: constitutive COX (COX-1) and inducible COX (COX-2) [14]. The COX-2 gene, encoding the inducible isozyme, is activated by growth factors, inflammatory stimuli, or carcinogenic factors, and its expression is usually undetectable in most normal tissues [15,16]. Furthermore, COX-2 is involved in the regulation of cell proliferation and apoptosis, stimulation of angiogenesis, and invasion [17-19]. Some studies suggest that COX-2 expression may correlate with the development and progression of several types of human malignancies [20-22]. The

expression of COX-2 is related to an increased risk of skin cancer [23], as well as to tumor size, lymph node metastasis, and poor prognosis in breast [24] and colorectal and ovarian cancers [25,26]. Nevertheless, no significant role of the expression of COX-2 was found on the survival of patients with non-small-cell lung cancer [27]. Importantly, COX-2 may regulate the genesis and progression of osteosarcoma [28] and COX-2 expression has been reported in patients with osteosarcoma [29–32]. In earlier studies, the percentage of osteosarcoma patients with COX-2-positive expression ranged from 16.7% to 81.2% [33,34]. Therefore, further investigations on the clinical significance of COX-2 expression in osteosarcoma patients are required.

Benign bone tumors include osteochondroma, osteoma, osteoid osteoma, osteoblastoma, giant cell tumor, aneurysmal bone cyst, fibrous dysplasia, and enchondroma [35]. Osteochondroma is the most common nonmalignant bone tumor, accounting for approximately 35% of all benign bone tumors [35]. However, the malignant transformation into osteosarcoma has not been sufficiently studied, and its mechanism is still unclear [36]. The sample sizes of the individual studies were generally small [29–34], which might have led to a lack of power in the statistical analysis.

In this study, the present meta-analysis integrated all available publications in a larger population to



Fig. 1. Flow diagram of the study selection.

evaluate whether COX-2 expression is associated with an increased risk of osteosarcoma in a comparison between osteosarcoma and benign osteochondroma cases. Additionally, we analyzed the possible clinicopathological and prognostic significance of COX-2 expression in patients with osteosarcoma.

# **Materials and methods**

#### Search strategy

A systematic search was performed of different electronic databases, including PubMed, Embase, EBSCO, Wanfang, and CNKI, for eligible papers published before June 2017. The following key words and search terms were used: (COX-2 OR COX2 OR Cyclooxygenase-2 OR PTGS2 OR Prostaglandin Synthase) AND (expression OR expressed) AND (osteosarcoma OR osteogenic sarcoma). Moreover, the reference lists of the included publications were also retrieved to find other potentially relevant studies.

### **Selection criteria**

The following inclusion criteria were applied in this meta-analysis: (a) confirmation of osteosarcoma by

histopathological examination; (b) COX-2 protein expression tissue analyses using immunohistochemistry (IHC); (c) studies with complete information concerning the rate of COX-2 protein expression in malignant and benign osteochondroma; (d) studies with sufficient information to assess the correlation of COX-2 expression with clinical characteristics of patients with osteosarcoma; and (e) studies providing sufficient information to evaluate the prognostic effect of COX-2 expression in osteosarcoma by multivariate analysis. Only the complete publications with more extensive information or larger populations were included when the authors had published more than one article using overlapping study populations.

### Data extraction and study selection

The following data were extracted from the eligible publications: the last name of the first author; year of publication; country; ethnicity; case number (osteosarcoma and benign osteochondroma); immunohistochemical staining patterns; cutoff values; median or mean years; tumor stage; frequency of COX-2 expression; clinical characteristics, such as age ( $\geq$ 20 years vs.  $\leq$ 20 years), gender (male vs. female), tumor location (femur vs. nonfemur), cancer histology (osteogenic osteosarcoma vs. nonosteogenic osteosarcoma), tumor grade (grade 3–4 vs. grade 1–2), clinical stage (stage

Table 1. Basic characteristics of the included publications. E+, positive expression status; IHC, immunohistochemistry; MA, multivariate analysis; N, study population; NA, not applicable.

First author	Country	Ethnicity	Age	Stage	Location	Osteosarcoma N (E+ %)	Osteochondroma N (E+ %)	Clinical features	MA- survival	IHC cutoff (positivity)
Dickens [46]	USA	Caucasians	NA	NA	Cytoplasm	99 (66.7)		Yes	No	25%
Jiang [47]	China	Asians	NA	NA	Cytoplasm	31 (67.7)		Yes	No	5%
Zhu [48]	China	Asians	25.5	NA	Cytoplasm	31 (61.3)	30 (6.7)	Yes	No	5%
Hosono [34]	Japan	Asians	29	NA	Cytoplasm	30 (16.7)		Yes	No	10%
Masi [33]	Italy	Caucasians	19	NA	Cytoplasm	42 (59.5)		Yes	No	20%
Ou [49]	China	Asians	18.6	1–3	Cytoplasm	49 (77.6)	20 (35)	Yes	No	5%
Wang [50]	China	Asians	18.68	NA	Cytoplasm	40 (67.5)	20 (15)	No	No	5%
Rodriguez [32]	USA	Mix	17	NA	NA	36 (47.2)		Yes	No	10%
Geng [51]	China	Asians	19.3	1–3	Cytoplasm	59 (69.5)		Yes	No	10%
Zhan [52]	China	Asians	19	1–3	Cytoplasm	38 (57.9)	20 (25)	Yes	No	5%
Urakawa [31]	Japan	Asians	15	2	NA	51 (23.5)		Yes	Yes	80%
Liu [53]	China	Asians	21.8	2–3	Cytoplasm	50 (70)	20 (30)	Yes	No	5%
Liao [54]	China	Asians	42.1	1–3	Cytoplasm	57 (70.2)	18 (11.1)	Yes	No	10%
Huang [55]	China	Asians	NA	1–3	Cytoplasm	37 (86.5)		Yes	No	30%
Boulytcheva [30]	Russia	Caucasians	NA	1–4	Cytoplasm	34 (32.4)		Yes	Yes	10%
Li [56]	China	Asians	19	1–3	Cytoplasm	85 (62.4)	20 (15)	Yes	No	10%
Xu [57]	China	Asians	20	1–3	Cytoplasm	28 (78.6)	25 (36)	Yes	No	5%
Ma [58]	China	Asians	NA	1–3	Cytoplasm	45 (80)		Yes	No	10%
Duan [29]	China	Asians	NA	NA	Cytoplasm	30 (73.3)	20 (25)	Yes	No	0%
Chen [59]	China	Asians	18.5	2–3	Cytoplasm	49 (49)		Yes	No	10%
Meng [60]	China	Asians	NA	NA	Cytoplasm	52 (71.2)	40 (35)	No	No	10%
Lian [61]	China	Asians	21.1	2–3	Cytoplasm	35 (82.9)	14 (14.3)	Yes	No	0%
Zhu [62]	China	Asians	27.5	NA	Cytoplasm	76 (68.4)		Yes	No	5%

Dickens [46] 11/24 15/21 Jiang [47] Zhu [48] Hosono [34] 1/16 4/14 4/16 1/14 4/18 Masi [33] Ou [49] Rodriguez [32] Cou [49] Rodriguez [32] Rodriguez [32] Rodr	15/21 6/9 1/14 4/18 3/7 9/21	1/12 8/15		E/N	E/N	E/N	3-4 E/N	1–2 E/N	Necrosis E/N	No E/N	Metastasis E/N	No E/N
Janug 14-1 Zhu [48] 1/16 4/14 4/16 1/14 4/18 Hosono [34] 1/16 4/14 4/16 1/14 4/18 Masi [33] Ou [49] 7/8 3/7 9/21 Geng [51] 5/20 7/31 9/33 3/18 Liu [53] Liu [53] Liu [53] Liu [53] Liu [53] Liu [53] Liu [53] Liu [53] Liu [53] Liu [53] X/18 1/18 1/18 1/18 1/18 1/18 1/18 1/18	6/9 1/14 4/18 3/7 9/21	1/12 8/15							6/9	8/18	24/32 14/17	23/35
Hosono [34] 1/16 4/14 4/16 1/14 4/18 Masi [33] Ou [49] Nou [49] Rodriguez [32] Geng [51] Zhan [52] Urakawa [31] 5/20 7/31 9/33 3/18 Liu [53] Liu [53] Liu [53] Liu [53] Liao [54] Huang [55] Boulytcheva [30] Li [56] Xu [57]	1/14 4/18 3/7 9/21	1/12 8/15			9/10	10/21					14/17 14/17	5/14
Masi [33] Ou [49] Rodriguez [32] Geng [51] Zhan [52] Urakawa [31] 5/20 7/31 9/33 3/18 Liu [53] Liu [53] Liao [54] Huang [55] Boulytcheva [30] Li [56] Xu [57]	3/7 9/21	8/15	0/16	5/14	5/26	0/4						
Ou [49] Rodriguez [32] 7/8 3/7 9/21 Geng [51] Zhan [52] 5/20 7/31 9/33 3/18 Liu [53] Liu [53] Liao [54] Huang [55] Boulytcheva [30] Li [56] Xu [57]	3/7 9/21	8/15			23/32	2/10						
Rodriguez [32] 7/8 3/7 9/21 Geng [51] Zhan [52] Urakawa [31] 5/20 7/31 9/33 3/18 Liu [53] Liao [54] Huang [55] Boulytcheva [30] Li [56] Xu [57]	3/7 9/21	8/15	15/19	23/30			12/13	26/36			12/13	26/36
Geng [51] Zhan [52] Urakawa [31] 5/20 7/31 9/33 3/18 Liu [53] Liao [54] Huang [55] Boulytcheva [30] Li [56] Xu [57]	050		6/8	11/28					7/14	10/22	6/10	11/26
Zhan [52] Urakawa [31] 5/20 7/31 9/33 3/18 Liu [53] Liao [54] Huang [55] Boulytcheva [30] Li [56] Xu [57]	0110		23/35	18/24							7/8	34/51
Urakawa [31] 5/20 7/31 9/33 3/18 Liu [53] Liao [54] Huang [55] Boulytcheva [30] Li [56] Xu [57]	01/0						8/11	14/27				
Liu [53] Liao [54] Huang [55] Boulytcheva [30] Li [56] Xu [57]	0/10								7/18	5/33	8/18	4/33
Liao [54] Huang [55] Boulytcheva [30] Li [56] Xu [57]							14/15	21/35				
Huang [55] Boulytcheva [30] Li [56] Xu [57]			15/22	25/35			22/23	18/34				
Boulytcheva [30] Li [56] Xu [57]			12/14	19/23			10/11	22/26			15/16	17/21
Li [56] Xu [57]											6/11	5/23
Xu [57]					25/31	28/54	21/25	32/60			36/48	17/37
			11/14	11/14			8/8	14/20				
Ma [58] 29/37 7/8	7/8		23/31	12/14			12/13	24/32				
Duan [29]			9/13	13/17	6/2	15/21						
Chen [59]									12/29	12/20		
Lian [61] 18/22 11/13	11/13						12/12	17/23			22/24	7/11
Zhu [62]					23/25	29/51						

Table 2. Basic characteristics of the eligible studies with clinical characteristics.

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Fig. 2. Forest plot of the association of COX-2 immunoexpression between osteosarcoma and benign osteochondroma (OR = 7.66, P < 0.001).

3–4 vs. stage 1–2), necrosis ( $\geq$ 90% vs. <90%), and metastasis (yes vs. no); and the prognosis from multivariate analysis. We analyzed data from 2 × 2 tables. The quality of the eligible studies was in accordance with the guidelines of the Cochrane Collaboration and the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) statement criteria [37]. A total number of 391 publications were found: 83 records in PubMed, 96 records in Embase, 77 records in EBSCO, 75 records in Wanfang, and 60 records in CNKI. Final 23 studies were identified in this meta-analysis. To categorize a patient as COX-2-positive or COX-2-negative, the COX-2 expression, determined using IHC staining, was considered positive or negative based on the cutoff values of the original articles.

### Statistical analysis

Data analysis was performed using the STATA software (version 12.0; Stata Corporation, College Station, TX, USA). The combined odds ratios (ORs) and their 95% confidence intervals (95% CIs) were calculated to estimate the relationships of COX-2 expression in osteosarcoma and benign osteochondroma. The correlations of COX-2 expression with the clinical characteristics of osteosarcoma patients were also analyzed using the overall ORs and their 95% CIs. The overall hazard ratios (HRs) with their 95% CIs were determined to evaluate the prognostic effect of COX-2 expression in osteosarcoma patients for multivariate analysis. Heterogeneity among the studies was detected using the Cochran's Q statistic [38]. The random-effects model was used in this meta-analysis. Subgroup, meta-regression, and sensitivity analyses were conducted of the results with substantial heterogeneity (P < 0.1), to explain the potential sources of heterogeneity [39,40]. The possible publication bias was measured using Egger's test in more than eight studies [41].

#### **Trial sequential analysis**

Trial sequential analysis (TSA) was performed to reduce type I error and to assess the required information size for determination of the statistical significance [42,43]. In the metaanalysis, the type I and type II errors were considered to be 5% and 20%, respectively. The relative risk reduction (RRR) was set to be of 20% for the outcome, and a statistical test power of 80% was defined. The cumulative Z-curve crossed the trial sequential monitoring boundary or the

Study			%
ID		OR (95% CI)	Weight
Tumor grade			
Zhu (2005)		9.90 (1.06, 92.66)	4.25
Hosono (2007)	*	2.30 (0.11, 49.51)	2.26
Masi (2007)	· · ·	10.22 (1.81, 57.69)	7.10
Li (2010)	<b>—</b> •—	3.87 (1.37, 10.93)	19.72
Duan (2012)	•	1.40 (0.22, 8.77)	6.32
Zhu (2016)	•	8.72 (1.86, 41.00)	8.88
Subtotal ( $I^2 = 0.0\%$ , $P = 0.573$ )	$\diamond$	4.81 (2.48, 9.32)	48.55
Clinical stage			
Ou (2007) –		4 62 (0 53, 40 28)	4.53
Zhang (2008)	•	2.48 (0.54, 11.40)	9.13
Liu (2009)		9.33 (1.10, 79.21)	4.65
Liao (2009)	•	19.56 (2.36, 161.97)	4.76
Huang (2009) —	•	1.82 (0.18, 18.41)	3.97
Li (2010)		4.59 (1.41, 15.00)	15.19
Xu (2010) —	*	7.62 (0.38, 152.83)	2.37
Ma (2011) —	•	4.00 (0.45, 35.79)	4.43
Lian (2013) —		9.29 (0.48, 180.38)	2.42
Subtotal ( $I^2 = 0.0\%, P = 0.870$ )	$\diamond$	4.89 (2.57, 9.30)	51.45
Overall			100.00
NOTE: Weights are from random-effects	analysis		
0.00554	1 18	0	

Fig. 3. Forest plot of the correlation of COX-2 immunoexpression with the tumor grade (OR = 4.81, P < 0.001) and clinical stage (OR = 4.89, P < 0.001).

required study population information, which suggested that the conclusion drawn in the meta-analysis was positive; otherwise, more studies with larger sample sizes were indicated to be necessary for consistency in the evidence [44,45].

# Results

# Characteristics of the relevant publications

Figure 1 illustrates the above-described search strategy. After the search of the relevant databases, 23 studies with a total of 1084 patients with osteosarcoma and 247 patients with benign osteochondroma were eventually included in the current meta-analysis. All these studies published from 2003 to 2016 had used IHC analysis of COX-2 expression [29–34,46– 62]. Eleven of these eligible studies compared the expression of COX-2 between osteosarcoma and benign osteochondroma. The associations between COX-2 expression and the clinical features of patients with osteosarcoma were examined in 21 studies. Two original studies reported the prognostic role of COX-2 expression using multivariate analysis in patients with osteosarcoma. The general characteristics of the eligible papers are listed in Table 1 and Table 2.

# Association between COX-2 expression and osteosarcoma development

The data from 11 comparative studies, including of 495 patients with osteosarcoma vs. 247 benign



Fig. 4. Forest plot of the correlation of COX-2 immunoexpression with metastasis (OR = 3.53, P < 0.001).

osteochondroma patients, revealed that COX-2 expression was significantly more increased in osteosarcoma than in benign osteochondroma (OR = 7.66, 95% CI = 5.25-11.17, P < 0.001) (Fig. 2).

# Association of COX-2 expression with tumor grade and clinical stage in osteosarcoma

Six studies with 294 osteosarcoma patients found a significant relationship between COX-2 expression and tumor grade (high grade vs. low grade: OR = 4.81, 95% CI = 2.48-9.32, P < 0.001) (Fig. 3).

Additionally, nine studies with 424 osteosarcoma patients discovered a significant correlation was observed between COX-2 expression and clinical stage (stage 3–4 vs. stage 1–2: OR = 4.89, 95% CI = 2.57–9.30, P < 0.001) (Fig. 3).

# Association of COX-2 expression with metastasis in osteosarcoma

The results obtained in 11 studies with 515 osteosarcoma patients demonstrated that COX-2 expression was significantly associated with metastasis in osteosarcoma (OR = 3.53, 95% CI = 2.27–5.51, P < 0.001) (Fig. 4).

## Prognostic role of COX-2 expression for multivariate analysis in osteosarcoma

Cyclooxygenase-2 expression was linked to worse metastasis-free survival (P = 0.029) in 51 osteosarcoma patients [31]. Moreover, Boulytcheva *et al.* [30] reported that COX-2 expression in 34 patients with osteosarcoma was associated with poor overall survival and relapse-free survival (P < 0.05). Nonetheless, additional studies are needed to further validate the findings concerning the prognostic effect of COX-2 expression established by multivariate survival analysis in osteosarcoma.

### Heterogeneity and publication bias

No significant heterogeneity was detected in our metaanalysis (all Ps > 0.1) (Figs 2–6).



**Fig. 5.** Forest plot of the possible publication bias determined using Egger's test (osteosarcoma vs. benign osteochondroma: P < 0.001), cancer histology (P = 0.958), clinical stage (P = 0.397), and metastasis (P = 0.186).



Fig. 6. Trial sequential analysis of COX-2 immunoexpression in osteosarcoma vs. benign osteochondroma. The cumulative Z-curve crossed the trial sequential monitoring boundary, suggesting that the result was conclusive.

Egger's test was applied to measure the publication bias, and significant heterogeneity was found in the comparison of osteosarcoma and benign osteochondroma (P < 0.001) (Fig. 5). However, no evidence of publication bias was found between COX-2 expression and metastasis, clinical stage,



Fig. 7. Trial sequential analysis of COX-2 immunoexpression in relation to tumor grade. The cumulative Z-curve crossed the trial sequential monitoring boundary, suggesting that the result was reliable.

or cancer histology in osteosarcoma (P > 0.05) (Fig. 5).

### **Trial sequential analysis**

Based on the a priori anticipated information size (APIS) method, when osteosarcoma was compared to benign osteochondroma (Fig. 6), the patients with

high-grade cancer were compared to patients with lowgrade cancer (Fig. 7), advanced-stage patients were compared to early-stage patients (Fig. 8), and the patients with metastasis were compared to patients without metastasis (Fig. 9). The cumulative Z-curve crossed the trial sequential monitoring boundary for the above analyses, which suggested that these results were reliable and firm.



Fig. 8. Trial sequential analysis of COX-2 immunoexpression in relation to clinical stage. The cumulative Z-curve crossed the trial sequential monitoring boundary, suggesting that the result was reliable.



Fig. 9. Trial sequential analysis of COX-2 immunoexpression in relation to metastasis. The cumulative Z-curve crossed the trial sequential monitoring boundary, suggesting that the result was conclusive.

# Association of COX-2 expression with age, gender, or tumor location in osteosarcoma

We explored whether the expression of COX-2 was correlated with the clinical characteristics of patients with osteosarcoma. The results showed that COX-2 expression was not associated with age (two studies with 81 cases:  $\geq 20$  years vs.  $\leq 20$  years), gender (seven studies with 252 cases: male vs. female), or tumor location (two studies with 66 cases: femur vs. nonfemur) of osteosarcoma (OR = 0.56, 95% CI = 0.09–3.49, P = 0.534; OR = 1.05, 95% CI = 0.47–2.37, P = 0.903; OR = 1.06, 95% CI = 0.26–4.41, P = 0.933, respectively) (Fig. 10).

# Association of COX-2 expression with histology and necrosis in osteosarcoma

The results showed no significant correlation between COX-2 expression and cancer histology (nine studies with 371 cases: osteogenic osteosarcoma vs. nonosteogenic osteosarcoma: OR = 0.87, 95% CI = 0.52–1.45, P = 0.583) (Fig. 11) or necrosis (four studies with 163 cases:  $\geq 90\%$  vs. < 90%: OR = 1.39, 95% CI = 0.54–3.56, P = 0.491) (Fig. 11).

# Discussion

Using the IHC method, the authors of a number of earlier studies have demonstrated that COX-2 is frequently expressed in a variety of human cancers, including lung cancer [30], breast carcinoma [20],

colorectal cancer [63], and nasopharyngeal carcinoma [64]. COX-2 immunoexpression was also detected in osteosarcoma [29,30]. The meta-analysis of Jiao et al. [65] involving 14 studies reported that COX-2 expression was correlated with the clinical stage and metastasis of osteosarcoma. However, the meta-analysis of Wang et al. involving nine studies established that COX-2 expression was not associated with the metastasis and clinical stage of osteosarcoma [66]. To compare whether COX-2 expression was different in osteosarcoma and benign osteochondroma, the results from 11 studies (495 osteosarcomas vs. 247 benign osteochondromas) demonstrated that COX-2 expression in osteosarcoma was notably higher than in benign osteochondroma (OR = 7.66, P < 0.001), which suggested that COX-2 expression as a potential marker using the IHC method could distinguish osteosarcoma and benign osteochondroma. TSA revealed that true-positive results were obtained in the comparative analysis of osteosarcoma and benign osteochondroma.

On further evaluation, the relationship between COX-2 expression and the clinical characteristics of patients with osteosarcoma showed no significant correlation of COX-2 expression with age, gender, tumor location, cancer histology, and necrosis across each study (Figs 10 and 11). And these results from all pooled studies remained constant. COX-2 expression was not associated with tumor grade in osteosarcoma [29,34], but Masi *et al.* [33] reported that COX-2 expression was correlated with tumor grade. Pooled

Study		%
ID	OR (95% CI)	Weight
Age ( $\geq$ 20 years vs $\leq$ 20 years)		
Hosono (2007)	0.17 (0.02, 1.72)	5.43
Urakawa (2009)	1.14 (0.31, 4.26)	14.02
Subtotal ( <i>I</i> <sup>2</sup> = 50.1%, <i>P</i> = 0.157)	0.56 (0.09, 3.49)	19.45
Gender (male vs female)		
Dickens (2003)	0.34 (0.10, 1.17)	15.28
Zhu (2005)	0.72 (0.14, 3.67)	10.10
Hosono (2007)	4.33 (0.42, 44.43)	5.45
Rodriguez (2008)	9.33 (0.71, 122.57)	4.54
Urakawa (2009)	1.88 (0.44, 8.05)	12.02
Ma (2011) • • •	0.52 (0.06, 4.85)	5.85
Lian (2013)	0.82 (0.13, 5.23)	8.10
Subtotal (/ <sup>2</sup> = 30.2%, P = 0.198)	1.05 (0.47, 2.37)	61.34
Tumor location (femur vs nonfemur)		
Hosono (2007)	3.14 (0.31, 32.28)	5.44
Rodriguez (2008)	0.66 (0.17, 2.49)	13.77
Subtotal (/2 = 24.2%, P = 0.251)	1.06 (0.26, 4.41)	19.21
Overall		100.00
NOTE: Weights are from random-effects analysis		
0.00816 1 12	3	

Fig. 10. Forest plot of the association of COX-2 immunoexpression with age factor, gender, and tumor location (P > 0.1).

data from six studies with 294 cases revealed that COX-2 expression was positively linked to tumor grade (OR = 4.81) and clinical stage (OR = 4.89). Dickens et al. [46] found no significant association between COX-2 expression and metastatic status. Rodriguez 2008 et al. also reported that no correlation existed between COX-2 expression and metastasis in osteosarcoma [32]. Conversely, Urakawa et al. [31] reported that the expression of COX-2 was significantly related to the metastatic status of osteosarcoma. Further analysis from 11 studies with 515 osteosarcoma patients revealed the presence of a positive association between COX-2 expression and the metastatic status of osteosarcoma in a larger population. The above-mentioned analyses suggest that the expression of COX-2 may play an important role in disease progression and metastasis in patients with osteosarcoma. No evidence of substantial heterogeneity was available concerning the clinical features. Further TSA revealed

that future additional studies are not essential. Therefore, the results of our analysis are conclusive.

The multivariate analysis showed that COX-2 expression was associated with worse prognosis in metastasis-free survival (5 years) [31], overall survival, and relapse-free survival [30], which indicated that COX-2 expression might become a potential prognostic marker. Additional clinical research of larger sample sizes using multivariate analysis in patients with osteosarcoma is necessary to confirm the prognostic significance of COX-2 expression.

The present meta-analysis has several limitations. First, the patients included in the analyses were mainly Asians and Caucasians, but the shares of other ethnic groups, such as the African population, were insufficient. Second, publication bias was detected in the comparison of osteosarcoma and benign osteochondroma. Papers with positive conclusions were more easily published than articles with

Study		%
ID	OR (95% CI)	Weight
Tumor histology		
Hosono (2007) 🗲 💌	0.05 (0.00, 1.05)	2.21
Ou (2007) —	<b>1.14 (0.28, 4.58)</b>	8.70
Rodriguez (2008)	4.64 (0.79, 27.24)	5.80
Geng (2008) —	• 0.64 (0.20, 2.03)	11.53
Liao (2009) —	0.86 (0.27, 2.73)	11.52
Huang (2009) —	• 1.26 (0.20, 7.99)	5.40
Xu (2010) —	1.00 (0.16, 6.08)	5.61
Ma (2011) ———	• 0.48 (0.09, 2.62)	6.23
Duan (2012) —	0.69 (0.14, 3.52)	6.72
Subtotal ( $I^2 = 0.0\%$ , $P = 0.434$ )	0.87 (0.52, 1.45)	63.70
Necrosis (≥ 90% vs < 90%)		
Dickens (2003)	2.50 (0.47, 13.27)	6.42
Rodriguez (2008) -	• 1.20 (0.31, 4.59)	9.20
Urakawa (2009)	<b>3.56 (0.93, 13.65)</b>	9.19
Chen (2012) —	• 0.47 (0.15, 1.50)	11.49
Subtotal ( $I^2 = 47.5\%$ , $P = 0.126$ )	1.39 (0.54, 3.56)	36.30
Overall		100.00
NOTE: Weights are from random-effects and	alysis	
0.0026	1 385	

Fig. 11. Forest plot of the correlation of COX-2 immunoexpression with cancer histology and necrosis (P > 0.1).

negative conclusions. In addition, publications of other styles, such as conference abstracts, were lacking due to insufficient information. Third, more studies using multivariate analysis of COX-2 expression are needed to confirm the prognostic effect in different ethnic groups. Finally, the cutoff values of COX-2 expression of the eligible studies were different. Thus, in the future, whether COX-2 expression is positive or negative should be defined based on common standards set.

In conclusion, the present findings suggest that COX-2 immunoexpression is significantly higher in osteosarcoma than in benign osteochondroma. Additionally, COX-2 expression is associated with tumor grade, clinical stage, and metastasis of osteosarcoma but is not correlated with age, gender, tumor location, histology, or necrosis. The expression of COX-2 may serve as a prognostic indicator for the multivariate analysis of metastasis-free, overall, and relapse-free

survival. Conducting additional, well-designed, prospective studies that investigate large populations in the future is essential to further validate the prognostic role of COX-2 expression.

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## Author contributions

ZG contributed to the conception and design of this study. SW, HG, and JZ contributed to acquisition, analysis, and interpretation of data, and drafting and revising the article. All authors gave final approval of the version to be published and agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

# **Conflict of interest**

The authors declare no conflict of interest.

# References

- Fox MG and Trotta BM (2013) Osteosarcoma: review of the various types with emphasis on recent advancements in imaging. *Semin Musculoskelet Radiol* 17, 123–136.
- 2 Mirabello L, Troisi RJ and Savage SA (2009) Osteosarcoma incidence and survival rates from 1973 to 2004: data from the Surveillance, Epidemiology, and End Results Program. *Cancer* **115**, 1531–1543.
- 3 Mutsaers AJ, Ng AJ, Baker EK, Russell MR, Chalk AM, Wall M, Liddicoat BJ, Ho PW, Slavin JL, Goradia A *et al.* (2013) Modeling distinct osteosarcoma subtypes in vivo using Cre:lox and lineage-restricted transgenic shRNA. *Bone* **55**, 166–178.
- 4 Egas-Bejar D, Anderson PM, Agarwal R, Corrales-Medina F, Devarajan E, Huh WW, Brown RE and Subbiah V (2014) Theranostic profiling for actionable aberrations in advanced high risk osteosarcoma with aggressive biology reveals high molecular diversity: the human fingerprint hypothesis. *Oncoscience* **1**, 167–179.
- 5 Fang Y, Zhang Z, Wang Q and Zhao J (2015) Expression and clinical significance of cyclooxygenase-2 and microRNA-143 in osteosarcoma. *Exp Ther Med* **9**, 2374–2378.
- 6 Luetke A, Meyers PA, Lewis I and Juergens H (2014) Osteosarcoma treatment – where do we stand? A state of the art review. *Cancer Treat Rev* **40**, 523–532.
- 7 Chou AJ, Geller DS and Gorlick R (2008) Therapy for osteosarcoma: where do we go from here? *Paediatr Drugs* 10, 315–327.
- 8 Bajpai J, Chandrasekharan A, Simha V, Talreja V, Karpe A, Pandey N, Singh A, Rekhi B, Vora T, Ghosh J et al. (2018) Outcomes in treatment-naive patients with metastatic extremity osteosarcoma treated with OGS-12, a novel non-high-dose methotrexate-based, dose-dense combination chemotherapy, in a tertiary care cancer center. J Glob Oncol 4, 1–10.
- 9 Taran SJ, Taran R and Malipatil NB (2017) Pediatric osteosarcoma: an updated review. *Indian J Med Paediatr Oncol* 38, 33–43.
- 10 Li Z, Jia M, Wu X, Cui J, Pan A and Li L (2015) Overexpression of Trps1 contributes to tumor angiogenesis and poor prognosis of human osteosarcoma. *Diagn Pathol* 10, 167.
- Ren L and Khanna C (2014) Role of ezrin in osteosarcoma metastasis. *Adv Exp Med Biol* 804, 181– 201.
- 12 Jones KB, Salah Z, Del Mare S, Galasso M, Gaudio E, Nuovo GJ, Lovat F, LeBlanc K, Palatini J, Randall RL et al. (2012) miRNA signatures associate with

pathogenesis and progression of osteosarcoma. *Cancer Res* **72**, 1865–1877.

- 13 Entz-Werle N, Schneider A, Kalifa C, Voegeli AC, Tabone MD, Marec-Berard P, Marcellin L, Pacquement H, Terrier P, Boutard P *et al.* (2003) Genetic alterations in primary osteosarcoma from 54 children and adolescents by targeted allelotyping. *Br J Cancer* 88, 1925–1931.
- 14 Smith WL, Garavito RM and DeWitt DL (1996) Prostaglandin endoperoxide H synthases (cyclooxygenases)-1 and -2. *J Biol Chem* 271, 33157– 33160.
- 15 Bernard MP, Bancos S, Sime PJ and Phipps RP (2008) Targeting cyclooxygenase-2 in hematological malignancies: rationale and promise. *Curr Pharm Des* 14, 2051–2060.
- 16 Dannenberg AJ and Subbaramaiah K (2003) Targeting cyclooxygenase-2 in human neoplasia: rationale and promise. *Cancer Cell* 4, 431–436.
- 17 Lee EJ, Choi EM, Kim SR, Park JH, Kim H, Ha KS, Kim YM, Kim SS, Choe M, Kim JI *et al.* (2007) Cyclooxygenase-2 promotes cell proliferation, migration and invasion in U2OS human osteosarcoma cells. *Exp Mol Med* **39**, 469–476.
- 18 Wendum D, Masliah J, Trugnan G and Flejou JF (2004) Cyclooxygenase-2 and its role in colorectal cancer development. *Virchows Arch* 445 (4), 327–333.
- 19 Tsujii M, Kawano S, Tsuji S, Sawaoka H, Hori M and DuBois RN (1998) Cyclooxygenase regulates angiogenesis induced by colon cancer cells. *Cell* 93, 705–716.
- 20 Sicking I, Rommens K, Battista MJ, Bohm D, Gebhard S, Lebrecht A, Cotarelo C, Hoffmann G, Hengstler JG and Schmidt M (2014) Prognostic influence of cyclooxygenase-2 protein and mRNA expression in node-negative breast cancer patients. *BMC Cancer* 14, 952.
- 21 Sinicrope FA and Gill S (2004) Role of cyclooxygenase-2 in colorectal cancer. *Cancer Metastasis Rev* 23, 63–75.
- 22 Howe LR, Subbaramaiah K, Brown AM and Dannenberg AJ (2001) Cyclooxygenase-2: a target for the prevention and treatment of breast cancer. *Endocr Relat Cancer* **8**, 97–114.
- 23 Shujiao L, Lilin H and Yong S (2016) Cyclooxygenase-2 expression and association with skin cancer: a meta-analysis based on Chinese patients. *J Cancer Res Ther* 12 (Supplement), C288–C290.
- 24 Xu F, Li M, Zhang C, Cui J, Liu J, Li J and Jiang H (2017) Clinicopathological and prognostic significance of COX-2 immunohistochemical expression in breast cancer: a meta-analysis. *Oncotarget* 8, 6003–6012.
- 25 Kunzmann AT, Murray LJ, Cardwell CR, McShane CM, McMenamin UC and Cantwell MM (2013) PTGS2 (Cyclooxygenase-2) expression and survival

among colorectal cancer patients: a systematic review. Cancer Epidemiol Biomarkers Prev 22 (9), 1490–1497.

- 26 Lee JY, Myung SK and Song YS (2013) Prognostic role of cyclooxygenase-2 in epithelial ovarian cancer: a meta-analysis of observational studies. *Gynecol Oncol* **129**, 613–619.
- 27 Zhan P, Qian Q and Yu LK (2013) Prognostic value of COX-2 expression in patients with non-small cell lung cancer: a systematic review and meta-analysis. *J Thorac Dis* 5, 40–47.
- 28 Qu L and Liu B (2015) Cyclooxygeanse-2 promotes metastasis in osteosarcoma. *Cancer Cell Int* **15**, 69.
- 29 Duan DP, Dang XQ, Wang KZ, Wang YP, Zhang H and You WL (2012) The cyclooxygenase-2 inhibitor NS-398 inhibits proliferation and induces apoptosis in human osteosarcoma cells via downregulation of the survivin pathway. *Oncol Rep* 28, 1693–1700.
- 30 Boulytcheva IV, Soloviev YN, Kushlinskii NE and Mahson AN (2010) Expression of molecular markers in the tumor and survival prognosis in osteosarcoma. *Bull Exp Biol Med* 150, 237–242.
- 31 Urakawa H, Nishida Y, Naruse T, Nakashima H and Ishiguro N (2009) Cyclooxygenase-2 overexpression predicts poor survival in patients with high-grade extremity osteosarcoma: a pilot study. *Clin Orthop Relat Res* 467, 2932–2938.
- 32 Rodriguez NI, Hoots WK, Koshkina NV, Morales-Arias JA, Arndt CA, Inwards CY, Hawkins DS, Munsell MF and Kleinerman ES (2008) COX-2 expression correlates with survival in patients with osteosarcoma lung metastases. *J Pediatr Hematol Oncol* 30, 507–512.
- 33 Masi L, Recenti R, Silvestri S, Pinzani P, Pepi M, Paglierani M, Brandi ML and Franchi A (2007) Expression of cyclooxygenase-2 in osteosarcoma of bone. *Appl Immunohistochem Mol Morphol* 15, 70–76.
- 34 Hosono A, Yamaguchi U, Makimoto A, Endo M, Watanabe A, Shimoda T, Kaya M, Matsumura T, Sonobe H, Kusumi T *et al.* (2007) Utility of immunohistochemical analysis for cyclo-oxygenase 2 in the differential diagnosis of osteoblastoma and osteosarcoma. *J Clin Pathol* **60**, 410–414.
- 35 Hakim DN, Pelly T, Kulendran M and Caris JA (2015) Benign tumours of the bone: a review. *J Bone Oncol* **4**, 37–41.
- 36 Kawashima H, Ogose A, Hotta T, Imai C, Imamura M and Endo N (2015) Secondary osteosarcoma arising from osteochondroma following autologous stem cell transplantation with total-body irradiation for neuroblastoma: a case report. *Oncol Lett* 10, 1026– 1030.
- 37 Liberati A, Altman DG, Tetzlaff J, Mulrow C, Gotzsche PC, Ioannidis JP, Clarke M, Devereaux PJ, Kleijnen J and Moher D (2009) The PRISMA statement for reporting systematic reviews and

meta-analyses of studies that evaluate health care interventions: explanation and elaboration. *PLoS Med* **6**, e1000100.

- 38 Zintzaras E and Ioannidis JP (2005) HEGESMA: genome search meta-analysis and heterogeneity testing. *Bioinformatics* 21, 3672–3673.
- 39 Han S, Zong S, Shi Q, Li H, Liu S, Yang W, Li W and Hou F (2017) Is Ep-CAM expression a diagnostic and prognostic biomarker for colorectal cancer? A systematic meta-analysis. *EBioMedicine* 20, 61–69.
- 40 Zhu C, Feng X, Ye G and Huang T (2017) Metaanalysis of possible role of cadherin gene methylation in evolution and prognosis of hepatocellular carcinoma with a PRISMA guideline. *Medicine (Baltimore)* **96**, e6650.
- 41 Egger M, Davey Smith G, Schneider M and Minder C (1997) Bias in meta-analysis detected by a simple, graphical test. *BMJ (Clinical research ed)* 315, 629– 634.
- 42 Kulinskaya E and Wood J (2014) Trial sequential methods for meta-analysis. *Res Synth Methods* 5, 212– 220.
- 43 Brok J, Thorlund K, Gluud C and Wetterslev J (2008) Trial sequential analysis reveals insufficient information size and potentially false positive results in many metaanalyses. J Clin Epidemiol 61, 763–769.
- 44 Holst LB, Petersen MW, Haase N, Perner A and Wetterslev J (2015) Restrictive versus liberal transfusion strategy for red blood cell transfusion: systematic review of randomised trials with meta-analysis and trial sequential analysis. *BMJ* **350**, h1354.
- 45 Wetterslev J, Thorlund K, Brok J and Gluud C (2009) Estimating required information size by quantifying diversity in random-effects model meta-analyses. *BMC Med Res Methodol* **9**, 86.
- 46 Dickens DS, Kozielski R, Leavey PJ, Timmons C and Cripe TP (2003) Cyclooxygenase-2 expression does not correlate with outcome in osteosarcoma or rhabdomyosarcoma. *J Pediatr Hematol Oncol* 25, 282– 285.
- 47 Jiang Y, Yang S and Liu Y (2004) Expression of cyclooxygenase 2 in osteosarcoma and its significance. *Cent China Med J* 28, 109–110.
- 48 Zhu Y, Chi Y and Wang Y (2005) Study of expression and their relationship of COX-2 and MMP-2 tn osteosarcoma. J Practical Oncol 16, 91–94.
- 49 Ou YZ, Wang SK, Wang CF, Kang XW and Guo HZ (2007) Expression of cyclooxygenase-2 and the relation between cyclooxygenase-2 and CD105 in osteosarcoma. *Cancer Res Clinic* **19**, 602–604.
- 50 Wang XB, Hu WH and Li HJ (2007) Expressions and clinical significances of cyclooxygenase-2, p53 in osteosarcoma. *Orthopedic J China* **15**, 1822–1824.
- 51 Geng YH, Wang CX and Chen PH (2008) Expressions of Cox-2 and HIF-1α and their relationship with

clinicopathologic characteristics of osteosarcoma. *Tumor* **28**, 427–430.

- 52 Zhan XQ (2008) Significance of expression of Cox-2 and Fas in cases of osteosarcoma. J Clin Exper Med 7, 45–46.
- 53 Liu YM, Zhou MH and Zhang DF (2009) Expressions and clinicopathologic significance of MMP-7 and COX-2 in osteosarcoma. *Med J National Defending Forces Northwest China* **30**, 280–282.
- 54 Liao H, Feng XR and Mao XF (2009) Relationships between expressions of COX-2, MMP-9 and tumor microvessel density in patients with osteosarcoma. *Med J National Defending Forces Southwest China* 19, 377–379.
- 55 Huang JJ, Shi Y, Xiang ZG, Tian ZL and Luo JS (2009) The expression and clinical significance of cox-2 in osteosarcoma and giant cell tumor of bone. *J Chin Physician* **11**, 1345–1347.
- 56 Yong-Hao LI and Xiao YZ (2010) Expression of COX-2 and Ki-67 in osteosarcoma and their clinical significance. *Chinese J Ge Practice* 8, 274–276.
- 57 Xu XZ, Huang G, Liu JF, Li LQ and Luo Y (2010) Expression of PTEN, COX-2 and MMP-9 in osteosarcoma and its clinical significance. *J Clin Res* 27, 820–823.
- 58 Jun MA, Chen XM, Zhong YP, Dan-Rong LI and Xiao ZM (2011) Expression and significance of COX-2 and VEGF in osteosarcoma. *Shandong Med J* 51, 7–9.
- 59 Chen Y, Wang CM, Shi YQ and Yang Y (2012) Expression of hypoxia-inducible factor 1alpha in osteosarcoma and its value in predicting chemosensitivity. *Zhonghua Zhong Liu Za Zhi* 34, 899–904.

- 60 Meng XW, Yan HN, Zhu M, Liu X, Gao LY and Qiu W (2013) Expressions of nuclear factor, matrix metalloproteinase-9 and cyclooxygenase-2 in osteosarcoma and their relationship with angiogenesis. *J Bengbu Med Coll* 38, 270–272.
- 61 Lian H, Yuqiang LI and Min Z (2013) Expression of COX-2, MMP-9 in osteosarcoma and their clinical significance. *Hebei J TCM* 35, 1105–1107.
- 62 Zhu Y, Yue YL, Wang YF, Zhao MH, Department TO (2016) The expression and clinical significance of COX-2 and P-gp in osteosarcoma. *Progress Anatom Sci* 22, 583–586.
- 63 Elzagheid A, Emaetig F, Alkikhia L, Buhmeida A, Syrjanen K, El-Faitori O, Latto M, Collan Y and Pyrhonen S (2013) High cyclooxygenase-2 expression is associated with advanced stages in colorectal cancer. *Anticancer Res* 33, 3137–3143.
- 64 Pan J, Tang T, Xu L, Lu JJ, Lin S, Qiu S, Chen G and K Tham IW (2013) Prognostic significance of expression of cyclooxygenase-2, vascular endothelial growth factor, and epidermal growth factor receptor in nasopharyngeal carcinoma. *Head Neck* 35, 1238–1247.
- 65 Jiao G, Ren T, Lu Q, Sun Y, Lou Z, Peng X, Liang W and Guo W (2013) Prognostic significance of cyclooxygenase-2 in osteosarcoma: a meta-analysis. *Tumour Biol* 34, 2489–2495.
- 66 Wang Z, He M, Xiao Z, Wu H and Wu Y (2013) Quantitative assessment of the association of COX-2 (Cyclooxygenase-2) immunoexpression with prognosis in human osteosarcoma: a meta-analysis. *PLoS ONE* **8**, e82907.