

## Research Article

# Compound Mutations of the COL4A3 including a Novel Allele Identified in a Patient with Alport Syndrome

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Alport syndrome (AS) is a hereditary nephropathy which is characterized by molecular abnormalities in collagen IV. Here, we report compound mutations of the COL4A3 gene including a novel allele identified in a patient with Alport syndrome. The patient was a 25-year-old Chinese woman. She has a history of proteinuria and hematuria with cleft lip and palate. The pathologic results were consistent with Alport syndrome. The patient received ACEI treatment but did not respond well to the treatment. Sequencing results revealed that the patient carried two heterozygous mutations in the COL4A3 gene, including a known mutation (c.4243G>C, p.G1415R), which was inherited from her father, and a previously undescribed allele (c.4216G>A, p.G1406R) inherited from her mother. To date, at least 294 different variants of COL4A3 have been reported according to the Human Gene Mutation Database (HGMD). Identification of c.4216G>A as a new AS-related mutation may contribute to both genetic diagnosis of AS and further functional study of COL4A3.

## 1. Introduction

Alport syndrome (AS) is a familial hereditary nephropathy caused by mutations in the type IV collagen genes COL4A3, COL4A4, and COL4A5 [1]. These genes encode  $\alpha 3$ ,  $\alpha 4$ , and  $\alpha 5$  collagen chains, which are the major structural components of the glomerular basement membrane as well as the basement membranes in the eye and cochlea. There are three types of heredity of Alport syndrome. The X-linked dominant inheritance of mutations in the COL4A5 gene accounts for 80%~85% of AS. The autosomal recessive inheritance of COL4A3 or COL4A4 mutations accounts for 10%~15% of AS. Dominant mutations in COL4A3 and COL4A4 have only been reported in about 5% cases [2–5]. High-throughput next-generation sequencing (NGS) technology is an efficient and appropriate form of genetic test for Alport syndrome, and more gene mutations are being discovered [6, 7]. Although the relationship between genotypes and phenotypes

has not been fully understood in Alport syndrome, more discovery of genetic mutations would contribute to genetic diagnosis and counseling of Alport syndrome.

## 2. Materials and Methods

**2.1. Case Report.** The proband is a 25-year-old woman with proteinuria and hematuria. She has had otitis media for more than 20 years and had a cleft lip and palate operation when she was 5 years old. She went to the hospital for hematuria over the past year. No one else in her family suffers from proteinuria, hematuria, or renal hypofunction or deafness. Urine analysis showed 3+ proteinuria and 3+ hematuria (77 red blood cells per high-power field) and a urinary protein/creatinine ratio (P/Cr) of 4.56 g/g Cr. Albumin level was 32.5 g/L, and antinuclear antibodies were negative. She had renal biopsies. Recently, she took perindopril medicine by mouth.

Informed written consent was obtained from all participants. The study was approved by the Research Ethics Committee of the Qilu Hospital of Shandong University.

## 2.2. Methods

**2.2.1. Whole-Exome Sequencing.** Peripheral blood was collected from EDTA anticoagulant vessels, and blood DNA was extracted with the extraction kit. DNA libraries were constructed with KAPA Library Preparation Kit. The vacuum-enriched DNA library samples were hybridized and captured in SureSelect hybridized buffer liquid series. Illumina NovaSeq high-throughput sequencing was performed on the captured DNA samples. The sequenced data were evaluated by Illumina Sequence Control Software (SCS), and bioinformatics analysis was performed. The data interpretation rules are based on guidelines from the American College of Medical Genetics and Genomics (ACMG) [8]. Variables named according to the rules of HGVS (<http://www.hgvs.org/mutnomen>) are given.

**2.2.2. Bioinformatics Analysis.** To predict the pathogenesis of Col4A3 mutation, we use Mutation Taster software (<http://www.mutationtaster.org/>). This program can be used to predict whether the mutation is pathogenic or harmless. Similarly, Col4A3 amino acid sequences were analyzed using PolyPhen-2 (<http://genetics.bwh.harvard.edu/pph2/>) and SIFT software (<http://sift.jcvi.org/>). Based on straightforward physical and comparative considerations, the possible effects of amino acid substitution on the structure and function of human proteins were predicted by PolyPhen-2. Similarly, whether the protein function is affected can be predicted by SIFT. We checked the conservation status of this exonic variant, namely, Clustal X software in 6 species including a mouse, pig, sheep, rabbit, human, and pan.

**2.2.3. Sanger Sequencing.** All the filtered mutations of this family were validated by Sanger sequencing. Specific primers were used to amplify the regional DNA fragments at the mutant sites. The sequences of the polymerase chain reaction (PCR) products were determined using the ABI 3730XL DNA Genetic Analyzer.

## 3. Results

**3.1. Renal Biopsies.** The renal biopsies of patient are shown in Figures 1(a) and 1(b). We did H&E and PAS staining on the patient's renal biopsies. Electron microscopy (EM) was also performed. One out of the six examined glomeruli showed mild mesangial hyperplasia, thickened glomerular capillary walls, and swelling podocytes. EM demonstrated markedly irregular subendothelial GBM surface with splitting and "basket weaving" (Figure 1(c)).

**3.2. Whole-Exome Sequencing.** We detected two mutated sites in the COL4A3 gene (Table 1). The first one is a missense mutation c.4243G>C in the COL4A3 gene, resulting in amino acid change p.G1415R (glycine > arginine), which has been reported as a pathogenic mutation [9]. The other

one is an undescribed missense mutation c.4216G>A, leading to replacement of the amino acid G1406 to R.

**3.3. Bioinformatics Analysis.** Mutation Taster software analysis revealed that COL4A3 c.4216G>A (p.G1406R) mutation could be a disease-causing mutation with a score of 0.810. Also, SIFT software analysis revealed that COL4A3 c.4216G>A (p.G1406R) mutation could be a damaging mutation with a score of 0.912. Analysis with the PolyPhen-2 software mutation supports the probably damaging role of this amino acid change with a high score of 0.970. Clustal X software showed that the novel exon variant COL4A3 c. 4216G>A (p.G1406R) was conservative in primates (Figure 2), suggesting the possible pathogenicity of this exon variant in human beings.

**3.4. Sanger Sequencing.** The father and two sisters carry COL4A3 c.4243G>C mutation, and the mother is a carrier of COL4A3 c.4216G>A mutation. Since the parents and sisters do not have any symptoms, we conclude that the genetic mode is autosomal recessive inheritance (Figure 3).

## 4. Discussion

Although the patient does not have a family history of hematuria and chronic renal failure and no eye lesions in this study, she had hematuria and albuminuria, which gradually progressed to renal dysfunction. In EM, the basement membrane was segmentally thickened, shrunken, and broken, which is the clinical diagnostic criteria of Alport syndrome, according to Fliter [10] and Gregory [11]. Through clinical manifestation, family history, and renal pathological examination, Alport syndrome was highly considered.

In order to further clarify the diagnosis, we sequenced the patients with high-throughput next-generation sequencing. The results showed that there were two heterozygous mutations in the COL4A3 gene, c.4243G>C (p. G1415R) and c. 4216G>A (p.G1406R), which may cause the disease. The COL4A3 c.4216G>A (p.G1406R) mutation has not been reported and linked to Alport syndrome. Another mutation c.4217G>A (p.G1406E), which leads to amino acid change in the same glycine as c.4216G>A, was identified from two Japanese families [12], indicating this glycine may be critical for the protein function. The COL4A3 gene encodes a chain of 1670 amino acids. Missense mutations account for about 45% of COL4A3 mutations, among which about 85% are glycine substitutions in the conserved Gly-X-Y repeat sequence in the collagen domain of  $\alpha 3$  (IV collagen) chain [4]. Glycine is the only amino acid without side chain and small enough to fit into the center of the triple helix structure. Substitutions of glycine in COL4A3 p.G3499A [13], p.G1406E [12], and p.G4235T [14] have been reported as pathogenic mutations. In this study, the novel missense mutation c.4216G>A causes substitution p.G1406R. Because arginine is larger and more polar than glycine, this substitution may reduce the stability of the complex and impair correct alignment of the type IV collagen triple helix [13, 15–17], leading to the decline of basement membrane filtration barrier function, which in turn causes Alport syndrome [13, 15]. The clinical symptoms

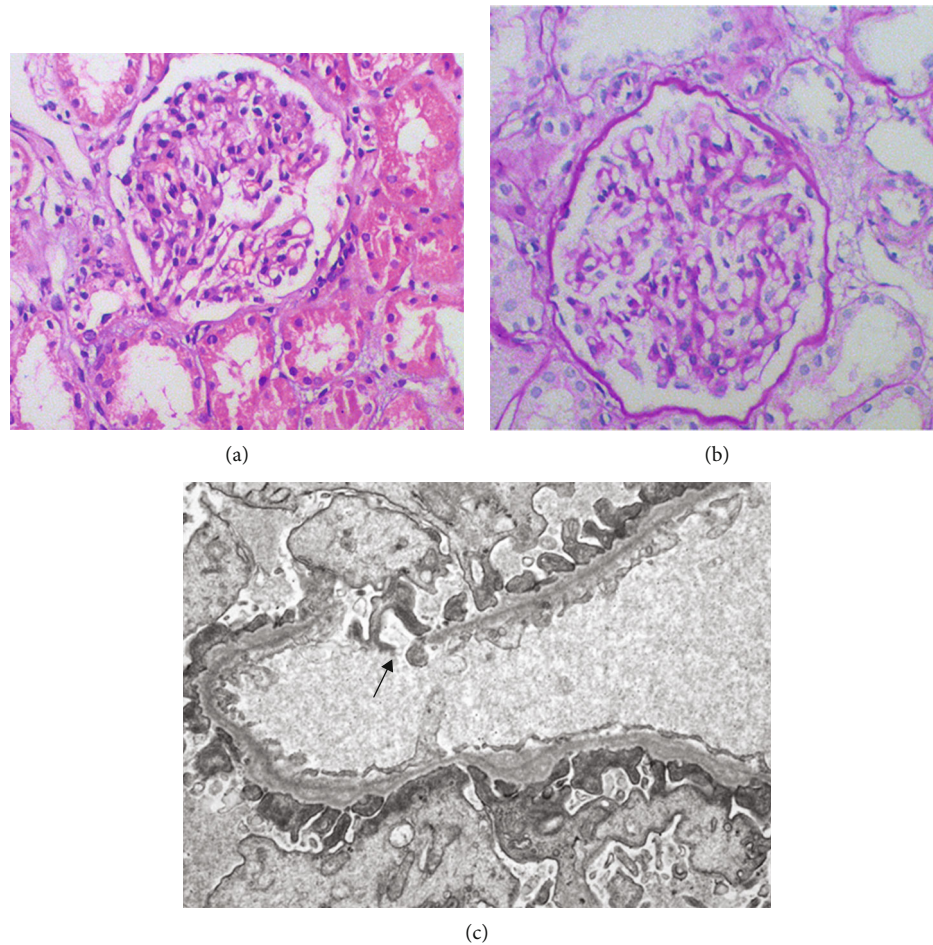


FIGURE 1: Light microscopy (a, b) and electron microscopy (c) of the patient’s renal biopsy. One glomerulus showed mild mesangial hyperplasia, thickened glomerular capillary walls, and swelled podocytes. Electron microscopy demonstrated markedly irregular subendothelial GBM surface with splitting and “basket weaving” (black arrow).

TABLE 1: Detected mutations.

Gene	Nucleotide changes	Amino acid change	Heterozygosity	SIFT	Polyphen-2	Mutation taste
Col4A3	c.4243G>C	p.G1415R	Het	Damaging	Probably damaging	Damaging
Col4A3	c.4216G>A	p.G1406R	Het	Damaging	Probably damaging	Damaging

Mouse	LPGIPGPCGPRGKPGKDGKPGTPGPA	TKGNKGLKGGQPPGLDGLPGLKGNPDRGTPA	1437
Pig	PQGVPESCGPRGKPLDGIPTGPI	EKGNKCKGEQPPGYDGLPGLKGRPGETGPPA	1437
Sheep	PPGDGPGCGPRGKPGEDGPPGTPGPT	EKGNKSKGEQPPGSDGLPGLKGRPGDTGPPA	1438
Rabbit	PPGSPGCGPKGKPGKDGKPGVPGPA	EKGNKSKGEQPPGSEGLPGMKGKPGDTGPPA	1262
Humam	PPGNLPGCGPRGKPGKDGKPGTPGPA	EKGNKSKGEPGAGSDGLPGLKGRGDSGSPA	1439
Pan	PPGNLPGCGPRGKPGKDGKPGTPGPA	EKGNKSKGEPGAGSDGLPGLKGRGDSGSPA	1410

FIGURE 2: Glycine 1406 is conserved in collagen alpha-3 (IV) chain across different mammalian species.

are consistent with this prediction. In this patient, the basement membrane of the glomerular capillary wall was slightly thickened, shrunken, and broken. The segmental changes were reticulated in EM.

It is reported that 90% of men and 12% of women with XLAS develop end-stage kidney failure at the age of 40 [18]. ARAS is more serious and progresses faster than XLAS, and the median age of patients with renal failure is 21 years old

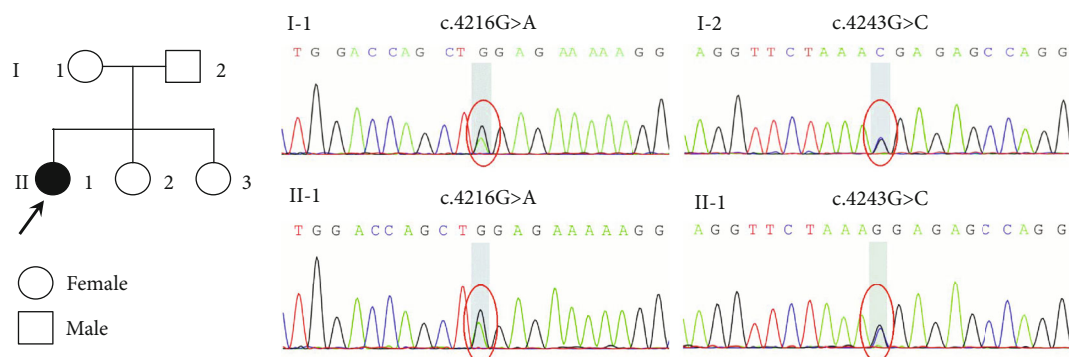


FIGURE 3: Pedigree and Sanger sequencing results of the patient's family.

[19]. This patient has mild symptoms (slight glomerular lesions and no renal failure) which may be related to the younger age of the patient or the shorter time of onset or that the mutation of the gene locus itself is less pathogenic and has little effect on the renal function of the patient.

Alport syndrome is often accompanied by hematuria, sensorineural deafness, and ocular abnormalities [20]. No abnormality in the eye, blood system, and heart ultrasound was observed in this case. The patient had congenital cleft lip. So far, there are no report suggesting mutation in COL4A3 may cause cleft lip and palate. The relationship between cleft lip and palate and COL4A3 gene needs further study.

## 5. Conclusion

In conclusion, via whole-exome sequencing in the combination of bioinformatics analysis strategy, we have identified a novel COL4A3 mutation (c. 4216G>A p.G1406R) in a suspected AS patient from China. Our study may expand the spectrum of COL4A3 mutations and contribute to genetic diagnosis and counseling of Alport syndrome.

## Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

## Conflicts of Interest

The authors declare that they have no conflicts of interest.

## Authors' Contributions

Zhendong Wang and Baichun Jiang contributed equally to this work.

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## References

- [1] F. A. Flinter, C. Chantler, J. S. Cameron, I. Houston, and M. Bobrow, "Genetics of classic Alport's syndrome," *The Lancet*, vol. 332, no. 8618, pp. 1005–1007, 1988.
- [2] B. G. Hudson, "The molecular basis of Goodpasture and Alport syndromes: beacons for the discovery of the collagen IV family," *Journal of the American Society of Nephrology*, vol. 15, no. 10, pp. 2514–2527, 2004.
- [3] C. Fallerini, L. Dosa, R. Tita et al., "Unbiased next generation sequencing analysis confirms the existence of autosomal dominant Alport syndrome in a relevant fraction of cases," *Clinical Genetics*, vol. 86, no. 3, pp. 252–257, 2014.
- [4] J. M. Hertz, M. Thomassen, H. Storey, and F. Flinter, "Clinical utility gene card for: Alport syndrome - update 2014," *European Journal of Human Genetics*, vol. 23, no. 9, article 1269, 2015.
- [5] V. Morinière, K. Dahan, P. Hilbert et al., "Improving Mutation Screening in Familial Hematuric Nephropathies through Next Generation Sequencing," *Journal of the American Society of Nephrology*, vol. 25, no. 12, pp. 2740–2751, 2014.
- [6] F. Lin, F. Bian, J. Zou et al., "Whole exome sequencing reveals novel COL4A3 and COL4A4 mutations and resolves diagnosis in Chinese families with kidney disease," *BMC Nephrology*, vol. 15, no. 1, p. 175, 2014.
- [7] L. Papazachariou, P. Demosthenous, M. Pieri et al., "Frequency of COL4A3/COL4A4 mutations amongst families segregating glomerular microscopic hematuria and evidence for activation of the unfolded protein response. Focal and segmental glomerulosclerosis is a frequent development during ageing," *PLoS One*, vol. 9, no. 12, article e115015, 2014.
- [8] S. Richards, on behalf of the ACMG Laboratory Quality Assurance Committee, N. Aziz et al., "Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology," *Genetics in Medicine*, vol. 17, no. 5, pp. 405–423, 2015.
- [9] Y. Zhang, F. Wang, J. Ding et al., "Genotype-phenotype correlations in 17 Chinese patients with autosomal recessive Alport syndrome," *American Journal of Medical Genetics Part A*, vol. 158A, no. 9, pp. 2188–2193, 2012.
- [10] F. Flinter, "Alport's syndrome," *Journal of Medical Genetics*, vol. 34, no. 4, pp. 326–330, 1997.
- [11] M. C. Gregory, D. A. Terreros, D. F. Barker, P. N. Fain, J. C. Denison, and C. L. Atkin, "Alport Syndrome—Clinical

- phenotypes, incidence, and Pathology,” *Contributions to Nephrology*, vol. 117, pp. 1–28, 1996.
- [12] N. Kamiyoshi, K. Nozu, X. J. Fu et al., “Genetic, clinical, and pathologic Backgrounds of patients with autosomal dominant Alport syndrome,” *Clinical Journal of the American Society of Nephrology*, vol. 11, no. 8, pp. 1441–1449, 2016.
- [13] L. Heidet, C. Arrondel, L. Forestier et al., “Structure of the human type IV collagen gene *COL4A3* and mutations in autosomal Alport syndrome,” *Journal of the American Society of Nephrology*, vol. 12, no. 1, pp. 97–106, 2001.
- [14] B. T. Vega, C. Badenas, E. Ars et al., “Autosomal recessive Alport's syndrome and benign familial hematuria are collagen type IV diseases,” *American Journal of Kidney Diseases*, vol. 42, no. 5, pp. 952–959, 2003.
- [15] K. Tryggvason, J. Zhou, S. L. Hostikka, and T. B. Shows, “Molecular genetics of Alport syndrome,” *Kidney International*, vol. 43, no. 1, pp. 38–44, 1993.
- [16] C. E. Kashtan, “Alport syndrome: abnormalities of type IV collagen genes and proteins,” *Renal Failure*, vol. 22, no. 6, pp. 737–749, 2000.
- [17] S. Gunwar, F. Ballester, M. E. Noelken, Y. Sado, Y. Ninomiya, and B. G. Hudson, “Glomerular basement membrane. Identification of a novel disulfide-cross-linked network of alpha3, alpha4, and alpha5 chains of type IV collagen and its implications for the pathogenesis of Alport syndrome,” *The Journal of Biological Chemistry*, vol. 273, no. 15, pp. 8767–8775, 1998.
- [18] J. P. Jais, B. Knebelmann, I. Giatras et al., “X-linked Alport syndrome: natural history and genotype-phenotype correlations in girls and women belonging to 195 families: a “European Community Alport Syndrome Concerted Action” study,” *Journal of the American Society of Nephrology*, vol. 14, no. 10, pp. 2603–2610, 2003.
- [19] M. Oka, K. Nozu, H. Kaito et al., “Natural history of genetically proven autosomal recessive Alport syndrome,” *Pediatric Nephrology*, vol. 29, no. 9, pp. 1535–1544, 2014.
- [20] P. Karki and J. K. Shrestha, “Alport syndrome,” *Nepalese Journal of Ophthalmology*, vol. 1, no. 2, pp. 139–140, 2009.