W J H World Journal of Henatology

Submit a Manuscript: https://www.f6publishing.com

World J Hepatol 2022 August 27; 14(8): 1562-1575

DOI: 10.4254/wjh.v14.i8.1562

**Basic Study** 

ISSN 1948-5182 (online)

ORIGINAL ARTICLE

# Assessment of circulating levels of microRNA-326, microRNA-424, and microRNA-511 as biomarkers for hepatocellular carcinoma in Egyptians

Samar Samir Youssef, Asmaa Elfiky, Mohamed M Nabeel, Hend Ibrahim Shousha, Tamer Elbaz, Dalia Omran, Mohammad Saeed Marie, Mohammad A Elzahry, Amr Abul-Fotouh, Ahmed Hashem, Mohamed F Guda, Ashraf O Abdelaziz

Specialty type: Tropical medicine

# Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review model: Single blind

#### Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): 0 Grade C (Good): C Grade D (Fair): D Grade E (Poor): 0

**P-Reviewer:** Tsoulfas G, Greece; Yao HR, China

Received: October 22, 2021 Peer-review started: October 22, 2021

First decision: December 27, 2021 Revised: January 14, 2022 Accepted: July 27, 2022 Article in press: July 27, 2022 Published online: August 27, 2022



Samar Samir Youssef, Department of Microbial Biotechnology, National Research Centre, Cairo 1211, Egypt

Asmaa Elfiky, Department of Environmental and Occupational Medicine, National Research Centre, Cairo 1211, Egypt

Mohamed M Nabeel, Hend Ibrahim Shousha, Tamer Elbaz, Mohammad Saeed Marie, Ashraf O Abdelaziz, Department of Endemic Medicine and Hepatogastroenterology, Faculty of Medicine, Cairo University, Cairo 11562 Egypt

Dalia Omran, Mohammad A Elzahry, Amr Abul-Fotouh, Ahmed Hashem, Department of Endemic Medicine, Faculty of Medicine, Cairo University, Cairo 1256, Egypt

Mohamed F Guda, Theodor Bilharz Research Institute, Cairo 1256, Egypt

**Corresponding author:** Samar Samir Youssef, PhD, Professor, Department of Microbial Biotechnology, National Research Centre, El Behoos st, Cairo 1211, Egypt. samaryoussef67@gmail.com

# Abstract

# BACKGROUND

Hepatocellular carcinoma (HCC) is the fifth most common cancer. Differential expression of microRNAs (miRNAs)-326, miRNA-424, and miRNA-511 has been associated with the diagnosis and prognosis of HCC in different populations. However, limited information is available regarding their expression in Egyptian HCC patients.

#### AIM

To assess the role of circulating miRNAs-326, miRNA-424, and miRNA-511 in Egyptian HCC patients.

# METHODS

This prospective observational study included 70 HCC patients and 25 healthy controls. The circulating levels of these three miRNAs were evaluated by real-time



PCR. Receiver operating characteristic curve analysis was used to test the diagnostic accuracy of microRNA expression levels.

#### RESULTS

All miRNAs were differentially expressed in HCC patients; miRNAs326 and miRNA-424 were upregulated, while miRNA-511 was downregulated. Both miRNA-326 and miRNA-424 showed sensitivity and specificity of 97%, 71.4%, and 52%, 60%, respectively, to differentiate HCC from controls. Moreover, miRNA-326 was associated with survival and could differentiate between Child grades (A vs B); miRNA-424 significantly differentiated early vs intermediate stages of HCC; while miRNA-511 was significantly correlated with response to modified Response Evaluation Criteria in Solid Tumors (mRECIST).

#### **CONCLUSION**

We conclude that miRNA-326, miRNA-424, and miRNA-511 have diagnostic and prognostic roles in Egyptian patients with hepatitis C virus-related HCC and should be considered for better disease management.

Key Words: Hepatocellular carcinoma; miRNAs-326; miRNA-424; miRNA-511; Modified response evaluation criteria in solid tumors

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Hepatocellular carcinoma (HCC) is the sixth most common malignancy worldwide. Recent cancer incidence data confirmed that the global age normalization rate of primary liver cancer is 10.1/100000, with a male/female ratio of 3:1. The diagnosis of HCC usually occurs in the late stages, resulting in an elevated death rate; this makes HCC the third deadliest malignancy. We examined whether circulating miRNAs326, miRNA-424, and miRNA-511 could serve as promising candidate non-invasive biomarkers for HCC. Such biomarkers could be used for targeted gene therapy research and may assist in monitoring the severity of HCC.

Citation: Youssef SS, Elfiky A, Nabeel MM, Shousha HI, Elbaz T, Omran D, Marie MS, Elzahry MA, Abul-Fotouh A, Hashem A, Guda MF, Abdelaziz AO. Assessment of circulating levels of microRNA-326, microRNA-424, and microRNA-511 as biomarkers for hepatocellular carcinoma in Egyptians. World J Hepatol 2022; 14(8): 1562-1575

URL: https://www.wjgnet.com/1948-5182/full/v14/i8/1562.htm DOI: https://dx.doi.org/10.4254/wjh.v14.i8.1562

#### INTRODUCTION

Liver cancer is the fifth most common cancer in men and is the second cause of cancer-related deaths worldwide[1]. Hepatocellular carcinoma (HCC) is a primary liver cancer and constitutes 80%-90% of all primary tumors of the liver [2,3]. It has a high incidence and mortality rate, with approximately 662.000 deaths every year<sup>[4]</sup>. Most HCC patients are diagnosed at advanced stages, contributing to the high rates of recurrence and development of metastasis[5]. In addition, the commonly used biomarker for HCC screening and diagnosis is alpha-fetoprotein (AFP), which has modest sensitivity and specificity[6] and is actually influenced by tumor size and cancer stage[7]. Due to these factors, it is crucial to identify more efficient biomarkers that can be used for the early diagnosis and prognosis of HCC.

MicroRNAs (miRNAs) are short, non-coding RNAs that regulate the transcription or degradation of certain target mRNAs[8-10]. They mediate important physiological processes such as cell differentiation, proliferation, and survival[11,12]. Aberrant regulation of miRNAs is associated with the initiation and progression of numerous cancers, including HCC[13]. Moreover, survival and response to chemotherapeutic drugs have been found to be linked to miRNAs[14-16]. Experimentally, a number of miRNAs have been proven to be related to HCC, and have been proposed as diagnostic and prognostic markers of HCC[17,18].

Aberrant expression of miRNA-424 has been documented in HCC tissues and cell lines[19,20]. It plays a tumor suppressor role[19]. A similar correlation was found for miRNA-326[21] and miRNA-511 [22], and both were significantly associated with survival[23]. Using the Cancer Genome Atlas, Lu and colleagues studied the prognostic and diagnostic roles of 33 miRNA signatures and found 11 downregulated and 22 upregulated miRNAs by comparing cancerous and non-cancerous samples. They

observed the good role of miRNA-326, -424, and -511[24].

Most miRNA studies in HCC are experimental and are based on HCC tissues and cell lines. They were mainly performed to identify the etiopathogenetic role of miRNAs in HCC. However, few studies have shown their fundamental role in the diagnosis and prognosis of HCC. Hence, this cross-sectional study with prospective follow-up of HCC patients was performed to search for three miRNAs (326, 424, and 511) in blood to highlight their potential role in the diagnosis and prognosis of HCC.

## MATERIALS AND METHODS

#### Patients and study design

This prospective observational study included 70 adult Egyptian patients who developed HCC on top of hepatitis C virus (HCV)-related liver cirrhosis and 25 healthy age- and sex-matched participants who were seronegative for HCV and HBV, and served as controls. HCC was diagnosed according to the American Association for the Study of Liver Diseases updated practice guidelines<sup>[25]</sup>, or was histopathologically confirmed. All the patients were recruited from the multidisciplinary HCC clinic, the Endemic Medicine Department, Kasr al Ainy School of Medicine, Cairo University, and were HCCtreatment naïve at the time of enrollment. Exclusion criteria included; HBV co-infection, any cause of chronic liver disease other than HCV, any associated malignancies other than HCC, and prior treatment for HCV or HCC.

#### Data collection

All patients were subjected to clinical assessment, laboratory investigations (complete blood count, prothrombin time and concentration, liver and kidney function tests, AFP, hepatitis markers), ultrasound examination, and triphasic CT with documentation of HCC site, size, and number as well as the presence of portal vein thrombosis (PVT). At the time of enrollment into the study, blood samples were obtained from each participant, and Child-Pugh-score[26] and Barcelona Clinic Liver Cancer (BCLC) stage[27] were assessed based on the clinical, laboratory, and imaging data obtained.

#### Collection of blood samples

Whole blood samples were collected from each participant in 5-mL sterile RNAase-free vacutainer tubes containing EDTA. Blood samples were collected on ice and processed within 30 min of collection. To separate the plasma, each blood sample was centrifuged for 10 min at 1900 g at 4 °C (Heraeus, Labofuge 400 R; Germany). Plasma samples were carefully transferred into sterile RNase-free tubes, and then centrifuged for 10 min at 12000 g at 4 °C to remove cellular nucleic acid contamination, and haemolysed plasma samples were excluded. Samples were separated into aliquots and stored at 80 °C until processed.

#### RNA extraction

Total RNA was extracted from 200 µL plasma using the miRNeasy Serum/plasma cell lysates kit (Qiagen, Germany), according to the manufacturer's instructions. RNA concentration and purity were monitored using a Nanodrop spectrophotometer device (ThermoScientific, Wilmington, DE, United States).

#### Quantitative RT-PCR

The reverse transcription reaction was performed using the TaqMan™ MicroRNA Reverse Transcription Kit, Cat number # 4366596, (Applied Biosystems, Foster City, CA, United States) according to the manufacturer's instructions. miRNA-326, miRNA-511, and miRNA-424 quantification was carried out using quantitative real-time PCR (qRT-PCR) (Stratagene Mx3000p; Agilent Technologies, Germany). The qRT-PCR for each sample was carried out in duplicate using TaqMan 2x universal master mix II (Applied Biosystems, Foster City, CA, United States) and TaqMan microRNA Assay Mix containing PCR primers and TaqMan probes for each miRNA. The expression level of RNU6B was used as an endogenous control for normalization. To determine miRNA relative expression, it was reported as a fold change (  $\triangle$  Ct and  $\triangle$   $\triangle$  Ct calculations).

#### Patient management and follow-up

After blood sampling, all patients were managed according to the BCLC guidelines<sup>[27]</sup> after a case-bycase discussion. All HCC patients were followed up for a period of 24 mo. HCC response to treatment was evaluated according to the Modified Response Evaluation Criteria in Solid Tumors (mRECIST)[28]. The overall survival time of the patients was defined as the period from the initial presentation to the last follow-up or death.

#### Statistical analysis

Microsoft Excel 2016 and the Statistical Package for Social Science (SPSS; Windows, version 26, IBM



Corp., Armonk, N.Y., United States) were used to analyze the data. Continuous, normally distributed variables were presented as mean ± SD, with a 95% confidence interval (CI), while non-normally distributed variables were summarized as the median and interquartile range (IQR). Categorical data were presented as frequencies and percentages. A *P* value of less than 0.05 was considered statistically significant. The Student *t*-test was performed to compare the means of normally distributed variables between groups, and the Mann-Whitney U test was used for non-normal variables. The Chi-square test and Fisher's exact test were used to determine the distribution of categorical variables between groups. The diagnostic performance of the studied miRNAs was assessed by receiver operating characteristic (ROC) curves. The area under the ROC (AUROC) was used as an index to compare the accuracy of tests. The optimal cut-off point value was taken from the maximum combined sensitivity and specificity. The sensitivity and specificity of relevant cut-offs are also displayed. The survival analysis was conducted using the "Log Rank (Mantel-Cox) Kaplan-Meier test".

# RESULTS

#### Demographic results

The mean age of the studied cohort was  $62.0 \pm 7.6$  years. Most of the patients were male (68.8%), nondiabetic (74.3%), non-smokers (80%), and Child A (82.9%). According to the BCLC staging system, most patients were in the early stage (48.6%). Most of the HCC lesions were single (70%), present in the right hepatic lobe (84.3%), and not associated with PVT (92.9%). That is why most of the patients were subjected to hepatectomy and microwave ablation (MWA), and most of them showed complete response according to mRECIST. All HCC patients were followed up for a period of 24 mo or until death. The mean survival time was 367.1±173.9 days. The median values of miRNA-326, miRNA-511, and miRNA-424 were 35, 1.2, and 5.1, respectively (Table 1).

#### Correlation of miRNAs with different parameters in HCC patients

There was no significant association between the studied miRNAs levels and gender, smoking, or the patients' performance status. The serum level of miRNA-424 was significantly elevated in diabetic patients. There was a significant difference in miRNA-326 between Child grades A and B. Moreover, there was a significant difference in miRNA-424 between early and intermediate HCC (Table 2).

#### Diagnostic efficiency of miRNAs in our patients

On comparing the miRNAs levels between healthy participants and HCC patients, it was found that HCC patients showed significantly higher levels of miRNA-326 (P = 0.001) and significantly lower levels of miRNA-511 (*P* = 0.02) (Table 3).

ROC analysis revealed the diagnostic performance of the studied miRNAs. miRNA-326 showed the best diagnostic performance, diagnosing HCC at a cut-off value of 1.165 with a sensitivity of 97.1%, specificity of 52%, positive predictive value (PPV) of 85%, and negative predictive value (NPV) of 86.7%. The area under curve (AUC) was 78.4% (67.7- 89.1), and the overall accuracy was 85.3% (P < 0.001) (Table 4, Figure 1).

#### Association of miRNAs with response to treatment and survival

Despite finding no statistically significant differences in the studied miRNAs between the surviving and non-surviving HCC patients, we found that miRNA-326 >1.165 was significantly associated with overall survival (P = 0.001) (Table 5, Figure 2). Moreover, there was a significant association between the BCLC stage as well as the response to treatment according to mRECIST and overall survival (Figure 2). It was also found that miRNA-511 was significantly associated with the response to treatment according to mRECIST (Table 6).

# DISCUSSION

Several experimental studies have shown that miRNAs play a regulatory role in the progression of HCC by controlling cell cycle progression, cell growth, and apoptosis. Our study focused on three important miRNAs: miRNA-326, miRNA-424, and miRNA-511. miRNA-326 upregulation is known to inhibit cell proliferation and colony formation, and influence the invasiveness and migratory properties of HCC. It has a tumor suppressor role, and its low expression was found to be associated with tumor, node, metastasis (TNM) staging, tumor differentiation, and lymph node metastases in HCC patients. Regarding miRNA-424, experimental studies found that it is downregulated in HCC and has a role in inhibiting tumor migration, proliferation, and invasion. It was associated with AFP, TNM, multinodularity, vascular invasion, and intrahepatic metastases, as well as poor survival [29,30] and predicted tumor recurrence in HCC patients following liver transplantation[31]. Few studies have mentioned the role of miRNA-511 in HCC proliferation and invasiveness.



# Table 1 Descriptive data of the studied hepatocellular carcinoma patients

			НСС
			N = 70
Demographic data	Age (yr)		$62.0 \pm 7.6$
	Gender (%)	Female	22 (31.4)
		Male	48 (68.6)
	Smoker (%)	No	56 (80.0)
		Yes	14 (20.0)
	DM	No	52 (74.3)
	(%)	Yes	18 (25.7)
	BMI (kg/m <sup>2</sup> )		27.7 (23.6- 31.9)
Laboratory investigation	Hb (g/dL)		$13.2 \pm 1.9$
	WBC (× $10^3/\mu$ L)		6.0 (4.6-8.0)
	Platelets (× $10^3/\mu$ L)		$169.5 \pm 77.9$
	ALT (U/mL)		52.5 (32.0-74.3)
	AST (U/mL)		60.5 (38.0- 83.3)
	Alb (g/dL)		$3.7 \pm 0.5$
	Bil.T (mg/dL)		0.9 (0.6- 1.4)
	AFP (ng/dL)		38.8 (10.0- 370.0)
	Creatinine (mg/dL)		$0.9 \pm 0.2$
BCLC (%)	Early		34 (48.6)
	Intermediate	23 (32.9)	
	Late		13 (18.6)
Performance status (%)	0		43 (61.4)
	1		24 (34.3)
	2		2 (2.9)
	3		1 (1.4)
Child grade (%)	А		58 (82.9)
	В		11 (15.7)
	С		1 (1.4)
Child score			$5.8 \pm 1.0$
Number of HCCs (%)	Single		49 (70.0)
	Two	12 (17.1)	
	≥3		9 (12.9)
Site of HCC (%)	Right lobe		59 (84.3)
	Left lobe		7 (10.0)
	Both lobes		4 (5.7)
HCC size (cm)			4.0 (2.7- 5.0)
Portal vein	Patent		65 (92.9)
(%)			
	Thrombosed		5 (7.1)
Abdominal lymphadenopathy	No		65 (92.9)
(%)	Yes		5 (7.1)



Decision of treatment	Best supportive care	7 (10.0)
	Combined therapy	1 (1.4)
(%)	Hepatectomy	3 (4.3)
	Microwave ablation	20 (28.5)
	Radioembolization	3 (4.3)
	Sorafinib	3 (4.3)
	TACE	29 (41.4)
Response to treatment according to mRECIST (%)	Stationary	4 (7.8)
	Partial response	7 (13.7)
	Complete response	34 (66.7)
	Progressive disease	6 (11.8)
Clinical decompensation (%)	Yes	9 (12.9)
	No	61 (54.5)
Alive or dead (%)	Dead	34 (48.6)
	Alive	36 (51.4)
Survival (days)		367.1 ± 173.9
miRNA-326		35.0 (12.5- 162.3)
miRNA-511		1.2 (0.3- 3.2)
miRNA-424		5.1 (2.9- 10.3)

DM: Diabetes mellitus; Hb: Hemoglobin; WBCs: White blood cells; ALT: Alanine aminotransferase; AST: Aspartate aminotransferase; INR: International nationalized ratio; AFP: a-fetoprotein; HCC: Hepatocellular carcinoma; BCLC: Barcelona clinic liver cancer; MWA: Microwave ablation; TACE: Transarterial chemoembolization; mRECIST: Modified response evaluation criteria in solid tumors; miRNA: MicroRNA.





#### Figure 1 Receiver operating characteristic curve of the studied miRNAs.

In our study, miRNA-326 was upregulated, in contrast to the aforementioned studies in which miRNA-326 was down-regulated. Importantly, these studies were on HCC tissues and cell lines, while our study adopted an easier methodology of testing miRNA-326 in the blood. It seems that blood tests could give paradoxical results. The study by Moya et al [32] assessed different miRNAs as biomarkers for prostate cancer and found a similar paradox. This difference is attributed to the possibility of preferential retainment of oncomirs (these are miRNAs that are overexpressed in cancers) and the release of tumor suppressor miRNAs into the circulation to promote oncogenesis. Added to this, the authors highlighted the physiological cancer conditions that can cause leakage of molecules.

Table 2 Studied miRNA associations with different parameters in patients with hepatocellular carcinoma					
	miRNA-326	miRNA-511	miRNA-424		
Gender					
Male 48 (68.6%)	33.6 (11.3-157.4)	1.1 (0.4-3.4)	5.4 (3.2-10.4)		
Females 22 (31.4%)	42.3 (11.9-168.2)	1.1 (0.4-3.4)	4.4 (1.9-8.3)		
<i>P</i> value	0.7	0.8	0.3		
Smoker					
No 56 (80.0%)	27.4 (8.4-159.0)	1.1 (0.3-2.7)	5.4 (2.9-10.6)		
Yes 14 (20.0%)	67.0 (28.0-177.0)	1.2 (0.7-5.9)	4.1 (2.7-7.0)		
<i>P</i> value	0.3	0.3	0.3		
Diabetes mellitus					
No 52 (74.3%)	29.4 (9.1-119.1)	0.9 (0.3-2.5)	4.6 (2.6-9.0)		
Yes 18 (25.7%)	70.4 (16.6- 218.4)	1.5 (0.9-5.1)	6.5 (4.1-13.7)		
<i>P</i> value	0.1	0.08	0.04 <sup>a</sup>		
Child grade					
A 58 (82.9%)	64.4 (15.3-177.0)	1.2 (0.4-3.8)	5.1 (3.0-10.3)		
B 11 (15.7%)	16.1 (4.1-110.7)	0.9 (0.0-1.4)	5.2 (2.6-11.9)		
C 1 (1.4%)	5.7 (5.7-5.7)	0.03 (0.03-0.03)	0.9 (0.9-0.9)		
<i>P</i> value					
B vs A					
C vs A	0.03 <sup>a</sup>	0.1	0.7		
C vs B	0.2	0.1	0.1		
	0.7	0.3	0.3		
HCC stages					
Early	32.0 (7.5-121.0)	1.0 (0.3-2.6)	4.5 (2.3-8.0)		
Intermediate	69.8 (16.1-185.0)	1.4 (0.6-4.6)	5.3 (3.9-13.2)		
Late	26.2 (14.5-179.8)	0.9 (0.1-2.9)	5.9 (2.5-10.1)		
<i>P</i> value					
Intermediate vs Early	0.2	0.1	0.04 <sup>a</sup>		
Late vs Early	0.6	0.8	0.5		
Late vs Intermediate	0.7	0.2	0.6		
Performance status					
0; 43 (61.4%)	64.4 (15.8-153.3)	1.2 (0.5-3.5)	5.7 (3.1-10.2)		
1; 24 (34.3%)	18.9 (7.1-184.5)	0.6 (0.0-2.5)	4.0 (1.5-10.4)		
2; 2 (2.9%)	7.2 (0.5-7.8)	3.1 (0.4-3.5)	5.3 (4.8-5.5)		
3; 1 (1.4%)	160.9 (160.9- 160.9)	9.6 (9.6-9.6)	22.0 (22.0-22.0)		
<i>P</i> value	0.2	0.2	0.2		

Studied miRNAs are represented as Median with Interquartile range (25% -75%), the data were analyzed by Mann-Whitney U test except for the Studied miRNAs associations with Performance status which is analysed by the Kruskal Wallis Test. HCC: Hepatocellular carcinoma. <sup>a</sup>P value < 0.05 is significant.

> In our study, a statistically significant difference was noted while revising the miRNA signature of miRNA-326 and miRNA-424 between our HCV-related HCC patients and healthy controls. A clear diagnostic role was identified for both markers. We would like to document another crucial issue. We found no single miRNA to correlate with the different HCC parameters. For example, miRNA-424



Baishideng® WJH | https://www.wjgnet.com

Table 3 Studied miRNAs in healthy controls vs hepatocellular carcinoma					
	Controls	HCC patients	<i>P</i> value		
	<i>N</i> = 25	<i>N</i> = 70			
miRNA-326	1.2 (0.3- 30.8)	35.0 (12.5- 162.3)	0.001 <sup>b</sup>		
miRNA-511	2.1 (0.9-7.7)	1.2 (0.3-3.2)	0.02 <sup>a</sup>		
miRNA-424	6.1 (0.7-12.4)	5.1 (2.9-10.3)	0.4		
miRNA-424	6.1 (0.7-12.4)	5.1 (2.9- 10.3)	0.4		

Studied miRNAs are represented as median with interquartile range (25% -75%), the data were analyzed by the Mann-Whitney *U* test. HCC: Hepatocellular carcinoma.

<sup>a</sup>*P* value < 0.05 is significant.

<sup>b</sup>*P* value < 0.01 is highly significant.

Table 4 Diagnostic performance of the studied miRNAs									
	Cut off	Sensitivity	Specificity	PPV	NPV	Accuracy	AUC	95%CI	P value
miRNA-326	> 1.165	0.971	0.52	0.85	0.867	0.853	0.784	67.7-89.1	< 0.001 <sup>b</sup>
miRNA-511	< 2.063	0.714	0.6	0.833	0.429	0.684	0.654	53.1-77.7	0.01 <sup>a</sup>
miRNA-424	> 2.462	0.829	0.48	0.817	0.5	0.737	0.559	40.4-71.5	0.4

PPV: Positive predictive value; NPV: Negative predictive value; AUC: Area under curve; 95% CI: 95% Confidence interval.

<sup>a</sup>P value < 0.05 is significant.

<sup>b</sup>*P* value < 0.01 is highly significant.

differentiated early vs intermediate HCC significantly, miRNA-326 was a major factor related to survival, and miRNA-511 significantly correlated with response to treatment. miRNA-326 cut-off level > 1.165 showed statistically significant sensitivity and specificity when upregulated, while miRNA-511 cut-off < 2.063 showed statistically significant sensitivity and specificity when downregulated. This provides the main clue that no single miRNA can be used for the diagnosis and prognosis of HCC. Wang and Lei identified an eleven long non-coding RNA (lncRNA) signature for predicting HCC prognosis and incorporated seven miRNAs (including miRNA-326 and -424)[33]. In another study, the authors mentioned seven miRNAs that were found to have different expressions between tumors and in-vicinity non-tumorous tissues and found that they were significantly associated with survival (including miRNA-326 and -511)[23]. Similarly, a multidimensional signature stratified the HCC patients into low-risk and high-risk groups (P < 0.001) in the training set, validation set, and independent set, and all were statistically significant. It showed better survival prediction power when compared to TNM staging and included three miRNAs (-149, -424, and -579)[34]. In the study by Lu et al [24], five biomarkers (among 33 miRNAs) significantly correlated with patient survival. These markers were miRNA-326, -421, -511, -3677, and -424. They divided patients into low-risk and high-risk groups according to the miRNA signatures. None of the high-risk patients survived for 5 years. Thus, it seems that using a combination of miRNAs gives better results than using one biomarker.

The mechanism of action of miRNA in HCC has been proposed in several experimental studies. Concerning miRNA-326, different mechanisms of action were proposed, including NF-kB expression [35], targeting MAPK1 and CSF1 as regulated by circASAP1[21], and targeting LIM and SH3 protein 1 (LASP1)[36]. Also, it acts by suppression of PDK1 in the PDK1/AKT/C-myc axis[37]. It acts as a sponge for circular RNA circ-0000517[38] and a regulator for the SMAD 6 axis[39]. HCC proliferation was found to be promoted via CircRNA-PTN that also sponges miRNA-326 and affects ErbB/PI3K in HCC cells [40]. Finally, HOXD-AS1 binds directly to miRNA-326, thereby targeting gene SLC27A4, which also plays a role in HCC progression and metastasis promotion[41]. Proposed mechanisms of action of miRNA-424 include the direct targeting of C-Myb[19], AKT3 and E2F3[30], E2F7 expression[42], and acts as a sponge to different lncRNAs such as lncRNA CASC9[43], LINL 00657[44], lncRNA CDKN2B-AS1[45], and lncRNA LINC00511[34]. All these targets are correlated with poor prognosis, promotion of cell viability, and migration. Also, it affected angiogenesis by activating the VEGFR-2 signaling pathway [46]. Serum clinical samples from HCC patients and healthy volunteers showed that miRNA-424 significantly decreased in HCC patients and correlated with poor overall survival and disease-free survival[47]. With regard to miRNA-511, studies proposed its action through sponging LINC01559[22], regulation of the AKT1 axis[48], and targeting PIK3R3 in the PIK3R3/AKT/mTOR signaling pathway [49].

Table 5 Survival analysis					
		Dead, <i>N</i> = 34	Median (95%CI)of the EstimatedSurvival Time	Log rank (Mantel- Cox)	P value
Gender	Female (%)	9 (26.5)	315.0 (134.4- 495.6)	0.094	0.8
	Male (%)	25 (73.5)	441.0 (329.7- 552.3)		
Smoker	No (%)	29 (85.3)	395.0 (299.9- 490.1)	1.595	0.2
	Yes (%)	5 (14.7)	581.0 (336.4- 825.6)		
DM	No (%)	27 (79.4)	413.0 (341.7- 484.3)	0.259	0.6
	Yes (%)	7 (20.6)	429.0 (257.4- 600.6)		
BCLC stages	Early (%)	12 (35.3)	532.0 (401.2- 662.8)	5.594	0.05 <sup>a</sup>
	Intermediate (%)	14 (41.2)	393.0 (225.7- 560.3)		
	Late (%)	8 (23.5)	284.0 (196.6- 371.4)		
Performance status	0 (%)	21 (61.8)	395.0 (314.6 - 475.4)	1.188	0.6
	1 (%)	12 (35.3)	516.0 (404.0- 628.0)		
	2 (%)	1 (2.9)	278.0 (278.0- 278.0)		
	3	0 (0.0)	-		
Child score	A (%)	26 (76.5)	429.0 (316.6- 541.4)	0.574	0.8
	B (%)	7 (20.6)	375.0 (241.9- 508.1)		
	C (%)	1 (2.9)	516.0 (516.0- 516.0)		
Response to treatment	Stationary	4 (11.8%)	284.0 (215.4- 352.6)	2.9	0.4
mRECISI	Partial response	1 (2.9%)	412.0 (412.0- 412.0)		
	Complete response	19 (55.9%)	456.0 (346.7- 565.3)		
	Progressive disease	5 (14.7%)	128.0 (0.0- 285.2)		
miRNA-326 >1.165			441.0 (336.7- 545.3)	17.1	0.001 <sup>b</sup>
miRNA-511<2.063			441.0 (329.5- 552.5)	0.626	0.4
miRNA-424>2.462			395.0 (300.7- 489.3)	2.707	0.1

mRECIST: Modified Response Evaluation Criteria in Solid Tumors; BCLC: Barcelona Clinic Liver Cancer; DM: Diabetes mellitus; 95% CI: 95% Confidence interval.

<sup>a</sup>*P* value < 0.05 is significant.

<sup>b</sup>*P* value < 0.01 is highly significant.

Finally, we would like to highlight the aggressive behavior of HCC. Our patients were predominantly BCLC-A, Child-Pugh score A, performance status 0-1, and the majority had single lesions without evidence of vascular invasion or lymph node metastases. Moreover, we had good overall response rates (complete and partial response rates were nearly 60%). Nearly half of patients died during the follow-up period despite all these facts. This indicates the importance of primary prevention of HCV-related HCC through early management of its risk factors rather than managing HCC after its development.

Nonetheless, our study has limitations. We could not study more miRNAs that are correlated with HCC, and we could not correlate blood tests with tissue tests. However, our study is a clinical (not an experimental) one, and this helped to better correlate miRNA with different co-existing potentials and factors that could be present in actual life scenarios. In conclusion, we found a good diagnostic and prognostic role for our studied biomarkers (miRNA-326, -424, and -511) in HCC patients.

# CONCLUSION

We conclude that miRNA-326, miRNA-424, and miRNA-511 have diagnostic and prognostic roles in Egyptian patients with HCV-related HCC and should be considered for better disease management.



Table 6 Studied miRNA associations with response to treatment according to modified response evaluation criteria in solid tumors							
Response to treatment mRECIST	Stationary, <i>n</i> = 4 (5.7%)	Partial response, <i>n</i> = 7 (10.0%)	Complete response, <i>n</i> = 34 (48.6%)	Progressive disease, <i>n</i> = 6 (8.6%)	P value		
miRNA-326	11.5 (2.1- 142.6)	31.1 (6.9- 209.4)	27.4 (10.0- 155.2)	69.6 (17.7-197.9)	0.5		
miRNA-511	3.8 (0.8- 10.5)	1.2 (0.3- 4.2)	1.0 (0.2- 1.5)	0.6 (0.1-1.3)	0.05 <sup>a</sup>		
miRNA-424	4.9 (3.0- 8.6)	4.8 (2.1-5.1)	5.4 (2.6- 13.1)	4.6 (3.1-15.2)	0.8		

mRECIST: Modified Response Evaluation Criteria in Solid Tumors.

<sup>a</sup>P value < 0.05 is significant.



Figure 2 Survival curve among the studied parameters. A: Survival curve in relation to stage; B: Survival curve in relation to response to treatment according to mRECIST; C: Survival curve in relation to miRNA-326.

# **ARTICLE HIGHLIGHTS**

#### Research background

Hepatocellular carcinoma (HCC) is lethal and is the fifth most common cancer. Differential expression of microRNAs (miRNAs)-326, miRNA-424, and miRNA-511 has been associated with the diagnosis and prognosis of HCC in different populations. However, limited information is available regarding their expression in Egyptian HCC patients.

#### **Research motivation**

This study aimed to assess the role of circulating miRNAs-326, miRNA-424, and miRNA-511 from plasma as a non-invasive route of detection and to explore the impact of these miRNAs in Egyptian HCC patients.

#### **Research objectives**

The study objectives were to assess the role of circulating miRNAs-326, miRNA-424, and miRNA-511 in Egyptian HCC patients by a non-invasive method using plasma samples.

#### **Research methods**

This prospective observational study included 70 adult Egyptian patients who developed HCC on top



of HCV-related liver cirrhosis and 25 healthy age- and sex-matched participants who were seronegative for HCV and HBV, and served as controls. HCC was diagnosed according to the American Association for the Study of Liver Diseases updated practice guidelines, or was histo-pathologically confirmed. All patients were recruited from the multidisciplinary HCC clinic, the Endemic Medicine Department, Kasr al Ainy School of Medicine, Cairo University, and were HCC-treatment naïve at the time of enrollment. Exclusion criteria included; HBV co-infection, any cause of chronic liver disease other than HCV, any associated malignancies other than HCC, and prior treatment for HCV or HCC. Data collection: All patients were subjected to clinical assessment, laboratory investigations [complete blood count, prothrombin time and concentration, liver and kidney function tests, alpha-fetoprotein (AFP), hepatitis markers], ultrasound examination, and triphasic CT with documentation of HCC site, size, and number as well as the presence of PVT. At the time of enrolment into the study, blood samples were obtained from each participant, and Child-Pugh-score and BCLC stage were assessed based on the clinical, laboratory, and imaging data obtained. Whole blood samples were collected from each participant in 5mL sterile RNAase-free vacutainer tubes containing EDTA. Blood samples were collected on ice and processed within 30 min of collection. To separate the plasma, each blood sample was centrifuged for 10 min at 1900g at 4 °C. Plasma samples were carefully transferred into sterile RNase-free tubes. Plasma samples were then centrifuged for 10 min at 12000g at 4 °C to remove cellular nucleic acid contamination, and haemolysed plasma samples were excluded. Samples were separated into aliquots and immediately stored at 80 °C until processed. RNA extraction: Total RNA was extracted from 200 µL plasma using the miRNeasy Serum/plasma cell lysates kit, according to the manufacturer's instructions. RNA concentration and purity were monitored using a Nanodrop spectrophotometer device. Quantitative RT-PCR: The reverse transcription reaction was performed using TaqMan<sup>™</sup> MicroRNA Reverse Transcription Kit, according to the manufacturer's instructions. miRNA-326, miRNA-511, and miRNA-424 quantifications were carried out using quantitative real-time PCR. The qRT-PCR for each sample was carried out in duplicate using TaqMan 2x universal master mix II and TaqMan microRNA Assay Mix containing PCR primers and TaqMan probes for each miRNA. The expression level of RNU6B was used as an endogenous control for normalization. To determine miRNA relative expression, it was reported as a fold change ( $\triangle$  Ct and  $\triangle$   $\triangle$  Ct calculations). Patient management and followup: After blood sample withdrawal, all the patients were managed according to the BCLC guidelines after a case-by-case discussion. All HCC patients were followed up for a period of 24 mo. The response of HCC to treatment was evaluated according to mRECIST. The overall survival time of the patients was defined as the period from the initial presentation to the last follow-up or death.

#### Research results

The mean age of the studied cohort was  $62.0 \pm 7.6$  years. Most of the patients were male (68.8%), nondiabetic (74.3%), non-smokers (80%), and Child A (82.9%). According to the BCLC staging system, most patients were in the early stage (48.6%). Most of the HCC lesions were single (70%), present in the right hepatic lobe (84.3%), and not associated with PVT (92.9%). That is why most of the patients were subjected to hepatectomy and microwave ablation (MWA), and most of them showed complete response according to mRECIST. All HCC patients were followed up for a period of 24 mo or until death. The mean survival time was 367.1 ± 173.9 days. The median values of miRNA-326, miRNA-511, and miRNA-424 were 35, 1.2, and 5.1, respectively. Correlation of miRNAs with different parameters in HCC patients: There was no significant association between the studied miRNAs levels and gender, smoking, or the patients' performance status. The serum level of miRNA-424 was significantly elevated in diabetic patients. There was a significant difference in miRNA-326 between Child grades A and B. Moreover, there was a significant difference in miRNA-424 between early and intermediate HCC. Diagnostic efficiency of miRNAs in our patients: On comparing the miRNAs levels between healthy participants and HCC patients, it was found that HCC patients showed significantly higher levels of miRNA-326 (P = 0.001) and significantly lower levels of miRNA-511 (P = 0.02). ROC analysis revealed the diagnostic performance of the studied miRNAs. miRNA-326 showed the best diagnostic performance, diagnosing HCC at a cut-off value of 1.165 with a sensitivity of 97.1%, specificity of 52%, PPV of 85%, and NPV of 86.7%. The AUC was 78.4% (67.7-89.1), and the overall accuracy was 85.3% (P < 0.001). Association of miRNAs with response to treatment and survival: Despite finding no statistically significant differences in the studied miRNAs between surviving and non-surviving HCC patients, we found that miRNA-326 > 1.165 was significantly associated with overall survival (P = 0.001). Moreover, there was a significant association between the BCLC stage as well as the response to treatment according to mRECIST and overall survival. It was also found that miRNA-511 was significantly associated with response to treatment according to mRECIST.

#### Research conclusions

We conclude that miRNA-326, miRNA-424, and miRNA-511 have diagnostic and prognostic roles in Egyptian patients with HCV-related HCC and should be considered for better disease management.

#### Research perspectives

miRNA-326, miRNA-424, and miRNA-511 can be detected from plasma and have diagnostic and



prognostic roles in Egyptian patients with HCV-related HCC and should be considered for better disease management.

# ACKNOWLEDGEMENTS

The authors would like to acknowledge the National Research Centre, as well as Endemic Medicine and Hepato-gastroenterology Department, Faculty of Medicine, Cairo University, Cairo, Egypt, for providing laboratory space, equipment, and patient samples for achievement of the current work.

# FOOTNOTES

Author contributions: Youssef SS conceived the study; Youssef S and ElFiky A performed the experiments, data analysis, and statistical analysis; Elbaz T and Omran D wrote the first draft; Shousha HI performed clinical examinations, conceptualization, and patient data curation; Nabeel MM, Shousha HI, Marie MS, Elzahry MA, Hashem A, Guda MF, and Abdelaziz AO performed clinical examinations; all authors contributed to manuscript reviewing and editing.

Institutional review board statement: The study was approved by the local research ethics committee of the Endemic Medicine Department, Faculty of Medicine, Cairo University, and it was conducted according to guidelines of the Declaration of Helsinki 1975. Informed written consent was obtained from each participant. All patients signed a written informed consent before inclusion in the study.

Conflict-of-interest statement: The authors declare no competing interests.

Data sharing statement: The authors confirm that the data supporting the findings of this study are available within the article.

**ARRIVE guidelines statement:** The authors have read the ARRIVE guidelines, and the manuscript was prepared and revised according to the ARRIVE guidelines.

**Open-Access:** This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is noncommercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

#### Country/Territory of origin: Egypt

ORCID number: Samar Samir Youssef 0000-0001-7114-4325; Asmaa Elfiky 0000-0001-7730-1678; Mohamed M Nabeel 0000-0001-6002-9858; Hend Ibrahim Shousha 0000-0002-0928-4738; Tamer Elbaz 0000-0003-0816-9575; Dalia Omran 0000-0002-5513-6955; mohammad Saeed Marie 0000000258930513; Mohammad A Elzahry 000000193244917; Amr Abul-Fotouh 000000230314372; Ahmed Hashem 0000000000000000; Mohamed F Guda 0000000232324320; Ashraf O Abdelaziz 0000-0001-9168-7431.

S-Editor: Ma YJ L-Editor: Webster JR P-Editor: Ma YJ

# REFERENCES

- 1 Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. Int J Cancer 2015; 136: E359-E386 [PMID: 25220842 DOI: 10.1002/ijc.29210]
- 2 Astarci HM, Özyalvaçli G, Sertçelik A. Karaciğerin primer ve metastatik karsinomlarının ayrımında pCEA, mCEA, AFP ve CK19'un tanısal değeri. Abant Tıp Dergisi 2019; 5: 39-46 [DOI: 10.5505/abantmedj.2016.94557]
- 3 Llovet JM, Kelley RK, Villanueva A, Singal AG, Pikarsky E, Roayaie S, Lencioni R, Koike K, Zucman-Rossi J, Finn RS. Hepatocellular carcinoma. Nat Rev Dis Primers 2021; 7: 6 [PMID: 33479224 DOI: 10.1038/s41572-020-00240-3]
- 4 McGuire S. World Cancer Report 2014. Geneva, Switzerland: World Health Organization, International Agency for Research on Cancer, WHO Press, 2015. Adv Nutr 2016; 7: 418-419 [PMID: 26980827 DOI: 10.3945/an.116.012211]
- Baek KK, Kim JH, Uhm JE, Park SH, Lee J, Park JO, Park YS, Kang WK, Lim HY. Prognostic factors in patients with advanced hepatocellular carcinoma treated with sorafenib: a retrospective comparison with previously known prognostic models. Oncology 2011; 80: 167-174 [PMID: 21701230 DOI: 10.1159/000327591]
- Attwa MH, El-Etreby SA. Guide for diagnosis and treatment of hepatocellular carcinoma. World J Hepatol 2015; 7: 1632-6



1651 [PMID: 26140083 DOI: 10.4254/wjh.v7.i12.1632]

- 7 Kudo M. Surveillance, diagnosis, treatment, and outcome of liver cancer in Japan. Liver Cancer 2015; 4: 39-50 [PMID: 26020028 DOI: 10.1159/000367727]
- He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. Nat Rev Genet 2004; 5: 522-531 [PMID: 8 15211354 DOI: 10.1038/nrg1379]
- 9 Lu J, Getz G, Miska EA, Alvarez-Saavedra E, Lamb J, Peck D, Sweet-Cordero A, Ebert BL, Mak RH, Ferrando AA, Downing JR, Jacks T, Horvitz HR, Golub TR. MicroRNA expression profiles classify human cancers. Nature 2005; 435: 834-838 [PMID: 15944708 DOI: 10.1038/nature03702]
- 10 Guzel E, Okyay TM, Yalcinkaya B, Karacaoglu S, Goemen M, Akcakuyu MH. Tumor suppressor and oncogenic role of long non-coding RNAs in cancer. North Clin Istanb 2020; 7: 81-86 [PMID: 32232211 DOI: 10.14744/nci.2019.46873]
- Bueno MJ, Pérez de Castro I, Malumbres M. Control of cell proliferation pathways by microRNAs. Cell Cycle 2008; 7: 11 3143-3148 [PMID: 18843198 DOI: 10.4161/cc.7.20.6833]
- 12 Ivey KN, Srivastava D. MicroRNAs as regulators of differentiation and cell fate decisions. Cell Stem Cell 2010; 7: 36-41 [PMID: 20621048 DOI: 10.1016/j.stem.2010.06.012]
- 13 Garzon R, Calin GA, Croce CM. MicroRNAs in Cancer. Annu Rev Med 2009; 60: 167-179 [PMID: 19630570 DOI: 10.1146/annurev.med.59.053006.104707
- Zhang J, Wang Y, Zhen P, Luo X, Zhang C, Zhou L, Lu Y, Yang Y, Zhang W, Wan J. Genome-wide analysis of miRNA 14 signature differentially expressed in doxorubicin-resistant and parental human hepatocellular carcinoma cell lines. PLoS One 2013; 8: e54111 [PMID: 23359607 DOI: 10.1371/journal.pone.0054111]
- 15 Wang Y, Gao X, Wei F, Zhang X, Yu J, Zhao H, Sun Q, Yan F, Yan C, Li H, Ren X. Diagnostic and prognostic value of circulating miR-21 for cancer: a systematic review and meta-analysis. Gene 2014; 533: 389-397 [PMID: 24076132 DOI: 10.1016/j.gene.2013.09.038]
- Vaira V, Roncalli M, Carnaghi C, Faversani A, Maggioni M, Augello C, Rimassa L, Pressiani T, Spagnuolo G, Di Tommaso L, Fagiuoli S, Rota Caremoli E, Barberis M, Labianca R, Santoro A, Bosari S. MicroRNA-425-3p predicts response to sorafenib therapy in patients with hepatocellular carcinoma. Liver Int 2015; 35: 1077-1086 [PMID: 25040368 DOI: 10.1111/liv.12636]
- 17 Wang Y, Lee AT, Ma JZ, Wang J, Ren J, Yang Y, Tantoso E, Li KB, Ooi LL, Tan P, Lee CG. Profiling microRNA expression in hepatocellular carcinoma reveals microRNA-224 up-regulation and apoptosis inhibitor-5 as a microRNA-224-specific target. J Biol Chem 2008; 283: 13205-13215 [PMID: 18319255 DOI: 10.1074/jbc.M707629200]
- Jin Y, Wong YS, Goh BKP, Chan CY, Cheow PC, Chow PKH, Lim TKH, Goh GBB, Krishnamoorthy TL, Kumar R, Ng 18 TP, Chong SS, Tan HH, Chung AYF, Ooi LLPJ, Chang JPE, Tan CK, Lee CGL. Circulating microRNAs as Potential Diagnostic and Prognostic Biomarkers in Hepatocellular Carcinoma. Sci Rep 2019; 9: 10464 [PMID: 31320713 DOI: 10.1038/s41598-019-46872-8]
- 19 Yu L, Ding GF, He C, Sun L, Jiang Y, Zhu L. MicroRNA-424 is down-regulated in hepatocellular carcinoma and suppresses cell migration and invasion through c-Myb. PLoS One 2014; 9: e91661 [PMID: 24675898 DOI: 10.1371/journal.pone.0091661
- Zhang Y, Li T, Guo P, Kang J, Wei Q, Jia X, Zhao W, Huai W, Qiu Y, Sun L, Han L. MiR-424-5p reversed epithelial-20 mesenchymal transition of anchorage-independent HCC cells by directly targeting ICAT and suppressed HCC progression. Sci Rep 2014; 4: 6248 [PMID: 25175916 DOI: 10.1038/srep06248]
- 21 Hu ZQ, Zhou SL, Li J, Zhou ZJ, Wang PC, Xin HY, Mao L, Luo CB, Yu SY, Huang XW, Cao Y, Fan J, Zhou J. Circular RNA Sequencing Identifies CircASAP1 as a Key Regulator in Hepatocellular Carcinoma Metastasis. Hepatology 2020; 72: 906-922 [PMID: 31838741 DOI: 10.1002/hep.31068]
- Su Q, Wang H. Long non-coding RNA 01559 mediates the malignant phenotypes of hepatocellular carcinoma cells through targeting miR-511. Clin Res Hepatol Gastroenterol 2021; 45: 101648 [PMID: 33588099 DOI: 10.1016/j.clinre.2021.101648
- Zhang J, Chong CC, Chen GG, Lai PB. A Seven-microRNA Expression Signature Predicts Survival in Hepatocellular 23 Carcinoma. PLoS One 2015; 10: e0128628 [PMID: 26046780 DOI: 10.1371/journal.pone.0128628]
- 24 Lu M, Kong X, Wang H, Huang G, Ye C, He Z. A novel microRNAs expression signature for hepatocellular carcinoma diagnosis and prognosis. Oncotarget 2017; 8: 8775-8784 [PMID: 28060739 DOI: 10.18632/oncotarget.14452]
- 25 Marrero JA, Kulik LM, Sirlin CB, Zhu AX, Finn RS, Abecassis MM, Roberts LR, Heimbach JK. Diagnosis, Staging, and Management of Hepatocellular Carcinoma: 2018 Practice Guidance by the American Association for the Study of Liver Diseases. Hepatology 2018; 68: 723-750 [PMID: 29624699 DOI: 10.1002/hep.29913]
- 26 Pugh RN, Murray-Lyon IM, Dawson JL, Pietroni MC, Williams R. Transection of the oesophagus for bleeding oesophageal varices. Br J Surg 1973; 60: 646-649 [PMID: 4541913 DOI: 10.1002/bjs.1800600817]
- 27 Forner A, Reig ME, de Lope CR, Bruix J. Current strategy for staging and treatment: the BCLC update and future prospects. Semin Liver Dis 2010; 30: 61-74 [PMID: 20175034 DOI: 10.1055/s-0030-1247133]
- 28 Lencioni R, Llovet JM. Modified RECIST (mRECIST) assessment for hepatocellular carcinoma. Semin Liver Dis 2010; **30**: 52-60 [PMID: 20175033 DOI: 10.1055/s-0030-1247132]
- Du H, Xu Q, Xiao S, Wu Z, Gong J, Liu C, Ren G, Wu H. MicroRNA-424-5p acts as a potential biomarker and inhibits 29 proliferation and invasion in hepatocellular carcinoma by targeting TRIM29. Life Sci 2019; 224: 1-11 [PMID: 30876939 DOI: 10.1016/j.lfs.2019.03.028]
- 30 Yang H, Zheng W, Shuai X, Chang R-M, Yu L, Fang F, Yang L-Y. MicroRNA-424 inhibits Akt3/E2F3 axis and tumor growth in hepatocellular carcinoma. Oncotarget 2015; 6: 27736
- 31 Wu L, Yang F, Lin B, Chen X, Yin S, Zhang F, Xie H, Zhou L, Zheng S. MicroRNA-424 expression predicts tumor recurrence in patients with hepatocellular carcinoma following liver transplantation. Oncol Lett 2018; 15: 9126-9132 [PMID: 29805644 DOI: 10.3892/ol.2018.8539]
- Moya L, Meijer J, Schubert S, Matin F, Batra J. Assessment of miR-98-5p, miR-152-3p, miR-326 and miR-4289 32 Expression as Biomarker for Prostate Cancer Diagnosis. Int J Mol Sci 2019; 20 [PMID: 30845775 DOI: 10.3390/ijms20051154]



- 33 Wang A, Lei J. Identification of an 11-IncRNA signature with high performance for predicting the prognosis of hepatocellular carcinoma using bioinformatics analysis. Medicine (Baltimore) 2021; 100: e23749 [PMID: 33592832 DOI: 10.1097/MD.00000000023749
- 34 Wang RP, Jiang J, Jiang T, Wang Y, Chen LX. Increased long noncoding RNA LINC00511 is correlated with poor prognosis and contributes to cell proliferation and metastasis by modulating miR-424 in hepatocellular carcinoma. Eur Rev Med Pharmacol Sci 2019; 23: 3291-3301 [PMID: 31081082 DOI: 10.26355/eurrev\_201904\_17691]
- 35 Bai ZZ, Li HY, Li CH, Sheng CL, Zhao XN. M1 Macrophage-Derived Exosomal MicroRNA-326 Suppresses Hepatocellular Carcinoma Cell Progression Via Mediating NF-κB Signaling Pathway. Nanoscale Res Lett 2020; 15: 221 [PMID: 33263825 DOI: 10.1186/s11671-020-03432-8]
- 36 Hu S, Ran Y, Chen W, Zhang Y, Xu Y. MicroRNA-326 inhibits cell proliferation and invasion, activating apoptosis in hepatocellular carcinoma by directly targeting LIM and SH3 protein 1. Oncol Rep 2017; 38: 1569-1578 [PMID: 28713953 DOI: 10.3892/or.2017.5810]
- 37 Mo Y, He L, Lai Z, Wan Z, Chen Q, Pan S, Li L, Li D, Huang J, Xue F, Che S. Gold nano-particles (AuNPs) carrying miR-326 targets PDK1/AKT/c-myc axis in hepatocellular carcinoma. Artif Cells Nanomed Biotechnol 2019; 47: 2830-2837 [PMID: 31298047 DOI: 10.1080/21691401.2018.1489266]
- He S, Yang J, Jiang S, Li Y, Han X. Circular RNA circ\_0000517 regulates hepatocellular carcinoma development via miR-38 326/IGF1R axis. Cancer Cell Int 2020; 20: 404 [PMID: 32863763 DOI: 10.1186/s12935-020-01496-1]
- 39 He S, Guo Z, Kang Q, Wang X, Han X. Circular RNA hsa\_circ\_0000517 modulates hepatocellular carcinoma advancement via the miR-326/SMAD6 axis. Cancer Cell Int 2020; 20: 360 [PMID: 32774154 DOI: 10.1186/s12935-020-01447-w]
- 40 Jia B, Yin X, Wang Y, Qian J, He Y, Yang C, Yu G, Guo B, Meng X. CircRNA-PTN Sponges miR-326 to Promote Proliferation in Hepatocellular Carcinoma. Onco Targets Ther 2020; 13: 4893-4903 [PMID: 32581550 DOI: 10.2147/OTT.S251300]
- Ji W, Wang Q, Yang J. LncRNA HOXD-AS1 promotes the metastasis of human hepatocellular carcinoma via modulating 41 miR-326/SLC27A4. Cancer Cell Int 2020; 20: 161 [PMID: 32425696 DOI: 10.1186/s12935-020-01217-8]
- 42 Zhao Y, Zhu C, Chang Q, Peng P, Yang J, Liu C, Liu Y, Chen X, Cheng R, Wu Y, Wu X, Hu L, Yin J. MiR-424-5p regulates cell cycle and inhibits proliferation of hepatocellular carcinoma cells by targeting E2F7. PLoS One 2020; 15: e0242179 [PMID: 33201900 DOI: 10.1371/journal.pone.0242179]
- 43 Yao J, Fu J, Liu Y, Qu W, Wang G, Yan Z. LncRNA CASC9 promotes proliferation, migration and inhibits apoptosis of hepatocellular carcinoma cells by down-regulating miR-424-5p. Ann Hepatol 2021; 23: 100297 [PMID: 33346094 DOI: 10.1016/j.aohep.2020.100297]
- 44 Cao X, Zhang G, Li T, Zhou C, Bai L, Zhao J, Tursun T. LINC00657 knockdown suppresses hepatocellular carcinoma progression by sponging miR-424 to regulate PD-L1 expression. Genes Genomics 2020; 42: 1361-1368 [PMID: 32996041 DOI: 10.1007/s13258-020-01001-v]
- Shen X, Li Y, He F, Kong J. LncRNA CDKN2B-AS1 Promotes Cell Viability, Migration, and Invasion of Hepatocellular 45 Carcinoma via Sponging miR-424-5p. Cancer Manag Res 2020; 12: 6807-6819 [PMID: 32801906 DOI: 10.2147/CMAR.S240000
- Teng F, Zhang JX, Chang QM, Wu XB, Tang WG, Wang JF, Feng JF, Zhang ZP, Hu ZQ. LncRNA MYLK-AS1 facilitates 46 tumor progression and angiogenesis by targeting miR-424-5p/E2F7 axis and activating VEGFR-2 signaling pathway in hepatocellular carcinoma. J Exp Clin Cancer Res 2020; 39: 235 [PMID: 33168027 DOI: 10.1186/s13046-020-01739-z]
- Yao H, Liu X, Chen S, Xia W, Chen X. Decreased expression of serum miR-424 correlates with poor prognosis of patients 47 with hepatocellular carcinoma. Int J Clin Exp Pathol 2015; 8: 14830-14835 [PMID: 26823812]
- Yang X, Liu L, Zou H, Zheng YW, Wang KP. circZFR promotes cell proliferation and migration by regulating miR-48 511/AKT1 axis in hepatocellular carcinoma. Dig Liver Dis 2019; 51: 1446-1455 [PMID: 31147216 DOI: 10.1016/j.dld.2019.04.012
- Cao G, Dong W, Meng X, Liu H, Liao H, Liu S. MiR-511 inhibits growth and metastasis of human hepatocellular carcinoma cells by targeting PIK3R3. Tumour Biol 2015; 36: 4453-4459 [PMID: 25608840 DOI: 10.1007/s13277-015-3085-z





# Published by Baishideng Publishing Group Inc 7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA Telephone: +1-925-3991568 E-mail: bpgoffice@wjgnet.com Help Desk: https://www.f6publishing.com/helpdesk https://www.wjgnet.com

