

Antidepressant Effect of Aminophylline After Ethanol Exposure

**Sarah Souza ESCUDEIRO¹, Paula Matias SOARES²,
Anália Barbosa ALMEIDA¹, Rodrigo de Freitas Guimarães LOBATO¹,
Dayane Pessoa de ARAUJO¹, Danielle Silveira MACEDO¹,
Francisca Cléa Florenço SOUSA¹, Manoel Cláudio Azevedo PATROCÍNIO³,
Silvânia Maria Mendes VASCONCELOS^{* 1}**

¹ Department of Physiology and Pharmacology, Federal University of Ceará, Rua Cel. Nunes de Melo 1127, CEP 60431-270, Fortaleza, Ceará, Brazil.

² Superior Institute of Biomedical Sciences, Academic Master in Physiological Sciences, State University of Ceará, Av. Paranjana 1700, CEP 60740-000, Campus do Itaperi, Fortaleza, Ceará, Brazil.

³ Christus Medicine Faculty, Rua Israel Bezerra 630, CEP 60135-460, Fortaleza, Ceará, Brazil.

* Corresponding author. E-mail: silvania_vasconcelos@yahoo.com.br (S. M. M. Vasconcelos)

Sci Pharm. 2013; 81: 211–222

doi:10.3797/scipharm.1208-17

Published: October 23rd 2012

Received: August 30th 2012

Accepted: October 23rd 2012

This article is available from: <http://dx.doi.org/10.3797/scipharm.1208-17>

© Escudeiro *et al.*; licensee Österreichische Apotheker-Verlagsgesellschaft m. b. H., Vienna, Austria.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/3.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

This work investigated the association of acute ethanol and aminophylline administration on behavioral models of depression and prefrontal monoamine levels (i.e. norepinephrine and dopamine) in mice. The animals received a single dose of ethanol (2 g/kg) or aminophylline (5 or 10 mg/kg) alone or in association. Thirty minutes after the last drug administration, the animals were assessed in behavioral models by the forced swimming and tail suspension tests. After these tests, the animals were sacrificed and the prefrontal cortices dissected to measure monoamine content. Results showed that ethanol presented depression-like activity in the forced swimming and tail suspension tests. These effects were reversed by the association with aminophylline in all tests. Norepinephrine and dopamine levels decreased, while an increase in the dopamine metabolite, (4-hydroxy-3-methoxyphenyl)acetic acid (DOPAC), after ethanol administration was observed. On the contrary, the association of ethanol and aminophylline increased the norepinephrine and dopamine content, while it decreased DOPAC when compared to the ethanol group, confirming the alterations observed in the behavioral tests. These data reinforce the involvement of the adenosinergic system on ethanol effects, highlighting the

importance of the norepinephrine and dopamine pathways in the prefrontal cortex to the effects of ethanol.

Keywords

Adenosine • Ethanol • Aminophylline • Monoamines • Behavior • Prefrontal cortex

Introduction

Ethanol differentially interferes with the transmission processes in the central nervous system, affecting neurotransmitters [1, 2] and leading to a variety of behavioral and physiological changes such as motor incoordination, memory impairment, anxiety reduction, and depression, among others [3–5]. A significant proportion of alcohol-dependent individuals suffer from affective disorders such as depression [6–10]. However, the onset of major depression after alcohol dependence/abuse does not necessarily imply a causative relationship, and the pathogenesis remains obscure.

Among the wide range of pathways in the central nervous system that are modified by ethanol, it is important to highlight those that underlie ethanol's diverse effects, like the ones related to the release of gamma-aminobutyric acid (GABA), glutamate, dopamine, and norepinephrine [11, 12]. Moreover, another pathway that is of increasing interest in research about ethanol's effects is the adenosine system [13–15].

Adenosine was described as a potent depressor of neuronal activity [16], and acts mainly via the A₁ receptor, which is a presynaptic inhibitor of the release of neurotransmitters such as dopamine, GABA, glutamate, acetylcholine, and norepinephrine [17–19]. Moreover, adenosine is involved in behavioral processes such as motor function, anxiety, depression, reward, and drug addiction, as well as in human disorders, for instance, Parkinson's disease and schizophrenia [20]. Indeed, adenosine plays an important role in affective disorders. Clinical and experimental evidence showed that reduced adenosinergic activity is involved in bipolar mania and aggressive behavior [21] and A_{2A} receptors have been implicated in panic disorders [22].

In addition, there is strong evidence for the involvement of the adenosinergic system on ethanol effects, including: i) the increase in extracellular adenosine levels after acute ethanol exposure [23, 24], ii) the potentiation or blockade of ethanol-induced motor incoordination provided by adenosine receptor agonists or antagonists, respectively [5, 25], and iii) the reduction of anxiogenic-like behavior after acute ethanol withdrawal [13]. Adenosine antagonists, like caffeine, are implicated in alcohol tolerance [26] and in the retrograde memory impairment caused by ethanol [27]. Thus, adenosine receptors seem to modulate some of the pharmacological properties of ethanol, interacting with this drug with a resulting blockade or potentiation of its properties.

The adenosine antagonist aminophylline is an established drug in the clinical treatment of asthma and consists of approximately 80% of theophylline, its major active compound, which shows a central excitatory effect [28]. Another clinical use of aminophylline, due to its stimulatory effect, occurs in anesthesia to accelerate the consciousness recovery process after general anesthesia [29].

Because ethanol causes depression-like behavior, the present study investigated the ability of aminophylline, a non-selective adenosine receptor antagonist, to reverse ethanol's behavioral alteration using the tail suspension and forced swimming tests. Monoamine levels (i.e. norepinephrine and dopamine) in mice prefrontal cortices were also evaluated.

Materials and methods

Animals and drugs

Male Swiss mice (n= 6–14) weighing 25–30 g from the Animal House of the Federal University of Ceará were used. The animals had free access to a commercial diet (Purina, Brazil) and water, and were housed in groups of 10 in a room with a 12 h on-and-off lighting schedule. Experiments were performed according to the Guide for the Care and Use of Laboratory Animals, from the US Department of Health and Human Services, and approved by the Ethics Committee for Animal Use of the State University of Ceará (protocol nº 08476336-1).

A 20 percent ethanol (Merck, Germany) solution (w/v) was administered orally (p.o.), with an intragastric cannula, at 2 g/kg body weight. Aminophylline (Brazilian Teuto Laboratory S/A, Brazil) was administered intraperitoneally (i.p.), at 5 or 10 mg/kg body weight. All drugs were diluted in distilled water.

Experimental procedure

Mice were treated with distilled water (controls), ethanol (E: 2 g/kg, p.o.), or aminophylline (A: 5 or 10 mg/kg, i.p.). For the association protocol (E/A), mice were pre-treated with ethanol 30 minutes before aminophylline (5 or 10 mg/kg, i.p.) administration. All drug doses, as well as the sequence of administration, were chosen according to previous studies in our laboratory [5, 30].

The effects of ethanol and aminophylline alone or in association were studied in mice behavioral models (forced swimming and tail suspension tests) 30 min after the last drug administration, and the monoamine concentrations were determined after the tests. Different groups of animals were used for each test.

Forced swimming test

To evaluate the antidepressant activity of the treatment with ethanol and/or aminophylline, the Porsolt protocol [31] was used which includes two exposures to a water tank, spaced 24 hours apart. For these experiments, the tank size was 22 cm in diameter and 40 cm in height. The tank had a rounded lid and contained 20 cm high fresh water at 25 °C. During the first exposure, mice were placed in the tank and left there for 15 min. During the second exposure (test session), mice were placed in the tank and left there for 5 min during which immobility time was registered. A mouse was considered immobile when it remained floating in the water, without struggling, making only very slight movements necessary to keep its head above water. Each animal was tested once.

Tail suspension test

The tail suspension test is a screening procedure to detect antidepressant activity of drugs in rodents. The total duration of the test (6 min), as originally proposed by Steru, Chermat [32], can be divided into periods of agitation and immobility. Antidepressant drugs decrease the duration of immobility time, as do psychostimulants. In the present protocol, mice were suspended on the edge of a shelf, 58 cm above a table top, by an adhesive tape placed approximately 1 cm from the tip of their tails. The duration of immobility was recorded for a 6 min period.

Determination of monoamine concentrations

For determination of monoamine concentrations, the groups were sacrificed after the tests, and the prefrontal cortex was dissected on ice for the preparation of 10% homogenates (10% w/v) that were sonicated in 0.1 M HClO₄, for 30 sec and centrifuged at 4 °C for 20 min at 14000 RPM. A 20 µL sample of the supernatant was then analyzed by high performance liquid chromatography (HPLC). The mobile phase was 0.163 M citric acid (pH 3.0) containing 0.02 mM EDTA, with 0.69 mM sodium octanesulfonic acid (SOS), as an ion pairing reagent, 4% v/v acetonitrile, and 1.7% v/v tetrahydrofuran.

Prefrontal cortex concentrations of norepinephrine (NE), dopamine (DA), and its metabolite (4-hydroxy-3-methoxyphenyl)acetic acid (DOPAC), were detected electrochemically using an amperometric detector (Shimadzu, Japan) by oxidation on a glassy carbon electrode at 0.85 V relative to the Ag–AgCl reference electrode and results were expressed as ng/g wet tissue.

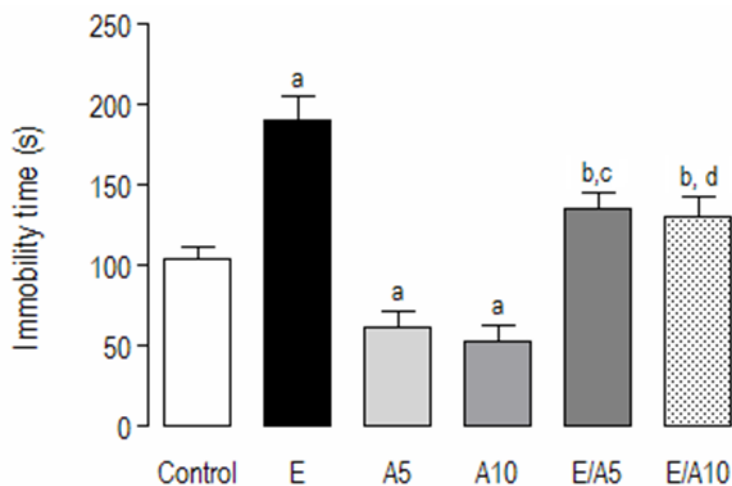


Fig. 1. Evaluation of the potential antidepressant-like activity of the association of ethanol and aminophylline using the forced swimming test. Animals were pre-conditioned 24h before the test, being exposed for 15 min in a water tank, without drugs. On the test day, 30 minutes after the drug administration, animals were tested in the forced swimming test, with the immobility time as the parameter observed over 5 minutes. a, b, c, d mean statistically significant differences, as related to the Control, E, A5, and A10 groups, respectively. $p < 0.05$ (ANOVA followed by Tukey as the *post hoc* test). (Abbreviations: E: ethanol, A: aminophylline).

Statistical analyses

In the present study, all results are presented as the mean \pm standard error media (S.E.M). Data were analyzed by One-Way ANOVA followed by Tukey as a *post hoc* test. Results were considered significant at $p < 0.05$. All tests were performed using the GraphPad Prism 5.0 software package.

Results

Forced swimming test

Figure 1 shows an increase in the immobility time in E-treated animals, while aminophylline in both doses studied decreased this parameter as compared to the controls (C: 104.8 ± 6.9 ; E: 190.8 ± 14.3 ; A5: 62.3 ± 9.4 ; A10: 53.7 ± 8.9). In the groups that received the association (E/A), the immobility time was reduced (E/A5: 135.6 ± 10.2 ; E/A10: 130.4 ± 12.4) as compared to the ethanol group, but increased when compared with aminophylline alone [F (5,69) = 23,84; $p < 0.0001$].

Tail suspension test

In the tail suspension test, an increase in immobility time was observed in the E group, while aminophylline (5 and 10 mg/kg) decreased this parameter when compared to the controls (C: 78.5 ± 7.6 ; E: 112.8 ± 11.6 ; A5: 40.7 ± 7.4 ; A10: 56.9 ± 7.1). Furthermore, both association groups presented a reversal of ethanol's depression-like effect, thus showing a decrease in the immobility time as compared to E and the control groups (E/A5: 39.4 ± 7.4 ; E/A10: 23.5 ± 4.4) [F (5,79) = 13,51; $p < 0.0001$] (Figure 2).

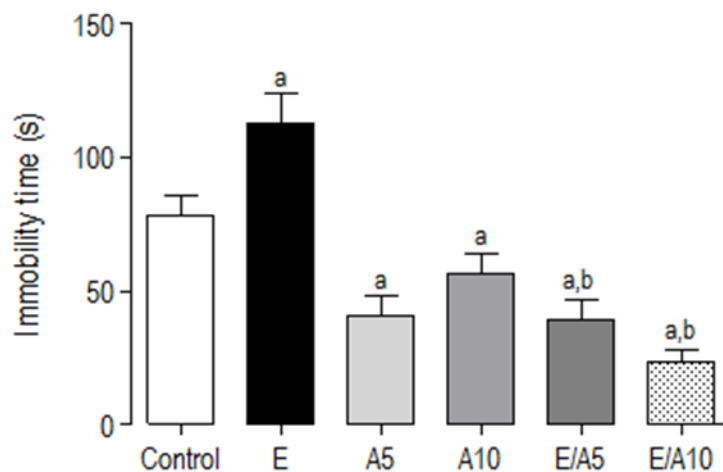


Fig. 2. Evaluation of the potential antidepressant activity of the association of ethanol and aminophylline using the tail suspension test. Thirty minutes after the drug administration, the animals were tested in the tail suspension test, with the immobility time as the parameter observed over 6 minutes. a, b mean statistically significant differences, as related to the Control and E groups, respectively. $p < 0.05$ (ANOVA followed by Tukey as the *post hoc* test). (Abbreviations: E: ethanol, A: aminophylline).

Monoamine levels

The levels of NE, DA, and DOPAC in the prefrontal cortex are presented in Figure 3. The results showed that NE and DA content decreased in ethanol-treated groups as compared to the control group [$F(5,64) = 14,85$; $p < 0.0001$]. The decrease in NE and DA content seen in the ethanol group was reversed by the association with aminophylline only in the lower dose (E/A5: 2820 ± 266.9), which increased the levels of these monoamines by 70% (NE) and 97% (DA) when compared to the ethanol group (NE: 1657 ± 131.1 ; DA: 1180 ± 103.8). Aminophylline alone, in both doses, did not alter the levels of these monoamines.

Analyzing the monoamines, metabolites, and ethanol (677.6 ± 59.1) increased the DOPAC levels as compared to the control (C: 362.8 ± 67.2) and the association groups (E/A5 or E/A10). This drug was also effective in the reversal of the metabolite alterations caused by ethanol, reducing the DOPAC concentration when compared to the ethanol group (E: 677.6 ± 59.1 ; E/A5: 322.6 ± 39.9 ; E/A10: 241.0 ± 63.2) [$F(5,54) = 5,42$; $p < 0.0005$].

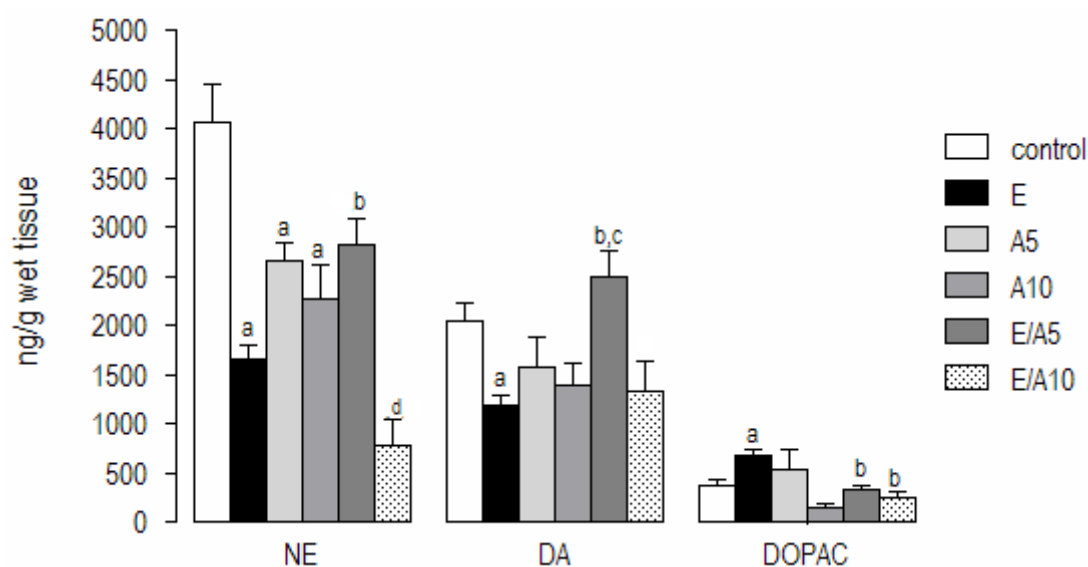


Fig. 3. Effects of ethanol associated with aminophylline on monoamine levels in mice prefrontal cortices. Data are presented as the mean \pm S.E.M. ($n = 4-14$). a, b, c, d, and e mean statistically significant differences with $p < 0.05$ as compared to the controls, A5, A10, and E/A5 groups, respectively. NE, norepinephrine; DA, dopamine; DOPAC, 4-hydroxy-3-methoxy-phenylacetic acid.

Discussion

We investigated the effects of the association of acute ethanol and aminophylline administration in behavioral models for the assessment of depression-like activity, i.e. forced swimming and tail suspension tests, which are predictive models of clinical antidepressant activity. Because regulations in monoamine levels are related to depression-like activity, we also assessed monoamine levels in the prefrontal cortex.

In the present work, the administration of ethanol in mice induced a depression-like behavior, as indicated by a significant increase in the immobility time in the forced swimming and tail suspension tests. The comorbidity of substance use disorders (for

example ethanol) and depression is highly prevalent in the general population [7, 33, 34]. On the other hand, aminophylline alone demonstrated an antidepressant-like action, thus presenting an opposite effect when compared to ethanol alone in almost all behavioral parameters analyzed. Worthy of mention, is our results have demonstrated that aminophylline is able to reverse ethanol depression-like behavior.

Aminophylline is a non-selective adenosine receptor antagonist [35, 36]. There is evidence that adenosine is a neuromodulator which takes part in a variety of processes in both physiological and pathological conditions. Adenosine and its analogues tend to produce depression-like behavior in animal models [37]. Indeed, adenosine and 2-chloroadenosine increased the immobility time of mice subjected to the forced swim test [38], while classical antidepressants have been found to reverse adenosine-mediated immobility [39]. It was shown that adenosine A_{2A} receptor knockout mice displayed reduced immobility in functional assays *in vivo*, such as in the tail suspension and forced swim tests [37].

In addition, an increasing body of evidence points to a direct role of adenosine in mediating some of the cellular and behavioral responses to ethanol [15, 40]. Caffeine and selective adenosine receptor antagonists reduce the duration of ethanol-induced loss of the righting reflex [38, 41], block the motor incoordination promoted by ethanol [5, 25, 42], and reverse retrograde memory impairment caused by a high dose of ethanol (3 g/kg) [27]. Indeed, adenosine A_1 receptors modulate the anxiolytic-like actions of ethanol [13], and it has been suggested that the reinforcing properties of ethanol are in part mediated via A_2 activation of cyclic adenosine monophosphate/phosphokinase A signaling in the nucleus accumbens, predicting that the administration of the A_2a antagonist might reduce ethanol reward and consumption [15].

Considering the alterations in monoamine levels, the results showed that the depression-like behavior produced by ethanol in the behavioral tests was followed by a decrease in norepinephrine and dopamine content in the prefrontal cortex, and this effect was reversed by aminophylline treatment, only in the lower dose.

According to the monoamine theory of depression, depressive disorders could be a result of low concentrations of monoamines such as norepinephrine, dopamine, and serotonin in putative brain areas [43, 45]. The depression-like behavior observed in our results after ethanol administration could be a consequence of a decrease in monoamine levels. However, it is noteworthy that ethanol's action in the brain is complex with a wide range of neurotransmitters involved in what can lead to a variety of behavioral and physiological alterations like sedation, amnesia, motor incoordination, depression mood, seizure, and others [3, 5, 46].

Adenosine is an important neuromodulator of the CNS. In fact, this purine nucleoside can modulate a variety of neurotransmitters such as norepinephrine, dopamine, and serotonin [19]. The modulation of the dopaminergic system occurs through its interaction with dopaminergic receptors, and it is thought that this antagonistic interaction between adenosine A_{2A} /dopamine D_2 and adenosine A_1 /dopamine D_1 receptors is at least partly responsible for the motor stimulant effects of adenosine receptor antagonists such as caffeine, and for the motor depressant actions of adenosine receptor agonists [47]. Dopaminergic neurons projecting to the prefrontal cortex provide direct and indirect inhibition of excitatory output to subcortical regions thought to be involved in the initiation

of motor activity. Thus, decreased dopamine transmission would enhance, while increased transmission would attenuate the response to psychostimulants [48]. According to this evidence, from our results we could observe that ethanol decreased dopamine levels, and this effect was reversed by the association with aminophylline only in the lower dose, also suggesting that aminophylline's effect could be dose-related.

Regarding the metabolites analyzed, we observed that DOPAC levels, the main dopamine metabolite in rodents, were increased after ethanol administration. A similar finding was presented by Myung *et al.* [4] in a study that evaluated the memory-enhancing abilities in ethanol-treated animals, in which the levels of some neurotransmitters were significantly changed by ethanol. This data suggests that there was an increase in dopamine's metabolic rate induced by ethanol. However, in the association group, DOPAC levels returned to the control ones, thus showing that aminophylline treatment is able to reverse the increase in dopamine metabolic rate induced by ethanol.

Conclusion

Ethanol produced a depression-like behavior as well as decreased the levels of NE and DA in the prefrontal cortex of mice, and these effects were reversed by the administration of aminophylline, a non-selective adenosine receptor antagonist.

Acknowledgement

This work was supported by grants from the National Council for Scientific and Technological Development (CNPq), Brazilian Coordination for Professorship Improvement for Higher Education (CAPES), and Research Foundation of the State of Ceará (FUNCAP), all from Brazil.

Authors' Statements

Competing Interests

The authors declare no conflict of interest.

Animal Rights

The institutional and international guide for the care and use of laboratory animals was followed. See the 'materials and methods' part for details.

References

- [1] Dahchour A, De Witte P. Ethanol and amino acids in the central nervous system: assessment of the pharmacological actions of acamprosate. *Progr Neurobiol.* 2000; 60: 343–362. [http://dx.doi.org/10.1016/S0301-0082\(99\)00031-3](http://dx.doi.org/10.1016/S0301-0082(99)00031-3)
- [2] Vasconcelos SM, Sales GT, Lima NM, Soares PM, Pereira EC, Fonteles MM, Sousa FC, Viana GS. Determination of amino acid levels in the rat striatum, after administration of ethanol alone and associated with ketamine, a glutamatergic antagonist. *Neurosci Lett.* 2008; 444: 48–51. <http://dx.doi.org/10.1016/j.neulet.2008.08.007>

- [3] Prediger RD, Batista LC, Takahashi RN. Adenosine A1 receptors modulate the anxiolytic-like effect of ethanol in the elevated plus-maze in mice. *Eur J Pharmacol.* 2004; 499: 147–154. <http://dx.doi.org/10.1016/j.ejphar.2004.07.106>
- [4] Myung CS, Shin HC, Bao HY, Yeo SJ, Lee BH, Kang JS. Improvement of memory by dieckol and phlorofucofuroeckol in ethanol-treated mice: possible involvement of the inhibition of acetylcholinesterase. *Arch Pharm Res.* 2005; 28: 691–698. <http://dx.doi.org/10.1007/BF02969360>
- [5] Soares PM, Patrocínio MC, Assreuy AM, Siqueira RC, Lima NM, Arruda Mde O, de Souza Escudeiro S, de Carvalho KM, Sousa FC, Viana GS, Vasconcelos SM. Aminophylline (a theophylline-ethylenediamine complex) blocks ethanol behavioral effects in mice. *Behav Pharmacol.* 2009; 20: 297–302. <http://dx.doi.org/10.1097/01.FBP.0000358355.88022.fa>
- [6] Gilder DA, Wall TL, Ehlers CL. Comorbidity of select anxiety and affective disorders with alcohol dependence in southwest California Indians. *Alcohol Clin Exp Res.* 2004; 28: 1805–1813. <http://dx.doi.org/10.1097/01.ALC.0000148116.27875.B0>
- [7] Schuckit MA, Tipp JE, Bergman M, Reich W, Hesselbrock VM, Smith TL. Comparison of induced and independent major depressive disorders in 2,945 alcoholics. *Am J Psychiatry.* 1997; 154: 948–957. <http://www.ncbi.nlm.nih.gov/pubmed/9210745>
- [8] Kessler RC, Crum RM, Warner LA, Nelson CB, Schulenberg J, Anthony JC. Lifetime co-occurrence of DSM-III-R alcohol abuse and dependence with other psychiatric disorders in the National Comorbidity Survey. *Arch Gen Psychiatry.* 1997; 54: 313–321. <http://dx.doi.org/10.1001/archpsyc.1997.01830160031005>
- [9] Hasin DS, Grant BF. Major depression in 6050 former drinkers: association with past alcohol dependence. *Arch Gen Psychiatry.* 2002; 59: 794–800. <http://dx.doi.org/10.1001/archpsyc.59.9.794>
- [10] Grant BF, Harford TC. Comorbidity between DSM-IV alcohol use disorders and major depression: results of a national survey. *Drug Alcohol Depend.* 1995; 39: 197–206. [http://dx.doi.org/10.1016/0376-8716\(95\)01160-4](http://dx.doi.org/10.1016/0376-8716(95)01160-4)
- [11] Vasconcelos SM, Cavalcante RA, Aguiar LM, Sousa FC, Fonteles MM, Viana GS. Effects of chronic ethanol treatment on monoamine levels in rat hippocampus and striatum. *Braz J Med Biol Res.* 2004; 37: 1839–1846. <http://dx.doi.org/10.1590/S0100-879X2004001200009>
- [12] Kaneyuki T, Morimasa T, Okada H, Shohmori T. The effect of acute and repeated ethanol administration on monoamines and their metabolites in brain regions of rats. *Acta Med Okayama.* 1991; 45: 201–208. <http://www.ncbi.nlm.nih.gov/pubmed/1962527>
- [13] Prediger RD, da Silva GE, Batista LC, Bittencourt AL, Takahashi RN. Activation of adenosine A1 receptors reduces anxiety-like behavior during acute ethanol withdrawal (hangover) in mice. *Neuropsychopharmacology.* 2006; 31: 2210–2220. <http://dx.doi.org/10.1038/sj.npp.1301001>

- [14] Vasconcelos SMM, Escudeiro SS, Martin AL, Soares PM, Vieira Filho A, Silva L, Dias KC, Macêdo D, Sousa FC, Fonteles M.
Ethanol Interference on Adenosine System.
In: Gallelli L, ed.
Pharmacology.
Croatia: InTech; 2012.
<http://dx.doi.org/10.5772/33791>
- [15] Thorsell A, Johnson J, Heilig M.
Effect of the adenosine A2a receptor antagonist 3,7-dimethyl-propargylxanthine on anxiety-like and depression-like behavior and alcohol consumption in Wistar Rats.
Alcohol Clin Exp Res. 2007; 31: 1302–1307.
<http://dx.doi.org/10.1111/j.1530-0277.2007.00425.x>
- [16] Dunwiddie TV, Haas HL.
Adenosine increases synaptic facilitation in the in vitro rat hippocampus: evidence for a presynaptic site of action.
J Physiol. 1985; 369: 365–377.
<http://www.ncbi.nlm.nih.gov/pubmed/3005559>
- [17] Fredholm BB, AP IJ, Jacobson KA, Klotz KN, Linden J.
International Union of Pharmacology. XXV. Nomenclature and classification of adenosine receptors.
Pharmacol Rev. 2001; 53: 527–552.
<http://www.ncbi.nlm.nih.gov/pubmed/11734617>
- [18] Brand A, Vissienon Z, Eschke D, Nieber K.
Adenosine A(1) and A(3) receptors mediate inhibition of synaptic transmission in rat cortical neurons.
Neuropharmacology. 2001; 40: 85–95.
[http://dx.doi.org/10.1016/S0028-3908\(00\)00117-9](http://dx.doi.org/10.1016/S0028-3908(00)00117-9)
- [19] Dunwiddie TV, Masino SA.
The role and regulation of adenosine in the central nervous system.
Ann Rev Neurosc. 2001; 24: 31–55.
<http://dx.doi.org/10.1146/annurev.neuro.24.1.31>
- [20] Moreau JL, Huber G.
Central adenosine A(2A) receptors: an overview.
Brain Res Rev. 1999; 31: 65–82.
[http://dx.doi.org/10.1016/S0165-0173\(99\)00059-4](http://dx.doi.org/10.1016/S0165-0173(99)00059-4)
- [21] Machado-Vieira R, Lara DR, Souza DO, Kapczinski F.
Purinergeric dysfunction in mania: an integrative model.
Med Hypotheses. 2002; 58: 297–304.
<http://dx.doi.org/10.1054/mehy.2001.1543>
- [22] Lam P, Hong CJ, Tsai SJ.
Association study of A2a adenosine receptor genetic polymorphism in panic disorder.
Neurosci Lett. 2005; 378: 98–101.
<http://dx.doi.org/10.1016/j.neulet.2004.12.012>
- [23] Nagy LE, Diamond I, Casso DJ, Franklin C, Gordon AS.
Ethanol increases extracellular adenosine by inhibiting adenosine uptake via the nucleoside transporter.
J Biol Chem. 1990; 265: 1946–1951.
<http://www.ncbi.nlm.nih.gov/pubmed/2298733>
- [24] Krauss SW, Ghirnikar RB, Diamond I, Gordon AS.
Inhibition of adenosine uptake by ethanol is specific for one class of nucleoside transporters.
Mol Pharmacol. 1993; 44: 1021–1026.
<http://www.ncbi.nlm.nih.gov/pubmed/7902530>

- [25] Dar MS.
Modulation of ethanol-induced motor incoordination by mouse striatal A(1) adenosinergic receptor.
Brain Res Bull. 2001; 55: 513–520.
[http://dx.doi.org/10.1016/S0361-9230\(01\)00552-4](http://dx.doi.org/10.1016/S0361-9230(01)00552-4)
- [26] Fillmore MT.
Alcohol tolerance in humans is enhanced by prior caffeine antagonism of alcohol-induced impairment.
Exp Clin Psychopharmacol. 2003; 11: 9–17.
<http://dx.doi.org/10.1037/1064-1297.11.1.9>
- [27] Spinetta MJ, Woodlee MT, Feinberg LM, Stroud C, Schallert K, Cormack LK, Schallert T.
Alcohol-induced retrograde memory impairment in rats: prevention by caffeine.
Psychopharmacology. 2008; 201: 361–371.
<http://dx.doi.org/10.1007/s00213-008-1294-5>
- [28] Niemand D, Martinell S, Arvidsson S, Ekstrom-Jodal B, Svedmyr N.
Adenosine in the inhibition of diazepam sedation by aminophylline.
Acta Anaesthesiol Scan. 1986; 30: 493–495.
<http://dx.doi.org/10.1111/j.1399-6576.1986.tb02462.x>
- [29] Sakurai S, Fukunaga A, Fukuda K, Kasahara M, Ichinohe T, Kaneko Y.
Aminophylline reversal of prolonged postoperative sedation induced by propofol.
J Anesth. 2008; 22: 86–88.
<http://dx.doi.org/10.1007/s00540-007-0587-x>
- [30] Vasconcelos SM, Macedo DS, Lima LO, Sousa FC, Fonteles MM, Viana GS.
Effect of one-week ethanol treatment on monoamine levels and dopaminergic receptors in rat striatum.
Braz J Med Biol Res. 2003; 36: 503–509.
<http://dx.doi.org/10.1590/S0100-879X2003000400013>
- [31] Porsolt RD, Anton G, Blavet N, Jalfre M.
Behavioural despair in rats: a new model sensitive to antidepressant treatments.
Eur J Pharmacol. 1978; 47: 379–391.
[http://dx.doi.org/10.1016/0014-2999\(78\)90118-8](http://dx.doi.org/10.1016/0014-2999(78)90118-8)
- [32] Steru L, Chermat R, Thierry B, Simon P.
The tail suspension test: a new method for screening antidepressants in mice.
Psychopharmacology. 1985; 85: 367–370.
<http://dx.doi.org/10.1007/BF00428203>
- [33] Boden JM, Fergusson DM.
Alcohol and depression.
Addiction. 2011; 106: 906–914.
<http://dx.doi.org/10.1111/j.1360-0443.2010.03351.x>
- [34] Hauser SR, Getachew B, Taylor RE, Tizabi Y.
Alcohol induced depressive-like behavior is associated with a reduction in hippocampal BDNF.
Pharmacol Biochem Behav. 2011; 100: 253–258.
<http://dx.doi.org/10.1016/j.pbb.2011.08.014>
- [35] Nakada T, Kwee IL, Lerner AM, Remler MP.
Theophylline-induced seizures: clinical and pathophysiologic aspects.
West J Med. 1983; 138: 371–374.
<http://www.ncbi.nlm.nih.gov/pubmed/6858124>
- [36] Narimatsu E, Aoki M.
Transient depression of excitatory synaptic transmission induced by adenosine uptake inhibition in rat hippocampal slices.
Brain Res. 2000; 862: 284–287.
[http://dx.doi.org/10.1016/S0006-8993\(00\)02123-5](http://dx.doi.org/10.1016/S0006-8993(00)02123-5)

- [37] El Yacoubi M, Ledent C, Parmentier M, Bertorelli R, Ongini E, Costentin J, Vaugeois JM. Adenosine A2A receptor antagonists are potential antidepressants: evidence based on pharmacology and A2A receptor knockout mice. *Br J Pharmacol.* 2001; 134: 68–77. <http://dx.doi.org/10.1038/sj.bjp.0704240>
- [38] Porsolt RD, Bertin A, Jalfre M. Behavioral despair in mice: a primary screening test for antidepressants. *Arch Int Pharmacodyn Ther.* 1977; 229: 327–336. <http://www.ncbi.nlm.nih.gov/pubmed/596982>
- [39] Kulkarni SK, Mehta AK. Purine nucleoside--mediated immobility in mice: reversal by antidepressants. *Psychopharmacology.* 1985; 85: 460–463. <http://dx.doi.org/10.1007/BF00429665>
- [40] Houchi H, Warnault V, Barbier E, Dubois C, Pierrefiche O, Ledent C, Daoust M, Naassila M. Involvement of A2A receptors in anxiolytic, locomotor and motivational properties of ethanol in mice. *Genes Brain Behav.* 2008; 7: 887–898. <http://dx.doi.org/10.1111/j.1601-183X.2008.00427.x>
- [41] El Yacoubi M, Ledent C, Parmentier M, Costentin J, Vaugeois JM. Caffeine reduces hypnotic effects of alcohol through adenosine A2A receptor blockade. *Neuropharmacology.* 2003; 45: 977–985. [http://dx.doi.org/10.1016/S0028-3908\(03\)00254-5](http://dx.doi.org/10.1016/S0028-3908(03)00254-5)
- [42] Barwick VS, Dar MS. Adenosinergic modulation of ethanol-induced motor incoordination in the rat motor cortex. *Prog Neuropsychopharmacol Biol Psychiatry.* 1998; 22: 587–607. [http://dx.doi.org/10.1016/S0278-5846\(98\)00025-6](http://dx.doi.org/10.1016/S0278-5846(98)00025-6)
- [43] Coppen A. Indoleamines and affective disorders. *J Psychiatr Res.* 1972; 9: 163–171. [http://dx.doi.org/10.1016/0022-3956\(72\)90018-0](http://dx.doi.org/10.1016/0022-3956(72)90018-0)
- [44] Aguiar CC, Castro TR, Carvalho AF, Vale OC, Sousa FC, Vasconcelos SM. [Antidepressant drugs]. *Acta Med Port.* 2011; 24: 91–98. <http://www.ncbi.nlm.nih.gov/pubmed/21672446>
- [45] Schildkraut JJ. The catecholamine hypothesis of affective disorders: a review of supporting evidence. *Am J Psychiatry.* 1965; 122: 509–522. <http://www.ncbi.nlm.nih.gov/pubmed/5319766>
- [46] Hunt WA, Majchrowicz E. Studies of neurotransmitter interactions after acute and chronic ethanol administration. *Pharmacol Biochem Behav.* 1983; 1: 371–374. <http://www.ncbi.nlm.nih.gov/pubmed/6138773>
- [47] Fuxe K, Ferre S, Genedani S, Franco R, Agnati LF. Adenosine receptor-dopamine receptor interactions in the basal ganglia and their relevance for brain function. *Physiol Behav.* 2007; 92: 210–217. <http://dx.doi.org/10.1016/j.physbeh.2007.05.034>
- [48] Steketee JD. Neurotransmitter systems of the medial prefrontal cortex: potential role in sensitization to psychostimulants. *Brain Res Rev.* 2003; 41: 203–228. [http://dx.doi.org/10.1016/S0165-0173\(02\)00233-3](http://dx.doi.org/10.1016/S0165-0173(02)00233-3)