



Comparison of the Humphrey Field Analyzer and Photopic Negative Response of Focal Macular Electroretinograms in the Evaluation of the Relationship Between Macula Structure and Function

Kazuyuki Hirooka*, Kenji Yokoyama, Kana Tokumo and Yoshiaki Kiuchi

OPEN ACCESS

Edited by:

Michele Lanza, University of Campania Luigi Vanvitelli, Italy

Reviewed by:

Teresa Rolle, Università di Torino, Italy Luigi Di Perna, University of Campania Luigi Vanvitelli, Italy

> *Correspondence: Kazuyuki Hirooka khirooka9@gmail.com

Specialty section:

This article was submitted to Ophthalmology, a section of the journal Frontiers in Medicine

Received: 06 January 2021 Accepted: 08 February 2021 Published: 26 February 2021

Citation:

Hirooka K, Yokoyama K, Tokumo K and Kiuchi Y (2021) Comparison of the Humphrey Field Analyzer and Photopic Negative Response of Focal Macular Electroretinograms in the Evaluation of the Relationship Between Macula Structure and Function. Front. Med. 8:649971. doi: 10.3389/fmed.2021.649971 Department of Ophthalmology and Visual Science, Graduate School of Biomedical Sciences, Hiroshima University, Hiroshima, Japan

Purpose: To investigate the association between macular inner retinal layer thickness and macula visual field (VF) mean deviation as measured by the Humphrey Field Analyzer (HFA) or macular function as measured by focal macular electroretinograms (ERGs) in patients with glaucoma.

Methods: The participants in this cross-sectional study were 71 patients with glaucoma and 10 healthy controls. Macular inner retinal layer thickness and function were measured in all participants using optical coherence tomography (OCT) and HFA or focal macular ERGs, respectively. Macular OCT images were segmented into the macular retinal nerve fiber layer (mRNFL), macular ganglion cell layer/inner plexiform layer (GCL/IPL), and ganglion cell complex (GCC). Spearman correlation analysis was used to assess the relationship between macular inner retinal layer thickness and function.

Results: Focal macular ERGs were composed of a negative wave (N1), a positive wave (P1), and a slow negative wave (N2). The N2 response density was significantly reduced in eyes with glaucoma, and was significantly associated with the thickness of the mRNFL (R = 0.317), GCL/IPL (R = 0.372), or GCC (R = 0.367). The observed structure–function relationship was also significantly correlated with the HFA VF mean deviation for each thickness [mRNFL (R = 0.728), GCL/IPL (R = 0.603), or GCC (R = 0.754)].

Conclusions: Although a significant correlation was found between the N2 response density and the thickness of the macular inner layer, the observed structure-function relationship with the mean deviation of the HFA VF was higher than that of the N2 response density.

Keywords: visual field, optical coherance tomography, electroretinogram (ERG), glaucoma, structure-function

INTRODUCTION

Glaucoma is a group of ocular diseases known to be characterized by retinal ganglion cell (RGC) soma and axon loss (1, 2). As about 50% of the RGCs are within 4.5 mm of the foveal center (3), measuring macular RGC function could be useful for diagnosing glaucoma or predicting disease progression. Some studies investigated the relationship between local sensitivity loss on 10-2 visual field (VF) loss and macular ganglion cell/inner plexiform layer (GCL/IPL) thickness (4–7). Clarifying the relationship between macular GCL/IPL thickness and central visual function could help clinicians gain a better understanding of how to detect glaucomatous damage at the early stage and disease progression. The Humphrey Field Analyzer (HFA; Carl Zeiss Meditec, Dublin, CA) has been confirmed to have high test–retest variability, with fixation errors being one of the major factors (8).

The photopic negative response (PhNR), which originates from the activity of RGCs and their axons (9), is a negative wave that follows the photopic b-wave. Increasing evidence has shown that the PhNR can be useful in evaluating the functional condition of neurons in patients with glaucoma (10, 11). The amplitude of the focal PhNR has been shown to be significantly correlated with a reduction in both visual sensitivity as determined by standard automated perimetry (SAP) (11) and retinal nerve fiber layer (RNFL) thickness (10) in patients with glaucoma. The PhNR recorded from the macular area can be used to assess the function of associated RGCs (12). The PhNR recorded using multifocal electroretinograms (mfERGs) with pseudorandom sequence stimulation has been found to be reduced in patients with glaucoma compared with controls, and this reduction in multifocal PhNR (mfPhNR) amplitude was correlated with disease severity (13). Due to recent improvements in the mfERG technique, the pupil does not need to be dilated before recording the mfPhNR.

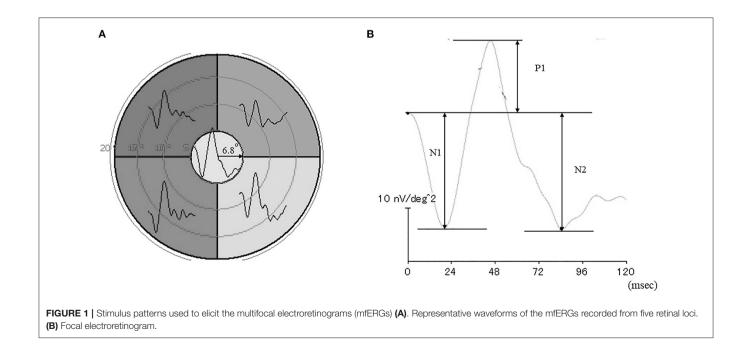
To improve the ability to detect the presence and progression of glaucomatous damage, numerous studies have applied spectral-domain optical coherence tomography (OCT) to examine the association between structural and functional damage. Given this background, the present study aimed to compare macular function measurements made by the HFA and focal macular PhNR, and to assess whether any potential relationships exist between these measurements and the thickness of the macular inner retinal layer.

MATERIALS AND METHODS

Patients

This cross-sectional study was carried out at Hiroshima University Hospital. The participants were all patients examined between November 2019 and August 2020. Before the study began, in accordance with the principles outlined in the Declaration of Helsinki, all participants were given a detailed explanation of the study purpose and methods and then asked to provide written informed consent. This study was approved by the Institutional Review Board of the Hiroshima University Faculty of Medicine.

First, all participants underwent a complete ophthalmic examination, which included visual acuity testing with refraction, intraocular pressure, gonioscopy, and a dilated fundus examination with stereoscopic biomicroscopy of the optic nerve head using indirect ophthalmoscopy and a slit lamp. Participants with a best-corrected visual acuity of $\geq 20/25$, a spherical error within a range of +4.0 and -6.0 diopters, a cylinder within \pm 2.0 diopters, an axial length <26 mm, and open angles (grades 3 and 4 according to the Shaffer grading



system) were included in the analysis. An optical biometer (IOLMaster, Carl Zeiss Meditec) was used to acquire axial length. Participants with a history of retinal pathology or neurologic disease or who had undergone a retinal laser procedure or either retinal or intraocular surgery were excluded. If both eyes met the inclusion criteria, the right eye was assessed. Control subjects were required to have an intraocular pressure ≤ 21 mmHg and a normal VF. All included eyes had to show the following structural glaucomatous changes to meet the definition of glaucoma: a vertical cup-disc asymmetry of ≥ 0.2 between the eves, a cup-to-disc ratio of >0.6, neuroretinal rim narrowing, notches, localized pallor, or RNFL defects with glaucomatous VF loss in the corresponding hemifield. To meet the definition of glaucomatous VF, the participant had to have undergone a glaucoma hemifield test outside of the normal limits in a minimum of two consecutive baseline tests, with at least three contiguous test points within the same hemifield on a pattern deviation plot at P < 1% and at least one contiguous test point at P < 0.5%, after excluding test points that were on the edge of the field or directly above and below the blind spot.

Measurement of Macular Inner Retinal Layer Thickness

Raster scanning [scan density of 512 (vertical) × 128 (horizontal) scans] of a 7 mm² area centered on the fovea was performed using a high-resolution fundus camera (Topcon 3D OCT-2000; Topcon, Inc., Tokyo, Japan). The built-in protocol measured a 6 × 6-mm area centered on the fovea using embedded software. The data were divided into 10 × 10 grids and exported by the Topcon software. Then, the mean thickness of the macular retinal nerve fiber layer (mRNFL), GCL/IPL, and ganglion cell complex (GCC), which consists of the mRNFL and GCL/IPL, were calculated. Images with a quality factor < 30 were excluded from the analysis.

Visual Sensitivity of the 10-2 HFA

Visual sensitivity was examined using static automated white-on-white threshold perimetry (HFA; 10-2 Swedish Interactive Threshold Algorithm Standard test). The VF results were considered reliable when the fixation losses and false-positive/false-negative rates were <20%. The subsequent analyses use only reliable test data.

mfERG Recordings

As described in a previous study on mfERG recordings (13) and shown in **Figure 1A**, stimuli consisting of five stimulus elements were generated on a cathode-ray tube monitor (VERIS? 7, Electro-Diagnostic Imaging, San Mateo, CA). mfERGs were elicited by a circular stimulus with a 6.8° radius centered on the fovea and a quarter of an annulus placed around the macula, with the radius of the outer border of the annulus set to 20° . White (200 cd/mm²) or black (4 cd/mm²) elements were presented in a pseudorandom binary m-sequence at a frequency of 6.25 Hz, with a steady background of 100 cd/m² surrounding the stimulus field.

A Burian-Allen bipolar contact lens electrode (Hansen Ophthalmic Laboratories, Coralville, IA) was placed on the cornea following corneal anesthesia. A chloride silver electrode as the ground electrode was placed on the left ear lobe. All responses were digitally band-pass filtered between 3 and 30 Hz. VERIS software (VERIS Science 4.1.1; Maya, Nagoya, Japan) was used to analyzed the mfERGs. The local retinal responses from the five different retinal loci were averaged to obtain the all-trace waveforms of the first-order kernels (**Figure 1A**) The response density of focal ERGs in the center area were evaluated as a macular function.

The N1 and P1 amplitudes were measured from the baseline to the trough of the first negative response and the peak of the following positive wave, respectively, and the N2 amplitude was measured from the baseline to the following trough (**Figure 1B**). The focal ERG amplitudes were expressed as response density (nV/deg²), which represents the amplitude as a function of the stimulus area.

Statistical Analysis

For continuous variables, variance equality was assessed using Levene's test. Based on the results obtained, a Student's *t*-test or Welch's test was used to assess differences between the control and glaucoma groups. The chi-square test for categorical parameters was used to assess differences between the control and glaucoma groups. Spearman rank order correlations were used to examine the correlation between mRNFL, GCL/IPL, and GCC thickness and VF mean deviation or N2 response density, and tests of equality of dependent correlation coefficients were used to evaluate comparisons of the strength of the structure– function association. All statistical values are presented as means

TABLE 1 | Clinical characteristics of the study population.

	Glaucoma (<i>n</i> = 71)	Normal (<i>n</i> = 10)	P-value
Age (y)	71.0 ± 1.5	68.2 ± 3.9	0.50
Gender (M/F)	39/32 6/4		0.76
Diagnosis			
POAG	46		
NTG	8		
EG	17		
Refraction (D)	-1.3 ± 0.2	-0.3 ± 0.6	0.13
Axial length (mm)	24.0 ± 0.1	23.5 ± 0.3	0.046

M, male; F, female; POAG, primary open-angle glaucoma; NTG, normal-tension glaucoma; EG, exfoliation glaucoma; D, diopter.

TABLE 2 | Macular inner retinal layer thickness and function.

Glaucoma	Normal	P-value
71.7 ± 1.2	101.0 ± 3.2	<0.001ª
51.6 ± 0.6	63.4 ± 1.5	<0.001ª
20.2 ± 0.9	37.8 ± 2.3	<0.001 ^b
14.01 ± 0.87	1.06 ± 2.31	<0.001 ^b
7.52 ± 0.76	14.14 ± 2.04	0.003 ^a
	71.7 ± 1.2 51.6 ± 0.6 20.2 ± 0.9 14.01 ± 0.87	71.7 ± 1.2 101.0 ± 3.2 51.6 ± 0.6 63.4 ± 1.5 20.2 ± 0.9 37.8 ± 2.3 14.01 ± 0.87 1.06 ± 2.31

GCC, ganglion cell complex; GCL, ganglion cell layer; IPL, inner plexiforma layer; mRNFL, macular retinal nerve fiber layer; MD, mean deviation.

^aStudent's t-test. ^bWelch test.

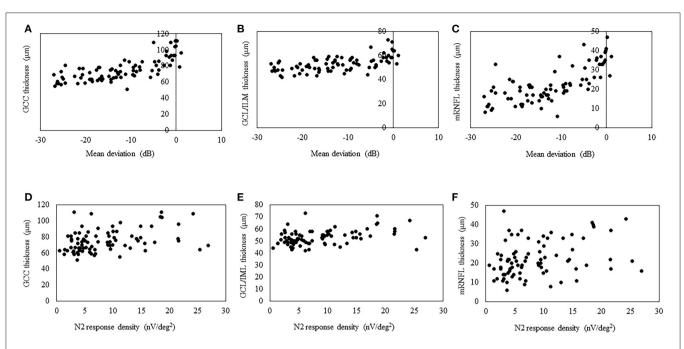


FIGURE 2 | Scatterplots showing the association between the 3D OCT-2000 thickness parameters and the corresponding retinal sensitivity (decibels) measured by the Humphrey Field Analyzer or center N2 response density. Association between the average thickness of the ganglion cell complex (GCC) (A), macular ganglion cell layer/inner plexiform layer (GCL/IPL) (B), or macular retinal nerve fiber layer (mRNFL) (C) and macular mean deviation. Association between the average thickness of the GCC (D), GCL/IPL (E), or mRNFL (F) and the center N2 response.

 \pm standard deviations (SDs), with P < 0.05 considered to be statistically significant. JMP software (version 15; SAS Inc., Cary, NC) was used for all statistical analyses.

RESULTS

Demographic Characteristics

The demographic characteristics of the 71 patients with glaucoma and 10 healthy controls who participated in the study are shown in **Table 1**. Disease grade in the glaucomatous eyes of the 71 patients, which was based on the standard VF severity grading scale (14), ranged from early to moderate, with 13 (18.3%), 17 (23.9%), and 41 (57.7%) eyes classified as early, moderate, and severe, respectively.

Comparison of the Normal and Glaucoma Groups

A significant difference in mRNFL, GCL/IPL, or GCC thickness was observed between the glaucoma and healthy control groups, as shown in **Table 2**. The mean deviation was significantly lower in the glaucomatous than in the healthy eyes, and the N2 response density was significantly reduced in the glaucomatous eyes, as shown in **Table 2**.

Correlation Between Macular Inner Layer Thickness and Mean Sensitivity and N2 Response Density

The structure-function relationship was evaluated based on the mRNFL, GCL/IPL, or GCC thickness and VF mean deviation

TABLE 3 | Comparison of the strength of the structure-function relationship

 between the Humphrey Field Analyzer and focal electroretinograms.

	Statistic	HFA	Focal ERG	P-value
GCC	Correlation coefficient	0.754	0.367	<0.001
	95% bootstrapped Cl	0.625–0.826	0.172-0.550	
GCL/IPL	Correlation coefficient	0.603	0.372	0.06
	95% bootstrapped Cl	0.444-0.727	0.155–0.537	
mRNFL	Correlation coefficient	0.728	0.317	< 0.001
	95% bootstrapped Cl	0.599–0.813	0.089–0.488	

HFA, humphrey field analyzer; ERG, electroretinogram; Cl, confidence interval; GCC, ganglion cell complex; GCL, ganglion cell layer; IPL, inner plexiform layer; mRNFL, macular retinal nerve fiber layer.

or N2 response density (**Figure 2** and **Table 3**). In each mRNFL, GCL/IPL, or GCC thickness, the structure–function relationship observed HFA VF mean deviation was higher than those of N2 response density. The Spearman correlation coefficient was the highest (0.754) for the GCC thickness-HFA VF mean sensitivity measurements. **Table 4** shows the structure–function relationship in each glaucoma type.

DISCUSSION

The retina contains about 1.07 million RGCs on average, approximately half of which are located within 4.5 mm of the foveal center (3, 15). During the early stages of glaucoma, RGC

TABLE 4 | Comparison of the strength of structure-function relationship between Humphrey Field Analyzer and multifocal electroretinogram in each glaucoma type.

		POAG (<i>n</i> = 46)		NTG (<i>n</i> = 8)		EG (<i>n</i> = 17)	
	Statistic	HFA	Focal ERG	HFA	Focal ERG	HFA	Focal ERG
GCC	Correlation coefficient	0.783	0.367	0.715	0.336	0.676	0.486
	95% bootstrapped Cl	0.661–0.864	0.087-0.538	0.270-0.908	0.263-0.749	0.405-0.838	0.138-0.727
GCL/IPL	Correlation coefficient	0.614	0.313	0.514	0.319	0.615	0.491
	95% bootstrapped Cl	0.429-0.750	0.067-0.524	0.051-0.830	0.281-0.740	0.314-0.803	0.144-0.703
mRNFL	Correlation coefficient	0.782	0.299	0.799	0.347	0.659	0.457
	95% bootstrapped Cl	0.661–0.864	0.052-0.513	0.444–0.938	0.252-0.754	0.380-0.829	0.102-0.709

POAG, primary open-angle glaucoma; NTG, normal-tension glaucoma; EG, exfoliation glaucoma; HFA, humphrey field analyzer; ERG, electroretinogram; CI, confidence interval; GCC, ganglion cell complex; GCL, ganglion cell layer; IPL, inner plexiform layer; mRNFL, macular retinal nerve fiber layer.

loss is evident around the fovea (16), which highlights the importance of assessing the central macular structure-function relationship. The N2 component of mfERGs recorded from the central area represents the RGC activity in the corresponding macular area (13).

Structure–Function Relationship of Glaucomatous Damage

Some researchers have recently reported on the diagnostic performance of mfERG in glaucoma patients (17, 18) or an animal model of glaucoma (19). The combination of mfERG and OCT improved diagnostic performance and monitoring of disease progression (17). By analyzing the mfPhNR/b-wave ratio, Al-Nosairy et al. (18) achieved the best performance for discriminating between controls and glaucoma suspects. The diagnostic performance and structure–function relationship were strongest for mfERG when compared with full-field flash ERG PhNR or pattern-reversal ERG in an experimental animal model of glaucoma (19). A combined approach using structural and functional assessment of glaucomatous retinal damage offers great promise for uncovering the interrelationship between the different components of ocular damage in glaucoma.

Correlation Between Macular Inner Layer Thickness and N2 Response Density

The high test-retest variability of SAP is often explained by poor patient vigilance and inattention in subjective examinations. By contrast, measurements of mfPhNR amplitude tend to show better test-retest reliability because of this is an objective test (20). Therefore, we hypothesized that the structure-function relationship for the observed N2 response would be higher than that for the HFA VA mean deviation. Although a significant correlation was found between N2 response density and macular inner layer thickness in this study, the structure-function relationship for the observed HFA VF mean deviation was higher than that for the N2 response density. Macular focal ERGs were elicited by a circular stimulus with a 6.8° radius centered on the fovea. The built-in protocol measured a 6×6 -mm area centered on the fovea corresponding to a 20° square of the retina in the macular area. The 3D-OCT used in this study and the 10-2 HFA measure similar macular areas (the 10-2 HFA analyzes 68 data points located within a central arc of 10°). Therefore, we assume that the results may be affected by the measurement area of each instrument. Moreover, the N2 may not represent the neural activity of RGCs only. In rodents, the PhNR has been shown to be affected by the neural activity of amacrine cells (21, 22).

In the present study, although a significant correlation was observed between the N2 response density and GCC thickness, the correlation coefficient was lower than that in a previous study (R = 0.363 vs. 0.575, respectively) (13), in which SD-OCT (RS-3000 Advance; Nidek Co., Ltd.) was used to obtain the GCC thickness. It is therefore difficult to compare the strength of the structure–function relationship in this study with that in their study because it can be affected by sample size, disease severity, and OCT instruments.

Limitations

This study did have some limitations. First, the present study did not include any patients with preperimetric glaucoma; such patients should be examined and compared with regard to the structure–function relationship in a future study. Second, although we observed no obvious differences in the structure– function relationship among patients with primary open-angle, normal tension, or exfoliation glaucoma, the sample size was small. Therefore, a large number of subjects will need to be closely examined for each glaucoma type in a future study.

CONCLUSIONS

The results of this study revealed that the N2 response density was affected by glaucoma in the central macular area. In addition, a significant correlation was found between the N2 amplitude and macular inner layer thickness; however, this correlation was weaker than that between the macular inner layer thickness and HFA VF mean deviation.

DATA AVAILABILITY STATEMENT

The datasets presented in this study can be found in online repositories. The names of the repository/repositories and accession number(s) can be found in the article/supplementary material.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by Institutional Review Board of the Hiroshima University Faculty of Medicine. The patients/participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

KH: conception of design of the work acquisition, analysis, and interpretation of the data final approval of the version to be published and agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy and integrity of any part of the work are appropriately investigated and resolved. KY: acquisition of the data final approval of the version to be published and agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy and integrity of any part of the work are appropriately investigated and resolved. KT: acquisition of the data final

REFERENCES

- Danias J, Shen F, Kavalarakis M, Chen B, Goldblum D, Lee K, et al. Characterization of retinal damage in the episcleral vein cauterization rat glaucoma model. *Exp Eye Res.* (2006) 82:219–28. doi: 10.1016/j.exer.2005.06.013
- Soto I, Pease ME, Son JL, Shi X, Quigley HA, Marsh-Armstrong N. Retinal ganglion cell loss in a rat ocular hypertension model is sectional and involves early optic nerve axon loss. *Invest Ophthalmol Vis Sci.* (2011) 52:434– 41. doi: 10.1167/iovs.10-5856
- Curcio CA, Allen KA. Topography of ganglion cells in human retina. J Comp Neurol. (1990) 300:5–25. doi: 10.1002/cne.903000103
- Raza AS, Cho J, de Moraes CG, Wang M, Zhang X, Kardon RH, et al. Retinal ganglion cell layer thickness and local visual field sensitivity in glaucoma. *Arch Ophthalmol.* (2011) 129:1529–36. doi: 10.1001/archophthalmol.2011.352
- Ohkubo S, Higashide T, Udagawa S, Sugiyama K, Hangai M, Yoshimura N, et al. Focal relationship between structure and function within the central 10 degrees in glaucoma. *Invest Ophthalmol Vis Sci.* (2014) 55:5269–77. doi: 10.1167/iovs.14-1 4153
- Hirooka K, Misaki K, Nitta E, Ukegawa K, Sato S, Tsujikawa A. Comparison of macular integrity assessment (MAIATM), MP-3, and the humphrey field analyzer in the evaluation of the relationship between the structure and function of the macula. *PLoS ONE.* (2016) 11:e0151000. doi: 10.1371/journal.pone.0151000
- Lee JW, Morales E, Sharifipour F, Amini N, Yu F, Afifi AA, et al. The relationship between central visual field sensitivity and macular ganglion cell/inner plexiform layer thickness in glaucoma. Br J Ophthalmol. (2017) 101:1052–8. doi: 10.1136/bjophthalmol-2016-309208
- Maddess T. Modeling the relative influence of fixation and sampling errors on retest variability in perimetry. *Graefes Arch Clin Exp Ophthalmol.* (2014) 252:1611–9. doi: 10.1007/s00417-014-2751
- Viswanathan S, Frishman LJ, Robson JG, Harwerth RS, Smith EL III. The photopic negative response of the macaque electroretinogram: reduction by experimental glaucoma. *Invest Ophthalmol Vis Sci.* (1999) 40:1124–36.
- Machida S, Toba Y, Ohtaki A, Gotoh Y, Kaneko M, Kurosaka D. Photopic negative response of focal electroretinograms in glaucomatous eyes. *Invest Ophthalmol Vis Sci.* (2008) 49:5636–44. doi: 10.1167/iovs.08-1946
- 11. Tamada K, Machida S, Oikawa T, Yokoyama D, Kaneko M, Kurosaka D. Correlation between photopic negative response of focal electroretinograms

approval of the version to be published and agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy and integrity of any part of the work are appropriately investigated and resolved. YK: final approval of the version to be published and agreement to be accountable for all aspects of the work in ensuring that questions related to the accuracy and integrity of any part of the work are appropriately investigated and resolved. All authors contributed to the article and approved the submitted version.

FUNDING

This work was supported by a Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science, and Technology of Japan (20K09827).

ACKNOWLEDGMENTS

We thank Forte Science Communications, Inc., for editing the draft of this manuscript.

and local loss of retinal neurons in glaucoma. *Curr Eye Res.* (2010) 35:155–64. doi: 10.3109/02713680903447926

- Machida S, Tamada K, Oikawa T, Yokoyama D, Kaneko M, Kurosaka D. Sensitivity and specificity of photopic negative response of focal electroretinogram to detect glaucomatous eyes. *Br J Ophthalmol.* (2010) 94:202–8. doi: 10.1136/bjo.2009.161166
- Kaneko M, Machida S, Hoshi Y, Kurosaka D. Alterations of photopic negative response of multifocal electroretinogram in patients with glaucoma. *Curr Eye Res.* (2015) 40:77–86. doi: 10.3109/02713683.2014.915575
- 14. Hodapp E, Parrish RK II, Anderson DR. *Clinical Detection in Glaucoma*. St. Louise, MO: Mosby (1993).
- Wässle H, Grünert U, Röhrenbeck J, Boycott BB. Retinal ganglion cell density and cortical magnification factor in the primate. *Vision Res.* (1990) 30:1897– 911. doi: 10.1016/0042-6989(90)90166-i
- Quigley HA, Dunkelberger GR, Green WR. Retinal ganglion cell atrophy correlated with automated perimetry in human eyes with glaucoma. *Am J Ophthalmol.* (1989) 107:453–64. doi: 10.1016/0002-9394(89) 90488-1
- Al-Nosairy KO, Prabhakaran GT, Pappelis K, Thieme H, Hoffmann MB. Damage with electroretinography and optical coherence tomography/angiogeaphy. *Trans Vis Sci Tech.* (2020) 9:7. doi: 10.1167//tvst.9.12.7
- Al-Nosairy KO, Thieme H, Hoffmann MB. Diagnostic performance of multifocal photopic negative responses, pattern electroretinogram and optical coherence tomography in glaucoma. *Exp Eye Res.* (2020) 200:108242. doi: 10.1016/j.exer.2020.10 8242
- Wilsey L, Gowrisankaran S, Cull G, Hardin C, Burgoyne CF, Fortune B. Comparing three different modes of electroretinography in experimental glaucoma: diagnostic performance and correlation to structure. *Doc Ophthalmol.* (2017) 134:111–28. doi: 10.1007//s10633-017-9 578-x
- reliability 20. Van Alstine AW, Viswanathan S. Test-retest of the multifocal photopic negative response. Doc Ophthalmol. (2017)134:25-36. doi: 10.1007/s10633-016-9 569-3
- Bui BV, Fortune B. Ganglion cell contributions to the rat full-field electroretinogram. J Physiol. (2004) 555:153–73. doi: 10.1113/jphysiol.2003.05 2738
- 22. Machida S, Raz-Prag D, Fariss RN, Sieving PA, Bush RA. Photopic ERG negative response from amacrine

cell signaling in RCS rat retinal degeneration. *Invest Ophthalmol Vis Sci.* (2008) 49:442–52. doi: 10.1167/iovs.07-0291

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Hirooka, Yokoyama, Tokumo and Kiuchi. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.