

# Divergent Modulation of Src-Family Kinase Regulatory Interactions with ATP-Competitive Inhibitors

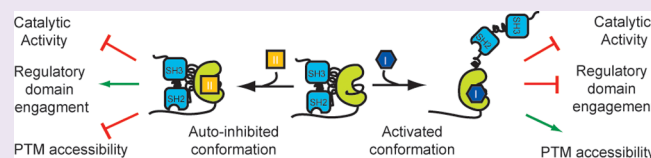
Stephen E. Leonard, A. C. Register, Ratika Krishnamurty, Gabriel J. Brighty, and Dustin J. Maly\*

Department of Chemistry, University of Washington, Seattle, Washington 98195, United States

## S Supporting Information

**ABSTRACT:** Multidomain protein kinases, central controllers of signal transduction, use regulatory domains to modulate catalytic activity in a complex cellular environment. Additionally, these domains regulate noncatalytic functions, including cellular localization and protein–protein interactions. Src-family kinases (SFKs) are promising therapeutic targets for a

number of diseases and are an excellent model for studying the regulation of multidomain kinases. Here, we demonstrate that the regulatory domains of the SFKs Src and Hck are divergently affected by ligands that stabilize two distinct inactive ATP-binding site conformations. Conformation-selective, ATP-competitive inhibitors differentially modulate the ability of the SH3 and SH2 domains of Src and Hck to engage in intermolecular interactions and the ability of the kinase–inhibitor complex to undergo post-translational modification by effector enzymes. This surprising divergence in regulatory domain behavior by two classes of inhibitors that each stabilize inactive ATP-binding site conformations is found to occur through perturbation or stabilization of the  $\alpha$ C helix. These studies provide insight into how conformation-selective, ATP-competitive inhibitors can be designed to modulate domain interactions and post-translational modifications distal to the ATP-binding site of kinases.



Protein kinases are critical mediators of cellular signaling through the propagation of phosphorylation cascades. For all kinases, a highly conserved bilobal domain containing an ATP-binding cleft is responsible for phosphotransfer activity.<sup>1,2</sup> In order to transmit signals with fidelity in the complex milieu of the cell, tight regulation of catalysis is required. This regulation is often achieved via fusion of the catalytic domain to regulatory or targeting domains.<sup>3</sup> These domains can allosterically regulate the activity of the kinase domain through intramolecular engagement and suppression of the catalytic domain.<sup>4,5</sup> Regulatory domains are not only important for modulating catalytic activity but also serve roles in other functions, including localization, DNA binding, and protein–protein interactions.<sup>6</sup> Often, these domains facilitate functions that are independent of kinase catalytic activity in the cell.

Src-family kinases (SFKs) are prototypical nonreceptor multidomain protein kinases consisting of regulatory SH2 and SH3 domains, a tyrosine kinase catalytic domain, and an N-terminal unique region. SFKs are involved in the regulation of important cellular processes including cell metabolism, proliferation, and differentiation.<sup>7–9</sup> Additionally, SFKs have prominent roles in invasion and tumor progression, angiogenesis, and metastasis, making them a promising target for cancer therapy.<sup>10–12</sup> More fundamentally, SFKs are a well-studied model for understanding how regulatory domains affect kinase catalysis.<sup>13,14</sup> SFK activity is allosterically suppressed by two intramolecular binding events: the SH2 domain's interaction with phospho-Tyr527 in the C-terminal tail and the SH3 domain's interaction with a proline-containing linker (SH2-kinase linker) that connects the SH2 domain with the catalytic domain.<sup>14–16</sup> Release of these interactions through dephosphorylation of pTyr527 or direct disruption of the

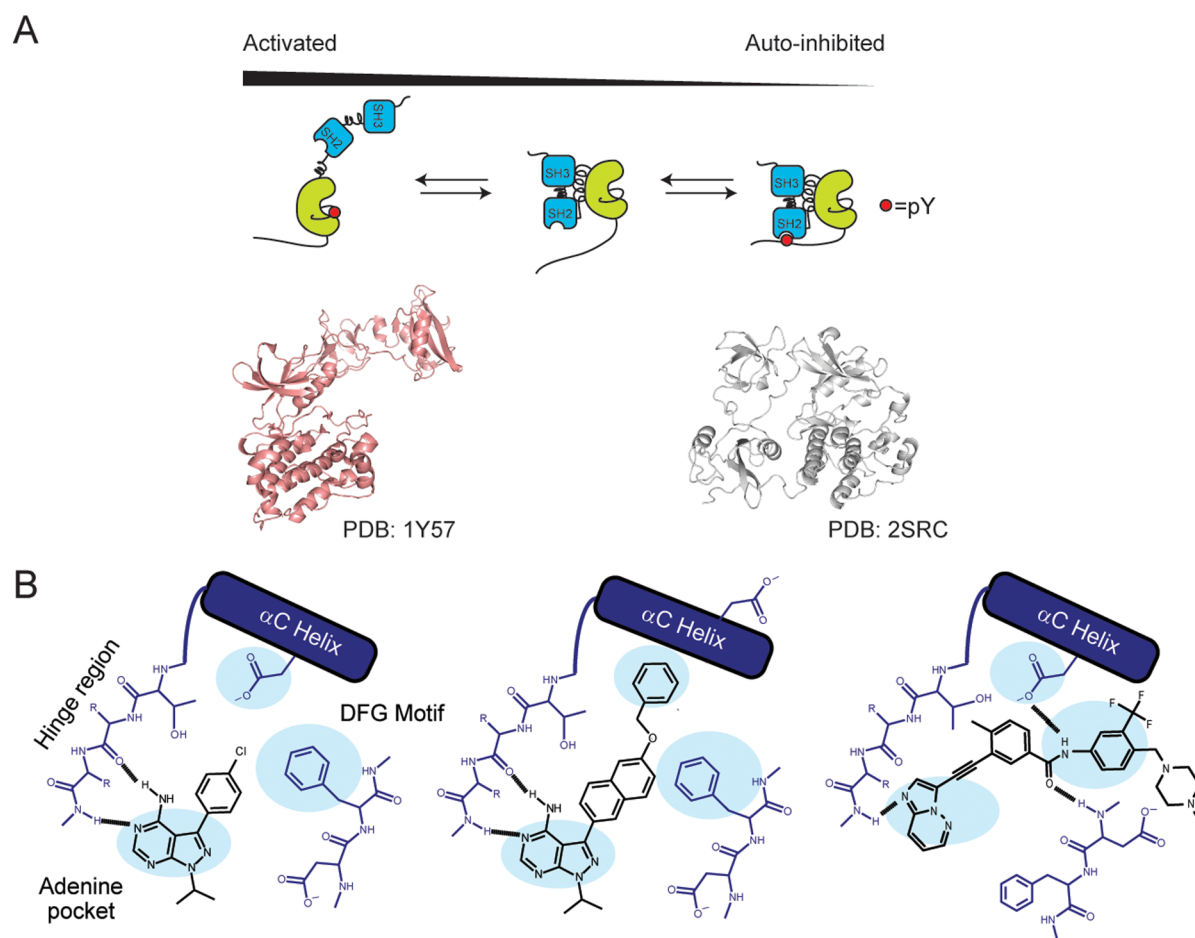
intramolecular SH2 and SH3 regulatory domain interactions leads to activation of the catalytic domain (Figure 1A). Full activation is achieved by phosphorylation of Tyr416 in the activation loop.<sup>15</sup>

Just as SFK regulatory domains undergo large conformational changes, their ATP-binding sites are also highly dynamic. The ATP-binding site of Src has been structurally characterized in three distinct conformations: one active and two inactive (DFG-out and  $\alpha$ C helix-out) forms (Figure 1). In the active conformation, all key catalytic residues are optimally positioned for catalysis, and two conserved networks of hydrophobic “spines” are aligned.<sup>1,17–19</sup> Both inactive ATP-binding site conformations are characterized by displacement of at least one conserved catalytic residue from the active site and disruption of the regulatory hydrophobic spine. The DFG-out inactive conformation involves flipping of the conserved Asp-Phe-Gly (DFG) motif at the base of the activation loop, which results in the displacement of the catalytic aspartic acid residue and removal of the phenylalanine side chain from the regulatory hydrophobic spine. Kinases in the  $\alpha$ C helix-out inactive conformation possess a disrupted salt bridge between the catalytic lysine (Lys295) and a conserved glutamic acid (Glu310) that is located on helix  $\alpha$ C (Figure 1B). Importantly, conformation-selective inhibitors that stabilize each of the three conformations described above have been identified for the SFKs.

Received: January 24, 2014

Accepted: June 19, 2014

Published: June 19, 2014



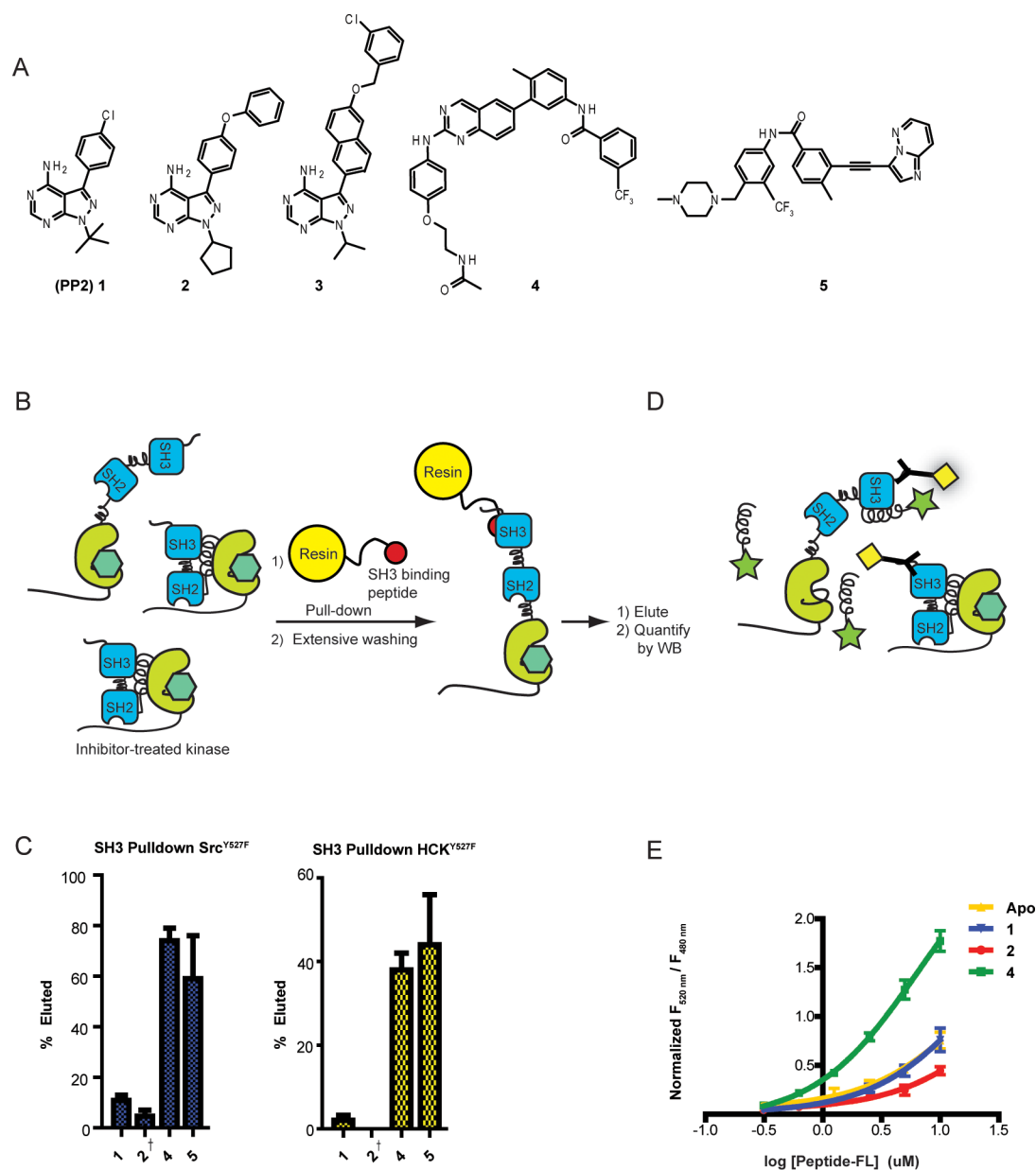
**Figure 1.** Regulation of SFK catalytic activity and ATP-binding site conformational accessibility. (A) SFK activity is allosterically modulated by engagement of the SH2 and SH3 regulatory domains (PDB: 2SRC). Release of these interactions through dephosphorylation of pTyr527 or intermolecular displacement of the regulatory domains leads to increased catalytic activity (PDB: 1Y57). Phosphorylation of Tyr416 on the activation loop fully activates the catalytic domain. (B) ATP-binding site conformations in which SFKs have been observed. Left: The active ATP-binding site conformation of SFKs, where all conserved catalytic residues are aligned for facilitating phosphate transfer. Center: An inactive SFK ATP-binding site conformation, where the  $\alpha$ C helix is rotated out of the active site, displacing a catalytically important glutamic acid. This inactive conformation is often referred to as the  $\alpha$ C helix-out or Src/CDK-like inactive conformation. Right: The DFG-out inactive conformation, where flipping of a conserved tripeptide motif (DFG motif) at the base of the activation loop results in the displacement of an aspartate residue that is important for catalysis.

While the allosteric coupling between the regulatory and catalytic domains of SFKs has been extensively investigated, there are many aspects of their inter-relationship that are still not well understood. For example, the ATP-binding sites of Src and Hck constructs with fully engaged regulatory domains are in the  $\alpha$ C helix-out inactive conformation in crystal structures,<sup>14,20</sup> but it is unclear whether alternative inactive forms, such as DFG-out, are also accessible to autoinhibited SFKs. Furthermore, the full complement of regulatory interactions that are controlled by the conformation of SFK ATP-binding sites and the effects on SFK post-translational modification are still not known. Here, we biochemically profile how stabilizing the ATP-binding sites of the SFKs Src and Hck in two distinct inactive ATP-binding site conformations ( $\alpha$ C helix-out and DFG-out) affect their regulatory interactions and their ability to undergo post-translational modification. Surprisingly, these studies show that different inactive ATP-binding site conformations lead to divergent SH2 and SH3 regulatory domain behaviors. Furthermore, we find that different classes of conformation-selective, ATP-competitive inhibitors also divergently modulate the ability of kinases and

phosphatases to act upon inhibitor-bound SFKs as substrates. These new results give mechanistic insights into the regulation of SFKs and suggest strategies for the design of ATP-competitive inhibitors that are able to modulate protein-protein interactions and the post-translational modification of multidomain tyrosine kinases.

## RESULTS AND DISCUSSION

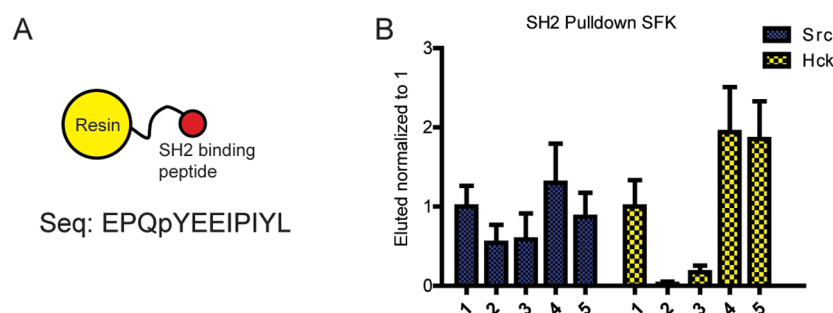
**The SH3 Domains of Src and Hck Are Divergently Affected by Ligands That Stabilize Different Inactive ATP-Binding Site Conformations.** To fully characterize how distinct inactive ATP-binding site conformations affect the regulatory domains of SFKs, we assembled a panel of conformation-selective inhibitors that stabilize both the  $\alpha$ C helix-out and DFG-out forms (Figure 2A). Inhibitors 2 and 3 contain extended hydrophobic substituents at the C-3 positions of their pyrazolopyrimidine scaffolds that are incompatible with the active conformation of helix  $\alpha$ C. Both inhibitors have been shown to stabilize the  $\alpha$ C helix-out inactive conformation of tyrosine kinases in crystal structures.<sup>21,22</sup> 4 and 5 are ligands that stabilize the DFG-out inactive conformation of protein



**Figure 2.** Conformation-selective inhibitors divergently modulate the SH3 domain accessibility of Src and Hck. (A) Panel of inhibitors that were used in this study. 2 and 3 stabilize the  $\alpha$ C helix-out inactive conformation. 4 and 5 stabilize the DFG-out inactive conformation. (B) SH3 domain accessibility pull-down assay. Src or Hck was incubated with an immobilized SH3 domain ligand in the presence of a saturating amount of an inhibitor (1, 2, 4, or 5). After incubation, beads were washed, and retained SFKs were eluted with SDS. Retained SFKs were quantified by immunoblotting. (C) Quantification of the pull-down experiments performed with Src<sup>Y527F</sup> and Hck<sup>Y527F</sup> in the presence of 1, 2, 4, or 5 (mean  $\pm$  SEM,  $n = 3$ ). † = Previously reported and shown for comparison.<sup>21</sup> (D) FRET assay to measure intermolecular SH3 domain accessibility. Hck was incubated with conformation-selective inhibitors in the presence of variable concentrations of a fluorophore-labeled SH3 peptide ligand (Peptide-FL). SH3 domain accessibility was determined by measuring FRET between the donor His-Tb, which is bound to the N-terminal His6 tag of Hck, and Peptide-FL. (E) Binding of fluorophore-labeled SH3 peptide ligand (Peptide-FL) in the presence of conformation-selective inhibitors (mean  $\pm$  SEM,  $n = 3$ ).

kinases.<sup>23</sup> Both ligands contain a 3-trifluoromethylbenzamide group that can be accommodated only in the ATP-binding sites of kinases with “flipped” DFG motifs. Furthermore, both inhibitors contain an amide linker between the substituent that occupies the adenine pocket and the 3-trifluoromethylbenzamide group, which provides a hydrogen bond donor/acceptor interaction that is observed in almost all ligands that stabilize the DFG-out form. Alternative pyrazolopyrimidine-based inhibitors that stabilize the DFG-out conformation, but contain a urea linker, have been reported. However, crystal structures

have shown that the ATP-binding site of Src can adopt either the DFG-out (PDB: 3EL8) or  $\alpha$ C helix-out (PDB: 3QLF) conformation when bound to these ligands, most likely due to the conformational flexibility of the urea linkage. The more conformationally rigid amide linkages of 4 and 5 are most likely compatible only with the DFG-out conformation. PP2 (1), which does not contain any functionalities that favor a particular ATP-binding site conformation, was used as a control for all studies.



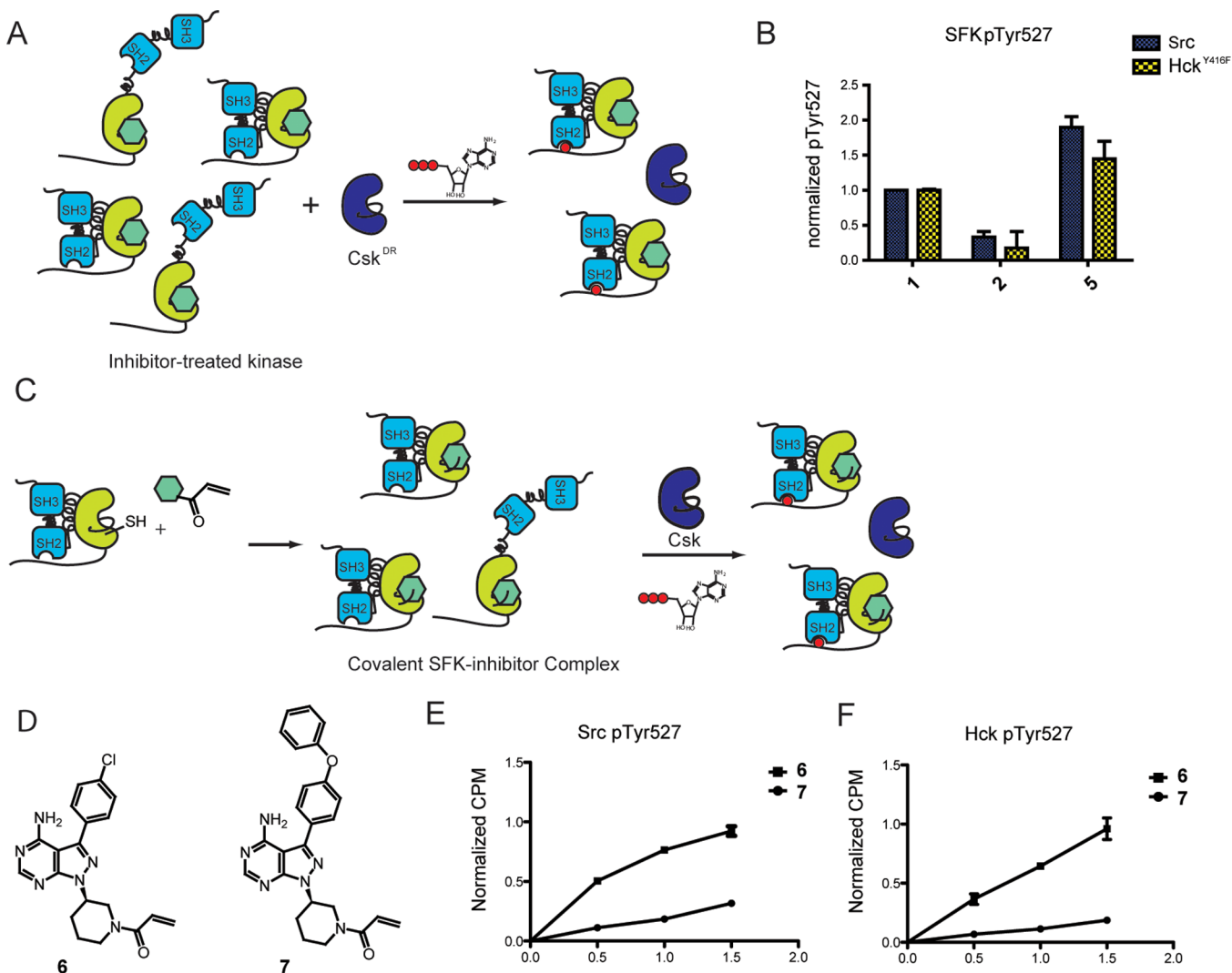
**Figure 3.** Conformation-selective inhibitors modulate SH2 domain accessibility. (A) SH2-binding peptide conjugated to resin. (B) Quantification of the SH2 pull-down assays performed with Src and Hck in the presence of various inhibitors (mean  $\pm$  SEM,  $n = 3$ ). The amount of kinase eluted from the beads is normalized to that of SFK-1.

The effect of stabilizing different inactive ATP-binding site conformations on SFK regulatory domain interactions was first probed using SH3 domain pull-down experiments (Figure 2B). In this assay, unphosphorylated SFK constructs in the presence of a saturating amount of an ATP-competitive inhibitor are incubated with resin containing an immobilized SH3 peptide ligand. The beads are then washed before eluting bound kinase with detergent, and the amount of retained SFK is determined using western blot analysis (Figure 2B). In the presence of control compound 1, 11% of total Src and 2% of total Hck were retained on beads (Figure 2C). Because we have previously demonstrated that stabilizing the ATP-binding sites of Src and Hck in the  $\alpha$ C helix-out inactive conformation with ligands 2 and 3 blocks the ability of their SH3 domains to engage in intermolecular interactions (Figure 2C), we were curious as to whether ligands 4 and 5, which stabilize the DFG-out conformation, would lead to the same effect. Surprisingly and in stark contrast to  $\alpha$ C helix-out stabilizing inhibitors, 4 and 5 lead to a dramatic increase in SH3 domain accessibility for both Src and Hck (Figure 2C), with 74% of Src-4 and 38% of Hck-4 retained by the immobilized SH3 domain ligand. A similar increase in intermolecular SH3 domain engagement is observed when Src and Hck are bound to 5. Because ligands that stabilize the  $\alpha$ C helix-out conformation lead to reduced SH3 domain accessibility by strengthening the intramolecular interaction between the SH3 domain and the SH2 linker, it is likely that the increased accessibility observed in the presence of DFG-out stabilizing inhibitors is due to destabilization of this same interaction. Thus, ligands 4 and 5 stabilize an inactive ATP-binding site conformation while stabilizing an active, extended global kinase conformation. Although this unexpected observation is novel for the SFKs, it has recently been observed that the DFG-out inhibitor, imatinib, stabilizes an open conformation of the kinase Abl, indicating that this phenomenon is not unique to SFKs and that it, in fact, may be general to many multidomain kinases.<sup>24</sup>

Next, a fluorescence resonance energy transfer (FRET)-based assay was employed to further characterize how conformation-selective inhibitors affect SH3 domain accessibility. To determine the ability of Hck's SH3 domain to intermolecularly engage a ligand, FRET between a terbium-conjugated  $\alpha$ -His6 antibody (Tb-His), which binds to the N-terminal His6 tag of Hck, and an Oregon Green-derivatized SH3 peptide ligand (Peptide-FL) was measured (Figure 2D). Relative differences in SH3 domain accessibility were determined by measuring FRET between Tb-His and variable concentrations of Peptide-FL in the presence of a saturating amount of conformation-selective inhibitors. Consistent with

the lack of ATP-binding site conformational preference of ligand 1, similar FRET was observed for the Hck-1 complex and *apo* Hck at all Peptide-FL concentrations tested (Figure 2E). Similar to the pull-down assays, the SH3 domain of Hck is almost completely inaccessible when this kinase is bound to  $\alpha$ C helix-out stabilizing inhibitor 2. However, ligand 4, which stabilizes the DFG-out inactive conformation, markedly increased the ability of Hck to bind Peptide-FL. Thus, the specific inactive ATP-binding site conformation that is stabilized, DFG-out or  $\alpha$ C helix-out, leads to a dramatic difference in the SH3 domain accessibility of Hck.

**Conformation-Selective Ligands Divergently Modulate SFK SH2 Domain Accessibility.** The SH3 and SH2 regulatory domains of SFKs are believed to act in a "snap-lock" fashion, where intramolecular binding of one domain causes concerted engagement of the second domain.<sup>25–27</sup> Therefore, increased intramolecular interactions between the SH3 domain and SH2 linker of SFKs should likely result in increased engagement of their SH2 domain with the C-terminal regulatory tail, reducing the capacity of the SH2 domain to partake in intermolecular binding events. To determine whether ATP-competitive ligands that decrease SH3 domain accessibility also reduce the ability of SFK SH2 domains to engage in intermolecular interactions, a series of SH2 pull down experiments was performed with an immobilized SH2 domain phospho-peptide ligand in the presence of saturating amounts of inhibitors (Figure 3A). Src-1 and Hck-1 complexes were used as references in the pull-down assay. A notable decrease in Hck SH2 domain accessibility was observed in the presence of inhibitors 2 or 3. Although the observed effect for inhibitor-bound Src was relatively small, 2% and 17% of Hck-2 and Hck-3 were retained by the immobilized SH2 ligand relative to that of Hck-1, respectively (Figure 3B). No change in retention was observed when Src was bound to DFG-out stabilizing ligands 4 or 5. However, a small increase in SH2 domain accessibility was observed for Hck-4 and Hck-5 relative to that of Hck-1. Therefore, stabilization of an  $\alpha$ C helix-out ATP-binding site conformation also modulates the SH2 regulatory domain of SFKs. However, the magnitude of this effect is different for Src and Hck. The binding affinities of the SH2 domains of Src and Hck for the pYEEI phosphopeptide have been shown to be very similar.<sup>28</sup> Therefore, a combination of differences in the SH2-linker sequence, residues linking the SH2 and SH3 domains, and residues comprising the C-terminal tail may account for these variations in coupled regulatory domain engagement.<sup>29–32</sup> Further study is needed to dissect the exact sequence determinants of these regulatory differences.

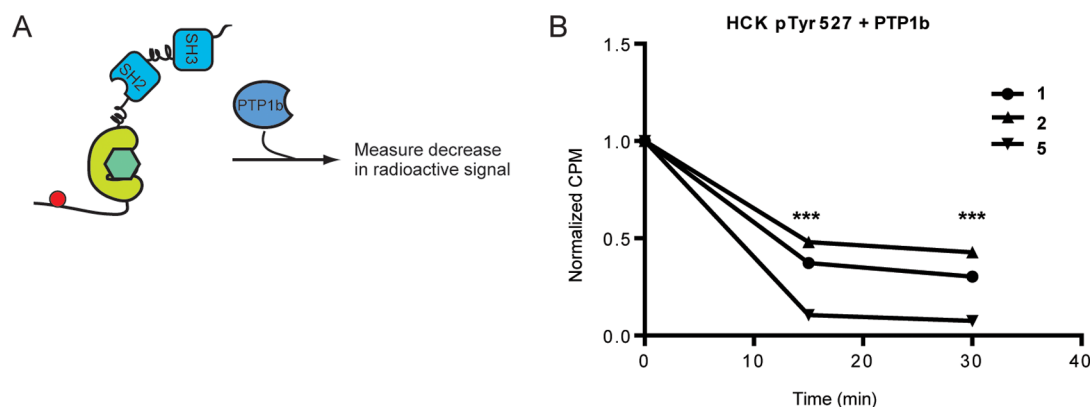


**Figure 4.** SFK inhibitors can modulate C-terminal tail phosphorylation. (A) Experimental schematic of SFK Tyr527 phosphorylation. Src or Hck<sup>Y416F</sup> was incubated with Csk<sup>DR</sup> in the presence of inhibitor. The concentration of each inhibitor is such that >85% of Src or Hck<sup>Y416F</sup> is bound to inhibitor and Csk<sup>DR</sup> is inhibited by <13%. ATP was added to the mixture to initiate Csk<sup>DR</sup>-mediated phosphorylation of the SFK–inhibitor complex. After 1 h of incubation, pTyr527 levels were determined by performing SDS-PAGE, followed by immunoblotting with an  $\alpha$ -phospho-527 Src antibody for Src or  $\alpha$ -pTyr antibody for Hck<sup>Y416F</sup>. All values are normalized to that of the SFK–1 complex. (B) Quantification of Src and Hck<sup>Y416F</sup> pTyr527 levels in the presence of inhibitors 1, 2, or 5. All values are normalized to that of the SFK–1 complex (mean  $\pm$  SEM,  $n = 3$ ). (C) Chemical genetic method for generating covalent SFK–inhibitor complexes to measure SFK Tyr527 phosphorylation. Covalent SFK–ligand complexes were generated by incubating SFK<sup>S245C</sup> with ligand 6 or 7. The resulting SFK–ligand substrate was purified and then incubated with Csk and  $\gamma$ -<sup>32</sup>P-ATP. Phosphorylation levels were measured at 0.5, 1, and 1.5 h. (D) Structures of electrophile-containing analogues. (E) Quantification of Src<sup>S245C</sup> pTyr527 levels in the presence of inhibitors 6 or 7 (mean  $\pm$  SEM,  $n = 3$ ). (F) Quantification of Hck<sup>S245C</sup> pTyr527 levels in the presence of inhibitors 6 or 7 (mean  $\pm$  SEM,  $n = 3$ ).

**Modulation of SFK Phosphorylation by ATP-Binding Site Occupancy.** To the best of our knowledge, the effects that ATP-binding site occupation has on SFK regulatory post-translational processing have not been determined. Given that ATP-competitive inhibitors can differentially modulate the accessibility of SFK regulatory domains, we were interested to determine whether ATP-binding site occupancy can also influence the activating and deactivating regulatory phosphorylation of these enzymes. It has recently been reported that the binding of ATP-competitive inhibitors can increase the level of activation loop phosphorylation of some kinases, such as Akt, RAF, and Jak2.<sup>33–36</sup> It has also been reported that when mitogen-activated protein kinases (MAPKs) are bound to inhibitors that stabilize the DFG-out inactive conformation their activation loops cannot be phosphorylated by upstream

activators.<sup>37,38</sup> Interestingly, this inhibitory effect is observed only when MAPKs are in the DFG-out inactive conformation. To determine whether conformation-selective inhibitors can likewise differentially influence the autoinhibitory phosphorylation site (Tyr527) of SFKs in a similar manner, a series of phosphorylation assays was performed with Src and Hck.

Although SFK C-terminal tail phosphorylation (Tyr527) can occur through *trans*-autophosphorylation, the major pathway for this post-translational modification is through the action of C-terminal Src kinase (Csk).<sup>39,40</sup> Csk is a highly specific tyrosine kinase known to selectively phosphorylate a single tail tyrosine in SFKs.<sup>41,42</sup> This phosphorylation event leads to increased intramolecular engagement between the C-terminal tail and the SH2 domain, resulting in SFK autoinhibition. In order to investigate the effects of ATP-binding site occupancy



**Figure 5.** ATP-competitive inhibitors regulate the dephosphorylation of pTyr527. (A) Experimental schematic of SFK dephosphorylation. Hck was radiolabeled by Csk phosphorylation ( $^{32}\text{P}$ pTyr527) with  $\gamma^{32}\text{P}$ -ATP. Phosphorylation was then blocked by the addition of inhibitor. PTP1b was then added to the  $^{32}\text{P}$ -labeled SFK-inhibitor complex. Loss of radioactive signal was measured over time. (B)  $^{32}\text{P}$ -labeled pTyr527 Hck $^{\text{Y416F}}$  was incubated with PTP1b in the presence of 1, 2, or 5, and the decrease in phosphorylated Hck was measured at 15 and 30 min (mean  $\pm$  SEM,  $n = 3$ ). \*\*\*  $p < 0.001$ .

on this inhibitory modification, an assay that allows uninhibited Csk to act upon an inhibitor-bound SFK as a substrate was needed. Because of the similarities of their ATP-binding sites, many SFK inhibitors also potently inhibit Csk as well.<sup>1</sup> Therefore, we developed a chemical genetic method that uses a drug-resistant Csk construct (Csk<sup>DR</sup>). Csk<sup>DR</sup> contains a T266 M gatekeeper mutation that provides resistance to many ATP-competitive inhibitors (Supporting Information Table S1). Src and Hck $^{\text{Y416F}}$ , which lacks the activation loop phosphorylation site, were used as inhibitor-bound substrates of Csk<sup>DR</sup> (Figure 4A).

Src or Hck $^{\text{Y416F}}$  was incubated with Csk<sup>DR</sup> and ATP in the presence of a concentration of inhibitor that leads to minimal inhibition of Csk<sup>DR</sup> (<13%) and >85% inhibitor-bound Src or Hck $^{\text{Y416F}}$  (see Supporting Information Table S2). Control activity assays were performed with a peptide substrate to verify that Csk<sup>DR</sup> activity was the same at all inhibitor concentrations used in the phospho-Tyr527 assays (data not shown).<sup>42,43</sup> Compound 1 was again used as a reference compound for the level of phosphorylation that is observed when an SFK construct is bound to an inhibitor that has minimal ATP-binding site conformational preference. The C-terminal tails of Src-1 and Hck $^{\text{Y416F}}$ -1 were phosphorylated by Csk<sup>DR</sup> 3- and 5.5-fold more than that of Src-2 and Hck-2, respectively. Therefore, the C-terminal tails of SFKs are less accessible to Csk phosphorylation when their ATP-binding sites are occupied by inhibitors stabilizing the  $\alpha\text{C}$  helix-out inactive conformation (Figure 4B). This correlates very well with our earlier observation that ATP-binding site occupation by this class of inhibitors results in decreased intermolecular accessibility of the SH2 domain, presumably through engagement of the C-terminal tail. Conversely, when Src or Hck $^{\text{Y416F}}$  was bound to ligands that stabilize the DFG-out conformation, an increase in C-terminal tail phosphorylation occurred. Src-5 and Hck-5 were phosphorylated 1.9- and 1.4-fold more than that of the SFK-1 substrate. Again, these results are consistent with the SH2 pull-down assay results shown in Figure 3, where ligands that stabilize the DFG-out conformation lead to an enhancement in the accessibility of SH2 domains to intermolecular binding events.

To provide a more quantitative comparison of how ATP-binding site occupancy affects C-terminal tail phosphorylation, we developed another chemical genetic method that allows

formation of a covalent inhibitor-SFK complex to be used as a substrate for Csk. In this system, >99% of each SFK is bound to inhibitor without any resultant partial inhibition of Csk. To facilitate the formation of covalent inhibitor-SFK complexes, a cysteine was engineered into the active site of Src and Hck (S245C) at a position that is rare among nonreceptor kinases (Figure 4C; found only in the TEC family kinases BTK and ITK).<sup>44</sup> Introduction of this mutation sensitizes the SFK constructs to pyrazolopyrimidine inhibitors containing an acryloyl functional group projecting from the N-1 position of the pyrazolopyrimidine ring (Figure 4D). Indeed,  $\alpha\text{C}$  helix-in ligand 6 and  $\alpha\text{C}$  helix-out ligand 7 inhibit SFK<sup>S245C</sup> constructs more potently than could be measured by activity assays, whereas  $K_i$  values measured for wild-type SFKs were at least 50-fold higher (Supporting Information Table S3).

Src- and Hck-ligand substrate complexes were generated by incubation of SFK<sup>S245C</sup> with 6 or 7. Excess small molecule was removed through Ni-affinity resin binding followed by desalting. Finally, unlabeled SFK was removed via incubation with resin displaying a potent kinase inhibitor, dasatinib, providing a single population of covalently labeled SFK. Activity assays were conducted to verify that all kinase activity was blocked (Supporting Information Table S4). We then verified that covalent modification of SFKs with 6 or 7 stabilized an  $\alpha\text{C}$  helix-in or -out ATP-binding site conformation, respectively, by conducting an SH3 pull-down experiment. Indeed, similar to our studies with noncovalent analogues, the SH3 domain of the Src<sup>S245C</sup>-6 complex was 3-fold more accessible than when Src<sup>S245C</sup> was covalently modified by 7 (Supporting Information Figure S1). Next, we measured the ability of covalent SFK-ligand complexes to be phosphorylated by Csk. The covalent inhibitor-SFK complex substrate incubated with Csk and  $\gamma^{32}\text{P}$ -ATP, and pTyr527 levels were determined at 0.5, 1, and 1.5 h (Figure 4C). No phosphorylation was observed when Csk or SFK substrate was absent from the reaction. Indeed, similar to the previous experiment, at 1.5 h the Src-6 complex was phosphorylated 3-fold more than the Src-7 substrate (Figure 4E). Likewise, Hck-6 was phosphorylated by Csk 5-fold more than Hck-7 (Figure 4E). Unfortunately a covalent ligand to stabilize a DFG-out conformation has not yet been developed, but these results corroborate our findings that stabilizing the ATP-

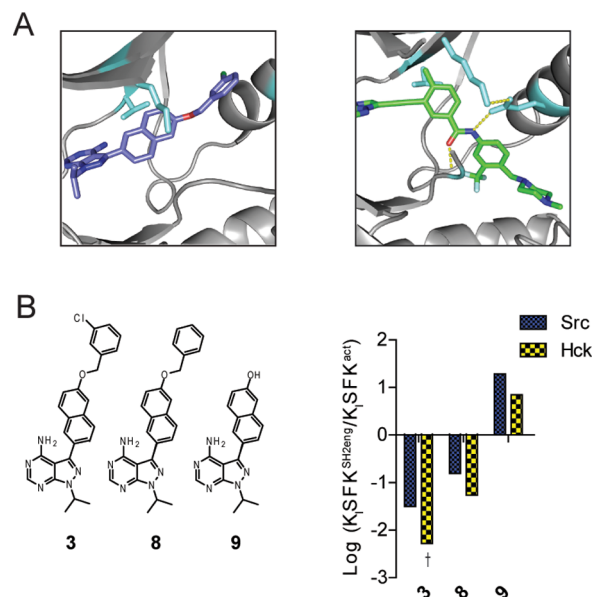
binding site in an  $\alpha$ C helix-out inactive conformation shields the C-terminal tail of SFKs from Csk phosphorylation.

**Modulation of SFK Dephosphorylation by ATP-Binding Site Occupancy.** SFK activity is regulated through both phosphorylation and dephosphorylation. Given that ATP-competitive inhibitors can differentially modulate the phosphorylation of SFKs on Tyr527, we were interested to determine whether ATP-binding site occupancy can also influence dephosphorylation of this site. Intracellular SFK dephosphorylation is mediated by several protein tyrosine phosphatases.<sup>45</sup> In order to probe the effects of stabilizing the ATP-binding sites of Src and Hck in the DFG-out or  $\alpha$ C helix-out conformations on the modification of pTyr527, an assay that looks at the dephosphorylation of inhibitor-bound SFK complexes by tyrosine phosphatase PTP1b was developed. This phosphatase has been shown to regulate SFK phosphorylation in cells.<sup>46</sup> In order to monitor dephosphorylation, SFKs were first radiolabeled at Tyr527 using  $\gamma$ -<sup>32</sup>P-ATP. Following labeling, kinase activity was halted by the addition of inhibitors at a concentration that results in >98% SFK–inhibitor complex. No increase in phosphorylation was observed after inhibitor addition. PTP1b was added to the preformed SFK–inhibitor complex, and dephosphorylation was monitored by loss of <sup>32</sup>P-labeled SFK over time (Figure 5A).

To investigate the dephosphorylation of pTyr527, <sup>32</sup>P-labeled Hck<sup>Y416F</sup> was generated by incubating Hck<sup>Y416F</sup> with Csk and  $\gamma$ -<sup>32</sup>P-ATP. Using a construct that lacks Tyr416 ensured that all radioactive signal was due to Csk phosphorylation of Tyr527 on Hck. Phosphorylation was stopped, and SFK–inhibitor complexes were formed by the addition of 1, 2, or 5. PTP1b was then added, and loss of radioactive signal was monitored over time (Figure 5B). Compound 1 was used as a reference compound for the level of dephosphorylation that is observed when an Hck<sup>Y416F</sup> construct is bound to an inhibitor that has minimal ATP-binding site conformational preference. Binding of inhibitor 2 shielded the C-terminal tail of the Hck<sup>Y416F</sup>–2 complex from dephosphorylation relative to that of 1. Shielding of pTyr527 from dephosphorylation in the Hck–2 complex agrees with the decreased rate of Tyr527 phosphorylation by Csk in this same complex (Figure 4B,E). 2's occupation of the ATP-binding site of Hck strengthens the intramolecular interaction between the SH2 domain and the C-terminal tail, blocking not only phosphorylation but also dephosphorylation. In contrast, Hck<sup>Y416F</sup>–5 is dephosphorylated much more rapidly than the Hck<sup>Y416F</sup>–1 complex. Thus, an inhibitor that stabilizes the DFG-out inactive conformation greatly increases the accessibility of pTyr527 to phosphatases. These results further highlight how different inactive ATP-binding site conformations can lead to divergent regulatory domain behaviors.

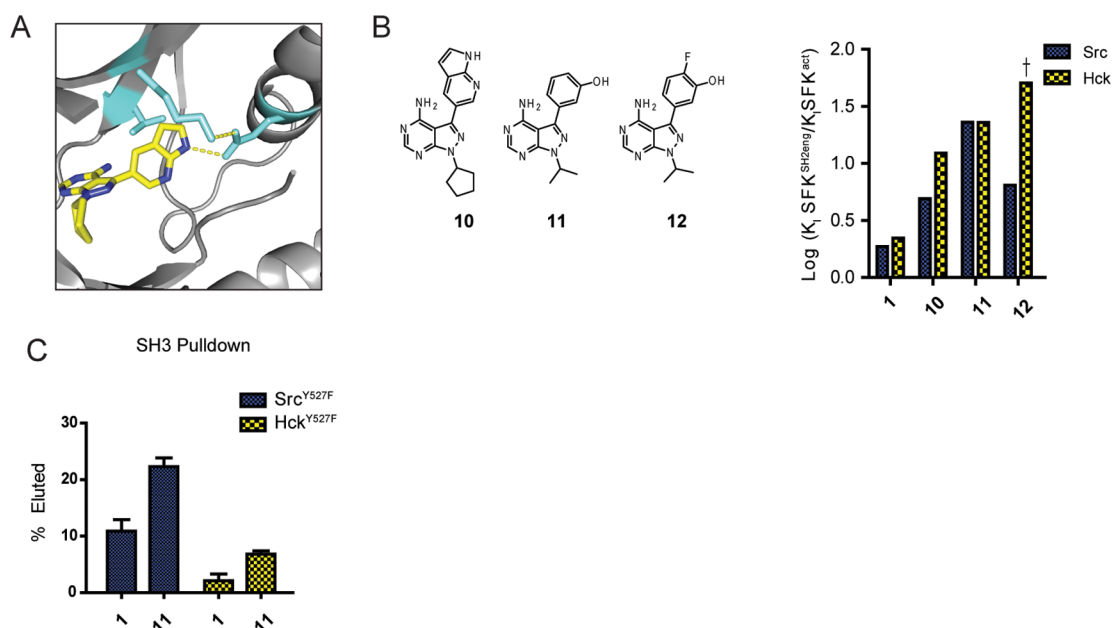
**Molecular Determinants of ATP-Binding Site Conformation Selectivity.** Stabilizing the ATP-binding sites of SFKs in an  $\alpha$ C helix-out inactive conformation induces a closed global conformation, which results in decreased accessibility to intermolecular binding events and reduced susceptibility to post-translational modifications. However, inhibitors that stabilize an alternative inactive ATP-binding site alignment, the DFG-out conformation, result in an extended and open global kinase conformation that is reminiscent of fully activated SFKs. In this conformation, SFKs are free to engage intermolecular binding partners and are highly susceptible to phosphorylating and dephosphorylating enzymes. Molecular dynamics studies have suggested that the DFG-out inactive

conformation plays a role in the catalytic cycle of tyrosine kinases.<sup>47</sup> Therefore, it is not surprising that DFG-out stabilizing ligands do not act entirely like inhibitors that are selective for the  $\alpha$ C helix-out form. However, we were quite surprised that these two inactive forms have opposite effects on the regulatory interactions of SFKs. Inhibitors that stabilize the DFG-out and  $\alpha$ C helix-out conformations make a number of different interactions with the ATP-binding sites of protein kinases that may account for their differing behaviors (Figure 6A). Therefore, we undertook a systematic analysis of each inhibitor class in order to understand the source of this divergence.



**Figure 6.** Binding preferences of conformation-selective, ATP-competitive inhibitors. (A) Ligands that stabilize the DFG-out and  $\alpha$ C helix-out conformations make a number of different interactions with the ATP-binding sites of kinases. Left: Src bound to 3 with the ATP-binding site in an inactive,  $\alpha$ C helix-out conformation (PDB: 4DGG). The  $\alpha$ C helix is rotated out of the active site, disrupting the interaction between Lys295 and Glu310. Right: Abl bound to 5 with the active site in an inactive, DFG-out conformation (PDB: 3OXZ). The DFG motif is flipped out of the active site, but Glu310 in the  $\alpha$ C helix maintains a salt bridge with the catalytic lysine (the distance between the NH of 5 and the Glu310 side chain is 3.0 Å). The gatekeeper (Thr338), catalytic lysine (Lys295), and catalytic glutamic acid (Glu310) residues are shown in light blue (Src numbering). (B) Analogues of 3 that were tested in activity assays against Src<sup>act</sup>, Hck<sup>act</sup>, Src<sup>SH2eng</sup>, and Hck<sup>SH2eng</sup> constructs. Quantitative comparison of the fold differences in  $K_i$  values between activated SFKs ( $K_i^{act}$ ) and their respective autoinhibited constructs ( $K_i^{SH2eng}$ ) allow systematic analysis of ATP-binding site conformational preference. Raw data for all assays are shown in Supporting Information Table S5.

As a preliminary screen of these interactions, the bidirectional nature of the interplay between the ATP-binding sites and regulatory domains of SFKs was exploited.<sup>21</sup> Just as ATP-binding site conformation can affect regulatory domain engagement, conversely, the engagement of regulatory domains can influence ATP-binding site conformation. Therefore, the relative affinities of inhibitors for SFKs with different regulatory domain interactions is a reflection of the importance of active site features and inhibitor substituents toward specific ATP-binding site interactions. Autoinhibited SFK constructs



**Figure 7.** Biochemical profiling of ligands predicted to stabilize an active,  $\alpha$ C helix-in ATP-binding site conformation. (A) Src bound to **8**, with the ATP-binding site in an active conformation (PDB: 3EN4). The hydrogen bond between **8** and Glu310 is shown (the distance between NH and the Glu310 side chain is 2.7 Å). The gatekeeper threonine (Thr338), catalytic lysine (Lys295), and catalytic glutamic acid (Glu310) residues are shown in light blue (Src numbering). (B) Chemical structures of the ATP-competitive SFK inhibitors that can potentially hydrogen bond with the  $\alpha$ C helix. Quantitative comparison of the fold differences in  $K_i$  values between an activated SFK (Src<sup>act</sup> and Hck<sup>act</sup>) and their respective SH2 domain-engaged construct (Src<sup>SH2eng</sup> and Hck<sup>SH2eng</sup>). † = The absolute fold difference in  $K_i$  value could not be determined because inhibitor affinity is less than the enzyme concentration used in the assay. (C) Quantification of the SH3 pull-down assays performed with Src and Hck in the presence of various inhibitors (mean  $\pm$  SEM,  $n = 3$ ).

(SFK<sup>SH2eng</sup>) that stabilize an  $\alpha$ C helix-out ATP-binding site were generated by introducing mutations that strengthen intramolecular regulatory interactions between the C-terminal tail and the SH2 domain.<sup>48–51</sup> Fully activated constructs (SFK<sup>act</sup>) that stabilize an  $\alpha$ C helix-in ATP-binding site were generated via autophosphorylation of SFK<sup>Y527F</sup> on Tyr416.<sup>15,52</sup> As predicted, Src<sup>act</sup> and Hck<sup>act</sup> are substantially more catalytically active than their autoinhibited counterparts, Src<sup>SH2eng</sup> and Hck<sup>SH2eng</sup> (Supporting Information Figure S2). The  $K_i$  values for various ligands against the SFK<sup>SH2eng</sup> and SFK<sup>act</sup> constructs were determined, and the  $K_i$  ratios were analyzed.

To determine the importance of specific interactions toward stabilizing the  $\alpha$ C helix-out conformation and inducing a closed SFK regulatory state, analogues of inhibitor **3** were tested (Figure 6B). Consistent with its effects on global kinase conformation, **3** is highly selective for autoinhibited SFKs over their fully activated forms. Ligand **8**, which also contains a 6-benzyloxy-2-naphthyl substituent but is missing the 3-chloro group of **3**, is also selective for autoinhibited Src and Hck, although to a lesser degree. However, an analogue of **3** that lacks a hydrophobic substituent at the 6-position of the naphthyl ring, ligand **9**, is selective for activated SFKs. Thus, the presence of a hydrophobic substituent to fill the pocket created by movement of the  $\alpha$ C helix is necessary to stabilize this inactive conformation.

Ligands **4** and **5** are only sterically compatible with the ATP-binding sites of protein kinases that possess a flipped DFG motif (the DFG-out conformation). Unlike the  $\alpha$ C helix-out conformation, the alignment of the  $\alpha$ C helix in the DFG-out conformation is minimally perturbed from an active alignment. In fact, almost all ligands that stabilize the DFG-out ATP-binding site alignment in SFKs form a hydrogen bond with Glu310 in the  $\alpha$ C helix that is possible only when this structural

motif is in an active conformation (Figure 6A, right). Therefore, we predicted that the unexpected abilities of inhibitors that stabilize the DFG-out inactive conformation to induce an active global SFK conformation may be due to the interactions that these ligands make with the  $\alpha$ C helix. Unfortunately, because of their highly potent inhibition of Src and Hck, we were unable to determine the selectivity of inhibitor **4** or **5** toward activated or autoinhibited SFK constructs. However, on the basis of their propensity to induce an open and active SFK global conformation, these inhibitors are most likely selective for activated SFKs over their autoinhibited forms.

Although not a DFG-out stabilizing ligand, as noted above, **9** is selective for activated SFKs. We hypothesized that this selectivity is likely due to hydrogen bonding with the  $\alpha$ C helix. The 6-hydroxyl group of the naphthyl substituent should be able to hydrogen bond with Glu310 in an  $\alpha$ C helix-in conformation, which is reflected by its pronounced selectivity toward activated SFKs. In fact, a pyrazolopyrimidine-based ligand, **10**, whose azaindole C-3 substituent contains a hydrogen-bond donor, has been crystallized bound to Src, revealing an active ATP-binding site conformation with hydrogen bonding to Glu310 of the  $\alpha$ C helix (Figure 7A).

To further probe the importance of hydrogen bonding with Glu310 in the  $\alpha$ C helix, we assembled a small panel of pyrazolopyrimidine-based ligands that contain substituents at the C-3 position that should be capable of hydrogen bonding (Figure 7A,B). Because of the small size of their C-3 substituents, these ligands cannot stabilize the  $\alpha$ C helix-out or DFG-out conformation but should interact with an  $\alpha$ C helix-in ATP-binding site conformation. Consistent with this notion, each of these ligands was also selective for activated SFKs over the autoinhibited constructs. Furthermore, ligand **1**, which is structurally very similar to **11** and **12** but lacks a hydrogen-



bond donor, shows little selectivity for either SFK construct, highlighting the importance of hydrogen bonding for activated SFK preference.

On the basis of the bidirectional relationship observed between the ATP-binding sites and regulatory domains of SFKs, inhibitors that are selective for activated SFK constructs should promote a more open global SFK conformation. Furthermore, if  $\alpha$ C helix alignment is the molecular switch regulating the divergent outcomes observed between the two inactive conformation-stabilizing ligand classes, then active conformation-selective,  $\alpha$ C helix-in ligands should behave similarly to that of DFG-out ligands. To test this, biochemical profiling of Src and Hck in complex with an active conformation-selective ligand was performed. Ligand **11** was chosen as a representative member of this class of inhibitors because of its structural similarity to **1**. In SH3 pull-down experiments, DFG-out inhibitors dramatically increased SH3 accessibility. For active-conformation selective inhibitors, SH3 domain accessibility was increased 2- and 3.2-fold for Src and Hck, respectively, relative to that of **1** (Figure 7C). SFKs bound to active-conformation selective inhibitors do, in fact, behave similarly to DFG-out inhibitors and not like  $\alpha$ C helix-out inhibitors. These data indicate that a ligand's ability to control the conformation of the  $\alpha$ C helix either through hydrogen bonding or steric perturbation is the major determinant of its influence on noncatalytic SFK regulation. This helix acts as a switch regulating both global kinase conformation and ability to undergo post-translational modification.

## CONCLUSIONS

In this study, we have determined that two distinct inactive ATP-binding site conformations can differentially modulate the engagement or disengagement of the SH2 and SH3 regulatory domains of SFKs through perturbation or stabilization of the  $\alpha$ C helix. Additionally, these conformations can regulate the accessibility of the C-terminal tail towards post-translational modification. Concisely, SFKs can be divergently modulated through the use of inhibitors that stabilize an  $\alpha$ C helix-in or -out conformation.

These results have direct implications on the pharmacological inhibition of multidomain kinases. Differential modulation of noncatalytic roles, post-translational modification, and catalytic activity through stabilization of discrete ATP-binding sites with particular inhibitor classes allows for greater control of aberrant signaling enzymes. Recent studies have shown that kinase inhibition in different ATP-binding site conformations can greatly alter downstream signaling.<sup>33–36,53,54</sup> Better understanding of how ATP-binding site stabilization affects intermolecular interactions is critical to predicting off-target effects of kinase inhibition. The potential to develop kinase inhibitors that selectively or cooperatively inactivate catalytic and noncatalytic functions should be of great use in both dissecting signaling pathways and in designing future therapeutic inhibitors of aberrant signaling networks.

## METHODS

**Protein Expression.** The Src-family kinases, Src (residues 84–533) and Hck (residues 84–531), were expressed with YopH and GroEL and purified as previously described.<sup>21,55</sup> Csk and Csk<sup>DR</sup> plasmids were transformed into BL21 (DE3) competent cells and purified by GST resin following the manufacturer's protocol.

**Synthetic Methods.** All compounds were synthesized as previously described.<sup>56–60</sup>

**Pull-Down Assay To Determine SH3 Domain Accessibility. Formation of the Kinase–Inhibitor Complex.** The kinase of interest (100 nM) and mammalian lysate (0.2 mg mL<sup>-1</sup>) were diluted in immobilization buffer (50 mM Tris, 100 mM NaCl, and 1 mM DTT, pH 7). The inhibitor of interest (5  $\mu$ M) was added to this kinase dilution. The mixture was allowed to incubate for 30 min before loading on the resin.

**Pull-Down.** Forty microliters of a 50% slurry of SNAP-Capture pull-down resin (NEB) was placed in a microcentrifuge tube. The resin was washed (2 $\times$ , 10 bed volumes) with immobilization buffer. A SNAP tag–polyproline peptide fusion (VSLARRPLPLP) (10  $\mu$ M) was loaded onto the resin at a final volume of 100  $\mu$ L in buffer. The resin was shaken at rt for 90 min with agitation by a pipet every 15 min. After polyproline peptide immobilization, the resin was washed (2 $\times$ , 10 bed volumes), and 100  $\mu$ L of the kinase–inhibitor complex was loaded. The resin was allowed to shake at rt for 1 h. After incubation with the kinase–inhibitor complex, the flow through was collected, and the resin was washed (4 $\times$ , 10 bed volumes). To elute the retained kinase, 100  $\mu$ L of 1 $\times$  SDS loading buffer was added, and the beads were boiled at 90 °C for 10 min. All samples were separated by SDS-PAGE and visualized by western blotting using a His6-specific antibody (at a 1:5000 dilution (abm, HIS.H8)). The scanned blots were quantified with LI-COR Odyssey software to determine the percentage of kinase retained on the resin on the basis of the loaded and eluted fractions (mean  $\pm$  SEM,  $n = 2$ ).

**Pull-Down Assay To Determine SH2 Domain Accessibility. Formation of the Kinase–Inhibitor Complex.** Kinase inhibitor complexes were prepared as described above.

**Pull-Down.** Pull-down experiments were performed as described above using a resin conjugated to an SH2 affinity peptide (sequence: EPQpYEEIPIYL).

**FRET Assay.** SFK (25 nM) was incubated with inhibitor (10  $\mu$ M) and titrated with Peptide-FL in buffer (final volume = 30  $\mu$ L, 50 mM HEPES, pH 7.5, 10 mM MgCl<sub>2</sub>, 2.5 mM EGTA, and 100 mM NaCl). After 1 h, His-Tb was added, and FRET signal was monitored on a plate reader.

**Phosphorylation of pTyr527 Assay with Csk<sup>DR</sup>.** Src or Hck<sup>Y416F</sup> (25 nM) was incubated with inhibitor in the presence of Csk<sup>DR</sup> (25 nM) with BSA (0.1 mg mL<sup>-1</sup>) in activation buffer. Following 0.5 h incubation, ATP (1  $\mu$ M) was added. After 1 h, the reaction was quenched by the addition of 3 $\times$  loading dye. All samples were separated by SDS-PAGE and visualized by western blotting using pTyr527 antibody for Src (1:2000 P-Src (Tyr527) Cell Signaling) or a P-tyrosine antibody for Hck (1:2000 (P Tyr-100) Cell Signaling) and a His6-specific antibody (at a 1:5000 dilution (abm, HIS.H8)). The scanned blots were quantified with LI-COR Odyssey software to determine the phosphorylation levels of the SFK pTyr527 treated with inhibitors relative to that of **1** (mean  $\pm$  SEM,  $n = 3$ ). For inhibitor concentrations, see Supporting Information Table S2.

**Preparation of Covalent SFK–Inhibitor Substrate Complex.** SFK (2  $\mu$ M) was incubated with inhibitor (100  $\mu$ M) in buffer (50 mM Tris, 100 mM NaCl, and 0.5 mM DTT, pH 7) overnight at 4 °C. SFK–inhibitor complexes were bound to Ni-NTA resin to wash away excess inhibitor (ThermoScientific). The kinase was then eluted and desalted using a Zeba spin column following the manufacturer's protocol. The kinase was then incubated with 10  $\mu$ L of dasatinib-conjugated resin for 1 h at rt to remove any unmodified kinase. The supernatant was then used in the Csk Tyr527 phosphorylation assay.

**Phosphorylation of pTyr527 Assay with Covalent SFK–Inhibitor Substrates.** Covalent SFK–inhibitor substrate (25 nM) was incubated with Csk (50 nM) and BSA (0.1 mg mL<sup>-1</sup>) in activation buffer. Following the addition of  $\gamma$ <sup>32</sup>P-ATP (0.2  $\mu$ Ci/well), reactions were incubated at rt for 0.5 h and then spotted on a nitrocellulose membrane. Membranes were washed with 0.5% phosphoric acid (3 $\times$ , 10 min each wash) and dried, and the radioactivity was determined by phosphorimaging with a GE Typhoon FLA 9000 phosphor scanner. The scanned membranes were quantified with ImageQuant. Data was plotted using GraphPad Prism software.

**Dephosphorylation of pTyr527 Assay.** Hck<sup>Y416F</sup> (250 nM) was phosphorylated at Tyr527 by Csk (100 nM) in activation buffer

containing BSA (0.25 mg mL<sup>-1</sup>) and radiolabeled ATP. After 1 h, inhibitor (5  $\mu$ M) was added, and the mixture was incubated for 30 min. Following incubation, 10  $\mu$ L of PTP1b (500 nM) and DTT (15 mM) was added to 20  $\mu$ L of the hot reaction mixture. Reactions were spotted at 0, 15, and 30 min onto a nitrocellulose membrane. Membranes were washed with 0.5% phosphoric acid (3 $\times$ , 10 min each wash) and dried, and the radioactivity was determined by phosphorimaging with a GE Typhoon FLA 9000 phosphor scanner. The scanned membranes were quantified with ImageQuant. Data was plotted using GraphPad Prism software.

**Activity Assay.** Inhibitors (initial concentration = 10  $\mu$ M, 3-fold serial dilutions, 10 data points) were assayed in triplicate against Src ([Src<sup>SH2eng</sup>] = 20 nM and [Src<sup>act</sup>] = 0.5 nM) or Hck ([Hck<sup>SH2eng</sup>] = 10 nM and [Hck<sup>act</sup>] = 0.5 nM) in assay buffer containing 75 mM HEPES, pH 7.5, 15 mM MgCl<sub>2</sub>, 3.75 mM EGTA, 150 mM NaCl, 0.2 mg mL<sup>-1</sup> BSA,  $\gamma$ -<sup>32</sup>P-ATP (0.2  $\mu$ Ci/well), and an optimized Src peptide substrate of the sequence Ac-EIYGFEKFKK-OH (final concentration = 100  $\mu$ M). The final volume of each assay well was 30  $\mu$ L. The enzymatic reaction was run at rt for 2 h and then terminated by spotting 4.6  $\mu$ L of the reaction mixture onto a phosphocellulose membrane. Membranes were washed with 0.5% phosphoric acid (3 $\times$ , 10 min each wash) and dried, and the radioactivity was determined by phosphorimaging with a GE Typhoon FLA 9000 phosphor scanner. The scanned membranes were quantified with ImageQuant and converted to percent inhibition. Data was analyzed using GraphPad Prism software, and IC<sub>50</sub> values were determined using nonlinear regression analysis.

## ■ ASSOCIATED CONTENT

### ● Supporting Information

Quantification of the pull-down experiments performed with Src<sup>S245C</sup> in the presence of 6 or 7; catalytic activity of SFK constructs; K<sub>i</sub> values for drug resistant constructs, covalent inhibitors, and SFKs; pY527 chemical genetic assay parameters; and activity of SFK treated with covalent inhibitors for the pY527 assay. This material is available free of charge via the Internet at <http://pubs.acs.org>.

## ■ AUTHOR INFORMATION

### Corresponding Author

\*E-mail: [maly@chem.washington.edu](mailto:maly@chem.washington.edu).

### Notes

The authors declare no competing financial interest.

## ■ ACKNOWLEDGMENTS

This work was supported by the NIH (R01GM086858 to D.J.M. and F32CA174346 to S.E.L.) and by the Alfred P. Sloan and Camille and Henry Dreyfus Foundations (to D.J.M.).

## ■ REFERENCES

- (1) Taylor, S. S., and Kornev, A. P. (2011) Protein kinases: evolution of dynamic regulatory proteins. *Trends Biochem. Sci.* 36, 65–77.
- (2) Hubbard, S. R., and Till, J. H. (2000) Protein tyrosine kinase structure and function. *Annu. Rev. Biochem.* 69, 373–398.
- (3) Bhattacharyya, R. P., Remenyi, A., Yeh, B. J., and Lim, W. A. (2006) Domains, motifs, and scaffolds: the role of modular interactions in the evolution and wiring of cell signaling circuits. *Annu. Rev. Biochem.* 75, 655–680.
- (4) Kuriyan, J., and Eisenberg, D. (2007) The origin of protein interactions and allostery in colocalization. *Nature* 450, 983–990.
- (5) Pellicena, P., and Kuriyan, J. (2006) Protein–protein interactions in the allosteric regulation of protein kinases. *Curr. Opin. Struct. Biol.* 16, 702–709.
- (6) Rauch, J., Volinsky, N., Romano, D., and Kolch, W. (2011) The secret life of kinases: functions beyond catalysis. *Cell Commun. Signaling* 9, 23.

- (7) Frame, M. C. (2002) Src in cancer: deregulation and consequences for cell behaviour. *Biochim. Biophys. Acta* 1602, 114–130.
- (8) Chong, Y. P., Ia, K. K., Mulhern, T. D., and Cheng, H. C. (2005) Endogenous and synthetic inhibitors of the Src-family protein tyrosine kinases. *Biochim. Biophys. Acta* 1754, 210–220.
- (9) Parsons, S. J., and Parsons, J. T. (2004) Src family kinases, key regulators of signal transduction. *Oncogene* 23, 7906–7909.
- (10) Summy, J. M., and Gallick, G. E. (2003) Src family kinases in tumor progression and metastasis. *Cancer Metastasis Rev.* 22, 337–358.
- (11) Sen, B., and Johnson, F. M. (2011) Regulation of SRC family kinases in human cancers. *J. Signal Transduction* 2011, 865819.
- (12) Guarino, M. (2010) Src signaling in cancer invasion. *J. Cell. Physiol.* 223, 14–26.
- (13) Jura, N., Zhang, X., Endres, N. F., Seeliger, M. A., Schindler, T., and Kuriyan, J. (2011) Catalytic control in the EGF receptor and its connection to general kinase regulatory mechanisms. *Mol. Cell* 42, 9–22.
- (14) Xu, W., Doshi, A., Lei, M., Eck, M. J., and Harrison, S. C. (1999) Crystal structures of c-Src reveal features of its autoinhibitory mechanism. *Mol. Cell* 3, 629–638.
- (15) Osusky, M., Taylor, S. J., and Shalloway, D. (1995) Autophosphorylation of purified c-Src at its primary negative regulation site. *J. Biol. Chem.* 270, 25729–25732.
- (16) Sicheri, F., Moarefi, I., and Kuriyan, J. (1997) Crystal structure of the Src family tyrosine kinase Hck. *Nature* 385, 602–609.
- (17) Johnson, L. N., Noble, M. E., and Owen, D. J. (1996) Active and inactive protein kinases: structural basis for regulation. *Cell* 85, 149–158.
- (18) Kornev, A. P., Taylor, S. S., and Ten Eyck, L. F. (2008) A helix scaffold for the assembly of active protein kinases. *Proc. Natl. Acad. Sci. U.S.A.* 105, 14377–14382.
- (19) Kornev, A. P., and Taylor, S. S. (2010) Defining the conserved internal architecture of a protein kinase. *Biochim. Biophys. Acta* 1804, 440–444.
- (20) Hennequin, L. F., Allen, J., Breed, J., Curwen, J., Fennell, M., Green, T. P., Lambert-van der Brempt, C., Morgentin, R., Norman, R. A., Olivier, A., Otterbein, L., Ple, P. A., Warin, N., and Costello, G. (2006) N-(5-Chloro-1,3-benzodioxol-4-yl)-7-[2-(4-methylpiperazin-1-yl)ethoxy]-5-(tetrahydro-2H-pyran-4-yloxy)quinazolin-4-amine, a novel, highly selective, orally available, dual-specific c-Src/Abl kinase inhibitor. *J. Med. Chem.* 49, 6465–6488.
- (21) Krishnamurthy, R., Brigham, J. L., Leonard, S. E., Ranjitkar, P., Larson, E. T., Dale, E. J., Merritt, E. A., and Maly, D. J. (2013) Active site profiling reveals coupling between domains in SRC-family kinases. *Nat. Chem. Biol.* 9, 43–50.
- (22) Marcotte, D. J., Liu, Y. T., Arduini, R. M., Hession, C. A., Miatkowski, K., Wildes, C. P., Cullen, P. F., Hong, V., Hopkins, B. T., Mertsching, E., Jenkins, T. J., Romanowski, M. J., Baker, D. P., and Silvian, L. F. (2010) Structures of human Bruton's tyrosine kinase in active and inactive conformations suggest a mechanism of activation for TEC family kinases. *Protein Sci.* 19, 429–439.
- (23) Zhou, T., Commodore, L., Huang, W. S., Wang, Y., Thomas, M., Keats, J., Xu, Q., Rivera, V. M., Shakespeare, W. C., Clackson, T., Dalgarno, D. C., and Zhu, X. (2011) Structural mechanism of the pan-BCR-ABL inhibitor ponatinib (AP24534): lessons for overcoming kinase inhibitor resistance. *Chem. Biol. Drug Des.* 77, 1–11.
- (24) Skora, L., Mestan, J., Fabbro, D., Jahnke, W., and Grzesiek, S. (2013) NMR reveals the allosteric opening and closing of Abelson tyrosine kinase by ATP-site and myristoyl pocket inhibitors. *Proc. Natl. Acad. Sci. U.S.A.* 110, E4437–E4445.
- (25) Young, M. A., Gonfloni, S., Superti-Furga, G., Roux, B., and Kuriyan, J. (2001) Dynamic coupling between the SH2 and SH3 domains of c-Src and Hck underlies their inactivation by C-terminal tyrosine phosphorylation. *Cell* 105, 115–126.
- (26) Ulmer, T. S., Werner, J. M., and Campbell, I. D. (2002) SH3–SH2 domain orientation in Src kinases: NMR studies of Fyn. *Structure* 10, 901–911.

- (27) Cowan-Jacob, S. W., Fendrich, G., Manley, P. W., Jahnke, W., Fabbro, D., Liebetanz, J., and Meyer, T. (2005) The crystal structure of a c-Src complex in an active conformation suggests possible steps in c-Src activation. *Structure* 13, 861–871.
- (28) Sicilia, R. J., Hibbs, M. L., Bello, P. A., Bjorge, J. D., Fujita, D. J., Stanley, I. J., Dunn, A. R., and Cheng, H. C. (1998) Common *in vitro* substrate specificity and differential Src homology 2 domain accessibility displayed by two members of the Src family of protein-tyrosine kinases, c-Src and Hck. *J. Biol. Chem.* 273, 16756–16763.
- (29) Gonfloni, S., Williams, J. C., Hattula, K., Weijland, A., Wierenga, R. K., and Superti-Furga, G. (1997) The role of the linker between the SH2 domain and catalytic domain in the regulation and function of Src. *EMBO J.* 16, 7261–7271.
- (30) Briggs, S. D., and Smithgall, T. E. (1999) SH2-kinase linker mutations release Hck tyrosine kinase and transforming activities in Rat-2 fibroblasts. *J. Biol. Chem.* 274, 26579–26583.
- (31) LaFevre-Bernt, M., Sicheri, F., Pico, A., Porter, M., Kuriyan, J., and Miller, W. T. (1998) Intramolecular regulatory interactions in the Src family kinase Hck probed by mutagenesis of a conserved tryptophan residue. *J. Biol. Chem.* 273, 32129–32134.
- (32) Williams, J. C., Weijland, A., Gonfloni, S., Thompson, A., Courtneidge, S. A., Superti-Furga, G., and Wierenga, R. K. (1997) The 2.35 Å crystal structure of the inactivated form of chicken Src: a dynamic molecule with multiple regulatory interactions. *J. Mol. Biol.* 274, 757–775.
- (33) Hatzivassiliou, G., Song, K., Yen, I., Brandhuber, B. J., Anderson, D. J., Alvarado, R., Ludlam, M. J., Stokoe, D., Gloor, S. L., Vigers, G., Morales, T., Aliagas, I., Liu, B., Sideris, S., Hoeflich, K. P., Jaiswal, B. S., Seshagiri, S., Koeppen, H., Belvin, M., Friedman, L. S., and Malek, S. (2010) RAF inhibitors prime wild-type RAF to activate the MAPK pathway and enhance growth. *Nature* 464, 431–435.
- (34) Poulikakos, P. I., Zhang, C., Bollag, G., Shokat, K. M., and Rosen, N. (2010) RAF inhibitors transactivate RAF dimers and ERK signalling in cells with wild-type BRAF. *Nature* 464, 427–430.
- (35) Chan, T. O., Zhang, J., Rodeck, U., Pascal, J. M., Armen, R. S., Spring, M., Dumitru, C. D., Myers, V., Li, X., Cheung, J. Y., and Feldman, A. M. (2011) Resistance of Akt kinases to dephosphorylation through ATP-dependent conformational plasticity. *Proc. Natl. Acad. Sci. U.S.A.* 108, E1120–E1127.
- (36) Andraos, R., Qian, Z., Bonenfant, D., Rubert, J., Vangrevelinghe, E., Scheufler, C., Marque, F., Regnier, C. H., De Pover, A., Ryckelynck, H., Bhagwat, N., Koppikar, P., Goel, A., Wyder, L., Tavares, G., Baffert, F., Pissot-Soldermann, C., Manley, P. W., Gaul, C., Voshol, H., Levine, R. L., Sellers, W. R., Hofmann, F., and Radimerski, T. (2012) Modulation of activation-loop phosphorylation by JAK inhibitors is binding mode dependent. *Cancer Discovery* 2, 512–523.
- (37) Hari, S. B., Merritt, E. A., and Maly, D. J. (2014) Conformation-selective ATP-competitive inhibitors control regulatory interactions and noncatalytic functions of mitogen-activated protein kinases. *Chem. Biol.* 21, 628–625.
- (38) Sullivan, J. E., Holdgate, G. A., Campbell, D., Timms, D., Gerhardt, S., Breed, J., Breeze, A. L., Birmingham, A., Pauptit, R. A., Norman, R. A., Embrey, K. J., Read, J., VanScyoc, W. S., and Ward, W. H. (2005) Prevention of MKK6-dependent activation by binding to p38alpha MAP kinase. *Biochemistry* 44, 16475–16490.
- (39) Murphy, S. M., Bergman, M., and Morgan, D. O. (1993) Suppression of c-Src activity by C-terminal Src kinase involves the c-Src SH2 and SH3 domains: analysis with *Saccharomyces cerevisiae*. *Mol. Cell. Biol.* 13, 5290–5300.
- (40) Okada, M., Nada, S., Yamanashi, Y., Yamamoto, T., and Nakagawa, H. (1991) CSK: a protein-tyrosine kinase involved in regulation of src family kinases. *J. Biol. Chem.* 266, 24249–24252.
- (41) Cole, P. A., Shen, K., Qiao, Y., and Wang, D. (2003) Protein tyrosine kinases Src and Csk: a tail's tale. *Curr. Opin. Chem. Biol.* 7, 580–585.
- (42) Sondhi, D., Xu, W., Songyang, Z., Eck, M. J., and Cole, P. A. (1998) Peptide and protein phosphorylation by protein tyrosine kinase Csk: insights into specificity and mechanism. *Biochemistry* 37, 165–172.
- (43) Hill, Z. B., Perera, B. G., Andrews, S. S., and Maly, D. J. (2012) Targeting diverse signaling interaction sites allows the rapid generation of bivalent kinase inhibitors. *ACS Chem. Biol.* 7, 487–495.
- (44) Pan, Z., Scheerens, H., Li, S. J., Schultz, B. E., Sprengeler, P. A., Burrill, L. C., Mendonca, R. V., Sweeney, M. D., Scott, K. C., Grothaus, P. G., Jeffery, D. A., Spoerke, J. M., Honigberg, L. A., Young, P. R., Dalrymple, S. A., and Palmer, J. T. (2007) Discovery of selective irreversible inhibitors for Bruton's tyrosine kinase. *ChemMedChem* 2, 58–61.
- (45) Roskoski, R., Jr. (2005) Src kinase regulation by phosphorylation and dephosphorylation. *Biochem. Biophys. Res. Commun.* 331, 1–14.
- (46) Bjorge, J. D., Pang, A., and Fujita, D. J. (2000) Identification of protein-tyrosine phosphatase 1B as the major tyrosine phosphatase activity capable of dephosphorylating and activating c-Src in several human breast cancer cell lines. *J. Biol. Chem.* 275, 41439–41446.
- (47) Shan, Y., Seeliger, M. A., Eastwood, M. P., Frank, F., Xu, H., Jensen, M. O., Dror, R. O., Kuriyan, J., and Shaw, D. E. (2009) A conserved protonation-dependent switch controls drug binding in the Abl kinase. *Proc. Natl. Acad. Sci. U.S.A.* 106, 139–144.
- (48) Alvarado, J. J., Betts, L., Moroco, J. A., Smithgall, T. E., and Yeh, J. I. (2010) Crystal structure of the Src family kinase Hck SH3–SH2 linker regulatory region supports an SH3-dominant activation mechanism. *J. Biol. Chem.* 285, 35455–35461.
- (49) Ayrapetov, M. K., Wang, Y. H., Lin, X., Gu, X., Parang, K., and Sun, G. (2006) Conformational basis for SH2–Tyr(P)527 binding in Src inactivation. *J. Biol. Chem.* 281, 23776–23784.
- (50) Lerner, E. C., and Smithgall, T. E. (2002) SH3-dependent stimulation of Src-family kinase autophosphorylation without tail release from the SH2 domain *in vivo*. *Nat. Struct. Biol.* 9, 365–369.
- (51) Moarefi, I., LaFevre-Bernt, M., Sicheri, F., Huse, M., Lee, C. H., Kuriyan, J., and Miller, W. T. (1997) Activation of the Src-family tyrosine kinase Hck by SH3 domain displacement. *Nature* 385, 650–653.
- (52) Smart, J. E., Oppermann, H., Czernilofsky, A. P., Purchio, A. F., Erikson, R. L., and Bishop, J. M. (1981) Characterization of sites for tyrosine phosphorylation in the transforming protein of Rous sarcoma virus (pp60v-src) and its normal cellular homologue (pp60c-src). *Proc. Natl. Acad. Sci. U.S.A.* 78, 6013–6017.
- (53) Wang, L., Perera, B. G., Hari, S. B., Bhatarai, B., Backes, B. J., Seeliger, M. A., Schurer, S. C., Oakes, S. A., Papa, F. R., and Maly, D. J. (2012) Divergent allosteric control of the IRE1alpha endoribonuclease using kinase inhibitors. *Nat. Chem. Biol.* 8, 982–989.
- (54) Koppikar, P., Bhagwat, N., Kilpivaara, O., Manshour, T., Adli, M., Hricik, T., Liu, F., Saunders, L. M., Mullally, A., Abdel-Wahab, O., Leung, L., Weinstein, A., Marubayashi, S., Goel, A., Gonen, M., Estrov, Z., Ebert, B. L., Chiosis, G., Nimer, S. D., Bernstein, B. E., Verstovsek, S., and Levine, R. L. (2012) Heterodimeric JAK–STAT activation as a mechanism of persistence to JAK2 inhibitor therapy. *Nature* 489, 155–159.
- (55) Seeliger, M. A., Young, M., Henderson, M. N., Pellicena, P., King, D. S., Falick, A. M., and Kuriyan, J. (2005) High yield bacterial expression of active c-Abl and c-Src tyrosine kinases. *Protein Sci.* 14, 3135–3139.
- (56) Johnson, S. M., Murphy, R. C., Geiger, J. A., DeRocher, A. E., Zhang, Z., Ojo, K. K., Larson, E. T., Perera, B. G., Dale, E. J., He, P., Reid, M. C., Fox, A. M., Mueller, N. R., Merritt, E. A., Fan, E., Parsons, M., Van Voorhis, W. C., and Maly, D. J. (2012) Development of *Toxoplasma gondii* calcium-dependent protein kinase 1 (TgCDPK1) inhibitors with potent anti-toxoplasma activity. *J. Med. Chem.* 55, 2416–2426.
- (57) Apsel, B., Blair, J. A., Gonzalez, B., Nazif, T. M., Feldman, M. E., Aizenstein, B., Hoffman, R., Williams, R. L., Shokat, K. M., and Knight, Z. A. (2008) Targeted polypharmacology: discovery of dual inhibitors of tyrosine and phosphoinositide kinases. *Nat. Chem. Biol.* 4, 691–699.
- (58) Brigham, J. L., Perera, B. G., and Maly, D. J. (2013) A hexylchloride-based catch-and-release system for chemical proteomic applications. *ACS Chem. Biol.* 8, 691–699.

(59) Zapf, C. W., Gerstenberger, B. S., Xing, L., Limburg, D. C., Anderson, D. R., Caspers, N., Han, S., Aulabaugh, A., Kurumbail, R., Shakya, S., Li, X., Spaulding, V., Czerwinski, R. M., Seth, N., and Medley, Q. G. (2012) Covalent inhibitors of interleukin-2 inducible T cell kinase (Itk) with nanomolar potency in a whole-blood assay. *J. Med. Chem.* 55, 10047–10063.

(60) Chen, W., and Loury, D. J. (2013) Preparation of pyrazolo[3,4-d]pyrimidine and pyrrolo[2,3-d]pyrimidine derivatives as Btk kinase inhibitors, p191, Patent US8377946 B1.