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Genetic abnormalities associated with acute lymphoblastic leukemia

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Acute lymphoblastic leukemia (ALL) occurs with high frequency in childhood and is associated with high mortality in adults. Recent technical advances in next-generation sequencing have shed light on genetic abnormalities in hematopoietic stem/progenitor cells as the precursor to ALL pathogenesis. Based on these genetic abnormalities, ALL is now being reclassified into newly identified subtypes. Philadelphia chromosome-like B-lineage ALL is one of the new high-risk subtypes characterized by genetic alterations that activate various signaling pathways, including those involving cytokine receptors, tyrosine kinases, and epigenetic modifiers. Philadelphia chromosome-like ALL is essentially heterogeneous; however, deletion mutations in the IKZF1 gene encoding the transcription factor IKAROS underlie many cases as a key factor inducing aggressive phenotypes and poor treatment responses. Whole-genome sequencing studies of ALL patients and ethnically matched controls also identified inherited genetic variations in lymphoid neoplasm-related genes, which are likely to increase ALL susceptibility. These findings are directly relevant to clinical hematology, and further studies on this aspect could contribute to accurate diagnosis, effective monitoring of residual disease, and patient-oriented therapies.

ymphopoietic potential is an essential feature of authentic HSCs. Hematopoietic stem cells supply multiple lymphocytes, including B, T, and NK cells, which form an exquisite network in the immune system for protection from external pathogens and internal neoplastic cells. B and T lymphocytes, which play pivotal roles in acquired immunity, are supplied by HSCs throughout life and mature by a step-wise process in the BM or thymus (Fig. 1).^(1,2) The lymphopoietic potential of HSCs, however, is not durable. Although HSCs produce large quantities of lymphocytes during fetal and neonatal periods, lymphocyte production dramatically attenuates after adolescence.^(3,4) Murine HSCs obtained from aged BM do not produce lymphocytes effectively in vitro or in vivo.⁽⁵⁾ In humans, adult BM HSCs are far less capable of producing lymphocytes compared to those of cord blood.⁽⁶⁾ The attenuation of lymphopoietic potential after reproductive age might be intrinsically inherent to the HSC characteristics shared between most existing species.

Fetal and neonatal HSCs are equipped to establish the immune system within a short period by producing a tremendous number of lymphocytes, whereas adult and elderly HSCs maintain homeostatic states of lymphopoiesis and become myeloid-lineage biased. The incidence of ALL reflects, in part, such age-related changes in HSCs. Acute lymphoblastic leukemia and AML account for approximately 80% and 20% of acute leukemia cases, respectively, in childhood, whereas the ratio of ALL /AML begins to decrease after adolescence. Additionally, the

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biological features of ALL cells seem to differ between children and adults, and the age at diagnosis of ALL patients is negatively correlated with survival rates and response to chemotherapy.⁽⁷⁾ Although the application of pediatric chemotherapy regimens to young adults and elderly ALL patients has improved treatment outcomes, the significant difference in survival rates between childhood and adult ALL cases persists.^(8,9)

The earliest evidence indicating that ALL pathogenesis involved genetic abnormality in HSCs or their more primitive ancestors was likely the observation by Wiemels *et al.*⁽¹⁰⁾ that chromosomal translocations and rearrangements of the *TEL-AML1* fusion gene were already detectable in neonatal blood cells of identical twin children with ALL, even several years before clinical manifestation of the disease. In recent years, the development of next-generation sequencing methods has significantly advanced the knowledge regarding ALL pathogenesis.⁽¹¹⁾ This review describes the current understanding of ALL pathogenesis and its new classifications, particularly from the viewpoint of genetic abnormalities in HSCs and somatic stem cells. Furthermore, perspectives on how these findings can be applied to improve ALL treatment are also discussed.

Philadelphia Chromosome-like ALL Cases, a New ALL Category, and Their Genetic Abnormalities

Gene-expression profiling approaches divided ALL into several subcategories, in which prognoses and frequencies according

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Review Genetic abnormalities with ALL



Fig. 1. Early lymphoid development in bone marrow and thymus. This schematic shows early differentiation routes of B- and T-lineage cells from hematopoietic stem cells (HSCs). In the bone marrow, the most primitive progenitors with lymphoid specification are contained within the c-KIT^{High} FMS-like tyrosine kinase-3 (FLT3)⁺ fraction, termed lymphoid-primed multipotent progenitors (LMPPs). The LMPP population overlaps multipotent progenitors (MPPs) and early lymphoid progenitors (ELPs), and is thought to be a major source for thymusimmigrating progenitors, including early T-lineage progenitors (ETPs). In the thymus, ETPs differentiate to double negative (DN)-2 and DN-3 cells with the activation of NOTCH signaling. ELPs differentiate to common lymphoid progenitor (CLP) and ProB cells in the bone marrow with the activation of interleukin-7 receptor (IL7R) signaling. Transcription factors are indicated near the stages where they are most active. Note that these data were taken from mouse studies. Human counterparts to each progenitor category remain to be identified. EBF, early B cell factor; RAG, recombination-activating gene.

to age differ significantly.⁽¹²⁾ Herein, we introduce one of the new ALL categories, Ph-like ALL, which is related to high-risk ALL.

Philadelphia chromosome positivity signifies the result of a translocation that gives rise to the *BCR-ABL1* oncogene and is one of the most detrimental hallmarks observed in ALL patients. Two groups independently proposed that multiple patients with Ph-negative B-lineage ALL had gene-expression profiles similar to those of patients with Ph-positive ALL.^(13,14) Such Ph-negative ALL cases were categorized as Philadelphia chromosome-like ALL (Ph-like ALL). Philadelphia chromosome-like ALL comprises 10% and 13% of standard and high-risk childhood B-lineage ALL, respectively.⁽¹⁵⁾ The frequency of Ph-like ALL increases with age, accounting for >25% of young adult cases, whose event-free and overall survival expectation is extremely poor, similar to that of Ph-positive cases.⁽¹⁵⁾

Given that the definition of Ph-like ALL was based only on the similarity of gene-expression profiles to Ph-positive ALL, the genetic abnormalities associated with Ph-like ALL cases were unlikely to be homogeneous. Therefore, transcriptome and whole-genome sequencing was carried out to understand genetic alterations underlying Ph-like ALL.^(15,16) Among 1725 B-lineage ALL cases, 154 patients were determined as Ph-like ALL and underwent genomic analyses. These approaches subcategorized Ph-like ALL into seven groups: type I, ABL-class fusions (*ABL1, ABL2, CSF1R, PDGRB*); type II, erythropoietin-receptor (*EPOR*) or *JAK2* rearrangements; type III, cytokine receptor-like factor 2 (*CRLF2*) rearrangements (often



Fig. 2. Categorization of Philadelphia chromosome-like acute lymphoblastic leukemia (Ph-like ALL) according to genetic abnormalities. Genomic analyses have shown that Ph-like ALL is heterogeneous, but involves a high frequency of kinase gene alterations. The data summarized in this figure are from Roberts *et al.*⁽¹⁵⁾

accompanied by *JAK2* mutations and JAK-STAT signal activation); type IV, other mutations activating JAK-STAT signaling *(IL7R, FLT3, SH2B3, TYK2, IL2RB)*; type V, uncommon miscellaneous kinase mutations (*NTRK3, DGKH*); type VI, RASpathway mutations (*KRAS, NRAS, PTPN11, NF1*); and type VII, no mutations in kinase genes (Fig. 2).

The type I category with ABL-class fusions accounted for 22% of Ph-like ALL cases and was independent from the abnormalities associated with the JAK-STAT signaling pathway. The type II category accounted for 18% of Ph-like ALL, while type III cases showed the most frequently observed genetic abnormality (approximately 20%) in Ph-like ALL cases. Interestingly, more than half of the *CRLF2* rearrangements associated with type III cases also harbored missense or multiple mutations in *JAK2*, which activate JAK-STAT signaling. The type IV category was related with other JAK-STAT activating abnormalities, in which IL7R and/or FLT3 mutations were common. Cases involving the type V and type VII categories were infrequent and difficult to characterize, and type VI occurred in a minority of Ph-like ALL cases, exhibiting genomic abnormalities activating RAS signaling.

Importantly, the high frequency of such kinase-activating mutations in Ph-like ALL suggested that the treatment outcomes of Ph-like ALL might improve with specific inhibitors (Fig. 3). Indeed, the Mullighan group reported that the addition of imatinib induced remission in a refractory Ph-like ALL patient who had an activating rearrangement of *PDGFB*.⁽¹⁷⁾

IKZF1 Mutations as a Key Factor Influencing the Development of High-Risk ALL

Deletions, amplifications, mutations, and structural rearrangements in key transcription factors promoting early lymphoid differentiation (e.g., *TCF3/E2A*, *EBF*, *LEF1*, *IKZF1*, *IKZF3*, *PAX5*, and *BLNK*) were detected in 40% of B-lineage ALL cases.⁽¹⁸⁾ Among these, mutations in the *IKZF1* gene encoding the IKAROS transcription factor were frequently observed and more highly correlated with poor prognosis associated with ALL than were mutations in genes encoding other transcription factors. Notably, many Ph-like ALL cases, regardless of the subcategories mentioned above, revealed mutations in the *IKZF1* gene, which is also a common finding in Ph-positive ALL. *IKZF1* encodes the transcription factor IKAROS, which is indispensable for the induction of B-lineage differentiation in HSCs.^(19,20) Its mutations are also strongly associated with lymphoid blast crisis of CML.⁽²¹⁾ Therefore, here, we introduce accumulating data pertaining to *IKZF1* mutations associated with high-risk ALL (Table 1).

Aberrant *IKZF1* mutations are likely some of the most detrimental driver mutations, accounting for >80% of Ph-positive ALL. IKZF1 deletions were not detectable in chronic-phase CML, but emerged simultaneously when CML transformed to lymphoid blast crisis.⁽²¹⁾ IKZF1 alterations were also common in Ph-like ALL cases, regardless of the type of kinase gene mutation described above,⁽¹⁵⁾ and suggested significantly lower 5-year event-free survival rates of Ph-like ALL patients compared to those without an IKZF1 alteration. With respect to Tlineage ALL, IKZF1 mutations were also observed more frequently in ETP-ALL, the phenotype of which is characterized as T-lineage marker-negative and HSC/myeloid marker-positive, than in other T-lineage ALL cases.⁽²²⁾ Homozygous germline IKZF1-null mice lacked T, B, and NK lymphocytes and their early progenitors, and heterozygous dominant-negative *IKZF1*-mutated mice rapidly acquired T-lineage ALL.^(19,23,24) Dominant-negative IKZF1 mutations are more deleterious and oncogenic, likely due to cross-interference with other IKAROS family members.



Fig. 3. Kinase gene alterations and their inhibitors in Philadelphia chromosome-like acute lymphoblastic leukemia (Ph-like ALL). The tyrosine kinase inhibitors for each kinase mutation in Ph-like ALL are indicated as a possible therapy.

Table 1. Features of *IKZF1*-mutated acute lymphoblastic leukemia (ALL)

High-risk ALL (except for *ERG*-deleted cases^(38,39))

84% of Philadelphia chromosome-positive ALL⁽²¹⁾

68% of Philadelphia chromosome-like ALL⁽¹⁵⁾

High adhesion potential to hematopoietic stem cell niche through integrins^(30,32)

Possible response to focal adhesion kinase inhibitors and retinoid receptor agonists^(30,32)

Most *IKZF1* deletions identified in Ph-positive ALL were monoallelic and lacked exons 3–6 of the *IKZF1* gene, which encode the N-terminal zinc finger DNA-binding domain.⁽²¹⁾ This deletion results in dominant-negative isoforms that inhibit both wild-type IKAROS and other family members.⁽²⁵⁾ Interestingly, genomic breakpoints in the *IKZF1* gene are located in the vicinity of cryptic heptamer-recombination-signal sequences, which are recognized by the RAG enzyme complex.⁽²¹⁾ These observations suggested that *IKZF1* deletions likely occurred with RAG expression, which marked the specification of hematopoietic stem/progenitor cells toward the lymphoid lineage.^(26–28) Recent studies have indicated that histone H3, trimethylated on lysine at position 4, in proximity to cryptic recombination-signal sequences, might contribute to misleading the RAG complex into producing aberrant recombination and promoting oncogenes.⁽²⁹⁾ Thus, epigenetic instability in Ph-positive HSCs or their proximate progenitors could underlie the incidence of *IKZF1* mutations.

Although *IKZF1* alterations strongly correlate with refractory ALL, the underlying mechanism of which remains unknown, recent efforts to investigate the biological features of IKZF1mutated cells offered clues to overcome this intractable disease. The Georgopoulos group showed that induction of dominant-negative IKAROS isoforms in early pre-B cells arrested their differentiation at the proliferative large pre-B cell stage and culminated in oligoclonal expansion with the occurrence of B-lineage ALL in transplanted recipients.^(30,31) These IKAROS-deleted pre-leukemic and leukemic cells expressed higher levels of integrins than those in normal counterpart cells, and were more dependent on BM stromal cells through integrin-mediated adhesion for their growth and survival. Notably, inhibitors for focal adhesion kinase, which transduces integrin signaling into cells, significantly abrogated adhesion and induced apoptosis in IKAROS-deleted leukemic cells.⁽³⁰⁾

The Mullighan group also reported that *IKZF1* alterations induced adhesive potential and HSC-related characteristics in Ph-positive ALL cells, while reducing their responsiveness to tyrosine kinase inhibitors.⁽³²⁾ *IKZF1*-altered ALL cells infiltrated BM and interacted with perivascular mesenchymal cells and arterial endothelial cells, which are thought to comprise the HSC niche.⁽³³⁾ It is noteworthy that treatment with retinoid-receptor agonists enhanced *IKZF1* expression, reversed HSC-like features of *IKZF1*-altered Ph-positive ALL cells, and recovered their sensitivity to tyrosine kinase inhibitors.⁽³²⁾ While retinoids have long been known to affect the integrity of HSCs and the differentiation of lympho-hematopoietic progenitors by directly or indirectly regulating a number of transcription factors,^(34–36) numerous nuclear receptors also potentially regulate transcription factors in HSCs.⁽³⁷⁾ Thus, it is worth examining the types of nuclear-receptor signals that upregulate *IKZF1* expression.

Intragenic deletions of *ERG*, which encodes an ETS family member transcription factor, were recently identified in a subset of childhood B-lineage ALL.⁽³⁸⁾ Intriguingly, although dominant-negative *IKZF1* deletions were frequently associated with *ERG*-deleted ALL cases, the response to treatment and survival of these patients were unexpectedly positive. In fact, within B-lineage ALL cases involving *IKZF1* deletion, 8-year event-free survival improved to >85% in cases involving *ERG* deletions, whereas the rate was only 51% without the deletions.⁽³⁸⁾ An independent group also reported similar results, suggesting that *ERG* deletions might mitigate the negative impact of *IKZF1* deletion in ALL prognoses.⁽³⁹⁾ Future studies on the molecular mechanisms underlying such exceptional cases might discover more sophisticated strategies to overcome *IKZF1* alterations in ALL cells.

Inherited Germline Variations as a Risk Factor for ALL Development

Historically, germline variations inherited from ancestors were rarely considered to influence ALL incidence or features. Until a decade ago, ALL pathogenesis was thought to be mostly attributed to acquired mutations in HSCs or lymphoid progenitors. However, recent studies have identified multiple genomic variants that increase ALL susceptibility and affect prognosis.

Genome-wide association studies for childhood ALL cases identified multiple germline polymorphisms showing high association with ALL incidence and characteristics.⁽¹¹⁾ Some of the inheritable variants were found in lymphoid neoplasmrelated genes, including IKZF1, ARID5B, and CDKN2.(40-44) IKZF1 encodes the lymphoid-lineage transcription factor IKAROS mutation, which is intimately associated with ALL as previously discussed. Notably, this protein is also an integral component combining transcription factors with the chro-matin-remodeling network.⁽⁴⁵⁾ ARID5B is a member of the AT-rich DNA-interaction domain family,⁽⁴⁶⁾ and although the ARID5B function in lympho-hematopoiesis has not been well studied, it may be involved in epigenetic regulation of gene expression in HSCs and early lymphoid progenitors, similar to other AT-rich DNA-binding proteins. (47–50) Interestingly, ARID5B polymorphisms are associated with B-lineage hyperdiploid ALL and racial differences associated with ALL incidence.^(40,51) CDKN2 encodes INK4a/ARF, which regulates HSC self-renewal and differentiation under the control of the transcriptional repressor and polycomb group protein Bmi1.^(52,53) Therefore, hereditary predisposition to ALL may be related to the epigenetic instability of HSCs and lymphoid progenitors.

Heteroploid chromosomal abnormality is often observed in ALL cases. A hyperdiploid karyotype is more common in childhood ALL compared to adult ALL; however, the associated mechanisms remain unknown. Moriyama and colleagues studied familial ALL and identified a nonsense variant of ETV6, a member of the erythroblast transformation-specific family, which is involved in hematopoiesis and oncogenesis, with high prevalence among familial ALL cases.^(54–56) They undertook a broad survey for ETV6 mutations in >4405 ALL children and identified 31 somatic ETV6 variants potentially related to ALL susceptibility. Interestingly, ALL children with ETV6 variants were significantly older than those without such variants when diagnosed, and more often had a hyperdiploid karyotype. Additionally, a hypodiploid karyotype is a hallmark of poor ALL outcomes. Holmfeldt and colleagues used genomic profiling on 124 childhood ALL cases with a hypodiploid karyotype⁽⁵⁷⁾ and identified inherited alterations in *TP53*, as</sup> well as RAS-, DNA-repair-, and receptor tyrosine kinase signaling-related genes.

The higher frequency of ALL in children relative to adults suggests that the influence of genetic predisposition to the disease might be more profound at younger ages. However, a recent study showed that inherited *GATA3* variants strongly enhanced susceptibility to ALL in adolescents and young

References

- Pelayo R, Welner R, Perry SS *et al.* Lymphoid progenitors and primary routes to becoming cells of the immune system. *Curr Opin Immunol* 2005; 17: 100–7.
- 2 Zhang Q, Iida R, Yokota T, Kincade PW. Early events in lymphopoiesis: an update. *Curr Opin Hematol* 2013; **20**: 265–72.

adults.⁽⁵⁸⁾ The detected *GATA3* polymorphisms were also detectable in childhood Ph-like ALL,⁽⁵⁹⁾ and the frequency of ALL patients with those *GATA3* variations was positively correlated with patient age at diagnosis. Furthermore, *GATA3* variations were also associated with poor treatment response and high risk of relapse. These findings suggested that age-related differences in ALL biology might reflect, at least in part, the genetic variations resident in HSCs.

Conclusions and Perspectives

In the last decade, advances in gene-expression profiling and genome-wide sequencing have revolutionized our understanding of ALL pathogenesis. As described in this short review, accumulating information has revamped ALL classifications according to genetic variations. Many pathogenic ALL mutations have been identified in HSC-related genes, which are often associated with ALL treatment failure and early relapse. As some of the mutations are also associated with activation of certain kinase pathways, the invention of simple, convenient, and cost-effective sequencing technologies would enable earlier and more sophisticated therapeutic intervention with specific inhibitors. On the other hand, we must stress that leukemia-initiating mutations are still undetectable in >10% of childhood ALL patients, and that the genetic information of older adult ALL patients has not been catalogued at this stage. Nonetheless, we believe that studies in the coming decade will completely describe the genomic landscape of ALL across all generations and refine the therapeutic algorithm to be more targeted and individualized.

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Disclosure Statement

The authors have no conflicts of interest relating to the topic of this article.

Abbreviations

ALL	acute lymphoblastic leukemia
AML	acute myeloid leukemia
BM	bone marrow
CML	chronic myeloid leukemia
ETP	early T-lineage progenitor
FLT3	FMS-like tyrosine kinase-3
HSC	hematopoietic stem cell
IL7R	interleukin-7 receptor
NK	natural killer
Ph-like	Philadelphia chromosome-like
RAG	recombination-activating gene
STAT	signal transducer and activator of transcription

- 3 Montecino-Rodriguez E, Dorshkind K. Evolving patterns of lymphopoiesis from embryogenesis through senescence. *Immunity* 2006; 24: 659–62.
- 4 Geiger H, de Haan G, Florian MC. The ageing haematopoietic stem cell compartment. *Nat Rev Immunol* 2013; **13**: 376–89.
- 5 Sudo K, Ema H, Morita Y, Nakauchi H. Age-associated characteristics of murine hematopoietic stem cells. *J Exp Med* 2000; **192**: 1273–80.

- 6 Rossi MI, Yokota T, Medina KL *et al.* B lymphopoiesis is active throughout human life, but there are developmental age-related changes. *Blood* 2003; 101: 576–84.
- 7 Hunger SP, Lu X, Devidas M *et al.* Improved survival for children and adolescents with acute lymphoblastic leukemia between 1990 and 2005: a report from the children's oncology group. *J Clin Oncol* 2012; **30**: 1663–9.
- 8 Schafer ES, Hunger SP. Optimal therapy for acute lymphoblastic leukemia in adolescents and young adults. *Nat Rev Clin Oncol* 2011; **8**: 417–24.
- 9 Ram R, Wolach O, Vidal L, Gafter-Gvili A, Shpilberg O, Raanani P. Adolescents and young adults with acute lymphoblastic leukemia have a better outcome when treated with pediatric-inspired regimens: systematic review and meta-analysis. *Am J Hematol* 2012; **87**: 472–8.
- Wiemels JL, Cazzaniga G, Daniotti M et al. Prenatal origin of acute lymphoblastic leukaemia in children. Lancet 1999; 354: 1499–503.
- 11 Roberts KG, Mullighan CG. Genomics in acute lymphoblastic leukaemia: insights and treatment implications. *Nat Rev Clin Oncol* 2015; **12**: 344– 57.
- 12 Mullighan CG. The genomic landscape of acute lymphoblastic leukemia in children and young adults. *Hematology Am Soc Hematol Educ Program* 2014; 2014: 174–80.
- 13 Mullighan CG, Su X, Zhang J et al. Deletion of IKZF1 and prognosis in acute lymphoblastic leukemia. N Engl J Med 2009; 360: 470–80.
- 14 Den Boer ML, van Slegtenhorst M, De Menezes RX et al. A subtype of childhood acute lymphoblastic leukaemia with poor treatment outcome: a genome-wide classifi cation study. *Lancet Oncol* 2009; 10: 125–34.
- 15 Roberts KG, Li Y, Payne-Turner D *et al.* Targetable kinase-activating lesions in Ph-like acute lymphoblastic leukemia. *N Engl J Med* 2014; **371**: 1005–15.
- 16 Roberts KG, Morin RD, Zhang J *et al.* Genetic alterations activating kinase and cytokine receptor signaling in high-risk acute lymphoblastic leukemia. *Cancer Cell* 2012; **22**: 153–66.
- 17 Weston BW, Hayden MA, Roberts KG *et al.* Tyrosine kinase inhibitor therapy induces remission in a patient with refractory EBF1-PDGFRB-positive acute lymphoblastic leukemia. *J Clin Oncol* 2013; **31**: e413–6.
- 18 Mullighan CG, Goorha S, Radtke I et al. Genome-wide analysis of genetic alterations in acute lymphoblastic leukaemia. Nature 2007; 446: 758–64.
- 19 Georgopoulos K, Bigby M, Wang JH et al. The Ikaros gene is required for the development of all lymphoid lineages. Cell 1994; 79: 143–56.
- 20 Ng SY, Yoshida T, Zhang J, Georgopoulos K. Genome-wide lineage-specific transcriptional networks underscore Ikaros-dependent lymphoid priming in hematopoietic stem cells. *Immunity* 2009; **30**: 493–507.
- 21 Mullighan CG, Miller CB, Radtke I et al. BCR-ABL1 lymphoblastic leukaemia is characterized by the deletion of Ikaros. Nature 2008; 453: 110–4.
- 22 Zhang J, Ding L, Holmfeldt L et al. The genetic basis of early T-cell precursor acute lymphoblastic leukaemia. Nature 2012; 481: 157–63.
- 23 Winandy S, Wu P, Georgopoulos K. A dominant mutation in the Ikaros gene leads to rapid development of leukemia and lymphoma. *Cell* 1995; 83: 289– 99.
- 24 Wang JH, Nichogiannopoulou A, Wu L et al. Selective defects in the development of the fetal and adult lymphoid system in mice with an Ikaros null mutation. *Immunity* 1996; 5: 537–49.
- 25 Sun L, Liu A, Georgopoulos K. Zinc finger-mediated protein interactions modulate Ikaros activity, a molecular control of lymphocyte development. *EMBO J* 1996; 15: 5358–69.
- 26 Igarashi H, Gregory SC, Yokota T, Sakaguchi N, Kincade PW. Transcription from the RAG1 locus marks the earliest lymphocyte progenitors in bone marrow. *Immunity* 2002; 17: 117–30.
- 27 Yokota T, Kouro T, Hirose J et al. Unique properties of fetal lymphoid progenitors identified according to RAG1 gene expression. *Immunity* 2003; 19: 365–75.
- 28 Yokota T, Sudo T, Ishibashi T *et al.* Complementary regulation of early Blymphoid differentiation by genetic and epigenetic mechanisms. *Int J Hematol* 2013; **98**: 382–9.
- 29 Shimazaki N, Lieber MR. Histone methylation and V(D)J recombination. Int J Hematol 2014; 100: 230–7.
- 30 Joshi I, Yoshida T, Jena N et al. Loss of Ikaros DNA-binding function confers integrin-dependent survival on pre-B cells and progression to acute lymphoblastic leukemia. Nat Immunol 2014; 15: 294–304.
- 31 Yoshida T, Georgopoulos K. Ikaros fingers on lymphocyte differentiation. Int J Hematol 2014; 100: 220–9.
- 32 Churchman ML, Low J, Qu C et al. Efficacy of retinoids in IKZF1-Mutated BCR-ABL1 acute lymphoblastic leukemia. Cancer Cell 2015; 28: 343–56.
- 33 Morrison SJ, Scadden DT. The bone marrow niche for haematopoietic stem cells. *Nature* 2014; 505: 327–34.

- 34 Collins SJ. The role of retinoids and retinoic acid receptors in normal hematopoiesis. *Leukemia* 2002; 16: 1896–905.
- 35 Purton LE, Dworkin S, Olsen GH et al. RARgamma is critical for maintaining a balance between hematopoietic stem cell self-renewal and differentiation. J Exp Med 2006; 203: 1283–93.
- 36 Chen X, Esplin BL, Garrett KP, Welner RS, Webb CF, Kincade PW. Retinoids accelerate B lineage lymphoid differentiation. *J Immunol* 2008; 180: 138–45.
- 37 Chute JP, Ross JR, McDonnell DP. Minireview: nuclear receptors, hematopoiesis, and stem cells. *Mol Endocrinol* 2010; 24: 1–10.
- 38 Clappier E, Auclerc MF, Rapion J *et al.* An intragenic ERG deletion is a marker of an oncogenic subtype of B-cell precursor acute lymphoblastic leukemia with a favorable outcome despite frequent IKZF1 deletions. *Leukemia* 2014; 28: 70–7.
- 39 Zaliova M, Zimmermannova O, Dorge P *et al.* ERG deletion is associated with CD2 and attenuates the negative impact of IKZF1 deletion in childhood acute lymphoblastic leukemia. *Leukemia* 2014; **28**: 182–5.
- 40 Trevino LR, Yang W, French D et al. Germline genomic variants associated with childhood acute lymphoblastic leukemia. Nat Genet 2009; 41: 1001–5.
- 41 Sherborne AL, Hosking FJ, Prasad RB *et al.* Variation in CDKN2A at 9p21.3 influences childhood acute lymphoblastic leukemia risk. *Nat Genet* 2010; **42**: 492–4.
- 42 Yang JJ, Cheng C, Devidas M et al. Ancestry and pharmacogenomics of relapse in acute lymphoblastic leukemia. Nat Genet 2011; 43: 237-41.
- 43 Yang JJ, Cheng C, Devidas M et al. Genome-wide association study identifies germline polymorphisms associated with relapse of childhood acute lymphoblastic leukemia. *Blood* 2012; **120**: 4197–204.
- 44 Xu H, Zhang H, Yang W *et al.* Inherited coding variants at the CDKN2A locus influence susceptibility to acute lymphoblastic leukaemia in children. *Nat Commun* 2015; **6**: 7553.
- 45 Georgopoulos K. Haematopoietic cell-fate decisions, chromatin regulation and ikaros. Nat Rev Immunol 2002; 2: 162–74.
- 46 Wilsker D, Probst L, Wain HM, Maltais L, Tucker PW, Moran E. Nomenclature of the ARID family of DNA-binding proteins. *Genomics* 2005; 86: 242–51.
- 47 Webb CF, Bryant J, Popowski M *et al.* The ARID family transcription factor bright is required for both hematopoietic stem cell and B lineage development. *Mol Cell Biol* 2011; **31**: 1041–53.
- 48 Will B, Vogler TO, Bartholdy B et al. Satb1 regulates the self-renewal of hematopoietic stem cells by promoting quiescence and repressing differentiation commitment. Nat Immunol 2013; 14: 437–45.
- 49 Satoh Y, Yokota T, Sudo T *et al.* The Satb1 protein directs hematopoietic stem cell differentiation toward lymphoid lineages. *Immunity* 2013; 38: 1105–15.
- 50 Yokota T, Kanakura Y. Role of tissue-specific AT-rich DNA sequence-binding proteins in lymphocyte differentiation. Int J Hematol 2014; 100: 238–45.
- 51 Yang W, Trevino LR, Yang JJ *et al.* ARID5B SNP rs10821936 is associated with risk of childhood acute lymphoblastic leukemia in blacks and contributes to racial differences in leukemia incidence. *Leukemia* 2010; **24**: 894–6.
- 52 Jacobs JJ, Kieboom K, Marino S, DePinho RA, van Lohuizen M. The oncogene and Polycomb-group gene bmi-1 regulates cell proliferation and senescence through the ink4a locus. *Nature* 1999; **397**: 164–8.
- 53 Oguro H, Iwama A, Morita Y, Kamijo T, van Lohuizen M, Nakauchi H. Differential impact of Ink4a and Arf on hematopoietic stem cells and their bone marrow microenvironment in Bmi1-deficient mice. *J Exp Med* 2006; 203: 2247–53.
- 54 Moriyama T, Metzger ML, Wu G et al. Germline genetic variation in ETV6 and risk of childhood acute lymphoblastic leukaemia: a systematic genetic study. Lancet Oncol 2015; 16: 1659–66.
- 55 Wang LC, Swat W, Fujiwara Y et al. The TEL/ETV6 gene is required specifically for hematopoiesis in the bone marrow. *Genes Dev* 1998; 12: 2392–402.
- 56 Kar A, Gutierrez-Hartmann A. Molecular mechanisms of ETS transcription factor-mediated tumorigenesis. *Crit Rev Biochem Mol Biol* 2013; 48: 522– 43.
- 57 Holmfeldt L, Wei L, Diaz-Flores E et al. The genomic landscape of hypodiploid acute lymphoblastic leukemia. Nat Genet 2013; 45: 242–52.
- 58 Perez-Andreu V, Roberts KG, Xu H et al. A genome-wide association study of susceptibility to acute lymphoblastic leukemia in adolescents and young adults. *Blood* 2015; **125**: 680–6.
- 59 Perez-Andreu V, Roberts KG, Harvey RC et al. Inherited GATA3 variants are associated with Ph-like childhood acute lymphoblastic leukemia and risk of relapse. Nat Genet 2013; 45: 1494–8.