Synthesis of rigid *p*-terphenyl-linked carbohydrate mimetics

Maja Kandziora and Hans-Ulrich Reissig*

Full Research Paper

Takustraße 3. D-14195 Berlin, Germany

Email:

Hans-Ulrich Reissig* - hans.reissig@chemie.fu-berlin.de

Freie Universität Berlin, Institut für Chemie und Biochemie,

* Corresponding author

Keywords:

carbohydrate mimetics; hydrogenolysis; multivalent glycosystems; 1,2-oxazines; samarium diiodide; Suzuki cross-coupling

Beilstein J. Org. Chem. 2014, 10, 1749-1758.

doi:10.3762/bjoc.10.182

Received: 19 March 2014 Accepted: 23 June 2014 Published: 30 July 2014

This article is part of the Thematic Series "Multivalent glycosystems for

Open Access

nanoscience".

Guest Editor: B. Turnbull

© 2014 Kandziora and Reissig; licensee Beilstein-Institut.

License and terms: see end of document.

Abstract

An approach to β -D-2-aminotalose- and β -D-2-aminoidose-configured carbohydrate mimetics bearing a phenyl substituent is described. Unnatural divalent rigid p-terphenyl-linked C-aryl glycosides with 2.0 nm dimension are available using Suzuki cross-couplings. The key compound, a p-bromophenyl-substituted 1,2-oxazine, was prepared by a stereoselective [3 + 3]-cyclization of a D-isoascorbic acid-derived (Z)-nitrone and lithiated TMSE-allene. The Lewis acid-induced rearrangement of this heterocycle provided the corresponding bicyclic 1,2-oxazine derivative that may be regarded as internally protected amino sugar analogue. After subsequent reduction of the carbonyl group, the resulting bicyclic compound was used for Suzuki cross-couplings to form biphenyl aminopyran or p-terphenyl-linked dimers. Hydrogenolysis afforded new unnatural aminosugar mimetics. Zinc in the presence of acid or samarium diiodide were examined for the N–O bond cleavage in order to obtain the rigid p-terphenyl-linked C-glycosyl dimers.

Introduction

Carbohydrates are the class of biomolecules with the highest structural diversity [1,2]. Specific carbohydrates are responsible for cell-type specific interactions [3] and they are involved in different diseases such as cancer [4], inflammation [5], and infections [6]. However, the use of carbohydrates as drugs has been strongly limited due to the hydrolytic lability of the glycosidic bond [7] and the weak binding affinities of single molecules. With the development of artificial *C*-glycosides

which possess structural and functional aspects of the corresponding carbohydrates, these disadvantages can be overcome, resulting in an improved bioavailability, higher affinities and improved selectivities [8-13]. Recent results indicate that divalent rigid carbohydrate conjugates may have even higher binding affinities and specificities than their flexible multivalent equivalents [14,15]. The rigidity of the system is supposed to improve the overall activity of the ligands by over-

coming the entropic penalty of flexible multivalent scaffolds [16].

Cross-coupling reactions are among the best methods to prepare C-arylglycosides, C-nucleosides and C-glycosidic oligomers when new artificial pharmacophores are approached [17]. With Suzuki cross-couplings C-glycoside analogues of phloriain with antidiabetic properties [18] or aryl-scaffolded dimers and trimers were successfully prepared [19]. The Suzuki crosscoupling is particularly suitable for carbohydrate chemistry due to the mild reaction conditions and its tolerance to a variety of functional groups [20]. In addition, the reactions are easy to perform and the required boronic acids exhibit exceptional stabilities to heat, air and moisture compared to other organometallic reagents [21]. Up to now there are not many examples of nanorod-like carbohydrate dimers with aryl-linked divalent glycosides. A mannopyranoside dimer was generated by a palladium-catalyzed Ullmann-type reductive homocoupling [22] and biphenyl-linked dimers were prepared by Lewis acid-catalyzed glycosidations [23]. All these examples possess an acid-labile O-glycosidic bond and are labile to hydrolysis and enzymes. Therefore, new approaches to the synthesis of rigid multivalent C-arylglycosides should be a valuable extension of compounds with potential biological activity. In order to achieve this goal we investigated the synthesis of divalent compounds of general structure 1 and their monovalent analogues 2 (Scheme 1). Structurally similar aminopyrans without aryl groups have intensively been studied as carbohydrate mimetics in our group [24,25]. When they are coupled by amide bonds to gold nanoparticle and O-sulfated these conjugates gave extremely high binding affinities towards L- and P-selectin in sub-nanomolar concentrations. These results were achieved by a multivalent presentation (ca. 1000–1200 ligands per nanoparticle) of the sulfated pyrans [26,27]. We were therefore interested to prepare inhibitors offering only a small number of ligands to get better information about structure–activity relationships and to study the influence of the flexible and rigid spacer units.

In this report we present methods for the synthesis of divalent compounds 1 with p-terphenyl spacers and of β -D-2-aminotalose- or β -D-2-aminoidose-configured carbohydrate mimetics 2 (Scheme 1). These novel carbohydrate mimetics represent unique structures, combining the features of C-aryl-glycosides and aminosugars. The p-bromophenyl-substituted bicyclic 1,2-oxazine derivative 3 was used as key building block for the Suzuki cross-coupling reaction to synthesize p-terphenyl-linked derivatives 1. The key intermediate 3 was prepared by a Lewis acid-induced rearrangement of 3,6-dihydro-2H-oxazine 4, that origins from a stereoselective [3 + 3]-cyclization of D-isoascorbic acid-derived (Z)-nitrone 6 and lithiated TMSE-allene 5.

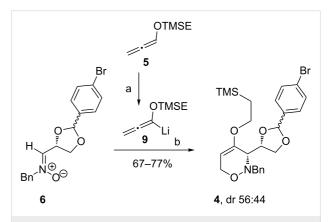
Results and Discussion

For our synthesis of new divalent carbohydrate mimetics we required 1,2-oxazine derivatives derived from (Z)-nitrone 6 and lithiated alkoxyallenes. The 4-bromophenyl group should allow transition metal-promoted coupling reactions to a variety of new compounds. For this purpose the D-erythrose-configured ester 7, easily available from D-isoascorbic acid [28], was converted into nitrone 6 in a three step procedure (Scheme 2). Its reduction with lithium aluminum hydride was performed under standard conditions providing diol 8 in excellent yield in multigram scale (up to 20 g). Attention should be paid to a possible reductive removal of the bromine substituent that can occur at higher temperature or longer reaction times as the resulting debrominated product is hard to remove from diol 8 by column chromatography. According to the protocol of Dondoni et al. [29] glycol cleavage of diol 8 afforded the corresponding aldehyde that was directly treated with N-benzylhydroxylamine to furnish the desired (Z)-nitrone 6. All compounds in this sequence of reactions are mixtures of the two diastereomers at the dioxolane C-2 (ratios close to 1:1).

Scheme 2: Synthesis of (*Z*)-nitrone 6. Conditions: a) LiAlH₄, THF, 1 h, rt; b) 1. NalO₄, CH₃CN/H₂O, 1 h, rt; 2. *N*-benzylhydroxylamine, MgSO₄, CH₂Cl₂, 18 h, rt.

The preparation of syn-1,2-oxazine 4 was achieved in good yields ranging from 67-77% by stereocontrolled addition of lithiated (trimethylsilyl)ethoxyallene 9 to (Z)-nitrone 6 at -78 °C (Scheme 3). Although the formation of four stereoisomers is possible only two were observed. Due to the complexation of lithiated allene 9 to the nitrone 6 an exclusive formation of the two syn-1,2-oxazines 4 was observed. This result suggests that the configuration at C-2 of the dioxolane moiety has no influence on the stereochemical outcome of the reaction. The model suggested by Dondoni et al. [30] can also be employed for this process to rationalize the observed diastereoselectivity of the addition step. The subsequent [3 + 3]-cyclization to 3,6dihydro-2H-1,2-oxazine 4 follows the previously reported mechanism [31]. In the presented sequence syn-1,2-oxazine 4 was successfully prepared in six steps with an overall yield of 46%. The diastereomers can easily be separated by column chromatography, but this turned out not to be mandatory. The

next step of our anticipated sequence, the Lewis acid-induced rearrangement, converts the dioxolane C-2 carbon into an sp²-hybridized carbon and hence the configuration of the precursor does not play a role for this reaction.



Scheme 3: [3 + 3]-Cyclization of (*Z*)-nitrone 6 with lithiated allene 9. Conditions: a) *n*-BuLi, THF, 15 min, -40 °C; b) 1. THF, 2 h, -78 °C; 2. H₂O, 1 h, -78 °C \rightarrow rt.

An alternative route to prepare 1,2-oxazine 4 is depicted in Scheme 4. The preparation of the 4-bromophenyl-1,3-dioxolane moiety started from diol 10 that has successfully been used earlier for the preparation of phenylthio-substituted 1,2oxazine derivatives [32]. Compound 10 is easily accessible by a mild cleavage of the corresponding acetonide by an indium trichloride-mediated hydrolysis [33]. By using cerium ammonium nitrate as Lewis acid [34] in high concentration (13 mmol/mL) as well as an excess of 1-bromo-4-(dimethoxymethyl)benzene enabled the synthesis of syn-1,2-oxazine 4. The conversion of this reaction was high (>80%) giving the two diastereomers of 4 (ca. 1:1), but only one diastereomer was isolated in pure form. The second diastereomer could hardly be separated from the excess of 1-bromo-4-(dimethoxymethyl)benzene by column chromatography or distillation. Besides, Brønsted acids like trifluoroacetic acid, p-toluenesulfonic acid,

 $\label{eq:Scheme 4: Synthesis of 1,2-oxazine 4 by acetal formation from 10. Conditions: a) 1-bromo-4-(dimethoxymethyl)benzene (10 equiv.), CAN, CH_2Cl_2, 3 d, rt.$

that are usually used to generate ketals, or weaker acids like pyridine/hydrogen fluoride led to a side product [35].

The Lewis acid-promoted rearrangement of 1,3-dioxolanylsubstituted 1,2-oxazines to bicyclic ketones has been described in many examples [24]. Gratifyingly, starting from 1,2-oxazine 4 with tin(IV) chloride as Lewis acidic promoter the corresponding ketone was obtained in excellent stereoselectivity. The subsequent protection of the primary hydroxy group as TBS ether under standard conditions provided 11 in very good yields of up to 82% (Scheme 5). In order to perform the Lewis acidpromoted rearrangement and the protection in one step, we also employed TBSOTf as Lewis acid for the rearrangement step [24], however, no product formation could be observed in this case. The mechanism of the rearrangement $4 \rightarrow 11$ can be described as an aldol-type cyclization process. The Lewis acid coordinates to the sterically less hindered oxygen atom of the dioxolane ring of 4 opening this ring and forming a carbenium ion that intramolecularly attacks the enol ether moiety. A sixmembered chair-like transition state with the bulky 4-bromophenyl group in an equatorial position for this crucial step rationalizes the product configuration as shown.

TBS-protected bicyclic ketone 11 was subsequently reduced with sodium borohydride at -40 °C to form alcohols 12a and 12b as 81:19 mixture of diastereomers in 72% yield (Scheme 5). In contrast, the reduction with L-selectride at -10 °C selectively furnished pure diastereomer 12a in 73%

yield. In accordance with previous observations of reductions of related phenylthio-substituted bicyclic compounds [25], a hydride attack from the side of the pyran moiety is assumed since the 1,2-oxazine side is more hindered by the bulky *N*-benzyl moiety. The secondary hydroxy group of **12a** was protected employing *t*-butyldimethylsilyl trifluoromethanesulfonate and 2,6-lutidine in quantitative yield.

The Lewis acid-promoted rearrangement of 1,2-oxazine derivative 4 and the direct reduction of the unpurified ketone 14 furnished bicyclic diol 15 in good overall yield of 58% (Scheme 6). The reduction of unprotected ketone 14 with L-selectride was less stereoselective and provided a 72:28 mixture of 15. The higher selectivity of the TBS-protected compound 11 may be explained by an indirect effect of the bulky TBS-group, possibly pushing the *N*-benzyl moiety to the top of the ring shielding the 1,2-oxazine side more efficiently. Separation of the two diastereomers and protection of the primary hydroxy group of 15a with trityl chloride provided compound 16 in 83% yield. This protecting group should allow its removal together with the benzyl group during hydrogenolysis. Surprisingly, a benzyl protection under the same conditions was not possible.

Before approaching divalent compounds such as 1 we wanted to convert our building blocks into simple monocyclic carbohydrate mimetics. To prepare phenyl-substituted aminopyrans the N-O bond of bicyclic compounds 15a and 15b was cleaved by

Scheme 5: Synthesis of bicyclic ketone 11 by Lewis acid-induced rearrangement and reduction to alcohols 12a and 12b and protection of 12a to 1,2-oxazine derivative 13. Conditions: a) 1. SnCl₄, CH₃CN, 4 h, -30 °C \rightarrow rt; 2. TBSCl, imidazole, THF, 4 h, rt; b) NaBH₄, EtOH, 4 h, -40 °C, 72%, dr 81:19; c) L-selectride, THF, 2 h, -10 °C, 73%, only 12a; d) TBSOTf, 2,6-lutidine, THF, 2 h, 0 °C.

Scheme 6: Synthesis of bicyclic diols 15 and of trityl-protected bicyclic 1,2-oxazine 16. Conditions: a) SnCl₄, CH₃CN, 18 h, -30 °C \rightarrow rt, b) 1) L-selectride, THF, 4 h, -15 °C, dr 72:28; 2) separation by column chromatography (silica gel, hexanes/EtOAc 1:1 \rightarrow 1:2); c) TrtCl, DMAP, pyridine, 3 d, rt.

hydrogenolysis. These reactions are challenging because the resulting aminopyrans are apparently poisoning the catalyst and hence large amounts of palladium on charcoal are required for full conversion. We did not add acid to diminish catalyst poisoning since we were afraid of other side reactions of the complex product. In addition, the resulting products are very polar and difficult to purify. In our recent report [28] related compounds were reduced in methanol as solvent providing several side products and the yield of the reactions were not fully reproducible. Nevertheless, 1,2-oxazine 15a was converted into aminopyran 17a (Scheme 7) by hydrogenolysis under standard conditions in methanol in a yield of 78%, but this yield was not fully reproducible and the conditions were optimized. We found that isopropanol as solvent and addition of one equivalent triethylamine were more reliable and the yield of 17a could be slightly improved. Triethylamine was added to neutralize the formed acid [36] that is generated in the first step by a very fast debromination. The debenzylation and the N-O bond cleavage occur as next steps. Under these improved conditions the isomeric bicyclic 1,2-oxazine 15b was converted into aminopyran 17b in a good yield of 77%. The formed aminopyrans 17a and 17b can be regarded as amino C-glycosides. Compound 17a is related to compounds with β-D-talose configuration that are rarely found in nature, an exception being the antibiotic amino glycoside hygromycin B [37]. Aminopyran 17b correlates to β-D-idopyranose; iduronic acid is a component of sulfated glycosamine glycans such as chrondroitin sulfate and heparan sulfate [38].

The prepared *p*-bromophenyl-substituted bicyclic 1,2-oxazine derivatives **12**, **13**, **15** and **16** provide options to perform cross-coupling reactions such as Buchwald/Hartwig, Heck, Hiyama, Kumada, Sonogashira or Stille couplings. In order to examine the conditions for Suzuki cross-couplings we subjected bicyclic compound **15a** to phenylboronic acid under standard conditions

Scheme 7: Hydrogenolyses of bicyclic 1,2-oxazine derivatives 15a and 15b. Conditions: a) H₂, Pd/C, MeOH, EtOAc, 20 h, rt; b) H₂, Pd/C, iPrOH, EtOAc, NEt₃, 18 h, rt.

of this reaction. The desired product 18 was obtained in 81% yield (Scheme 8) and the subsequent hydrogenolysis furnished carbohydrate mimetic 19 bearing a biphenyl substituent.

The smooth transformation of bicyclic compound **15a** into a biphenyl compound by Suzuki cross-coupling encouraged us to aim the synthesis of divalent compound **21**. 1,4-Phenylenediboronic acid (**20**) was used as precursor and coupled to two equivalents of **12a** to afford the desired *p*-terphenyl compound **21** in excellent 84% yield (Scheme 9). Unfortunately, the subsequent hydrogenolysis of this compound under conditions as above led to a complex product mixture and no product could be observed. An alternative method for N–O cleavage employs elemental zinc in the presence of acid [39], conditions that should simultaneously cleave the TBS protective groups. For the conversion of **21** into **22** long reaction times were required and the high acidity led to the formation of side products.

Scheme 8: Suzuki cross-coupling of 15a leading to biphenyl derivative 18 and hydrogenolysis to 19. Conditions: a) phenylboronic acid, Pd(PPh₃)₄, 2 M Na₂CO₃ aq, THF, 70 °C, 48 h; b) H₂, Pd/C, iPrOH, THF, rt, 24 h.

Scheme 9: Synthesis of N-benzylated p-terphenyl derivative 21 by Suzuki cross-coupling of 12a with 20 and subsequent reduction with zinc. Conditions: a) Pd(PPh₃)₄, 2 M Na₂CO₃ aq, THF/DMF (8:2), 70 °C, 48 h; b) Zn, AcOH, THF, 60 °C, 18 h.

Nevertheless, the target compound **22** was isolated in a moderate yield of 59%.

As a milder alternative, samarium diiodide was examined for the N-O bond cleavage [40-45]. With this selective reagent a TBS protected aminopyran dimer 23 was expected that should be well soluble in organic solvents and therefore more suitable for subsequent transformations (Scheme 10). Disappointingly, a samarium diiodide solution converted compound 21 in a complex reaction mixture. For a better understanding of this unexpected result the reductive cleavage was examined with the simpler bicyclic 1,2-oxazine derivative 12a. Here the unexpected bicyclic compound 24 was isolated as major product in 79% yield together with the desired aminopyran derivative 25 in 14% yield. It was observed by ¹H NMR spectroscopy that aminopyran 25 is not stable and slowly cyclizes to 24; after two days in CDCl₃ solution approximately 20% of aminopyran 25 were converted into 24. This result indicates that the N-O bond cleavage of 12a preceded as expected, but that the produced amino group of the pyran ring seems to be in close proximity to the C-5 hydroxy group and leads to a nucleophilic substitution

under formation of the pyrrolidine moiety. This process is possibly promoted by samarium(III) which can act as Lewis acid and by the steric demanding TBS group which decreases the distance between the two functional groups. This unexpected side reaction leading to 24 indicates that in the above mentioned unclean reaction of samarium diiodide with dimer 21 similar complications may lead to the observed mixture.

To overcome these difficulties dimer 21 was deprotected with tetra-n-butylammonium fluoride giving the poorly soluble polyhydroxylated compound 26 (Scheme 11). Due to the amphiphilic character this compound was only soluble in pyridine that makes purification and subsequent reactions fairly difficult. The conversion of the deprotection step was high but the yield after purification was only 30%. The reduction with samarium diiodide was then performed in a methanol/tetrahydrofuran mixture in which compound 26 was scarcely soluble. Gratifyingly, after one hour reaction time and purification by column chromatography aminopyran 22 was isolated in 30% yield. Interestingly, no cyclization product similar to 24 was detected in this case.

Scheme 10: Attempted reductive cleavage of the N–O bond of compound 21 by samarium diiodide and reaction of 12a. Conditions: a) Sml₂ (0.09 M in THF), MeOH, rt, 60 min; b) Sml₂ (0.1 M in THF), THF, rt, 45 min.

Scheme 11: Deprotection of compound 21 and samarium diiodide-mediated reaction of 26. Conditions: a) TBAF, THF, rt, 2 d; b) Sml₂ (0.09 M in THF), MeOH, rt, 60 min.

We also investigated the Suzuki cross-coupling of bicyclic compound 16 with 1,4-phenylenediboronic acid (20) (Scheme 12). Although the trityl group is quite far away from the reacting bromo substituent it led to longer reaction times and slightly lower yields, but the expected product 27 was obtained in 51% yield as a well soluble compound.

The trityl protective groups enabled the deprotection of dimer **27** in one step. The *O*-trityl and *N*-benzyl groups were removed by hydrogenolysis under acidic conditions to obtain a mixture of compounds **28** and **29** (Scheme 13). As solvent a 3:2 mixture of isopropanol/hexafluoro-2-propanol was used in order to combine high polarity with product solubility and to avoid side

Scheme 12: Suzuki cross-coupling of compound 16. Conditions: Pd(PPh₃)₂Cl₂, 2 M Na₂CO₃, DMF, 80 °C, 3 d.

Scheme 13: Hydrogenolysis of compound 27 and samarium diiodide-mediated reaction leading to compounds 30 and 31. Conditions: a) H₂, Pd/C, TFA, HFIP, iPrOH, 8 h, rt; b) Sml₂ (0.1 M in THF), MeOH, 30 min, rt. HFIP = hexafluoro-2-propanol.

product formation [26]. The ratio of mono-benzylated **28** and fully deprotected *p*-terphenyl derivative **29** was 59:41 as confirmed by ¹H NMR spectroscopy of the crude product. The reaction mixture was directly filtered through a pad of Celite[®] and the solvents were removed in vacuo to provide a crude product that was used for the subsequent samarium diiodide-mediated reduction without any purification. This reaction proceeded smoothly and furnished the very polar compounds **30** and **31**. Removal of the formed samarium salts by size exclusion chromatography provided the two divalent carbohydrate mimetics in very good overall yield. The considerably better yield in this samarium diiodide-mediated reaction is probably due to the good solubility of compounds **28** and **29** in methanol/THF.

In summary, by optimizing the protective group strategy and the reductive cleavage methods we were able to prepare the desired rigid *p*-terphenyl-linked carbohydrate mimetic **30** in twelve steps starting from D-isoascorbic acid, but in only six steps with respect to crucial intermediate **4**. The overall yields of 6% or 13% are quite respectable. The distance between the two terminal amino groups is in the range of 2.0 nm (according to optimized molecular geometry obtained by MM2 calculations performed by ChemBio3D Ultra 11.0 from ChemBioOffice 2008).

Conclusion

We successfully established methods for the efficient preparation of phenyl-substituted aminopyrans and rigid divalent

p-terphenyl-linked C-aryl glycoside using Suzuki crosscouplings as key method. Starting from the D-isoascorbic acidderived diol 8, which was converted into the corresponding p-bromophenyl-substituted (Z)-nitrone, a stereoselective [3 + 3]-cyclization with lithiated TMSE-allene provided the required 1,2-oxazine 4 in six steps with an overall yield of 46%. Alternatively, this 1,2-oxazine could be obtained under the formation of a 4-bromophenyl-1',3'-dioxolane moiety from diol 10 (actually derived from D-mannitol) which is a promising route to differently substituted 1,2-oxazines, introducing the dioxolane substituent at a late stage. The Lewis acid-induced rearrangement of the 1,2-oxazine 4 afforded the bicyclic ketone 11 which can be regarded as an internally protected amino sugar. After subsequent reduction of the carbonyl group the resulting bicyclic compound 12 was used as substrate for Suzuki cross-couplings to form a biphenyl-substituted aminopyran or rigid C-aryl dimers in good yields. These p-terphenyllinked aminopyran derivatives 22, 30 and 31 have a dimension of approximately 2.0 nm.

p-Bromophenyl-substituted intermediates such as **15** or **16** are also useful precursors for the synthesis of other *C*-aryl glycosides with potential biological activity. By simple hydrogenolysis of the bicyclic compounds, three new aminopyrans could be synthesized. Different methods for N–O bond cleavage like palladium-catalyzed hydrogenolysis, zinc/acid or samarium diiodide were tested. For each of the substrates the suitable method has to be found. By the N–O bond cleavage of compound **12a** with samarium diiodide an unexpected bicyclic

pyrrolidine derivative **24** was isolated. The prepared unnatural *C*-aryl glycosides could find applications in medicinal chemistry, e.g., as selectin inhibitors. After O-sulfation the biological activity of these divalent compounds will be studied together with that of related carbohydrate mimetics.

Experimental

For general methods: See Supporting Information File 1

Supporting Information

Supporting Information File 1

Experimental procedures.

[http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-10-182-S1.pdf]

Supporting Information File 2

Characterization data ¹H NMR and ¹³C NMR spectra of synthesized compounds.

[http://www.beilstein-journals.org/bjoc/content/supplementary/1860-5397-10-182-S2.pdf]

Acknowledgements

This work was generously supported by the Deutsche Forschungsgemeinschaft (SFB 765) and by Bayer HealthCare. We acknowledge valuable discussions and help during preparation of the manuscript by Dr. R. Zimmer and Dr. L. Bouché.

References

- Werz, D. B.; Ranzinger, R.; Herget, S.; Adibekian, A.; von der Lieth, C.-W.; Seeberger, P. H. ACS Chem. Biol. 2007, 2, 685–691. doi:10.1021/cb700178s
- Adibekian, A.; Stallforth, P.; Hecht, M.-L.; Werz, D. B.; Gagneux, P.; Seeberger, P. H. Chem. Sci. 2011, 2, 337–344. doi:10.1039/c0sc00322k
- Hakomori, S.-i. Pure Appl. Chem. 1991, 63, 473–482. doi:10.1351/pac199163040473
- Kannagi, R.; Izawa, M.; Koike, T.; Miyazaki, K.; Kimura, N. Cancer Sci. 2004, 95, 377–384. doi:10.1111/j.1349-7006.2004.tb03219.x
- Lasky, L. A. Annu. Rev. Biochem. 1995, 64, 113–140. doi:10.1146/annurev.bi.64.070195.000553
- Sharon, N. FEBS Lett. 1987, 217, 145–157. doi:10.1016/0014-5793(87)80654-3
- Levy, D. E.; Tang, C. The Chemistry of C-glycosides; Elsevier Science Ltd.: Oxford. 1995.
- Sears, P.; Wong, C.-H. Angew. Chem. 1999, 111, 2446–2471. doi:10.1002/(SICI)1521-3757(19990816)111:16<2446::AID-ANGE2446 >3.0.CO:2-4
 - Angew. Chem., Int. Ed. **1999**, 38, 2300-2324. doi:10.1002/(sici)1521-3773(19990816)38:16<2300::aid-anie2300>3.0. co:2-6
- Koester, D. C.; Holkenbrink, A.; Werz, D. B. Synthesis 2010, 3217–3242. doi:10.1055/s-0030-1258228

- Koester, D. C.; Werz, D. B. Beilstein J. Org. Chem. 2012, 8, 675–682. doi:10.3762/bjoc.8.75
- Koester, D. C.; Leibeling, M.; Neufeld, R.; Werz, D. B. Org. Lett. 2010, 12, 3934–3937. doi:10.1021/ol101625p
- Ernst, B.; Magnani, J. L. Nat. Rev. Drug Discovery 2009, 8, 661–677. doi:10.1038/nrd2852
- Koester, D. C.; Kriemen, E.; Werz, D. B. Angew. Chem. 2013, 125, 3059–3063. doi:10.1002/ange.201209697
 Angew. Chem., Int. Ed. 2013, 52, 2985-2989. doi:10.1002/anie.201209697
- Mackeviča, J.; Ostrovskis, P.; Leffler, H.; Nilsson, U. J.; Rudovica, V.;
 Viksna, A.; Belyakov, S.; Turksa, M. ARKIVOC 2014, No. (iii), 90–112.
 doi:10.3998/ark.5550190.p008.402
- 15. Pertici, F.; de Mol, N. J.; Kemmink, J.; Pieters, R. J. *Chem.–Eur. J.* **2013**, *19*, 16923–16927. doi:10.1002/chem.201303463
- Pertici, F.; Varga, N.; van Duijn, A.; Rey-Carrizo, M.; Bernardi, A.;
 Pieters, R. J. *Beilstein J. Org. Chem.* **2013**, *9*, 215–222.
 doi:10.3762/bjoc.9.25
- Merino, P.; Tejero, T.; Marca, E.; Gomollón-Bel, F.; Delso, I.; Matute, R. *Heterocycles* 2012, *86*, 791–820. doi:10.3987/REV-12-SR(N)3
- Link, J. T.; Sorensen, B. K. Tetrahedron Lett. 2000, 41, 9213–9217. doi:10.1016/S0040-4039(00)01709-3
- 19. Johnson, C. R.; Johns, B. A. Synlett 1997, 1406-1408.
- Kotha, S.; Lahiri, K.; Kashinath, D. Tetrahedron 2002, 58, 9633–9695. doi:10.1016/S0040-4020(02)01188-2
- Kotha, S.; Lahiri, K. Eur. J. Org. Chem. 2007, 1221–1236. doi:10.1002/ejoc.200600519
- Bergeron-Brlek, M.; Giguère, D.; Shiao, T. C.; Saucier, C.; Roy, R. J. Org. Chem. 2012, 77, 2971–2977. doi:10.1021/jo2025652
- Bergeron-Brlek, M.; Shiao, T. C.; Trono, M. C.; Roy, R. Carbohydr. Res. 2011, 346, 1479–1489.
 doi:10.1016/j.carres.2011.03.041
- 24. Al-Harrasi, A.; Pfrengle, F.; Prisyazhnyuk, V.; Yekta, S.; Koóš, P.; Reissig, H.-U. *Chem.–Eur. J.* **2009**, *15*, 11632–11641. doi:10.1002/chem.200900996
- 25. Pfrengle, F.; Reissig, H.-U. *Chem.–Eur. J.* **2010**, *16*, 11915–11925. doi:10.1002/chem.201001060
- Dernedde, J.; Enders, S.; Reissig, H.-U.; Roskamp, M.; Schlecht, S.;
 Yekta, S. Chem. Commun. 2009, 932–934. doi:10.1039/b818263a
- 27. Roskamp, M.; Enders, S.; Pfrengle, F.; Yekta, S.; Dekaris, V.; Dernedde, J.; Reissig, H.-U.; Schlecht, S. *Org. Biomol. Chem.* **2011**, *9*, 7448–7456. doi:10.1039/c1ob05583f
- Bouché, L.; Kandziora, M.; Reissig, H.-U. Beilstein J. Org. Chem. 2014, 10, 213–223. doi:10.3762/bjoc.10.17
- Dondoni, A.; Franco, S.; Junquera, F.; Merchán, F. L.; Merino, P.;
 Tejero, T. Synth. Commun. 1994, 24, 2537–2550.
 doi:10.1080/00397919408010565
- Dondoni, A.; Franco, S.; Junquera, F.; Merchán, F. L.; Merino, P.;
 Tejero, T.; Bertolasi, V. Chem.–Eur. J. 1995, 1, 505–520.
 doi:10.1002/chem.19950010804
- 31. Helms, M.; Schade, W.; Pulz, R.; Watanabe, T.; Al-Harrasi, A.; Fišera, L.; Hlobilová, I.; Zahn, G.; Reissig, H.-U. Eur. J. Org. Chem. 2005, 1003–1019. doi:10.1002/ejoc.200400627
- Pfrengle, F.; Lentz, D.; Reissig, H.-U. Angew. Chem. 2009, 121, 3211–3215. doi:10.1002/ange.200805724
 Angew. Chem., Int. Ed. 2009, 48, 3165-3169. doi:10.1002/anie.200805724
- Pfrengle, F.; Dekaris, V.; Schefzig, L.; Zimmer, R.; Reissig, H.-U.
 Synlett 2008, 2965–2968. doi:10.1055/s-0028-1083628

- Manzo, E.; Barone, G.; Parrilli, M. Synlett 2000, 887–889. doi:10.1055/s-2000-6700
- Bressel, B.; Egart, B.; Al-Harrasi, A.; Pulz, R.; Reissig, H.-U.; Brüdgam, I. Eur. J. Org. Chem. 2008, 467–474. doi:10.1002/ejoc.200700792
- Boerner, A.; Krause, H. J. Prakt. Chem. 1990, 332, 307–312. doi:10.1002/prac.19903320305
- Neuss, N.; Koch, K. F.; Molloy, B. B.; Day, W.; Huckstep, L. L.;
 Dorman, D. E.; Roberts, J. D. Helv. Chim. Acta 1970, 53, 2314–2319.
 doi:10.1002/hlca.19700530846
- Pellissier, H. Org. Prep. Proced. Int. 2002, 34, 441–465. doi:10.1080/00304940209355764
- Wuts, P. G. M.; Jung, Y. W. J. Org. Chem. 1988, 53, 1957–1965. doi:10.1021/jo00244a021
- Keck, G. E.; McHardy, S. F.; Wager, T. T. Tetrahedron Lett. 1995, 36, 7419–7422. doi:10.1016/0040-4039(95)01557-4
- Chiara, J. L.; Destabel, C.; Gallego, P.; Marco-Contelles, J. J. Org. Chem. 1996, 61, 359–360. doi:10.1021/jo951571q
- 42. Jung, S. H.; Lee, J. E.; Koh, H. Y. *Bull. Korean Chem. Soc.* **1998**, *19*, 33–35.
- 43. Pulz, R.; Al-Harrasi, A.; Reissig, H.-U. *Org. Lett.* **2002,** *4*, 2353–2355. doi:10.1021/ol0260573
- 44. Revuelta, J.; Cicchi, S.; Brandi, A. *Tetrahedron Lett.* **2004**, *45*, 8375–8377. doi:10.1016/j.tetlet.2004.09.050
- 45. Jasiński, M.; Watanabe, T.; Reissig, H.-U. *Eur. J. Org. Chem.* **2013**, 605–610. doi:10.1002/ejoc.201201210

License and Terms

This is an Open Access article under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/2.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The license is subject to the *Beilstein Journal of Organic Chemistry* terms and conditions:

 $(\underline{http://www.beilstein\text{-}journals.org/bjoc})$

The definitive version of this article is the electronic one which can be found at:

doi:10.3762/bjoc.10.182