

SHORT REPORT

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Screening and analysis of genes expressed upon infection of broad bean with *Clover yellow vein virus* causing lethal necrosis

Kenji S Nakahara^{*}, Hiroaki Kitazawa, Go Atsumi, Sun Hee Choi, Yuji Suzuki and Ichiro Uyeda

Abstract

Clover yellow vein virus (CIYV) causes lethal systemic necrosis in legumes, including broad bean (*Vicia faba*) and pea (*Pisum sativum*). To identify host genes involved in necrotic symptom expression after CIYV infection, we screened cDNA fragments in which expression was changed in advance of necrotic symptom expression in broad bean (*V. faba* cv. Wase) using the differential display technique and secondarily with Northern blot analysis. Expression changes were confirmed in 20 genes, and the six that exhibited the most change were analyzed further. These six genes included a gene that encodes a putative nitrate-induced NOI protein (*VfNOI*), and another was homologous to an *Arabidopsis* gene that encodes a glycine- and proline-rich protein GPRP (*VfGPRP*). We recently reported that necrotic symptom development in CIYV-infected pea is associated with expression of salicylic acid (SA)-dependent pathogenesis-related (PR) proteins and requires SA-dependent host responses. Interestingly, *VfNOI* and *VfGPRP* expression was correlated with that of the putative SA-dependent PR proteins in CIYV-infected broad bean. However, broad bean infected with a recombinant CIYV expressing the *VfGPRP* protein showed weaker symptoms and less viral multiplication than that infected with CIYV expressing the GFP protein. These results imply that *VfGPRP* plays a role in defense against CIYV rather than in necrotic symptom expression.

Findings

Virus infections induce a number of disease symptoms such as developmental abnormalities, chlorosis, and necrosis in plants. These symptom expressions not only result from disruption of cell activity by virus infection but also from the complex interplay between plant and virus. Although it is broadly true that symptom severity, e.g., latent to lethal, depends on both viral pathogenicity and plant susceptibility, elucidating how these symptoms are expressed is an enormous challenge to plant virologists.

Recent studies regarding expression analysis of a large number of genes with microarrays have shown that sets of defense-related genes are expressed upon infection of a susceptible plant with several different viruses [1,2], suggesting that even susceptible plants recognize virus infection and mount defense responses. Ironically, in several plant-virus pathosystems, close relationships between defense responses to virus infection and severe necrotic

symptoms have been reported [3-7]; these defense responses are thought to involve a hypersensitive response (HR) and programmed cell death (PCD), which are mediated by recognition of virus infection through a resistance (R) gene, which encodes a nucleotide-binding-leucine-rich repeat (NB-LRR) protein that detects a specific viral avirulence (Avr) gene. Although the way in which these defense responses are associated with necrotic symptoms largely remains to be examined, one likely explanation is that when a plant bearing an R gene is infected with a virus possessing a corresponding Avr gene and fails to control the spread of the virus throughout the entire plant, HR-PCD would necessarily occur wherever the virus infects, resulting in systemic necrosis [4,8]. We have recently shown that the lethal systemic necrosis observed in legumes infected with *Clover yellow vein virus* (CIYV) is one such case involving defense responses [9,10]. Therefore, it is hardly surprising that the screening of host genes whose expression is elicited by CIYV infection identifies genes involved in defense responses related to HR-PCD. The objective of the screening is, however, aimed at

* Correspondence: knakahar@res.agr.hokudai.ac.jp
Pathogen-Plant Interactions Group, Plant Breeding Science, Research Faculty of Agriculture, Hokkaido University, Sapporo 060-8589, Japan

understanding the molecular mechanism of necrotic symptom expression in legumes infected with CIYVV.

A CIYVV culture derived from a plasmid pCIYVV-Pst/CP clone, which carries full-length cDNA of CIYVV-No. 30 strain [11], was used for the screening of host genes. CIYVV usually induced necrotic symptoms in non-inoculated upper leaves of broad bean (*Vicia faba* cv. Wase) 5-6 days post-inoculation (dpi) and killed the plant following systemic induction of necrotic symptoms in 2 weeks (Figure 1). Western blot analysis with anti-CIYVV CP antibody revealed that CIYVV reached upper leaves at 4 dpi (data not shown).

To identify host genes whose expression changes are not just the outcome of necrotic symptoms but are a cause of the symptoms, gene expression was analyzed in broad bean before the development of CIYVV necrotic symptoms. Total RNAs were extracted from non-inoculated upper leaves of CIYVV and mock-inoculated plants at 1, 2, and 4 dpi with Trizol reagent (Invitrogen, Carlsbad, CA, USA), basically in accordance with the manufacturer's instructions (Figure 1), and their cDNA populations were compared using differential display. Fluorescent cDNAs were generated from the extracted total RNA by reverse transcription (RT) coupled with polymerase chain reaction (PCR) using an Enzyme Set-FDD and Fluorescence Differential Display Kit (TaKaRa, Shiga, Japan) according to the manufacturer's instructions. PCR products amplified using 96 different binary combinations with four fluorescent downstream primers and 24 upstream primers were fractionated in electrophoresis on 6% Long Ranger acrylamide gel (BioWhittaker Molecular Applications, Rockland, ME, USA) containing 8 M Urea. The fluorescent amplified

cDNA fragments were visualized using an FMBIO II image analyzer (TaKaRa, Ohtsu, Japan).

We cut off 53 of the cDNA fragments (m fragments) specifically detected in samples from CIYVV-infected broad bean at 1 dpi and 210 of k fragments from samples at 2 and 4 dpi, when compared with mock-inoculated samples (Figure 1). These fragments were then re-amplified and cloned. When the nucleotide sequences of the clones were determined, 245 independent sequences among the 585 clones were identified. We secondarily screened 67 genes of the assertively selected sequences by Northern blot analysis using DIG-cRNA probes according to the manufacturer's protocol (Roche Diagnostics GmbH, Mannheim, Germany). Expression changes in 20 genes of CIYVV-infected broad bean were confirmed by at least two Northern blot analyses (data not shown). Six of the 20 genes were further analyzed because of their relatively large expression changes (Table 1).

Further analyses focused on the involvement of these genes in necrotic symptom expression. We recently found that necrotic symptom development in CIYVV-infected pea was associated with expression of genes encoding pathogenesis-related (PR) proteins and required salicylic acid (SA)-dependent host responses [9]. Consistent with this report [9], expression of *acidic chitinase (aCHI)* and *hsr203J* genes, which were putatively dependent on SA accumulation and cell death, respectively, was induced in leaves infected with CIYVV at 6 dpi (Figure 2A), and was therefore correlated with necrotic symptom expression in broad bean (Figure 1). In contrast, expression of the *basic chitinase (bCHI)* gene, which was putatively dependent on ethylene signaling, was negatively correlated with necrotic

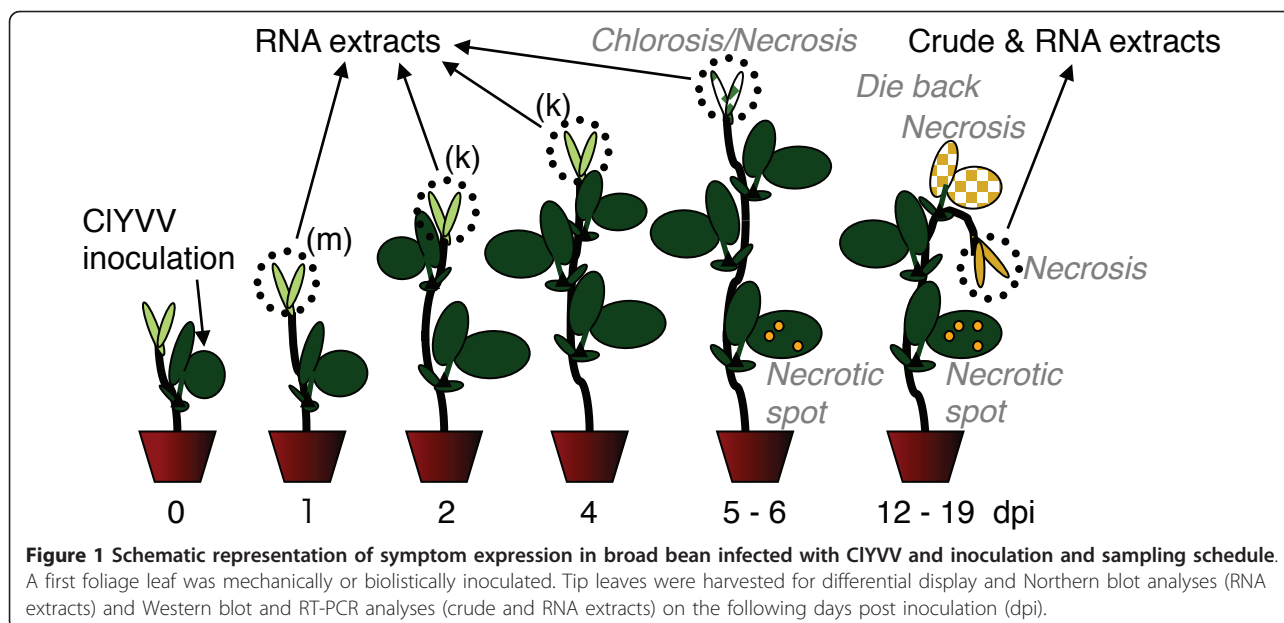


Table 1 The six cDNA fragments whose expressions changed upon infection of broad bean with CIYVV

1 Fragments (deduced amino acid sequence)	Homologous proteins (amino acid sequence)	Organisms	2 Identity (%)	Accession
k47 (199 aa)	Glutathione S-transferase GST 16 protein (221 aa)	<i>Glycine max</i>	66.8	AF243371
k58 (79 aa)	Putative NOI protein, nitrate-induced protein (79 aa)	<i>Arabidopsis thaliana</i>	70.9	AY065260
k158 (202 aa)	GPRP (177 aa)	<i>A. thaliana</i>	55.5	NM_123903
	AtGPRP2 (179 aa)	<i>A. thaliana</i>	59	NM_118040
m2 (112 aa)	Putative senescence-associated protein (279 aa)	<i>Pisum sativum</i>	84.7	AB049724
m4 (74 aa)	Putative thaumatin-like protein (224 aa)	<i>Medicago truncatula</i>	65.4	AM943532
m18 (34 aa)	Quinone reductase family protein (205 aa)	<i>A. thaliana</i>	68.4	NM_118861

1. The full length mRNAs of k58 and k158 [DDBJ: AB615378, AB615379] were obtained by 5' and 3' RACE and others seem to be partial.

2. The identity of each fragment with the homologous protein was calculated by GENETYX software (GENETYX, Tokyo, Japan).

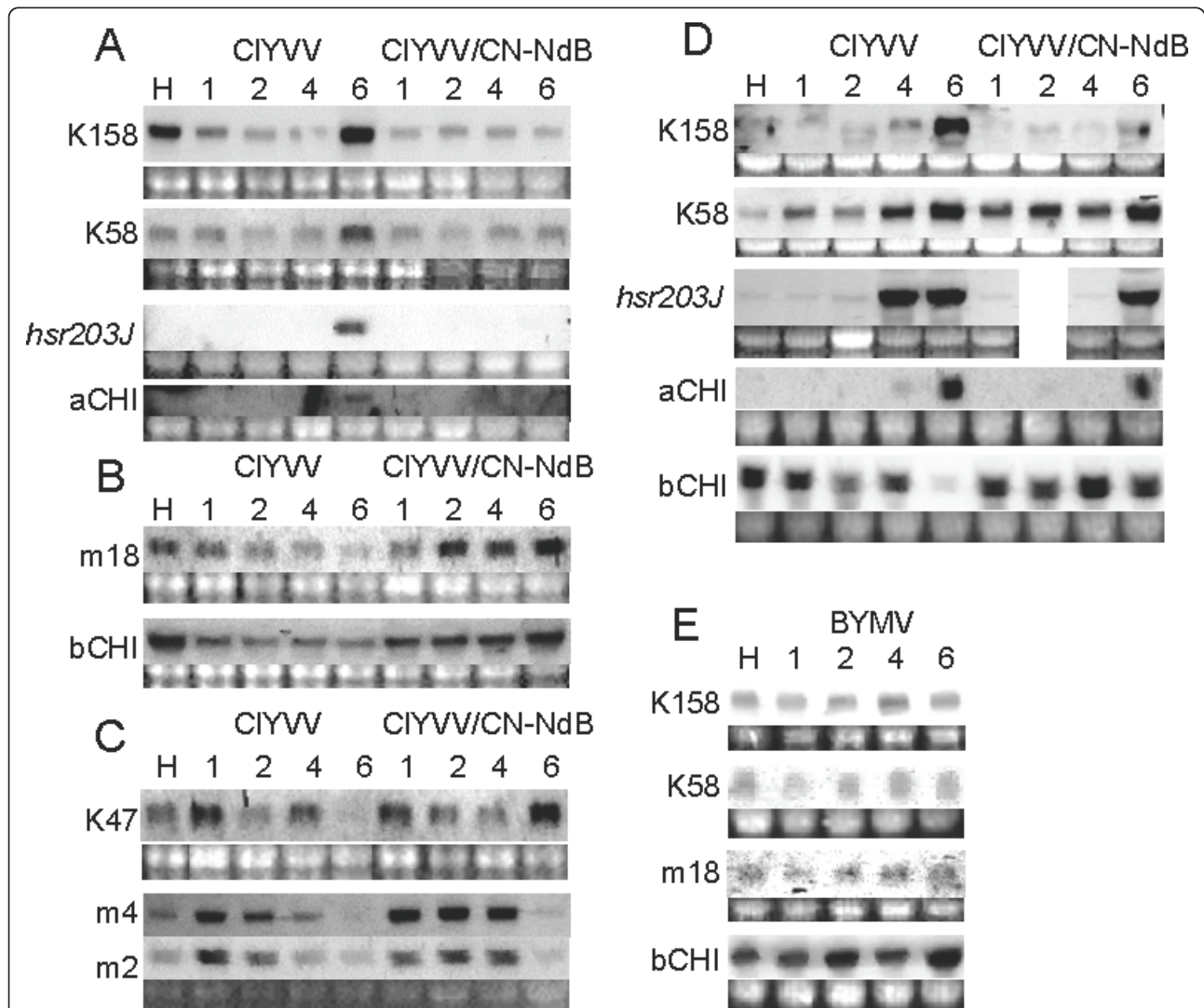


Figure 2 Time course of expression of genes identified in this study and pathogenesis-related (PR) genes in CIYVV-infected plants. mRNA accumulation was investigated by Northern blot analysis with DIG-labeled RNA probes. **(A, B, C)** RNAs were extracted from tip leaves of CIYVV- or CIYVV/CN-NdB-inoculated broad beans for Northern blot analysis (Figure 1). Hybridized probes were chemiluminescently detected with CDP-star reagent (New England Biolabs, Beverly, MA, USA). **(D)** The same experiment as in A, B, and C was performed with a pea line, PI 343958. **(E)** The same experiment as in D was performed with BYMV, which did not cause necrotic symptoms, but did result in mosaic symptoms in inoculated pea PI 343958.

symptom expression (Figure 2B). Then, when expression of the six genes identified in this study was compared with that of the PR genes in CIYVV-infected broad bean, k58 and k158 showed similar expression patterns to *aCHI* and *hsr203J*, and were thus correlated with necrotic symptom expression (Figure 2A). Consistently, k58, k158, *aCHI*, and *hsr203J* were not elicited in broad beans inoculated with a CIYVV mutant derived from an infectious plasmid clone pCIYVV/CN-NdB (CIYVV/CN-NdB) [12] that did not cause necrosis, but did cause weak chlorosis in broad bean. On the other hand, m18 expression was similar to that of *bCHI* and was negatively correlated with necrotic symptoms (Figure 2B). The other three genes, k47, m2, and m4, were elicited by CIYVV infection at 1 dpi, without reference to necrotic symptoms at earlier times. However, the expression of k47 was negatively correlated with necrotic symptoms at 6 dpi (Figure 2C).

The correlation of k58 and k158 expression with necrosis was further examined in a different host, pea (*Pisum sativum*). When the pea line PI 343958 expressed necrotic spots under either CIYVV or CIYVV/CN-NdB infection [9,10], k58 and k158 expression, as well as expression of *aCHI* and *hsr203J*, was also elicited by either infection (Figure 2D). Nevertheless, k58 and k158 were not elicited in the same pea line (PI 343958) inoculated with *Bean yellow mosaic virus* (BYMV)-CS [13], which is another potyvirus closely related to

CIYVV in terms of genome nucleotide sequences and host range, and does not cause necrosis, but does result in mosaic symptoms in pea (Figure 2E). Taken together, k58 and k158 expression was constantly correlated with *aCHI*, *hsr203J*, and necrotic symptom expression in legumes infected with CIYVV, raising the possibility of their involvement in the SA-dependent defense responses required for necrotic symptom expression.

In this study, the involvement of k158 in necrotic symptom expression in broad bean was examined. An 860-bp full-length cDNA of k158 was obtained by conventional 5' and 3' RACE methods and its nucleotide sequence was determined [DDBJ: AB615379]. A BLAST search with the sequence found that the k158 gene product had 55.5% identity at the amino acid sequence level with a glycine- and proline-rich protein (GPRP) in *Arabidopsis thaliana* [14]. GPRP is well conserved in plants, including rice, soybean, poplar, and spruce (Figure 3). Conserved motifs among GPRPs were found throughout the amino acid sequence: MGG at the N-terminal, YPP repeats, a central hydrophobic domain tentatively required for anchoring GPRP to a membrane [14], HG and YG repeats, and FKKWK at the C-terminal. Taking the sequence identity and conserved motifs into account, we conclude that k158 is the broad bean homolog of GPRP (*VfGPRP*). The *VfGPRP* association with necrotic symptoms was examined by ectopic

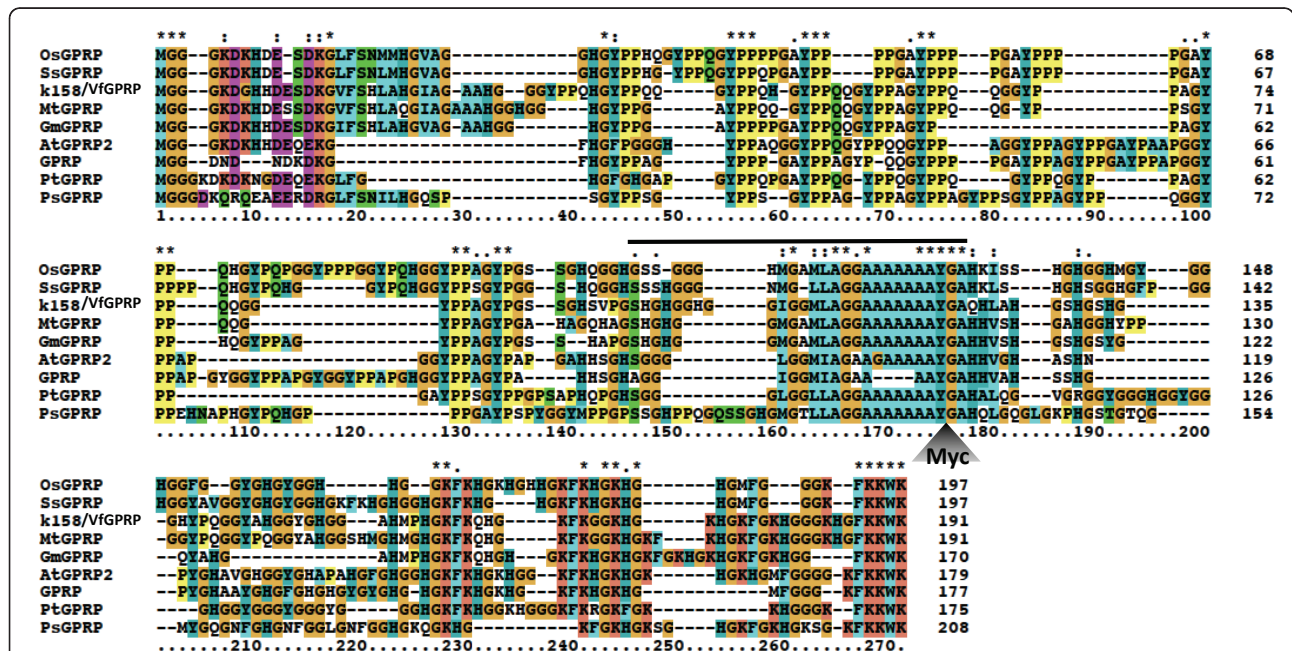


Figure 3 Alignment of deduced amino acid sequences of GPRPs in diverse plants. The GPRP sequences of *Oryza sativa* (OsGPRP) [DDBJ: NM_001061018], *Sporobolus stapfianus* (SsGPRP) [DDBJ:AJ242804] [16], broad bean (k158/VfGPRP, this study) [DDBJ:AB615379], *Medicago truncatula* (MtGPRP) [DFCI Medicago Gene Index TC:TC146460], *Glycine max* (GmGPRP) [DDBJ:BT089628], *Arabidopsis thaliana* (AtGPRP2), DDBJ: NM_118040], *A. thaliana* (GPRP) [DDBJ:NM_123903] [14], *Populus trichocarpa* (PtGPRP) [DDBJ:XM_002328813], and *Picea sitchensis* (PsGPRP) [DDBJ: EF083838] were aligned using the Clustal x program. The lined sequence was the central hydrophobic domain tentatively important for anchoring GPRP to a membrane [14]. A k158/VfGPRP mutant (VfGPRPmyc) used in Figure 4 was made by inserting a Myc tag sequence, EQKLISEEDL, into the position indicated by a Myc-labeled arrowhead.

expression of VfGPRP protein in CIYVV-infected cells using a recombinant infectious plasmid pCIYVV/VfGPRP derived from the virus expression vector pCIYVV [11]. We also made another recombinant pCIYVV/VfGPmycRP, which expressed a mutant VfGPRP protein with a Myc-tag peptide inserted into

the central hydrophobic domain to potentially disrupt VfGPRP function (Figure 3).

These two plasmids and pCIYVV/C3-S65T expressing GFP in infected cells [15] were biolistically inoculated into broad bean to compare their virulence (Figure 4A). CIYVV/C3-S65T caused lethal necrosis in infected broad

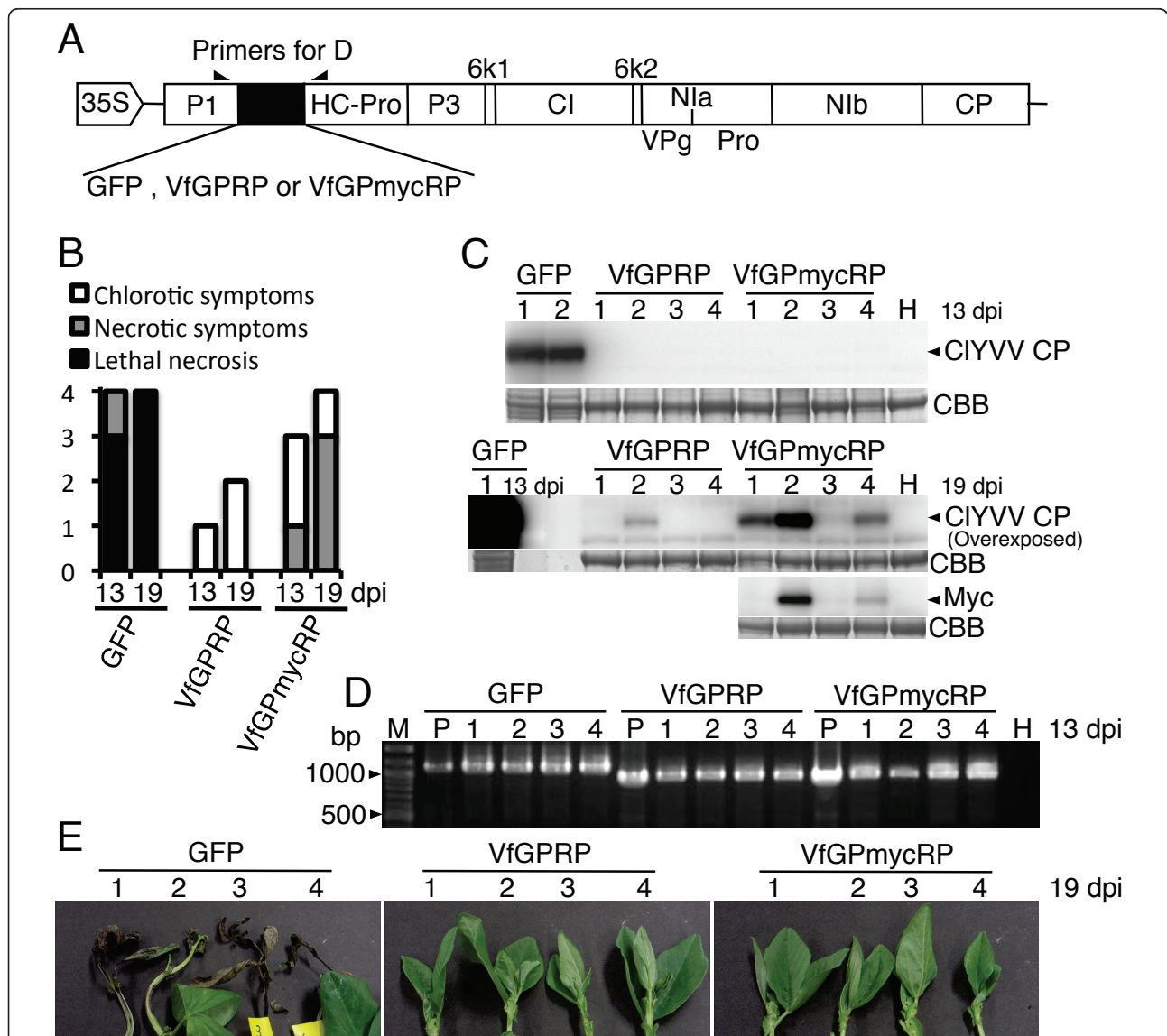


Figure 4 Effect of ectopically expressed GPRP on CIYVV virulence in infected broad bean. (A) Each of the recombinant CIYVV vectors possessing GFP, VfGPRP, or VfGPmycRP between P1 and HC-Pro cistrons was biolistically inoculated into broad bean leaves of four plants. **(B)** The reactions of broad beans inoculated with the recombinant CIYVV vectors, pCIYVV/C3-S65T (GFP), pCIYVV/VfGPRP (VfGPRP), or pCIYVV/VfGPmycRP (VfGPmycRP), were evaluated at 13 and 19 days post-inoculation (dpi). The vertical axis indicates the number of plants showing the indexed symptoms. **(C)** The coat protein (CP) of CIYVV was detected in extract from a third upper leaf from the inoculated leaf by Western blotting with anti-CP polyclonal antibodies at 13 and 19 dpi. Since an extract from CIYVV/C3-S65T was no longer available at 19 dpi because of lethal necrosis, the extract from 13 dpi was applied for comparison of CP accumulation. VfGPmycRP protein was also detected using anti-Myc monoclonal antibody (Sigma-Aldrich, St Louis, MO, USA). Lane H is extract from a healthy broad bean leaf. CBB stained gels are shown as loading controls. **(D)** Viral genome cDNA including that of transgenes was detected in RNA extracts from the same leaves as used for Western blotting by reverse transcription-coupled PCR (RT-PCR) using primers 5'-GATGTACACGTGTGTCCAATGTCTTTGG-3' and 5'-CTTATGGCATGCACATAATTGTTAACC-3', whose positions on the viral genome are shown in A. Lane P is PCR products amplified from the parental CIYVV vectors using the same primer pair. Lane H is the RT-PCR product with RNA extract from a healthy broad bean leaf. Lane M is a 100-bp ladder. **(E)** Photographs of tips of plants infected with recombinant CIYVVs.

beans about 2 weeks after inoculation and thus seems to have comparable virulence to wild-type CIYVV (Figure 4B and 4E). Compared with the CIYVV/C3-S65T-infected plants, drastically attenuated symptoms were observed in plants infected with CIYVV/VfGPRP. Additionally, accumulation of CIYVV/VfGPRP CP was far lower than that of CIYVV/C3-S65T (Figure 4C). RT-PCR nevertheless detected viral genomic cDNA fragments containing a transgene of an expected size in all tested RNA extracts from upper leaves (Figure 4D), indicating that CIYVV/VfGPRP at least infected and spread systemically and its progeny retained VfGPRP. Western blotting with anti-Myc antibody confirmed the expression of the VfGPRP protein in infected cells (Figure 4C). Taken together, these results suggest that VfGPRP represses CIYVV multiplication and/or spread in infected plants leading to attenuated symptoms. We cannot rule out the possibility of a *cis* effect of the inserted VfGPRP nucleotide and amino acid sequences on replication of the viral genome and/or translation of viral genes. However, the attenuated virulence was partly reversed by the additional insertion of a Myc tag peptide into the hydrophobic domain of VfGPRP on the recombinant CIYVV (CIYVV/VfGPRP-Myc; Figure 3, 4B, C, D and 4E), suggesting that the function of VfGPRP rather than its nucleotide and amino acid sequence is responsible for attenuation of CIYVV virulence. It is hard to obtain a mutant or transgenic broad bean in which VfGPRP expression is modified. Instead, we are preparing transgenic tobacco plants in which GPRP is overexpressed or knocked down by double-stranded RNA-mediated RNA silencing. We are now curious as to how GPRP is involved in not only the symptom expression of tobacco infected with poty- and other viruses but also internal viral replication and spread.

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Authors' contributions

IU designed and coordinated the study. HK and YS screened the broad bean genes using differential display techniques. HK performed cloning and sequencing of the identified genes and analyzed their expression. HK, GA, SHC, and KSN investigated the involvement of the identified genes in viral virulence in broad bean. KSN wrote the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

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References

1. Whitham SA, Quan S, Chang HS, Cooper B, Estes B, Zhu T, Wang X, Hou YM: Diverse RNA viruses elicit the expression of common sets of genes in susceptible *Arabidopsis thaliana* plants. *Plant J* 2003, **33**:271-283.

2. Whitham SA, Yang C, Goodin MM: Global impact: elucidating plant responses to viral infection. *Mol Plant Microbe Interact* 2006, **19**:1207-1215.
3. Komatsu K, Hashimoto M, Ozeki J, Yamaji Y, Maejima K, Senshu H, Himeno M, Okano Y, Kagiwada S, Namba S: Viral-induced systemic necrosis in plants involves both programmed cell death and the inhibition of viral multiplication, which are regulated by independent pathways. *Mol Plant Microbe Interact* 2010, **23**:283-293.
4. Dinesh-Kumar SP, Tham WH, Baker BJ: Structure-function analysis of the tobacco mosaic virus resistance gene *N*. *Proc Natl Acad Sci USA* 2000, **97**:14789-14794.
5. Hajimorad MR, Eggenberger AL, Hill JH: Loss and gain of elicitor function of *Soybean mosaic virus G7* provoking *Rsv1*-mediated lethal systemic hypersensitive response maps to *P3*. *J Virol* 2005, **79**:1215-1222.
6. Fisher ML, Kyle MM: Inheritance of resistance to potyviruses in *Phaseolus vulgaris* L.3. Cosegregation of phenotypically similar dominant responses to 9 potyviruses. *Theor Appl Genet* 1994, **89**:818-823.
7. Kim B, Masuta C, Matsuura H, Takahashi H, Inukai T: Veinal necrosis induced by turnip mosaic virus infection in *Arabidopsis* is a form of defense response accompanying HR-like cell death. *Mol Plant Microbe Interact* 2008, **21**:260-268.
8. Collmer CW, Marston MF, Taylor JC, Jahn M: The *I* gene of bean: a dosage-dependent allele conferring extreme resistance, hypersensitive resistance, or spreading vascular necrosis in response to the potyvirus *Bean common mosaic virus*. *Mol Plant Microbe Interact* 2000, **13**:1266-1270.
9. Atsumi G, Kagaya U, Kitazawa H, Nakahara KS, Uyeda I: Activation of the salicylic acid signaling pathway enhances *Clover yellow vein virus* virulence in susceptible pea cultivars. *Mol Plant Microbe Interact* 2009, **22**:166-175.
10. Ravelo G, Kagaya U, Inukai T, Sato M, Uyeda I: Genetic analysis of lethal tip necrosis induced by *Clover yellow vein virus* infection in pea. *J Gen Plant Pathol* 2007, **73**:59-65.
11. Masuta C, Yamana T, Tacahashi Y, Uyeda I, Sato M, Ueda S, Matsumura T: Development of clover yellow vein virus as an efficient, stable gene-expression system for legume species. *Plant J* 2000, **23**:539-546.
12. Yambao ML, Yagihashi H, Sekiguchi H, Sekiguchi T, Sasaki T, Sato M, Atsumi G, Tacahashi Y, Nakahara KS, Uyeda I: Point mutations in helper component protease of clover yellow vein virus are associated with the attenuation of RNA-silencing suppression activity and symptom expression in broad bean. *Arch Virol* 2008, **153**:105-115.
13. Kume H, Tanaka S, Murayama D: A strain of bean yellow mosaic virus isolated from mosaic-diseased red clover plant. *Mem Fac Agr Hokkaido Univ* 1970, **7**:435-449.
14. Marty I, Monfort A, Stiefel V, Ludevid D, Delseny M, Puigdomenech P: Molecular characterization of the gene coding for GPRP, a class of proteins rich in glycine and proline interacting with membranes in *Arabidopsis thaliana*. *Plant Mol Biol* 1996, **30**:625-636.
15. Sato M, Masuta C, Uyeda I: Natural resistance to *Clover yellow vein virus* in beans controlled by a single recessive locus. *Mol Plant Microbe Interact* 2003, **16**:994-1002.
16. Neale AD, Blomstedt CK, Bronson P, Le TN, Guthridge K, Evans J, Gaff DF, Hamill JD: The isolation of genes from the resurrection grass *Sporobolus stapfianus* which are induced during severe drought stress. *Plant Cell Environ* 2000, **23**:265-277.

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