

Acute Effects of Strength and Endurance Training on Bone Turnover Markers in Young Adults and Elderly Men

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Stunes AK, Brobakken CL, Sujan MAJ, Aagård N, Brevig MS, Wang E, Syversen U and Mosti MP (2022) Acute Effects of Strength and Endurance Training on Bone Turnover Markers in Young Adults and Elderly Men. Front. Endocrinol. 13:915241. doi: 10.3389/fendo.2022.915241 **Context:** Exercise is recognized as an important strategy to prevent bone loss, but its acute effects on bone turnover markers (BTMs) and related markers remain uncertain.

Objective: To assess the acute effects of two different exercise modes on BTMs and related markers in young adults of both sexes and elderly men.

Design, Setting, Participants: This was a three-group crossover within-subjects design study with a total of 53 participants—19 young women (aged 22–30), 20 young men (aged 21–30 years), and 14 elderly men (aged 63–74 years)—performing two different exercise sessions [strength training (ST) and high-intensity interval training (HIIT)] separated by 2 weeks, in a supervised laboratory setting.

Main Outcome Measures: Plasma volume-corrected serum measurements of the BTMs C-terminal telopeptide of type 1 collagen (CTX-I) and procollagen of type 1 N-terminal propeptide (P1NP), total osteocalcin (OC), sclerostin, and lipocalin-2 (LCN2) at baseline, immediately after, and 3 and 24 h after each of the two exercise modes were performed.

Results and Conclusion: Analyses revealed sex- and age-dependent differences in BTMs and related bone markers at baseline and time-, sex-, and age-dependent differences in response to exercise. No differences between exercise modes were observed for BTM response except for sclerostin in young men and LCN2 in elderly men. An acute, transient, and uniform increase in P1NP/CTX-1 ratio was found in young participants, demonstrating that beneficial skeletal effects on bone metabolism can be attained through both aerobic endurance and resistance exercise, although this effect seems to be attenuated with age. The acute effects of exercise on bone-related

1

biomarkers were generally blunted after 24 h, suggesting that persistent alterations following prolonged exercise interventions should be assessed at later time points.

Keywords: exercise, sclerostin, lipocalin-2, HIIT (high-intensity interval training), strength training, CTX-1, P1NP, osteocalcin

INTRODUCTION

Already in 1638, Galileo Galilei stated that "loading is required to preserve bone mass" (1). In the 1980s, the "mechanostat" theory, which describes the mechanism underlying the load-induced bone adaptation, was introduced by Harold Frost, who also identified the osteocyte as the "mechanostat" of bone (2, 3). Mechanical load causes dynamic fluid shifts in bone canalicular networks, which are sensed by bone-embedded osteocytes (4). The osteocytes convert the mechanical signal into biochemical signals transmitted to the effector cells, osteoclasts, and osteoblasts, impacting bone turnover (4).

Mechanical load can improve bone health and reduce the risk of osteoporosis; hence, physical activity and exercise are now regarded as beneficial interventions to prevent bone loss in the elderly (5) as well as in young subjects (6). In children, exercise/mechanical load is necessary to obtain an optimal peak bone mass to prevent future osteoporosis and fractures (7, 8). In the context of musculoskeletal health, strength training (ST) is widely advocated as a countermeasure for age- and lifestyle-related declines (9, 10). We have previously shown that maximal ST (MST) in hack squat provides a simple, low-volume training method to improve skeletal properties and neuromuscular function in young adults and postmenopausal women (11–13). Hack squat MST is a lowerextremity exercise that provides compressive load through the spine and the hip, which are sites particularly prone to osteoporotic fractures. Aerobic high-intensity interval training (HIIT) is a well-recognized intervention for cardiovascular and metabolic health that has become widely advocated in the last few years (14-16). HIIT is often performed as treadmill running, which has been shown to improve bone metabolism and bone mineral density (BMD), both acutely and long term (17, 18).

Most studies state a positive relationship between BMD and exercise in adults and the elderly and that exercise contributes to maintaining BMD and/or reducing bone loss (19). Conflicting results regarding the effects of training, as well as the load, intensity, and magnitude of necessary exercise, are present; however, high-impact activities (e.g., running, plyometrics, and jumping) are generally considered more osteogenic than lowerimpact activities (e.g., cycling and swimming). In impact exercises, such as running/HIIT, ground reaction forces constitute peak force exerted on the skeleton, whereas in lowimpact exercises, such as MST, the peak force is generated through muscle work (20, 21). Although HIIT and MST subsequently differ in skeletal loading characteristics, both training forms can be regarded as being osteogenic (12, 17, 18, 22). Circulating bone turnover markers (BTMs) reflect dynamic changes and provide a metabolic image of bone metabolism (23). The recommended BTMs in clinical practice are serum levels of resorption marker C-terminal telopeptide of type 1 collagen (CTX-I), a product of the enzymatic degradation of collagen fibers in bone, and total procollagen of type 1 N-terminal propeptide (P1NP), a sensitive marker for processes in proliferating osteoblast (23–25). Osteocalcin (OC), which is synthesized during bone formation by mature osteoblasts, is often regarded as a bone formation marker (26) but might also be released during bone resorption in both its intact and fragmented forms and may therefore indicate general bone remodeling (27).

Bone resorption and formation are orchestrated by many substances, and measurements of these may yield insight into the mechanisms for alterations in serum levels of BTMs. Sclerostin is an osteocyte-specific secreted potent inhibitor of bone formation *via* inhibition of the Wnt/ β -catenin pathway. Immobilization is associated with a rise in serum sclerostin and decreased BMD (28).

Lipocalin-2 (LCN2) has been proposed to act as an osteoblast mechano-responding gene, upregulated in osteoblasts during microgravity and correlated with poor osteoblast activity (29). Serum LCN2 was increased after 14 days in a bed rest study with male subjects, and Lcn2 expression was upregulated in bones in different mouse models of unloading, which could be counteracted by physical activity (30). In contrast, serum LCN2 was found to increase after high-intensity acute exercise in young and middle-aged male runners (31).

A systematic review of BTMs in subjects with osteoporosis revealed possible exercise benefits in terms of improving bone formation and decreasing bone resorption biomarkers (32). Whereas some studies have reported beneficial acute effects of exercise on BTMs (17, 22, 33), a direct comparison of heavy resistance training and aerobic running exercise has not yet been performed under strictly controlled conditions. Hack squat ST and treadmill HIIT are two well-studied training interventions that are highly relevant in the context of musculoskeletal and cardiometabolic health (11–16). Also, the acute response of these interventions on other mechano-responsive biomarkers, such as sclerostin and LCN2, has been less investigated, along with possible sex and age differences.

In the current study, we aimed to characterize the respective responses of hack squat ST and treadmill HIIT across age and gender. Specifically, we sought to investigate the acute exercise-induced effects of these two interventions on serum BTMs (CTX-1 and P1NP), OC, sclerostin, and LCN2 in men and women 21 to 30 years of age and elderly men 63 to 74 years of age.

METHODS

Participants and Study Design

This was a three-group crossover within-subjects design study. Recruitment of participants took place from September 2018 to February 2019, through posters on local gyms and at the University of Science and Technology, Trondheim, Norway. The study was approved by the Regional Ethics Committee for Medical and Health Research (REK2018/926) and was performed in accordance with the Declaration of Helsinki.

A total of 53 men and women volunteered to participate in this study (19 young women (age 22–31 years), 20 young men (age 21–30 years), and 14 elderly men (age 63–74 years). Inclusion criteria were as follows: 1) did not suffer any significant illness relevant to the study, 2) free from chronic conditions and injuries that could influence blood samples or prevent the participants in performing the physical tests, and 3) not taking dietary supplements known to affect bone metabolism. All participants agreed to take part in the study by signing a consent form.

A flowchart of the study is presented in **Figure 1**. Each participant performed two training sessions, ST and HIIT, separated by 2 weeks. During both training sessions, a total of four blood samples were collected from each participant: at baseline (pre), immediately (0–5 min, post), and 3 and 24 h after training. The training sessions were performed in a supervised laboratory setting, and participants were asked to fast overnight, avoid dietary supplements, and exercise 48 h before the sessions. At the first visit (between 08:00 and 09:00), participants signed an informed consent form and answered a questionnaire regarding lifestyle, medical history, and use of oral contraceptives (young women). Overnight fasting baseline



(ST_{baseline}) blood samples were drawn, and participants were tested for 1 repetition maximum (1RM) before the ST session, followed by post and 3 h after ST blood sampling. Between 08:00 and 10:00 the following day (second visit), blood samples were collected 24 h after ST, following an overnight fast. Body weight was obtained using an electronic scale, height was measured using a wall-mounted stadiometer, and body mass index (BMI; kg/m²) was calculated. Further, after participants had a meal, their maximal oxygen uptake (VO_{2max}) and maximal heart rate (HR) (HR_{max}) were measured. Two weeks later, between 08:00 and 09:00 (third visit), overnight fasting baseline (HIIT_{baseline}) blood samples were drawn, and participants took part in the HIIT session, followed by post and 3 h after HIIT blood sampling. Between 08:00 and 10:00 the following day (fourth visit), the blood samples after overnight fasting 24 h after HIIT were collected.

One Repetition Maximum

1RM was tested in a seated leg press apparatus (TechnoGym, Cesena, Italy), according to a protocol by the American College of Sports Medicine (ACSM) (34). The protocol consisted of two warm-up sets and a maximum of five attempts to determine the maximum load of a single dynamic leg press movement. Participants were instructed to perform a leg press with a knee joint angle of 90°. The first warm-up set consisted of 5–10 repetitions at 40%–60% of assumed 1RM. The second warmup set consisted of 3–5 repetitions at 60%–80% of assumed 1RM. After the second warm-up, the load was increased to the expected 1RM and the first attempt started. If the participant was able to complete one repetition with the given load, a new attempt with increased load was initiated after 3–5 min of rest, which was repeated until failure.

Maximal Oxygen Uptake and Maximal Heart Rate

Maximal oxygen uptake (\dot{VO}_{2max}) was measured on a treadmill (Woodway PPS Med, Weil am Rhein, Germany) with a Metamax II Portable device (Cortex, Leipzig, Germany). Participants warmed up for 10 min before the test. During the test, the minimum inclination on the treadmill was 5.3%, and the speed was increased every 2–3 min until the participant reached exhaustion. \dot{VO}_{2max} was accepted when it leveled off despite further increases in workload and verified by lactate levels ≥8.0 mmol/L and respiratory exchange ratio ≥1.05. Lactate was measured with a biochemical analyzer (EKF Biosen C-line Diagnostics, Barleben, Germany), and HR was assessed with a Polar M200 monitor (Kempele, Finland). Maximal HR (HR_{max}) was set as the highest HR registered + 3 beats per minute.

Blood Sampling and Biochemical Analyses

Blood samples were collected by standard venipuncture. Blood was collected in vacuum tubes, let sit for 30 min at room temperature, and centrifuged (3,000 $g/4^{\circ}C/10$ min) before serum was aliquoted and stored at $-80^{\circ}C$ until further analyses.

Acute Exercise and Bone Turnover

Serum albumin, creatinine, and total calcium (Ca) were analyzed with accredited analyses at the Department of Biochemistry, St. Olavs University Hospital, Trondheim, Norway. The bone turnover markers (BTMs) in serum resorption marker Cterminal telopeptide of type 1 collagen (CTX-I) and formation marker total procollagen of type 1 N-terminal propeptide (P1NP) were measured by accredited electrochemiluminescence immunoassays (Roche Cat# 11972308122, RRID:AB_2905599 and Roche Cat# 03141071190, RRID:AB 2782967, respectively), and total osteocalcin (OC) (intact (1-49) and large fragments (1-43)) by an accredited chemiluminescence assay (DiaSorin, Cat# 310950, RRID:AB_2917975;LIAISON® OSTEOCALCIN 1-49) at the Hormone Laboratory, Oslo University Hospital, Oslo, Norway. Serum sclerostin was analyzed by a human sclerostin ELISA kit (Biomedica Cat# BI-20492, RRID : AB_2894889), and serum LCN2 was analyzed by a human LCN2/NGAL DuoSet ELISA kit (R&D Systems Cat# DLCN20, RRID : AB_2894833). Analyses were done according to the manufacturers' instructions.

Strength Training

At the first visit, after the overnight fasting baseline_{ST} blood samples were drawn, participants were offered one pack of energy gel, containing pure carbohydrates (60 ml, 87 kcal) (Go Isotonic Energy, Science in Sport, London, UK) before the ST session. The training was conducted in the same leg press apparatus as used for the 1RM testing procedure, consisting of 4 sets of 8–10 repetitions until failure, with 2–3-min rest between sets. The load was 80% of the participants measured 1RM. The training session was performed with a slow eccentric movement down to a 90° knee angle and maximal intended velocity in the concentric movement. Participants' mean values of 1RM are presented in **Table 1**.

TABLE 1 Participants' baseline characteristics and measures

High-Intensity Interval Training

At the third visit, 2 weeks after the ST session, the participants took part in a HIIT session on a treadmill, as previously described (35). The participants were offered two packs of energy gel before this session. Participants warmed up for 10 min before performing 4×4 min intervals, with 3 min of active rest, on a treadmill with a 5.3% incline. During the intervals, the treadmill speed was set to a target intensity of 90%–95% of each participant's HR_{max}, and during active rest periods, an intensity was set corresponding to 70% of HR_{max}. Participants' mean values of HR_{max} are presented in **Table 1**.

Corrections for Plasma Volume Changes

Plasma volume changes (ΔPV) can occur during and after exercise due to transient fluid shifts (hemodilution and hemoconcentration) (36). The ΔPV can affect the interpretation of biochemical measurements in blood, and changes in the concentrations of biomarkers in blood should therefore be adjusted for ΔPV in exercise studies (36, 37). The standard method for calculation of % ΔPV has been the Dill and Costill equation (37), requiring hematocrit (Htc) and hemoglobin (Hb) measurements. Changes in serum total calcium levels after exercise have been found to correlate well with % ΔPV and can therefore be used as a hemoconcentration biomarker in exercise studies (38). In the current study, all serum analyses at post-test, 3 h, and 24 h were corrected for % ΔPV using % Δ in [Ca] from corresponding baseline values, before comparisons. The following equations for corrections were used:

A) %
$$\Delta PV_{t1} = 100 * \left(\frac{[Ca]_{t1} - [Ca]_{baseline}}{[Ca]_{baseline}}\right)$$

	I, Young women (n = 19)		II, Young men (n = 20)		III, Elderly men (n = 14)	
	Mean ± SD	Range	Mean ± SD	Range	Mean ± SD	Range
Age (years)	25.0 ± 2.3	22–30	25.5 ± 2.2	21–30	68.9 ± 3.7	63–74
Weight (kg)	65.6 ± 7.5	46.6-75.5	77.2 ± 10.3	62.8-103	79.2 ± 10.4	61.2-98.5
Height (cm)	169 ± 5.4	158–181	181 ± 6.2	168–193	180 ± 6.7	169–193
BMI (kg⋅m ⁻²)	23.0 ± 2.9	16–28	23.5 ± 2.5	19–28	24.4 ± 2.5	20–28
Ϋ́O _{2max}						
(L·min ^{−1})	3.3 ± 0.5	2.26-4.05	4.8 ± 0.6	3.76-5.85	3.5 ± 0.5	2.64-4.81
(mL·kg ⁻¹ ·min ⁻¹)	50.5 ± 6.3	35.7-60.3	62.6 ± 8.3	46.9-77.0	42.5 ± 5.4	34.0-51.3
HRmax (bmp)*	195 ± 9	179-210	196 ± 7	182-207	169 ± 17	143–197
1RM (kg)**	104 ± 25	64–140	134 ± 28	80-194	130 ± 17	98–164
sAlbumin (g/L)	44 ± 1.8	41-47	46 ± 1.8	42-49	43 ± 1.8	40-46
Reference***		36-48		36-48		36–45
sCreatinine (µmol/L)	73.8 ± 12.5	58-105	78.8 ± 7.8	63-90	84.3 ± 13.4	61-106
Reference***		45-90		60-105		60-105
sCalcium (nmol/L)	2.35 ± 0.04	2.26-2.46	2.40 ± 0.08	2.14-2.50	2.36 ± 0.06	2.22-2.44
Reference***		2.15-2.51		2.15-2.51		2.15-2.51

Data are in mean ± SD and range (minimum to maximum). Baseline serum levels are calculated mean values from ST_{baseline} and HIIT_{baseline}

BMI, body mass index; VO2max, maximal oxygen uptake; HRmax, maximal heart rate in beats per minute; 1RM, one repetition maximum; s, serum.

*Data of one young woman are missing.

**Data of one young woman and one young man are missing.

***Clinical reference ranges for validated analyses from Department of Biochemistry, St. Olavs University Hospital, Trondheim, Norway.

B) [bone marker]corrected_{t1}

$$= \begin{bmatrix} bone & marker \end{bmatrix}_{t1} * \left(1 - \left(\frac{\% \Delta PV_{t1}}{100} \right) \right)$$

where t_1 represents the time point for measurements (posttest, 3 h, or 24 h) after baseline in the applicable training mode. All serum analyses at post-tests and 3 and 24 h after the training intervention were performed with serum levels corrected for Δ PV as described (denoted as bone marker_c).

Statistical Analyses

Three missing values at individual time points (due to participant's absence) and one extreme outlier were replaced with the median value of the corresponding time point for analyses.

Data were assessed for normality using a Shapiro–Wilk test and checked for outliers by visual inspection by Q-Q plots. Skewness and kurtosis were analyzed by g1 and z-score.

A one-way ANOVA with a *post-hoc* Tukey's multiplecomparison test with computed multiplicity adjusted p-values for each comparison was used to examine group differences in mean baseline values between the two young groups and the two male groups (calculated as mean values from $ST_{baseline}$ and HIIT_{baseline}) of BTMs, OC, sclerostin, and LCN2.

Two-way repeated-measures ANOVA (two-way-RM-ANOVA) with Geisser–Greenhouse correction to adjust for lack of sphericity was used to examine time and group main effects for $CTX-1_{\odot}$ P1NP $_{\odot}$ P1NP $_{c}/CTX_{c}$ ratio, OC_{\odot} sclerostin $_{c}$, and LCN2 $_{c}$ in each training mode. In case of a significant simple main effect of time, a within-group *post-hoc* Tukey's multiple-comparison test with computed multiplicity adjusted p-values for each comparison was performed. Thereafter, a two-way-RM-ANOVA with Geisser–Greenhouse correction to adjust for lack of sphericity was used to examine the time and training mode main effects for each serum marker within each participant group. In case of significant training mode main effects, a Sidák multiple-comparison *post-hoc* test, with computed multiplicity adjusted p-values for each comparison for each comparison, was performed for comparison of training mode at each time point.

For serum markers that exhibited a significant simple main effect of time, Pearson's correlation coefficients were used to determine the relationship between percentage change in plasma volume-corrected serum marker concentrations from baseline and the participants' 1RM and $\dot{V}O_{2max}$.

All statistical analyses were performed using SPSS (IBM SPSS, Inc., version 27, 2020) and GraphPad Prism (GraphPad Software, Inc., version 9.2.0, 2021). Figures were made in GraphPad Prism.

RESULTS

Participants and Baseline Measurements

A flowchart of the study design and participants is shown in **Figure 1**. Values for serum BTMs and related markers were normally distributed, with the expectation of one extreme outlier value for measurements of $LCN2_c$, which were replaced with the median value of the corresponding time point for further analyses. The participants' baseline characteristics, $\dot{V}O_{2max}$, HR_{max}, and 1RM are presented in **Table 1**. All three groups of participants were healthy, meaning an average BMI within the accepted normal range and fasting mean baseline serum levels of albumin, creatinine, and calcium within the clinical reference ranges (**Table 1**). A total of 396 blood samples (n = 50 and 4 time points in ST and n = 49 and 4 time points in HIIT) were included in the analyses.

Serum values for each group and exercise mode of calcium [Ca] and the Δ [Ca] from the baseline used for correction of Δ PV (Δ PV) are shown in **Table 2**.

The mean baseline levels of BTMs, OC, sclerostin, and LCN2 are presented in **Figure 2**. Young men had higher baseline CTX-1 levels than young women (mean difference (MD) = 0.17 µg/L, **Figure 2A**). Elderly men had lower baseline BTMs (CTX-1 and P1NP) and OC levels as compared to the young male group (CTX-1, MD = 0.35 µg/L; P1NP, MD = 28.6 µg/L; OC, MD = 1.4 nmol/L, **Figures 2A–C**). Elderly men had higher baseline sclerostin and LCN2 levels as compared to young men (sclerostin, MD = 23.2 pmol/L; LCN2, MD = 17.4 µm/L, **Figures 2D, E**).

Sex/Age and Time Effects

There were significant main effects of group (sex/age)-by-time on plasma volume-corrected levels of CTX-1_c, P1NP_c/CTX-1_c ratio, and sclerostin_c in ST (F(6,141) = 5.48, 3.61, and 2.18, respectively, p < 0.05 for all) and for CTX-1_c and P1NP_c in HIIT (F(6,138) =

Calcium (nmol/L)	I, Young women		II, Young men		III, Elderly men	
	ST (n = 18)	HIIT (n = 17)	ST (n = 19)	HIIT (n = 19)	ST (n = 13)	HIIT (n = 13)
Baseline (BL)	2.37 ± 0.07	2.34 ± 0.05	2.39 ± 0.08	2.42 ± 0.08	2.36 ± 0.05	2.35 ± 0.08
Post test	2.43 ± 0.09	2.49 ± 0.06	2.52 ± 0.08	2.47 ± 0.08	2.44 ± 0.05	2.40 ± 0.07
%∆[Ca] from BL	6.62 ± 2.89	2.69 ± 2.40	5.25 ± 1.95	2.28 ± 1.97	3.61 ± 1.92	1.68 ± 1.93
3 h	2.40 ± 0.06	2.42 ± 0.05	2.45 ± 0.08	2.47 ± 0.09	2.38 ± 0.06	2.39 ± 0.08
%∆[Ca] from BL	3.59 ± 2.90	1.71 ± 1.96	2.26 ± '3.19	2.07 ± 2.7	0.92 ± 1.68	0.99 ± 1.90
24 h	2.38 ± 0.06	2.39 ± 0.07	2.41 ± 0.07	2.41 ± 0.07	2.34 ± 0.07	2.32 ± 0.06
%∆[Ca] from BL	2.10 ± 2.60	0.47 ± 2.05	0.52 ± 3.10	-0.52 ± 3.09	-0.95 ± 1.72	-1.37 ± 2.00

TABLE 2 | Participants' serum total calcium [Ca] and % [Ca] from baseline for correction of plasma volume changes.

Data are in mean \pm SD.

ST, strength training; HIIT, high-intensity interval training; BL, baseline.



7.81, p = 0.0001, and F(6,138) = 4.35, p = 0.0005, respectively). There was a significant simple main effect of group on all analyzed plasma volume-corrected serum markers (F(2,47) ranging from 3.40 to 39.9, p < 0.05 for all in ST and F(2, 44) ranging from 3.43 to 41.64, p < 0.05 for all in HIIT).

There was a significant simple main effect of time on most of the plasma volume-corrected serum markers in both exercise modes, and *post-hoc* Tukey multiple-comparison tests of the simple effect of time (from baseline) within-group and exercise mode showed the following.

Group I, Young Women

Serum $CTX-1_c$ decreased from baseline to post-test after ST training and after 3 h after both training modes, before

returning to baseline levels after 24 h (**Figure 3A**). Serum P1NP_c decreased from baseline to post-test and 3 h after ST training, increased from baseline to post-test after HIIT, and returned to baseline levels after 24 h for both exercise modes (**Figure 3B**). Serum P1NP_c/CTX-1_c ratio increased from ST baseline to post-test after 3 h and from HIIT baseline to post-test and returned to baseline levels after 24 h for both exercise modes (**Figure 3C**). Serum OC_c decreased from baseline ST to post-test after 3 and 24 h. After HIIT, serum OC_c decreased from baseline to 3 h but returned to baseline values after 24 h (**Figure 3D**). There were no significant time effects on the serum levels of neither sclerostin_c nor LCN2_c after either exercise mode in the young female group (**Figures 3E, F**).



FIGURE 3 | Plasma volume-corrected serum levels of C-terminal telopeptide of type 1 collagen (CTX-L) (**A**), total procollagen of type 1 N-terminal propeptide (P1NP_c) (**B**), PINP_c/CTX-1_c (**C**), total osteocalcin (OC_c) (**D**), sclerostin_c (**E**), and lipocalin-2 (LCN2_c) (**F**) in young women at baseline (pre), immediately after the strength training (ST) or high-intensity interval training (HIIT) sessions (post), and 3 and 24 h after exercise. Data are in mean \pm SEM. *p < 0.05, **p < 0.01, ***p < 0.001 versus baseline after ST and ^{##}p < 0.01, ^{###}p < 0.001 versus baseline after HIIT (two-way repeated-measures ANOVA and a within-group *post-hoc* Tukey's multiple-comparison test with computed multiplicity adjusted p-values for each comparison).

Group II, Young Men

There was a significant decrease in serum $CTX-1_c$ from ST baseline to post-test and 3 h and from HIIT baseline to 3 h (**Figure 4A**). Serum $CTX-1_c$ returned to baseline values after 24 h for both exercise modes. Serum $P1NP_c$ decreased from ST baseline to post-test and increased from HIIT to post-test (**Figure 4B**). After 24 h, $P1NP_c$ returned to baseline levels. The serum $P1NP_c/CTX-1_c$ ratio was increased from baseline after 3 h in ST and from HIIT baseline to post-test and after 24 h (**Figure 4C**). Serum OC_c decreased from ST baseline to post-test and after 3 h and returned to baseline to 3 h before returning to baseline values after 24 h (**Figure 4D**). Serum sclerostin_c increased from ST baseline to 24 h and increased from HIIT baseline to post-test

and after 24 h (**Figure 4E**). There were no significant time effects on serum LCN2_c after ST, but LCN2_c increased from HIIT baseline to post-test and was thereafter decreased compared to baseline values after 24 h (**Figure 4F**).

Group III, Elderly Men

There were no significant time effects on serum $CTX-1_c$, except for a significant increase from baseline to 24 h after HIIT (**Figure 5A**). No time effects were seen in serum $P1NP_c$ levels after ST, while there was a significant increase from HIIT baseline to post-test followed by a decrease after 3 h, before returning to baseline values after 24 h (**Figure 5B**). There were no significant time effects on serum $P1NP_c/CTX-1_c$ ratio or serum OC_c in neither of the two exercise modes (**Figures 5C, D**).



FIGURE 4 | Plasma volume-corrected serum levels of C-terminal telopeptide of type 1 collagen (CTX-l₂) (**A**), total procollagen of type 1 N-terminal propeptide (P1NP_c) (**B**), PINP_c/CTX-1_c (**C**), total osteocalcin (OC_c) (**D**), sclerostin_c (**E**), and lipocalin-2 (LCN2_c) (**F**) in young men at baseline (pre), immediately after the strength training (ST) or high-intensity interval training (HIIT) sessions (post), and 3 and 24 h after exercise. Data are in mean \pm SEM. *p < 0.05, **p < 0.01, ***p < 0.001 versus baseline after ST and [#]p < 0.05, ^{##}p < 0.001 versus baseline after HIIT (two-way repeated-measures ANOVA and a within-group *post-hoc* Tukey's multiple-comparison test with computed multiplicity adjusted p-values for each comparison). ^{SSS}p < 0.001 ST versus HIIT training modes (two-way repeated-measures ANOVA and a Šidák multiple-comparison *post-hoc* test for comparison of the two training modes at each time point).

Serum sclerostin_c was not affected after ST at any measured time points, unlike after HIIT, where sclerostin_c was significantly increased at post-test, after 3 and 24 h compared to baseline (**Figure 5E**). Serum LCN2_c was not significantly affected by time after exercise, except for a small increase after 24 h compared to ST baseline (**Figure 5F**).

Exercise Mode and Time Effects

Since there was a significant main effect of the group on all plasma volume-corrected serum markers, the effect of exercise mode by time was analyzed separately in the groups I, II, and III for each serum analysis.

Group I, Young Women:

The two-way RM ANOVA showed no significant main effects of the exercise modes ST and HIIT for any of the plasma volume-corrected measured markers (CTX-1_c, F(1,33) = 0.008, p = 0.93; P1NP_c, F(1,33) = 0.106, p = 0.75; P1NP_c/CTX-1_c ratio, F(1,33) = 0.224, p = 0.64; OC_c, F(1,33) = 0.842, p = 0.37; sclerostin, F (1,33) = 3.52, p = 0.07; LCN2_c, F(1,33) = 1.94, p = 0.17).

Group II, Young Men

There were no significant main effects of the exercise modes for any of the plasma volume-corrected serum markers, except for sclerostin_c (CTX-1_c, F(1,36) = 0.407, p = 0.53; $P1NP_{c}$, F(1,36) =

FIGURE 5 | Plasma volume-corrected serum levels of C-terminal telopeptide of type 1 collagen (CTX-I_c) (**A**), total procollagen of type 1 N-terminal propeptide (P1NP_c) (**B**), PINP_c/CTX-1_c (**C**), total osteocalcin (OC_c) (**D**), sclerostin_c (**E**), and lipocalin-2 (LCN2_c) (**F**) in elderly men at baseline (pre), immediately after the strength training (ST) or high-intensity interval training (HIIT) sessions (post), and 3 and 24 h after exercise. Data are in mean \pm SEM. *p < 0.05 versus baseline after ST and [#]p < 0.05, ^{##}p < 0.01 versus baseline after HIIT (two-way repeated-measures ANOVA and a within-group *post-hoc* Tukey's multiple-comparison test with computed multiplicity adjusted p-values for each comparison). ^{\$\$\$\$}p < 0.01, ^{\$\$\$\$}p < 0.001 ST versus HIIT training modes (two-way repeated-measures ANOVA and a Šidák multiple-comparison *post-hoc* test for comparison of the two training modes at each time point).

0.085, p = 0.77; $P1NP_c/CTX-1_c$ ratio, F(1,36) = 0.009, p = 0.92; OC_c , F(1,36) = 0.008, p = 0.93; sclerostin_c, F(1,36) = 5.837, p = 0.021; $LCN2_c$, F(1,36) = 0.874, p = 0.36).

A Sidák multiple-comparison *post-hoc* test for comparison of exercise mode at each time point showed significantly higher sclerostin_c in HIIT compared to ST at post-test immediately after exercise (MD 10.85 pmol/L) (**Figure 4E**).

Group III, Elderly Men

There was a significant main effect of exercise mode for LCN2_c, but not for CTX-1_c, P1NP_c, OC_c, and sclerostin_c (CTX-1_c, F (1,24) = 0.261, p = 0.61; P1NP_c, F(1,24) = 0.007, p = 0.93; P1NP_c/ CTX-1_c ratio, F(1,24) = 0.295, p = 0.66; OC_c, F(1,24) = 0.081, p = 0.78; sclerostin_c, F(1,24) = 3.243, p = 0.08; LCN2_c, F(1,24) = 3.567, p < 0.001).

A Śidák multiple-comparison *post-hoc* test for comparison of exercise mode at each time point showed significantly higher levels of LCN2_c in HIIT versus ST at post-test immediately after exercise (MD 27.21 μ g/L), as well as 3 h after exercise (MD 24.56 μ g/L) (**Figure 5F**).

Correlations

For plasma volume-corrected serum markers that exhibited a significant simple main effect of time, we analyzed the relationship between the percentage change and 1RM and $\dot{V}O_{2max}$. For young women, we found a negative correlation

between 1RM and CTX-1_c percentage changes from baseline to post-test and 3 h after ST (r = -0.571, p = 0.013, and r = -0.471, p = 0.049, respectively) and a positive correlation between 1RM and the percentage changes in P1NP_c from baseline to post-test after HIIT (r = 0.645, p = 0.013). The change in OC_c was negatively correlated to 1RM at post-test after ST (r = -0.664, p = 0.003) and positively correlated to 1RM at post-test and 3 h after HIIT (r = 0.596, p = 0.025 and r = 0.641, p = 0.014, respectively), while changes in sclerostin_c at post-test after HIIT were positively correlated to 1RM (r = 0.645, p = 0.013) in the young female group.

The plasma volume-corrected percentage changes in serum markers from baseline to post-test and 3 and 24 h after ST and HIIT, were not correlated to \dot{VO}_{2max} in neither of the three groups nor with 1RM in the young and elderly male groups.

DISCUSSION

In the current study, we investigated the acute effects of two wellrecognized training interventions for cardiometabolic and musculoskeletal health, namely, treadmill HIIT and hack squat ST. The respective responses to bone turnover and mechanoresponsive biomarkers in young men and women, and in elderly men, were explored.

The elderly men displayed significantly lower baseline values of the BTMs CTX-1 and P1NP, as well as OC, whereas LCN2 and sclerostin levels were higher, the latter by twofold, compared to those in the young male participants. Both training modes induced an acute decrease in plasma volume-corrected CTX-1_c in the young groups, whereas PINP_c levels were divergently affected, with HIIT promoting a transient increase and ST a transient decrease. Both training modes evoked a rise in the ratio of P1NP_c/CTX-1_c at post-test and/or after 3 h in the young groups, but not in elderly men. A significant increase in sclerostin_c was seen in young and elderly men after HIIT, which lasted for 24 h. Moreover, LCN2_c levels were elevated immediately after HIIT in young men but decreased after 24 h as compared to baseline. Correlations between simple time effects changes in BTMs and related bone markers and the participants' muscle strength and cardiovascular fitness were solely seen in the young female group.

Age and Sex Differences in Baseline Levels of Bone Turnover Markers, Osteocalcin, Sclerostin, and Lipocalin-2

In line with previous studies, we observed significantly lower baseline levels of BTMs and OC in elderly men compared to young men, reflecting an age-induced decline in bone turnover (39). This could partly be due to the high level of baseline sclerostin in this group, as sclerostin acts as a strong inhibitor of bone formation. The role of LCN2 in bone is not fully settled, but it has been shown to inhibit osteoclast generation (40) and is associated with reduced osteoblast activity (29), which could reduce bone turnover. Baseline levels of sclerostin and LCN2 were higher in the elderly men compared to the young male participants. Moreover, young men displayed higher levels of CTX-1 than young women, confirming previous studies (41, 42), while baseline levels of P1NP, OC, sclerostin, and LCN2 did not differ between the young groups.

Age and Sex Differences in Training-Induced Changes in Serum Markers

The alterations in BTMs after training were most pronounced in the young participants. This indicates that the acute effects of training on bone turnover decrease by age, as also suggested in a recent systematic review by Smith et al. (19). In both groups of young participants, we observed an immediate decrease in the serum bone resorption marker CTX-1_c after both training modes, which returned to baseline level after 24 h. A decline in P1NP_c levels was also seen in these groups after ST. Notably, P1NP_c levels increased transiently in all groups after HIIT. This was paralleled by a rise in sclerostin in the two male groups and by an increase in LCN2_c in the young male group. When calculating the ratio of P1NP_c/CTX-1_c, we observed an increase in both sexes, after both training modes, at post-test and/or after 3 h, suggesting a favorable acute effect of training on bone formation. In line with this, Whipple et al. found that resistance training with moderate intensity acutely reduced the serum bone resorption marker type I collagen N-telopeptide (NTX) for at least 8 h post-exercise in untrained young men (43). In contrast, Mieszkowski et al. reported that CTX-1 levels increased acutely after exercise in physically active young men, with no change in P1NP levels (44).

In contrast to the young participants, we observed no alterations after ST in elderly men, whereas HIIT induced altered CTX-1_c and P1NP_c levels. Moreover, no significant time effects were seen on P1NPc/CTX-1c ratio after either exercise mode. These results are partly in line with a study by Maïmoun et al., who found no significant change in CTX-1 in elderly participants after a maximal incremental exercise test (45); however, both elderly men and women were included, and P1NP was not measured in this study. In continuation of this, a more recent study reported beneficial effects of plyometric exercise on the P1NP/CTX-1 ratio in premenopausal women, an effect that was blunted in postmenopausal women (33). The latter study reported the effect of BTMs as a formation/ resorption ratio, using P1NP and CTX-1 (33). In the current study, we present P1NP/CTX-1 ratio, as part of a strategy to overcome the existing discrepancy between studies, as also suggested and used in several other reports (46-48).

We observed an acute decrease in OC_c at post-test and/or 3 h after both exercise modes in young participants, but not in the elderly men. Other studies show conflicting results regarding the effect of exercise on serum OC levels, as reviewed by Mohammad Rahimi et al. (49), making comparisons difficult. The interpretation of serum OC levels is further complicated by the fact that it exists in the circulation in two forms, namely, a carboxylated (cOC) form reflecting bone mineralization and an undercarboxylated (ucOC) form acting as a hormone, i.e., energy metabolism (50).

Few Differences in Exercise Mode Effects on Serum Markers

The hack squat ST and treadmill HIIT induced similar effects on BTMs in young women and men. Potential differences between these exercise modes in the mechanisms accounting for the BTM response are still unclear. Treadmill HIIT yields a substantially higher metabolic load than hack squat ST. In the context of bone strain rate, however, running exercise (as in HIIT) is commonly regarded as an impact exercise, where ground reaction forces constitute the peak forces exerted on the skeleton, whereas ST provides peak forces through muscle work, being referred to as low-impact exercise (20, 21). A substantial amount of research has demonstrated that distinct changes in BTMs can occur following both high- and low-impact exercise, despite differences in metabolic load (20-22, 51). The same has also been reported with BMD adaptations from long-term training interventions (18). Previous studies of acute exercise have reported similar BTM responses as the results of our study, following resistance, plyometric, and endurance exercise, when investigated separately (17, 22, 51). Collectively, these findings suggest that BTMs can be acutely secreted from bone cells, reflecting changes in bone homeostasis induced by both high- and low-impact exercise despite differences in the initiating stimulus.

The BTM response was essentially similar after ST and HIIT among the young participants, whereas the immediate response in serum sclerostin_c differed, with an increase in level after HIIT and a decrease after ST in young men. Kouvelitoti et al. investigated the acute response in BTMs after interval treadmill running versus interval cycling (low mechanical load) in young women and found no differences in CTX-1 but an acute increase in sclerostin immediately after both exercise modes, which contrasts with the findings in young women in the current study (52). Acute physical activity is postulated to inhibit sclerostin (53), and plyometric exercise has been found to cause an immediate and transient increase in serum sclerostin levels in men, but not in prepubertal boys and girls (54). We speculate that the high load and rapid execution (i.e., high rate of force development) in our ST model could be the decisive factor regarding the sclerostin response, as this type of execution is more similar to plyometric exercise. Still, sclerostin's response to exercise is not yet clear and requires further research, as conflicting reports exist regarding circulating sclerostin alterations, from elevation, no effect, and suppression (55).

In the elderly men, we observed a significant difference in $LCN2_c$ levels between the two exercise modes, as it was increased immediately and 3 h after HIIT before returning to baseline values but was increased at 24 h only after ST. Few studies have explored the effects of exercise on circulating LCN2 in humans, and since it is regarded as a mechanosensor (29), we find it interesting that the acute effects of the two exercise modes differ in the current study, even though in elderly men only. The increase in $LCN2_c$ after HIIT in the elderly men corresponds to the results from Ponzetti et al. describing increased serum LCN2 immediately after acute

high-intensity exercise in young and middle-aged male runners (31), suggesting that HIIT is a potent stimulus for LCN2 response. In the young male participants, we also found an acute simple time effect on $LCN2_c$, which increased immediately after HIIT and then decreased after 24 h, but these effects were not significantly different from ST.

Effect of Participants' Fitness Level and Acute Responses of Exercise

The participants included in this study all exhibited higher aerobic capacity, measured by VO_{2max} (between 8% and 18% above the average) for their sex and age compared to reference data from healthy Norwegian men and women aged 20-90 (56). The participants' fitness level might modulate BTMs and related bone marker responses to acute training. Mieszkowski et al. described that serum CTX-1 increased after a Wingate high-intensity anaerobic test in physically active men, but not in professional gymnastics athletes (44), while Scott et al. found no significant differences in β -CTX response between endurance-trained and recreationally active men after exhaustive running (57). Interestingly, we found no association between 1RM or VO_{2max} and changes in plasma volume-corrected BTMs and related markers among the male participants, indicating that in the current setting, neither muscle strength nor cardiovascular fitness affected the traininginduced changes in BTMs and related bone markers, in young and elderly men. In young women, muscle strength (1RM) was correlated with the significant main effect of time changes in all measured BTMs and related markers, but no correlations were detected with $\dot{V}O_{2max}$. The baseline levels of BTMs and related bone markers in the young women showed some variations between the two exercise modes' baseline levels, which could not be detected among the male participants. Circulating levels of CTX-1 might vary during the menstrual cycle (58), and the use of oral contraceptives is known to decrease bone turnover (59), which partly can explain the observed variations in baseline values in the female group. We did not collect data on the menstrual cycle in the current study, but ten of the 19 women included reported the use of oral contraceptives. When analyzing the baseline levels of BTMs between users/nonusers of oral contraceptives, we found no significant differences, possibly due to the relatively small sample size.

Implications for Long-Term Training Adaptations

We do not know how acute and transient changes to individual BTMs and related markers translate in the longterm regarding changes in BMD and microarchitecture and, ultimately, to bone strength and future fracture risk. A recent systematic review of the effects of acute exercise on BTMs in middle-aged and older adults concluded that acute exercise is an effective tool to modify BTMs, but the response appears to be dependent on exercise modality, intensity, age, and sex (19). The current study indicates that both high-intensity running exercise and lower-extremity ST can provide favorable effects on bone metabolism, at least in younger individuals, which agrees with our previous findings of improved skeletal properties following 12 weeks of hack squat MST (11–13). Further research is needed to unravel whether this also applies to treadmill HIIT. The observed acute effects in BTMs and related biomarkers returned to baseline levels after 24 h, indicating that persistent traininginduced changes from an intervention should be measured at later time points.

CONCLUSION

Despite being inherently different in skeletal loading characteristics, HIIT and ST induced similar effects on bone metabolism markers in young adults of both sexes. The two exercise modes did not diverge in BTM and OC_c responses but did for sclerostin_c in young men and $LCN2_c$ in elderly men. Baseline fitness measures were not correlated to changes in BTMs and related bone markers in the male groups, while muscle strength measured by 1RM was associated with changes in BTMs and related markers in the young female group.

Collectively, these findings demonstrate that beneficial skeletal effects on bone metabolism can be attained through both aerobic endurance and resistance exercise, although this effect seems to be attenuated with age. Mostly, these effects were blunted after 24 h, suggesting that persistent alterations following prolonged exercise interventions should be assessed at later time points.

STUDY STRENGTHS AND LIMITATIONS

Our study has several strengths. We included participants across sex and age. All baseline and 24 h after exercise blood samples were taken in the morning, after overnight fasting, to rule out diurnal variations. The energy intake was controlled and similar for all participants during exercise sessions, and sessions were supervised in a laboratory setting with close individual monitoring by qualified staff. All analyses of serum markers were performed with concentrations corrected for potential ΔPV , using changes in total calcium as a hemoconcentration biomarker. All participants were thoroughly tested regarding their muscular strength and cardiovascular fitness. The current study also has some limitations. We did not measure BMD in our participants, so we cannot rule out that the bone status by means of BMD could affect our results.

The baseline and 24-h blood samples were drawn in the morning after overnight fasting. Participants received equal amounts of carbohydrate energy gel before performing the exercise, and it cannot be ruled out that the serum BTMs measured at post-test and 3 h after exercise samples might have been affected by nutrition intake. Studies have demonstrated that intake of carbohydrates causes a decrease in serum BTMs, especially CTX-1, for up to 2 h after intake (60–62). However, Sale et al.

suggested that even though carbohydrate feeding during exercise reduces overall bone turnover in the hours following exercise, the balance between resorption and formation markers is maintained (63). A recent review stated circadian variation for BTMs CTX-1 and OC, with nighttime or early morning peak, but with less amplitude for P1NP and none for sclerostin (64). We tried to minimize the effects of circadian variation for BTMs by drawing blood at the same time of the day for all participants.

We measured serum tOC and were therefore unable to reveal a possible shift between the two forms of circulating OC (bone mineralization related carbonylated (cOC) and undercarboxylated (ucOC)). The optimal time for blood sampling for assessment of BTMs and related bone markers after training is not established. Ideally, blood tests should be taken every 30 min post-exercise for an extended period. For the feasibility of the study, we chose four time points for blood sampling (baseline, immediately after, 3, and 24 h after the exercise sessions). We cannot, however, rule out that we have "missed" the participants' individual peak responses in BTMs, OC, sclerostin, and LCN2 after exercise. Also, for the feasibility of the study and to minimize the number of visits for the participants, the order of the exercise sessions was not randomized. Preferably, both sexes should have been evaluated also in the elderly group, but we chose to prioritize men in an attempt to undermine the estrogen component related to aging. In the context of skeletal health, it could be argued that elderly women would be more relevant, although previous studies suggest that comparable results as in the present study can be expected (33, 65).

DATA AVAILABILITY STATEMENT

The original contributions presented in the study are included in the article/supplementary material. Further inquiries can be directed to the corresponding author.

ETHICS STATEMENT

The studies involving human participants were reviewed and approved by the Norwegian Regional Ethics Committee for Medical and Health Research (REK2018/926). The patients/ participants provided their written informed consent to participate in this study.

AUTHOR CONTRIBUTIONS

Conceptualization: MM, US, and EW. Data collection and curation: CB, NA, MB, MM, and EW. Analysis: MS and AS. Formal analyses, statistics, and visualization: AS. Funding acquisition: MM, US, and AS. Project administration: MM. Supervision: MM and EW. Validation: MM and AS. Writing original draft: AS. Writing—review and editing: all authors.

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REFERENCES

- Jastifer JR, Toledo-Pereyra LH, Gustafson PA. Galileo's Contribution to Modern Orthopaedics. J Invest Surg (2011) 24(4):141-4. doi: 10.3109/ 08941939.2011.585293
- 2. Frost HM. Bone "Mass" and the "Mechanostat": A Proposal. *Anat Rec* (1987) 219(1):1–9. doi: 10.1002/ar.1092190104
- Frost HM. The Mechanostat: A Proposed Pathogenic Mechanism of Osteoporoses and the Bone Mass Effects of Mechanical and Nonmechanical Agents. *Bone Mine* (1987) 2(2):73-85.
- 4. Bonewald LF. The Amazing Osteocyte. *J Bone Mine Res* (2011) 26(2):229–38. doi: 10.1002/jbmr.320
- Pinheiro MB, Oliveira J, Bauman A, Fairhall N, Kwok W, Sherrington C. Evidence on Physical Activity and Osteoporosis Prevention for People Aged 65+ Years: A Systematic Review to Inform the Who Guidelines on Physical Activity and Sedentary Behaviour. *Int J Behav Nutr Phys Act* (2020) 17(1):150. doi: 10.1186/s12966-020-01040-4
- Bielemann RM, Martinez-Mesa J, Gigante DP. Physical Activity During Life Course and Bone Mass: A Systematic Review of Methods and Findings From Cohort Studies With Young Adults. *BMC Musculoskelet Disord* (2013) 14:77. doi: 10.1186/1471-2474-14-77
- Min SK, Oh T, Kim SH, Cho J, Chung HY, Park DH, et al. Position Statement: Exercise Guidelines to Increase Peak Bone Mass in Adolescents. *J Bone Metab* (2019) 26(4):225–39. doi: 10.11005/jbm.2019.26.4.225
- Simões D, Craveiro V, Santos MP, Camões M, Pires B, Ramos E. The Effect of Impact Exercise on Bone Mineral Density: A Longitudinal Study on Non-Athlete Adolescents. *Bone* (2021) 153:116151. doi: 10.1016/j.bone.2021.116151
- Kohrt WM, Bloomfield SA, Little KD, Nelson ME, Yingling VR. American College of Sports M. American College of Sports Medicine Position Stand: Physical Activity and Bone Health. *Med Sci Sport Exerc* (2004) 36(11):1985– 96. doi: 10.1249/01.mss.0000142662.21767.58
- Garber CE, Blissmer B, Deschenes MR, Franklin BA, Lamonte MJ, Lee IM, et al. American College of Sports Medicine Position Stand. Quantity and Quality of Exercise for Developing and Maintaining Cardiorespiratory, Musculoskeletal, and Neuromotor Fitness in Apparently Healthy Adults: Guidance for Prescribing Exercise. *Med Sci Sport Exerc* (2011) 43(7):1334– 59. doi: 10.1249/MSS.0b013e318213fefb
- Mosti MP, Kaehler N, Stunes AK, Hoff J, Syversen U. Maximal Strength Training in Postmenopausal Women With Osteoporosis or Osteopenia. J Strength Cond Res (2013) 27(10):2879–86. doi: 10.1519/JSC.0b013e 318280d4e2
- Mosti MP, Carlsen T, Aas E, Hoff J, Stunes AK, Syversen U. Maximal Strength Training Improves Bone Mineral Density and Neuromuscular Performance in Young Adult Women. J Strength Cond Res (2014) 28(10):2935–45. doi: 10.1519/jsc.000000000000493
- Nygard M, Mosti MP, Brose L, Flemmen G, Stunes AK, Sorskar-Venaes A, et al. Maximal Strength Training Improves Musculoskeletal Health in Amphetamine Users in Clinical Treatment. Osteoporos Int (2018) 29 (10):2289–98. doi: 10.1007/s00198-018-4623-5
- Letnes JM, Berglund I, Johnson KE, Dalen H, Nes BM, Lydersen S, et al. Effect of 5 Years of Exercise Training on the Cardiovascular Risk Profile of Older Adults: The Generation 100 Randomized Trial. *Eur Heart J* (2021) 43 (21):2065–75. doi: 10.1093/eurheartj/ehab721
- Williams CJ, Gurd BJ, Bonafiglia JT, Voisin S, Li Z, Harvey N, et al. A Multi-Center Comparison of O2peak Trainability Between Interval Training and Moderate Intensity Continuous Training. *Front Physiol* (2019) 10:19. doi: 10.3389/fphys.2019.00019
- 16. Weston KS, Wisloff U, Coombes JS. High-Intensity Interval Training in Patients With Lifestyle-Induced Cardiometabolic Disease: A Systematic

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Review and Meta-Analysis. Br J Sport Med (2014) 48(16):1227-34. doi: 10.1136/bjsports-2013-092576

- Scott JP, Sale C, Greeves JP, Casey A, Dutton J, Fraser WD. The Role of Exercise Intensity in the Bone Metabolic Response to an Acute Bout of Weight-Bearing Exercise. J Appl Physiol (1985) (2011) 110(2):423-32. doi: 10.1152/japplphysiol.00764.2010
- Snow-Harter C, Bouxsein ML, Lewis BT, Carter DR, Marcus R. Effects of Resistance and Endurance Exercise on Bone Mineral Status of Young Women: A Randomized Exercise Intervention Trial. J Bone Mine Res (1992) 7(7):761– 9. doi: 10.1002/jbmr.5650070706
- Smith C, Tacey A, Mesinovic J, Scott D, Lin X, Brennan-Speranza TC, et al. The Effects of Acute Exercise on Bone Turnover Markers in Middle-Aged and Older Adults: A Systematic Review. *Bone* (2021) 143:115766. doi: 10.1016/ j.bone.2020.115766
- Kohrt WM, Barry DW, Schwartz RS. Muscle Forces or Gravity: What Predominates Mechanical Loading on Bone? *Med Sci Sport Exerc* (2009) 41 (11):2050–5. doi: 10.1249/MSS.0b013e3181a8c717
- 21. Judex S, Carlson KJ. Is Bone's Response to Mechanical Signals Dominated by Gravitational Loading? *Med Sci Sport Exerc* (2009) 41(11):2037-43. doi: 10.1249/MSS.0b013e3181a8c6e5
- Rogers RS, Dawson AW, Wang Z, Thyfault JP, Hinton PS. Acute Response of Plasma Markers of Bone Turnover to a Single Bout of Resistance Training or Plyometrics. J Appl Physiol (2011) 111(5):1353–60. doi: 10.1152/ japplphysiol.00333.2011
- 23. Shetty S, Kapoor N, Bondu JD, Thomas N, Paul TV. Bone Turnover Markers: Emerging Tool in the Management of Osteoporosis. *Indian J Endocrinol Metab* (2016) 20(6):846-52. doi: 10.4103/2230-8210.192914
- Delmas PD, Eastell R, Garnero P, Seibel MJ, Stepan J. The Use of Biochemical Markers of Bone Turnover in Osteoporosis. Committee of Scientific Advisors of the International Osteoporosis Foundation. *Osteoporos Int* (2000) 11 Suppl 6:S2–17. doi: 10.1007/s001980070002
- 25. Vasikaran S, Eastell R, Bruyère O, Foldes AJ, Garnero P, Griesmacher A, et al. Markers of Bone Turnover for the Prediction of Fracture Risk and Monitoring of Osteoporosis Treatment: A Need for International Reference Standards. *Osteoporos Int* (2011) 22(2):391–420. doi: 10.1007/s00198-010-1501-1
- Booth SL, Centi A, Smith SR, Gundberg C. The Role of Osteocalcin in Human Glucose Metabolism: Marker or Mediator? *Nat Rev Endocrinol* (2013) 9 (1):43–55. doi: 10.1038/nrendo.2012.201
- Dolan E, Varley I, Ackerman KE, Pereira RMR, Elliott-Sale KJ, Sale C. The Bone Metabolic Response to Exercise and Nutrition. *Exerc Sport Sci Rev* (2020) 48(2):49–58. doi: 10.1249/jes.00000000000215
- Spatz JM, Ellman R, Cloutier AM, Louis L, van Vliet M, Suva LJ, et al. Sclerostin Antibody Inhibits Skeletal Deterioration Due to Reduced Mechanical Loading. J Bone Mine Res (2013) 28(4):865–74. doi: 10.1002/ jbmr.1807
- Capulli M, Rufo A, Teti A, Rucci N. Global Transcriptome Analysis in Mouse Calvarial Osteoblasts Highlights Sets of Genes Regulated by Modeled Microgravity and Identifies a "Mechanoresponsive Osteoblast Gene Signature". J Cell Biochem (2009) 107(2):240–52. doi: 10.1002/ jcb.22120
- Rucci N, Capulli M, Piperni SG, Cappariello A, Lau P, Frings-Meuthen P, et al. Lipocalin 2: A New Mechanoresponding Gene Regulating Bone Homeostasis. *J Bone Mine Res* (2015) 30(2):357–68. doi: 10.1002/jbmr.2341
- Ponzetti M, Aielli F, Ucci A, Cappariello A, Lombardi G, Teti A, et al. Lipocalin 2 Increases After High-Intensity Exercise in Humans and Influences Muscle Gene Expression and Differentiation in Mice. J Cell Physiol (2021) 237 (1):551–65. doi: 10.1002/jcp.30501

- 32. Marini S, Barone G, Masini A, Dallolio L, Bragonzoni L, Longobucco Y, et al. The Effect of Physical Activity on Bone Biomarkers in People With Osteoporosis: A Systematic Review. Front Endocrinol (Lausanne) (2020)
- 11:585689. doi: 10.3389/fendo.2020.585689
 33. Nelson K, Kouvelioti R, Theocharidis A, Falk B, Tiidus P, Klentrou P. Osteokines and Bone Markers at Rest and Following Plyometric Exercise in Pre- and Postmenopausal Women. *BioMed Res Int* (2020) 2020:7917309. doi: 10.1155/2020/7917309
- Ratamess NA, Alvar BA, Evetoch TK, Housh TJ, Kibler WB, Kraemer WJ, et al. American College of Sports Medicine Position Stand. Progression Models in Resistance Training for Healthy Adults. *Med Sci Sport Exerc* (2009) 41(3):687–708. doi: 10.1249/MSS.0b013e3181915670
- Helgerud J, Høydal K, Wang E, Karlsen T, Berg P, Bjerkaas M, et al. Aerobic High-Intensity Intervals Improve Vo2max More Than Moderate Training. *Med Sci Sport Exerc* (2007) 39(4):665–71. doi: 10.1249/mss.0b013e3180304570
- 36. Kargotich S, Goodman C, Keast D, Morton AR. The Influence of Exercise-Induced Plasma Volume Changes on the Interpretation of Biochemical Parameters Used for Monitoring Exercise, Training and Sport. Sport Med (1998) 26(2):101–17. doi: 10.2165/00007256-199826020-00004
- Dill DB, Costill DL. Calculation of Percentage Changes in Volumes of Blood, Plasma, and Red Cells in Dehydration. J Appl Physiol (1974) 37(2):247–8. doi: 10.1152/jappl.1974.37.2.247
- Alis R, Sanchis-Gomar F, Primo-Carrau C, Lozano-Calve S, Dipalo M, Aloe R, et al. Hemoconcentration Induced by Exercise: Revisiting the Dill and Costill Equation. Scand J Med Sci Sport (2015) 25(6):e630–7. doi: 10.1111/sms.12393
- Szulc P, Kaufman JM, Delmas PD. Biochemical Assessment of Bone Turnover and Bone Fragility in Men. Osteoporos Int (2007) 18(11):1451–61. doi: 10.1007/s00198-007-0407-z
- Kim HJ, Yoon HJ, Yoon KA, Gwon MR, Jin Seong S, Suk K, et al. Lipocalin-2 Inhibits Osteoclast Formation by Suppressing the Proliferation and Differentiation of Osteoclast Lineage Cells. *Exp Cell Res* (2015) 334(2):301– 9. doi: 10.1016/j.yexcr.2015.03.008
- Szulc P, Garnero P, Munoz F, Marchand F, Delmas PD. Cross-Sectional Evaluation of Bone Metabolism in Men. J Bone Mine Res (2001) 16(9):1642– 50. doi: 10.1359/jbmr.2001.16.9.1642
- Verroken C, Zmierczak HG, Goemaere S, Kaufman JM, Lapauw B. Bone Turnover in Young Adult Men: Cross-Sectional Determinants and Associations With Prospectively Assessed Bone Loss. J Bone Mine Res (2018) 33(2):261–8. doi: 10.1002/jbmr.3303
- Whipple TJ, Le BH, Demers LM, Chinchilli VM, Petit MA, Sharkey N, et al. Acute Effects of Moderate Intensity Resistance Exercise on Bone Cell Activity. *Int J Sport Med* (2004) 25(7):496–501. doi: 10.1055/s-2004-820942
- 44. Mieszkowski J, Kochanowicz A, Piskorska E, Niespodziński B, Siódmiak J, Buśko K, et al. Serum Levels of Bone Formation and Resorption Markers in Relation to Vitamin D Status in Professional Gymnastics and Physically Active Men During Upper and Lower Body High-Intensity Exercise. J Int Soc Sport Nutr (2021) 18(1):29. doi: 10.1186/s12970-021-00430-8
- 45. Maïmoun L, Simar D, Malatesta D, Caillaud C, Peruchon E, Couret I, et al. Response of Bone Metabolism Related Hormones to a Single Session of Strenuous Exercise in Active Elderly Subjects. Br J Sport Med (2005) 39 (8):497–502. doi: 10.1136/bjsm.2004.013151
- Rantalainen T, Heinonen A, Linnamo V, Komi PV, Takala TE, Kainulainen H. Short-Term Bone Biochemical Response to a Single Bout of High-Impact Exercise. J Sport Sci Med (2009) 8(4):553–9.
- Fisher A, Srikusalanukul W, Fisher L, Smith PN. Lower Serum P1np/Bctx Ratio and Hypoalbuminemia Are Independently Associated With Osteoporotic Nonvertebral Fractures in Older Adults. *Clin Interv Aging* (2017) 12:1131–40. doi: 10.2147/cia.S141097
- Fisher A, Fisher L, Srikusalanukul W, Smith PN. Bone Turnover Status: Classification Model and Clinical Implications. *Int J Med Sci* (2018) 15 (4):323–38. doi: 10.7150/ijms.22747
- Mohammad Rahimi GR, Niyazi A, Alaee S. The Effect of Exercise Training on Osteocalcin, Adipocytokines, and Insulin Resistance: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. *Osteoporos Int* (2021) 32 (2):213–24. doi: 10.1007/s00198-020-05592-w
- 50. Sanchez-Enriquez S, Ballesteros-Gonzalez IT, Villafán-Bernal JR, Pascoe-Gonzalez S, Rivera-Leon EA, Bastidas-Ramirez BE, et al. Serum Levels of Undercarboxylated Osteocalcin Are Related to Cardiovascular Risk Factors in

Patients With Type 2 Diabetes Mellitus and Healthy Subjects. World J Diabetes (2017) 8(1):11-7. doi: 10.4239/wjd.v8.i1.11

- Scott JP, Sale C, Greeves JP, Casey A, Dutton J, Fraser WD. Effect of Exercise Intensity on the Cytokine Response to an Acute Bout of Running. *Med Sci Sport Exerc* (2011) 43(12):2297–306. doi: 10.1249/MSS.0b013e31822113a9
- Kouvelioti R, Kurgan N, Falk B, Ward WE, Josse AR, Klentrou P. Response of Sclerostin and Bone Turnover Markers to High Intensity Interval Exercise in Young Women: Does Impact Matter? *BioMed Res Int* (2018) 2018:4864952. doi: 10.1155/2018/4864952
- 53. Lombardi G, Barbaro M, Locatelli M, Banfi G. Novel Bone Metabolism-Associated Hormones: The Importance of the Pre-Analytical Phase for Understanding Their Physiological Roles. *Endocrine* (2017) 56(3):460–84. doi: 10.1007/s12020-017-1239-z
- Falk B, Klentrou P. Elevation in Sclerostin After Exercise: Is It Affected by Age and Sex? *Calcif Tissue Int* (2018) 102(3):380–1. doi: 10.1007/s00223-017-0348-2
- Gerosa L, Lombardi G. Bone-To-Brain: A Round Trip in the Adaptation to Mechanical Stimuli. Front Physiol (2021) 12:623893. doi: 10.3389/ fphys.2021.623893
- Loe H, Rognmo Ø, Saltin B, Wisløff U. Aerobic Capacity Reference Data in 3816 Healthy Men and Women 20-90 Years. *PloS One* (2013) 8(5):e64319. doi: 10.1371/journal.pone.0064319
- Scott JP, Sale C, Greeves JP, Casey A, Dutton J, Fraser WD. The Effect of Training Status on the Metabolic Response of Bone to an Acute Bout of Exhaustive Treadmill Running. J Clin Endocrinol Metab (2010) 95(8):3918– 25. doi: 10.1210/jc.2009-2516
- Gass ML, Kagan R, Kohles JD, Martens MG. Bone Turnover Marker Profile in Relation to the Menstrual Cycle of Premenopausal Healthy Women. *Menopause* (2008) 15(4 Pt 1):667–75. doi: 10.1097/gme.0b013e31815f8917
- Garnero P, Sornay-Rendu E, Delmas PD. Decreased Bone Turnover in Oral Contraceptive Users. *Bone* (1995) 16(5):499–503. doi: 10.1016/8756-3282(95) 00075-0
- Paldánius PM, Ivaska KK, Hovi P, Andersson S, Väänänen HK, Kajantie E, et al. The Effect of Oral Glucose Tolerance Test on Serum Osteocalcin and Bone Turnover Markers in Young Adults. *Calcif Tissue Int* (2012) 90(2):90–5. doi: 10.1007/s00223-011-9551-8
- Clowes JA, Hannon RA, Yap TS, Hoyle NR, Blumsohn A, Eastell R. Effect of Feeding on Bone Turnover Markers and Its Impact on Biological Variability of Measurements. *Bone* (2002) 30(6):886–90. doi: 10.1016/s8756-3282(02)00728-7
- Clowes JA, Allen HC, Prentis DM, Eastell R, Blumsohn A. Octreotide Abolishes the Acute Decrease in Bone Turnover in Response to Oral Glucose. J Clin Endocrinol Metab (2003) 88(10):4867–73. doi: 10.1210/jc.2002-021447
- Sale C, Varley I, Jones TW, James RM, Tang JC, Fraser WD, et al. Effect of Carbohydrate Feeding on the Bone Metabolic Response to Running. J Appl Physiol (2015) 119(7):824–30. doi: 10.1152/japplphysiol.00241.2015
- 64. Diemar SS, Dahl SS, West AS, Simonsen SA, Iversen HK, Jørgensen NR. A Systematic Review of the Circadian Rhythm of Bone Markers in Blood. *Calcif Tissue Int* (2022). doi: 10.1007/s00223-022-00965-1
- Vincent KR, Braith RW. Resistance Exercise and Bone Turnover in Elderly Men and Women. *Med Sci Sport Exerc* (2002) 34(1):17–23. doi: 10.1097/ 00005768-200201000-00004

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