

Supplementary Materials

In vitro characterization of the phage lysis protein MS2-L

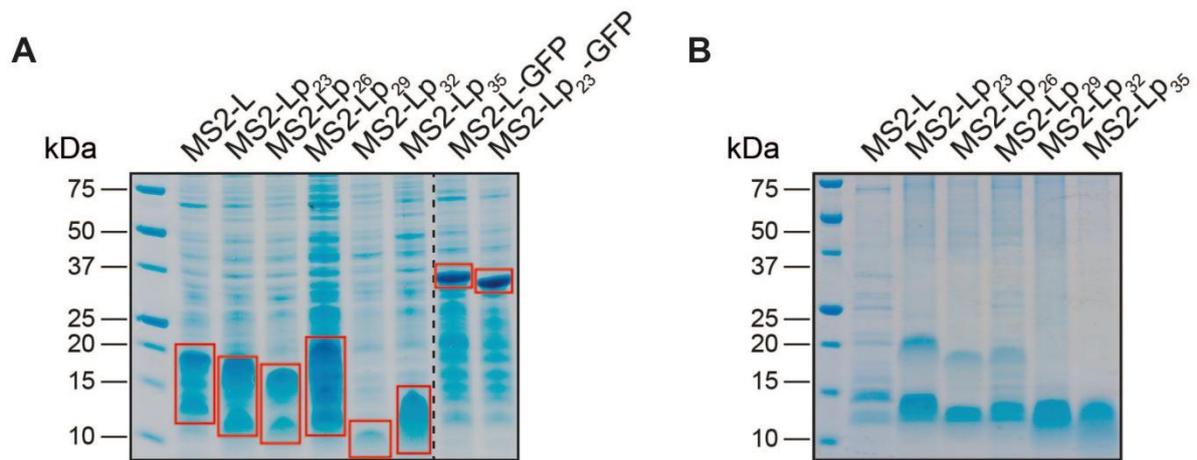
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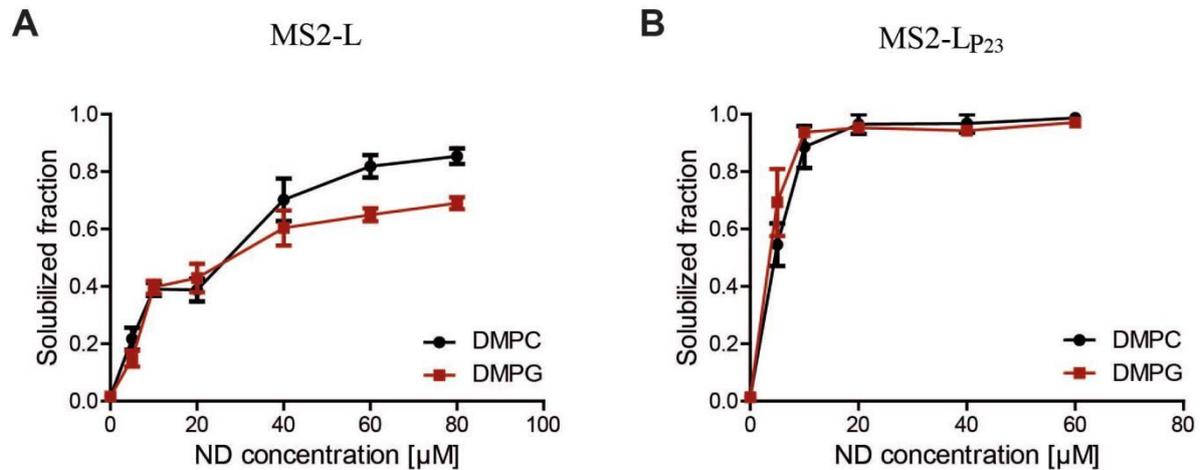
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MKTRFPQQSQ QTPASTNRRR PFKHEDYPCR RQQRSSLYV LIFLAIFLSK FTNQLLLSLL EAVIRTVTTL QQLLT MS2-L
      MKHEDYPCR RQQRSSLYV LIFLAIFLSK FTNQLLLSLL EAVIRTVTTL QQLLT MS2-Lp23
      MDYPCR RQQRSSLYV LIFLAIFLSK FTNQLLLSLL EAVIRTVTTL QQLLT MS2-Lp26
      MCR RQQRSSLYV LIFLAIFLSK FTNQLLLSLL EAVIRTVTTL QQLLT MS2-Lp29
      MQQRSSLYV LIFLAIFLSK FTNQLLLSLL EAVIRTVTTL QQLLT MS2-Lp32
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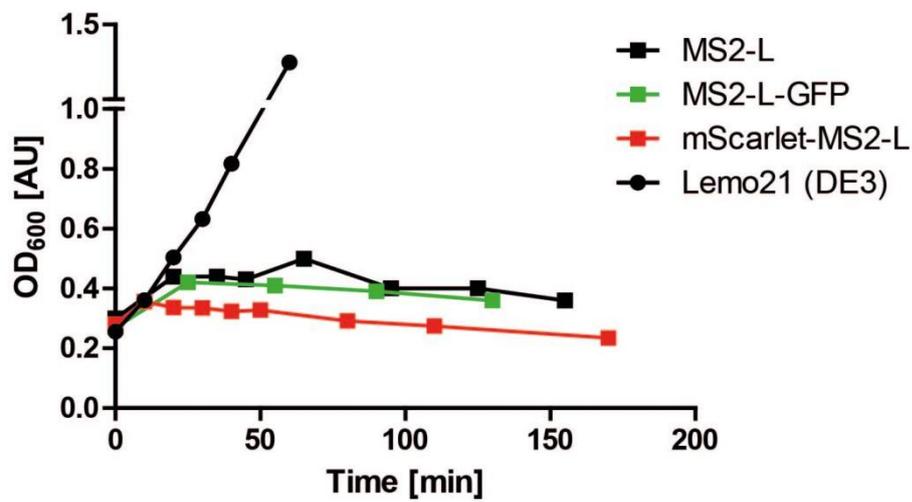
Supplementary Figure 1. Amino acid sequences of MS2-L derivatives. Full-length MS2-L was gradually truncated at the N-terminally located soluble domain. The transmembrane region, predicted by the interactive protein feature visualization tool PROTTER (ETH Zürich), is highlighted in yellow. All constructs were synthesized with a C-terminal StrepII-tag separated by a short linker (MS2-L-GTGG-WSHPQFEK).



Supplementary Figure 2. CF expression and purification of detergent solubilized MS2-L derivatives. All proteins were CF synthesized at a previously determined Mg^{2+} optimum of 20 mM. (A) Exemplary P-CF expression. After harvesting, the pellets were suspended in S30 buffer C in the respective RM volume and 1.5 μ L of the sample was analyzed on a discontinuous 4%-11 % Tris-Tricine SDS-PAGE. Overexpression bands are framed by red boxes. Other bands appearing on the SDS-PAGE correspond to co-precipitated proteins from the S30 lysate; (B) Exemplary StrepII-purified MS2-L derivatives after D-CF expression with detergent Brij78.



Supplementary Figure 3. Co-translational insertion of MS2-L and MS2-L_{P23} into NDs. CF reactions were supplemented with increasing concentrations of preformed NDs. After expression, the samples were centrifuged and pellets were suspended in S30 buffer C in the respective RM volume. 3 μL of pellet and supernatant fractions were separated by SDS-PAGE and the amount of peptide in both fractions was determined by densitometry of immunoblots. The combined signal was normalized to 1 and the relative amounts of solubilized peptide/ND complexes were calculated. Error bars represent the SEM. (A) MS2-L; DMPC ($n = 6$ for 5, 10, 20, 40, 60 μM; $n = 4$ for 0 μM; $n = 3$ for 80 μM), DMPG ($n = 4$ for 0, 20, 40, 60, 80 μM; $n = 3$ for 5, 10 μM); (B) MS2-L_{P23}; DMPC ($n = 3$ for all concentrations), DMPG ($n = 3$ for 0 μM, $n = 4$ for 10 μM, $n = 5$ for 20, 40, 60 μM and $n = 6$ for 5 μM).



Supplementary Figure 4. Representative growth curves of Lemo21 (DE3) cells expressing MS2-L derivatives. Liquid cultures were inoculated with freshly plated transformed cells and induced with 1 mM IPTG at early log phase ($t = 0$). Cell growth was monitored over time at the indicated time points after induction.