



In Vivo Positron Emission Tomography Imaging of Adenosine A_{2A} Receptors

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As an invasive nuclear medical imaging technology, positron emission tomography (PET) possess the possibility to imaging the distribution as well as the density of selective receptors via specific PET tracers. Inspired by PET, the development of radio-chemistry has greatly promoted the progress of innovative imaging PET tracers for adenosine receptors, in particular adenosine A2A receptors ($A_{2A}Rs$). PET imaging of A2A receptors play import roles in the research of adenosine related disorders. Several radio-tracers for A_{2A} receptors imaging have been evaluated in human studies. This paper reviews the recent research progress of PET tracers for A2A receptors imaging, and their applications in the diagnosis and treatment of related disease, such as cardiovascular diseases, autoimmune diseases, neurodegenerative and psychiatric disease. The future development of A2A PET tracers were also discussed.

OPEN ACCESS

Edited by:

Francisco Ciruela, University of Barcelona, Spain

Reviewed by:

Christa Elisabeth Müller, University of Bonn, Germany Masahiro Mishina, Nippon Medical School, Japan

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Specialty section:

This article was submitted to Experimental Pharmacology and Drug Discovery, a section of the journal Frontiers in Pharmacology

> Received: 28 August 2020 Accepted: 04 November 2020 Published: 26 November 2020

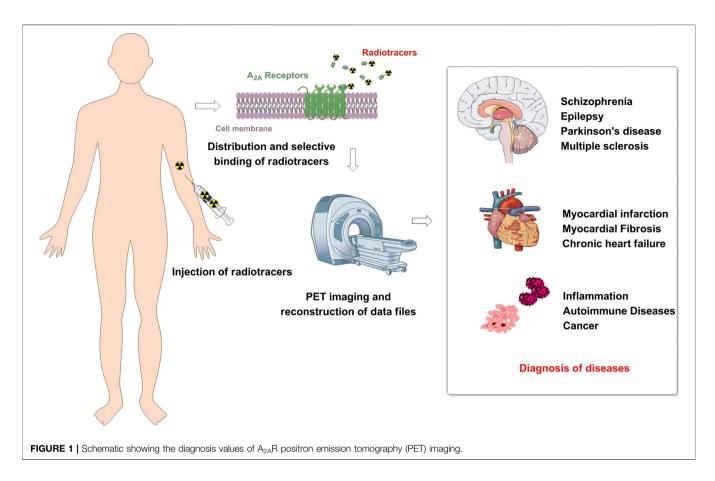
Citation:

Sun M-J, Liu F, Zhao Y-F, Wu X-A (2020) In Vivo Positron Emission Tomography Imaging of Adenosine A_{2A} Receptors. Front. Pharmacol. 11:599857. doi: 10.3389/fphar.2020.599857 Keywords: adenosine A2A receptors, positron emission tomography tracers, positron emission tomography imaging, neurodegenerative and psychiatric disease, autoimmune diseases, cardiovascular diseases

INTRODUCTION

As an extracellular endogenous messenger, adenosine play important roles in biochemical processes, signal transduction and neurotransmission (Estrela and Abraham, 2011). In physiological and pathological conditions, it acts as a cytoprotectant and a neuromodulator in response to organ and tissue stress (Khanapur et al., 2013). It also holds the capability to reduce energy demand or increase energy supply to organs or tissues which are damaged or disturbed. It is known that cytoprotective and neuromodulatory function in the brain are mediated by four adenosine receptors (ARs), namely A1, A2A, A2B, and A3 (Jacobson and Gao, 2006; Khanapur et al., 2013). A2ARs are ubiquitously distributed in brain, heart, lungs and spleen, and A2ARs mainly facilitates neurotransmissions and other physiological functions. A_{2A}Rs are involved in multiple physiological processes (Tang et al., 2019; Chen and Cunha, 2020), as well as in various pathological conditions (Illes et al., 2016; Burnstock, 2017). The dysfunction of $A_{2A}Rs$ are related to many diseases such as cardiovascular diseases, autoimmune Diseases, neurodegenerative and psychiatric disease. However, many of their functions in pathophysiological processes remain unknown, partly due to the lack of available techniques for spatial and temporal control of purinergic signaling. Positron emission tomography (PET) is a nuclear medical technology that allows in vivo imaging and quantification of specific targets, as well as molecular and cellular processes in the living body. For example, with specific brain-targeted radio-tracers, PET therefore enables the in vivo imaging of local brain function, including receptor-binding ability, cerebral blood flow, and molecular metabolism (Mishina and Ishiwata, 2014).

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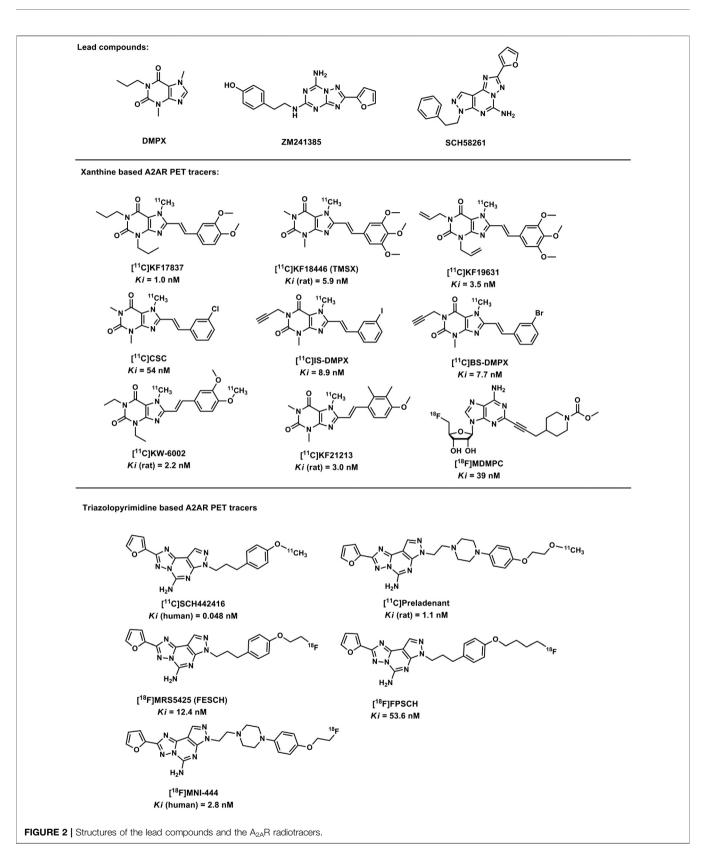
At present, PET imaging studies on adenosine receptors are mainly focused on A1 and A2A receptors, and for the diagnose of related diseases (**Figure 1**). In this paper, we will discuss the recent progress of lead compounds and related radio tracers for PET imaging for $A_{2A}Rs$. In addition, this review also outlines PET imaging for adenosine A_{2A} receptors in health and diseases subjects. Furthermore, the direction of future development of A_{2A} PET tracers were also discussed.

DEVELOPMENT OF ADENOSINE A_{2A} BASED POSITRON EMISSION TOMOGRAPHY TRACERS

In 1988, 3,7-dimethyl-1-propylxanthine (DMPX) was identified as an $A_{2A}R$ -targeted selective antagonists (Seale et al., 1988), several xanthine based radio-tracers were also successfully developed thereafter. In addition, shortly after the discovery and report of a novel pyrazolol-pyrimidine based compound as a potent and selective $A_{2A}R$ antagonist (Poucher et al., 1995; Baraldi et al., 1996; Zocchi et al., 1996; Baraldi et al., 1998), these compounds with a fused heterocycles were also regarded as lead compounds for $A_{2A}R$ PET tracers (**Figure 2**). Therefore, current PET tracers for A_{2A} receptors can be subdivided into the following two categories (**Figure 2**): 1. xanthine based $A_{2A}R$ PET tracers; 2. triazolopyrimidine based $A_{2A}R$ PET tracers.

Development of Xanthine Based A_{2A}R Positron Emission Tomography Tracers

With the similar chemical structure of A2AR endogenous ligand purine, xanthine and its derivatives showed promising properties in the A2AR PET imaging. Shimada et al. have identified that xanthine bearing the styryl group showed selective A2AR antagonistic properties (Schiffmann et al., 1991), and KF17837, a ligand with superior selectivity and potent affinity for A2ARs, was optimized for the development A2AR PET tracers (Seale et al., 1988). At present, several PET tracers were reported, such as [11C]TMSX ([11C]KF18446) (Ishiwata et al., 2000a; Ishiwata et al., 2003b), [11C]KF19631, [11C]CSC, [11C]BS-DMPX, [11C] IS-DMPX (Ishiwata et al., 2000b), [11C]KW-6002, [11C] KF21213 (Wang et al., 2000) and [18F]MDMPC (Lowe et al., 2017), and were investigated as promising PET agents (Ishiwata et al., 1996; Stone-Elander et al., 1997; Wang et al., 2000). In addition, [11C]TMSX (formally designated as [11C]KF18446) was selected for medical applications (Ishiwata et al., 2005). After the discovery of [11C]KW-6002 (Hirani et al., 2001), its reference standard KW-6002 (with commercial name istradefylline), was developed as an anti-PD agent (Hauser et al., 2003; Bara-Jimenez, W et al., 2003). Compared with [11C]TMSX, [11C]KF21213 showed a slightly higher affinity but an improved selectivity over other ARs (Wang et al., 2000). However, [11C]KF21213 has not been evaluated in human research. However, studies also indicated that the styryl group will lead to the photoisomerization



for almost all xanthine-type adenosine $A_{2A}R$ -selective ligands (Merskey, 1983; Ishiwata et al., 2003b).

Triazolopyrimidine Based A2 Adenosine Receptor Positron Emission Tomography Tracers

Based on the findings of antagonism for A2AR from triazolopyrimidine based compounds such as ZM241385 (Poucher et al., 1995) and SCH58261 (Zocchi et al., 1996), another class of A2AR PET tracers were developed based on the novel triazolopyrimidine structure. Todde et al. prepared [11C]SCH442416 by O-methylation (Todde et al., 2000), and this radio-ligand exhibits the highest selectivity and affinity among all A2A PET ligands reported as present. In addition, several nonxanthine heterocycles have also been synthesized and radiolabeled for A2AR PET imaging, including [18F]FESCH ([18F]MRS5425) (Bhattacharjee et al., 2011), [18F]FPSCH (Fastbom et al., 1998; Khanapur et al., 2017), [18F]MNI-444 (Barret et al., 2014, Barret et al., 2015), [18F]MDMPC (Lowe et al., 2017) and [11C]preladenant (Zhou et al., 2014). S. Khanapure et al. (Khanapur et al., 2017) reported the discovery of [18F]FESCH and [18F]FPSCH analogs and the evaluation in normal rats (Khanapur et al., 2014). Preliminary investigations of these tracers revealed a similar distribution pattern with the known expressions of A2AR in rat brain (Khanapur et al., 2017). Labeled with 18F, [18F]FPSCH provided more convenience in imaging protocols, as well as tracer kinetics files similar to [11C]preladenant. Compared with [18F]FESCH, [18F]FPSCH exhibited lower striatal SRTM BP_{ND} value. Dynamic PET imaging suggesting [18F]FESCH is the most favorable PET ligand for in vivo quantitation of A2AR distribution in the rodent brain.

Among all xanthine and non-xanthine based ligands, the most potent affinity for $A_{2A}Rs$ was observed in SCH442416. IS–DMPX, KF21213 and SCH442416 exhibited higher $A_{2A}R$ selectivity. The selective uptake in striatum was observed in validation studies in rodents for all radio-labeled compounds, which is correspond to $A_{2A}Rs$ expressions. However, most of the radioligands also showed a considerable degree of specific binding in the cerebral cortex and cerebrum, which is not observed with [11C]KF21213. Therefore, based on the uptake on the receptor poor cerebellum to receptorrich striatum, [11C]KF21213 was found to be the most selective for $A_{2A}R$, and followed by other representative $A_{2A}R$ PET tracers such as [11C]SCH442416 and [11C]TMSX (Bar-Yehuda et al., 2009; Mishina and Ishiwata, 2014).

POSITRON EMISSION TOMOGRAPHY IMAGING OF A2A RECEPTORS *In vivo* Positron Emission Tomography Imaging of Adenosine A_{2A} Receptors in Healthy Subjects

Based on the specific binding between the A_{2A} receptor ligand and the PET radioactive tracer, Ishiwata et al. directly visualized human brain adenosine A2A receptors using [11C]TMSX PET (Leussis et al., 2008; Bar-Yehuda et al., 2009; Ishiwata et al., 2010). Theophylline stimulation confirmed the specific binding of [11C] TMSX to adenosine A2A receptor (Ishiwata et al., 2005). Thus, the good reproducibility of [11C]TMSX PET in striatum was confirmed. The highest A2A receptor density was observed in nucleus putamen in [11C]TMSX PET images, followed by caudate head and thalamus. And the relative low density of A_{2A} receptor was observed in cerebral cortex and frontal lobe. (Ishiwata et al., 2005; Leussis et al., 2008). Autopsy and nonhuman studies have found that [11C]TMSX PET shows great binding potential in the adenosine A_{2A} receptor-rich striatum, but [11C]TMSX binds more strongly in the human thalamus than in other mammals (Mishina and Ishiwata, 2014). Moreover, as the first non-xanthine A2AR PET tracer, [11C]SCH442416 showed highest binding in putamen and the lowest binding in cerebellar was observed in unaffected people (Brooks et al., 2010). The specific binding of [11C]SCH442416 was also calculated with cerebellum as the reference region to study the different binding potentials in the putamen by Ramlackhansingh et al., (2011).

With a good maximal striatal to cerebellar ratio in rodents but low in primates, [11C]SCH442416 was not suitable for the receptor occupancy quantification studies. Barret et al. used 18F to label a new compound (Barret et al., 2014), namely [18F]MNI-444, to solve this problem. Barret et al. reported the first whole-body biodistribution and dose estimates of [18F]MNI-444 in healthy controls. The high accumulation of [18F]MNI-444 was observed in the caudate and putamen, where the density of A2A receptor is high, while the lower accumulation was discovered in the cortex and cerebellum. The distribution of MNI-444 in the brain is consistent with the known A2A expressions reported by autoradiography and with previous observations in non-human primate brains. Therefore, [18F]MNI-444 holds the possibility to be a good PET tracer for imaging of A2A receptors in the human brain (Barret et al., 2015). PET imaging with [18F]MNI-444 showed a rapid brain distribution, and the uptake pattern was consistent with known A2AR densities in the human brain. The favorable kinetic properties of [18F]MNI-444 may promote the PET imaging of A2AR in research related to neurodegenerative and psychiatric diseases.

What's more, the clinical study of [11C]preladenant showed the individual organ and total-body administration of [11C] preladenant were comparable with other 11C-labeled tracers. As is known to all, the highest signal level of $A_{2A}Rs$ was observed in the basal ganglia, followed by cerebral cortex and thalamus. [11C]preladenant's regional distribution in healthy human brain is consistent with $A_{2A}R$ density. [11C] preladenant provides a feasible approach for imaging of adenosine $A_{2A}R$ in the brain. Therefore, $A_{2A}R$ density can be quantified using the cerebellum as a reference tissue model for the reference region. Further inhibition studies in the human brain may be needed to fully verify the existence of reference regions.

Adenosine A_{2A} Receptor Positron Emission Tomography Imaging in Patients

As a novel and powerful imaging technology, PET and its clinical applications are expanding rapidly. Compared with other

imaging technologies, PET possess unique characteristics such as high target specificity, quantitative ability, and high sensitivity, which can visualize and characterize receptor expressions during the development of disease. Several radio-tracers for A_{2A} receptors imaging have also been evaluated in human studies for the diagnosis of disease, including [11C]TMSX, [11C] SCH442416, [11C]preladenant, and [11C]KW6002, the characteristics and availability of these ligands are discussed below.

[11C]TMSX

Dynamic PET imaging using A2AR-specific [11C]TMSX was performed on progressive multiple sclerosis (SPMS) patients (Merskey, 1983), Parkinson's disease (PD) (Hirani et al., 2001), multiple sclerosis (Rissanen et al., 2013; Rissanen et al., 2015) and healthy controls (Merskey, 1983). The upregulated A2AR expression was observed in the brain of those patients, and these studies indicates that the [11C]TMSX dynamic PET can be used in the diagnosis of central nervous system (CNS) disorders. Studies using PET on the normal appearing of white matter (NAWM) in SPMS patients found that increased A2AR expression was correlated with decreased anisotropy score. This highlights the potential usefulness of TMSX-PET imaging in helping to detect normal appearing of white matter in diffuse lesions associated with progressive disease. Therefore, this method can make up for the deficiency of traditional imaging in diffuse change. Studies have shown an increase in [11C]TMSX binding in the putamen after anti-parkinsonian treatment. This finding may reflect compensatory changes in dopamine reduction in PD patients. Therefore, this may suggest that in PD patients, after anti-parkinsonian treatment, the increase of A2ARs in the putamina precedes the occurrence of dyskinesia. The application [11C]TMSX PET will help to further investigate the unknown mechanisms of side effects of anti-Parkinson drugs. Thus, [11C] TMSX-PET affords a novel method to diagnosis the pathology in CNS disorders (Li et al., 2019).

Moreover, Ishiwata reported that the highly uptake of [11C] TMSX in the myocardium suggested the specific binding of myocardial $A_{2A}R$ and [11C]TMSX can be used for myocardial PET imaging (Ishiwata et al., 2003a). The differences between the PET images generated by [11C]TMSX and the non-xanthine ligands is the signal-to-noise ratios, as TMSX holds higher affinity than other non-xanthine radiotracers (Li et al., 2019).

In addition, [11C]TMSX showed specific binding in peripheral tissues that was not detected by non-xanthine radiooligomer [11C]SCH442416, or [11C]Preladenant. [11C]TMSX can be used for brown adipose tissue (BAT) related A_{2A} imaging in addition to the central neural system and cardiovascular system. Lahesmaa et al. discovered that cold exposure stimulates the release of noepinephrine and significantly reduced the concentrations of available A_{2A} R in BAT used for [11C]TMSX binding, demonstrating an increased endogenous adenosine release compared to baseline (Lahesmaa et al., 2019). [11C] TMSX binding with BAT decreases when BAT is exposed to cold, which indicates that endogenous adenosine and irradiated oligosaccharide competition receptors show high binding (Sousa and Diniz, 2017). Interestingly, the reduction of [11C]TMSX

binding is related to increased perfusion in BAT, further indicating that endogenous adenosine release in BAT is accompanied by the increased oxidative metabolism. This implies that adenosine and $A_{2A}R$ are significant in the BAT activation induced by cold, which provides a new therapeutic direction for the fight against obesity and diabetes.

[11C]SCH442416

In order to avoid photoisomerization generated by xanthine analogues, Todde et al. labelled the first non-xanthine A2A antagonist, [11C]SCH442416, whose kinetic behavior in rodents suggests that it may be used for in vivo imaging of the A_{2A} adenosine receptor in future (Todde et al., 2000). [11C] SCH442416, as an in vivo marker of A2A effectiveness, can selectively and reversibly bind to striatum A2A receptor with nanoscale affinity. PET imaging with [11C]SCH442416 was used to observe the expression of levodopa-induced dyskinesias (LIDs) in patients with Parkinson's disease (Ramlackhansingh et al., 2011). This implies that A2A antagonists may have value in levodopa-induced dyskinesias intervention while reducing levodopa dose. [11C]SCH442416 PET provides an efficient and robust approach for in vivo studies of the effectiveness of A2A. [11C]SCH442416 PET also can be used to determine the dose occupation of other A2A antagonists. In addition, [18F] FESCH and [18F]FPSCH are prepared as the analogs of SCH442416 (Khanapur et al., 2014; Khanapur et al., 2017).

[11C]Preladenant

[11C]SCH442416 and [11C]TMSX are the most favorable tracers for imaging A_{2A}Rs in brain. However, low target-to-nontarget ratios, high nonspecific binding and low binding potentials are the disadvantages of these tracers. Thus, the newly improved radioactive ligand [11C]preladenant was developed for imaging A_{2A}Rs in the living brain, including human brain, rat brain and monkey brain (Sakata et al., 2017; Zhou et al., 2017a; Zhou et al., 2017b; Zhou et al., 2017c). It is a non-xanthine heterocyclic compound with high selectivity, sufficient affinity for image receptors without affecting the quantification of receptors, and this compound also showed good pharmacokinetic properties (Zhou et al., 2014). With superior target-to-nontarget ratios and excellent pharmacokinetic properties, this tracer was advanced into human studies. Recently, studies have shown that [11C] preladenant is applied to healthy human brains in a manner consistent with A_{2A}R density. Thus, it indicated that [11C] preladenant is suitable for imaging of A2ARs in the living brain (Sakata et al., 2017). In addition, compared with other ARs, [11C]preladenant showed high affinity and significant selectivity for A2AR (Neustadt et al., 2007; Zhou et al., 2014). Recently, Ishibashi et al. reported [11C]preladenant PET can be used to calculate the occupancy rate of Istradefylline to A2AR (Ishibashi et al., 2018). These results demonstrated that [11C] preladenant is a suitable tracer to evaluate A2A receptor occupancy and quantify striatal A2A receptor density by A2A receptor-targeting molecules (Sakata et al., 2017; Zhou et al., 2017a; Zhou et al., 2017c). Thus, [11C] preladenant PET is suitable for non-invasive A2AR quantification and evaluation of A2AR occupation in $A_{2A}R$ abundant regions in living brain.

[11C]KW6002

In healthy rat, although [11C]KW-6002 shows some potential as a PET ligand, it also showed low cerebral cortex and cerebellar retention, and it may proved to be insufficiently selective to be a useful in vivo radio-tracer, at least in rodents; however, it also binds to the outer fissure region, so its potential as a PET tracer needs further studies (Hirani et al., 2001). In primate and rodent models, KW6002 offers symptomatic relief of Parkinson's motor deficits without causing or exacerbating previous motor deficits. A human study of KW6002 in advanced PD patients with levodopa-related motor complications yielded good results in the remission of motor symptoms without the side effects of exercise (Bar-Yehuda et al., 2009). The uptake of [11C]KW-6002 in the brain was characterized by a blood volume term in the twocompartment model and a 50% effective dose (ED50) of cold KWL-6002 in the striatum at 0.5 mg (Bar-Yehuda et al., 2009). In humans, [11C]KW-6002 blocks were observed in all brain regions studied, which may be caused by non-specific binding to A1R and A2BR. In addition, may be due to the non-specific binding, [11C]KW-6002 has not been further studied.

SUMMARY

Extracellular adenosine is an important regulatory molecule that interacts with four ARs: A1R, A2AR, A2BR and A3R through intracellular adenosine regulating the physiological function of the cell. Changes in function and expression in neurological disorder (Parkinson's disease, Alzheimer's disorder, epilepsy), inflammation, cardiovascular disease, autoimmune diseases, and cancer were studied. A series of PET tracers for ARs were developed. Of all the tracers listed, [11C]TMSX is the oldest ligand and has been widely evaluated in several mammal populations. Since the 1990s, several radioligands have been produced for brain A2ARs PET imaging. These ligands suitable for studying humans include [11C] TMSX, [11C]SCH442416, [11C]preladenant, [11C]KW-6002, [18F]MNI-444. It seems that the xanthine scaffold may provide

REFERENCES

- Bar-Yehuda, S., Synowitz, M., Powell, J. D., Klotz, K. N., Gessi, S., and Borea, P. A. (2009). Adenosine receptors and cancer. *Handb. Exp. Pharmacol.* 193, 399–441. doi:10.1007/978-3-540-89615-9
- Bara-Jimenez, W., Sherzai, A., Dimitrova, T., Favit, A., Bibbiani, F., Gillespie, M., et al. (2003). Adenosine A(2A) receptor antagonist treatment of Parkinson's disease. *Neurolog*, 61, 293–296. https://doi.org/10.1212/01.wnl.0000073136.00548.d4
- Baraldi, P. G., Cacciari, B., Spalluto, G., Bergonzoni, M., Dionisotti, S., Ongini, E., et al. (1998). Design, synthesis, and biological evaluation of a second generation of pyrazolo[4,3-e]-1,2,4-triazolo[1,5-c]pyrimidines as potent and selective A(2A) adenosine receptor antagonists. *J. Med. Chem.* 41, 2126–2133. doi:10. 1021/jm9708689
- Baraldi, P. G., Cacciari, B., Spalluto, G., Pineda De Las Infantas Y Villatoro, M. J., Zocchi, C., Dionisotti, S., et al. (1996). Pyrazolo[4,3-e]-1,2,4-triazolo[1,5-c] pyrimidine derivatives: potent and selective A2A adenosine antagonists. J. Med. Chem. 39, 1164–1171. doi:10.1021/jm9507461
- Barret, O., Hannestad, J., Alagille, D., Vala, C., Tavares, A., Papin, C., et al. (2014). Adenosine 2A receptor occupancy by tozadenant and preladenant in rhesus monkeys. J. Nucl. Med. 55, 1712–1718. doi:10.2967/jnumed.114.142067

efficient binding specificity for the A2AR subtype. However, photoisomerization should be taken into consideration when developing xanthine-type adenosine A2A receptor-selective ligands.

Although adenosine can also be tested by *in situ* hybridization and immunochemistry probes in recent years, PET imaging of $A_{2A}R$ can further be used to capture changes in $A_{2A}Rs$ distribution and density as the disease progresses, as well as to monitor treatment responses to these changes. In addition, PET can also determine the $A_{2A}R$ occupancy in the brain can be measured by PET, and hence providing a useful method for drug discovery (Tavares et al., 2013). The PET radio-tracers provided valuable information for the diagnosis and treatment of diseases associated with altered ARs expression, following of the summary picture.

Molecular imaging plays a crucial role in improving accuracy by quantifying, characterizing and visualizing biological processes at the molecular and cellular levels in living body, which provides an achievable basis for precision medicine. Therefore, how to realize the personalized diagnosis and treatment of A2A-related diseases with PET imaging technology will become an important research direction in the future. In addition, the application of PET molecular imaging technology in assessing A2A disease risk and understanding disease mechanisms would also make a significant contribution to the medical profession.

AUTHOR CONTRIBUTIONS

MS and YZ prepared the manuscript, review and editing by XW and FL.

FUNDING

This work is supported by Sichuan Science and Technology Program (No. 2017JY0324).

- Barret, O., Hannestad, J., Vala, C., Alagille, D., Tavares, A., Laruelle, M., et al. (2015). Characterization in humans of 18F-MNI-444, a PET radiotracer for brain adenosine 2A receptors. J. Nucl. Med. 56, 586–591. doi:10.2967/jnumed. 114.152546
- Bhattacharjee, A. K., Lang, L., Jacobson, O., Shinkre, B., Ma, Y., Niu, G., et al. (2011). Striatal adenosine A_{2A} receptor-mediated positron emission tomographic imaging in 6-hydroxydopamine-lesioned rats using [¹⁸F]-MRS5425. *Nucl. Med. Biol.* 38, 897–906. doi:10.1016/j.nucmedbio.2011. 01.009
- Brooks, D. J., Papapetropoulos, S., Vandenhende, F., Tomic, D., He, P., Coppell, A., et al. (2010). An open-label, positron emission tomography study to assess adenosine A2A brain receptor occupancy of vipadenant (BIIB014) at steadystate levels in healthy male volunteers. *Clin. Neuropharmacol.* 33, 55–60. doi:10. 1097/WNF.0b013e3181d137d2
- Burnstock, G. (2017). Purinergic signalling and neurological diseases: an update. CNS Neurol. Disord. - Drug Targets. 16, 257–265. doi:10.2174/ 1871527315666160922104848
- Chen, J. F., and Cunha, R. A. (2020). The belated US FDA approval of the adenosine A_{2A} receptor antagonist istradefylline for treatment of Parkinson's disease. *Purinergic signalling*, 16 (2), 167–174. doi:10.1007/ s11302-020-09694-2

- Estrela, A., and Abraham, W.-R. (2011). Adenosine in the inflamed gut: a janus faced compound. Curr. Med. Chem. 18, 2791–2815. doi:10.2174/ 092986711796011274
- Fastbom, J., Pazos, A., Palacios, J. M., Estrela, B. A., Abraham, W.-R., Congreve, M., et al. (1998). Functions, dysfunctions and possible therapeutic relevance of adenosine A2A receptors in Huntington's disease. *Prog. Neurobiol.* 111, 1970–1975. doi:10.1385/JMN/26
- Hauser, R. A., Hubble, J. P., and Truong, D. D. (2003). Randomized trial of the adenosine A(2A) receptor antagonist istradefylline in advanced PD. *Neurology* 61, 297–303. doi:10.1212/01.WNL.0000081227.84197.0B
- Hirani, E., Gillies, J., Karasawa, A., Shimada, J., Kase, H., Opacka-Juffry, J., et al. (2001). Evaluation of [4-O-methyl-11C]KW-6002 as a potential PET ligand for mapping central adenosine A2A receptors in rats. *Synapse* 42, 164–176. doi:10. 1002/syn.1110
- Illes, P., Verkhratsky, A., Burnstock, G., and Sperlagh, B. (2016). Purines in neurodegeneration and neuroregeneration. *Neuropharmacology*. 104, 1–3. doi:10.1016/j.neuropharm.2016.01.020
- Ishibashi, K., Miura, Y., Wagatsuma, K., Toyohara, J., Ishiwata, K., and Ishii, K. (2018). Occupancy of adenosine A_{2A} receptors by istradefylline in patients with Parkinson's disease using 11C-preladenant PET. *Neuropharmacology* 143, 106–112. doi:10.1016/j.neuropharm.2018.09.036
- Ishiwata, K., Kawamura, K., Kimura, Y., Oda, K., and Ishii, K. (2003a). Potential of an adenosine A2A receptor antagonist [11C]TMSX for myocardial imaging by positron emission tomography: a first human study. *Ann. Nucl. Med.* 17, 457–462. doi:10.1007/BF03006434
- Ishiwata, K., Kimura, Y., Oda, K., Ishii, K., Sakata, M., Kawasaki, K., et al. (2010). Development of PET radiopharmaceuticals and their clinical applications at the positron medical center. *Geriatr. Gerontol. Int.* 10, S180–S196. doi:10.1111/j. 1447-0594.2010.00594.x
- Ishiwata, K., Mishina, M., Kimura, Y., Oda, K., Sasaki, T., and Ishii, K. (2005). First visualization of adenosine A(2A) receptors in the human brain by positron emission tomography with [11C]TMSX. Synapse 55, 133–136. doi:10.1002/syn. 20099
- Ishiwata, K., Noguchi, J., Toyama, H., Sakiyama, Y., Koike, N., Ishii, S. I., et al. (1996). Synthesis and preliminary evaluation of [11C]KF17837, a selective adenosine A2A antagonist. *Appl. Radiat. Isot.* 47, 507–511. doi:10.1016/0969-8043(95)00295-2
- Ishiwata, K., Ogi, N., Shimada, J., Nonaka, H., Tanaka, A., Suzuki, F., et al. (2000a). Further characterization of a CNS adenosine A2a receptor ligand [11C] KF18446 with *in vitro* autoradiography and *in vivo* tissue uptake. Ann. Nucl. Med. 14, 81–89. doi:10.1007/BF02988585
- Ishiwata, K., Shimada, J., Wang, W. F., Harakawa, H., Ishii, S. I., Kiyosawa, M., et al. (2000b). Evaluation of iodinated and brominated [11C]styrylxanthine derivatives as *in vivo* radioligands mapping adenosine A2A receptor in the central nervous system. *Ann. Nucl. Med.* 14, 247–253. doi:10.1007/BF02988206
- Ishiwata, K., Wang, W. F., Kimura, Y., Kawamura, K., and Ishii, K. (2003b). Preclinical studies on [11C]TMSX for mapping adenosine A2A receptors by positron emission tomography. Ann. Nucl. Med. 17, 205–211. doi:10.1007/BF02990023
- Jacobson, K. A., and Gao, Z. G. (2006). Adenosine receptors as therapeutic targets. Nat. Rev. Drug Discov. 5, 247–264. doi:10.1038/nrd1983
- Khanapur, S., Paul, S., Shah, A., Vatakuti, S., Koole, M. J. B., Zijlma, R., et al. (2014). Development of [18F]-labeled pyrazolo[4,3-e]-1,2,4-triazolo[1,5-c]pyrimidine (SCH442416) analogs for the imaging of cerebral adenosine A 2A receptors with positron emission tomography. J. Med. Chem. 57, 6765–6780. doi:10.1021/ jm500700y
- Khanapur, S., Van Waarde, A., Dierckx, R. A. J. O., Elsinga, P. H., and Koole, M. J. B. (2017). Preclinical evaluation and quantification of 18F-Fluoroethyl and 18F-Fluoropropyl Analogs of SCH442416 as radioligands for PET imaging of the adenosine a2a receptor in rat brain. J. Nucl. Med. 58, 466–472. doi:10.2967/jnumed.116.178103
- Khanapur, S., Waarde, A., Ishiwata, K., Leenders, K., Dierckx, R., and Elsinga, P. (2013). Adenosine A_{2A} receptor antagonists as positron emission tomography (PET) tracers. *Curr. Med. Chem.* 21, 312–328. doi:10.2174/ 09298673113206660265
- Lahesmaa, M., Oikonen, V., Helin, S., Luoto, P., U Din, M., Pfeifer, A., et al. (2019). Regulation of human brown adipose tissue by adenosine and A 2A receptors – studies with [15 O]H 2 O and [11 C]TMSX PET/CT. Eur. J. Nucl. Med. Mol. Imaging 46, 743–750. doi:10.1007/s00259-018-4120-2

- Leussis, M., Lawson, K., Stone, K., and Andersen, S. L. (2008). The enduring effects of an adolescent social stressor on synaptic density, Part II: poststress reversal of synaptic loss in the cortex by adinazolam and MK-801. *Synapse* 61, 185–192. doi:10.1002/syn.2043
- Li, J., Hong, X., Li, G., Conti, P. S., Zhang, X., and Chen, K. (2019). PET imaging of adenosine receptors in diseases. *Curr. Top. Med. Chem.* 19, 1445–1463. doi:10. 2174/1568026619666190708163407
- Lowe, P.T., Dall'Angelo, S., Mulder-Krieger, T., IJzerman, A.P., Zanda, M., and O'Hagan, D. (2017). A new class of fluorinated A2A adenosine receptor agonist with application to last-step enzymatic [18F] fluorination for PET imaging. *ChemBioChem.* 18, 2156–2164. doi:10.1002/cbic.201700382
- Merskey, H. (1983). Communications to the editor. J. Psychosom. Res. 27, 85. doi:10.1016/0022-3999(83)90113-7
- Mishina, M., and Ishiwata, K. (2014). Adenosine receptor PET imaging in human brain. Int Rev Neurobiol. 119, 51–69. doi:10.1016/B978-0-12-801022-8.00002-7.
- Neustadt, B. R., Hao, J., Lindo, N., Greenlee, W. J., Stamford, A. W., Tulshian, D., et al. (2007). Potent, selective, and orally active adenosine A2A receptor antagonists: arylpiperazine derivatives of pyrazolo[4,3-e]-1,2,4-triazolo[1,5-c] pyrimidines. *Bioorganic Med. Chem. Lett.* 17, 1376–1380. doi:10.1016/j.bmcl. 2006.11.083
- Poucher, S. M., Palmer, T. M., Jacobson, K. A., and Stiles, G. L.. (1995). 125I-4-(2-[7-amino-2-[2-furyl][1,2,4]triazolo[2,3-a][1,3,5] triazin-5-yl-amino]ethyl) phenol, a high affinity antagonist radioligand selective for the A2a adenosine receptor. *Mol. Pharmacol.* 48, 970–974.
- Ramlackhansingh, A. F., Bose, S. K., Ahmed, I., Turkheimer, F. E., Pavese, N., and Brooks, D. J. (2011). Adenosine 2A receptor availability in dyskinetic and nondyskinetic patients with Parkinson disease. *Neurology* 76, 1811–1816. doi:10.1212/WNL.0b013e31821ccce4
- Rissanen, E., Tuisku, J., Luoto, P., Arponen, E., Johansson, J., Oikonen, V., et al. (2015). Automated reference region extraction and population-based input function for brain [11C]TMSX PET image analyses. J. Cerebr. Blood Flow Metabol. 35, 157–165. doi:10.1038/jcbfm.2014.194
- Rissanen, E., Virta, J. R., Paavilainen, T., Tuisku, J., Helin, S., Luoto, P., et al. (2013). Adenosine A2A receptors in secondary progressive multiple sclerosis: a [11 C] TMSX brain PET study. *J. Cerebr. Blood Flow Metabol.* 33, 1394–1401. doi:10. 1038/jcbfm.2013.85
- Sakata, M., Ishibashi, K., Imai, M., Wagatsuma, K., Ishii, K., Zhou, X., et al. (2017). Initial evaluation of an adenosine A2A receptor ligand, 11C-preladenant, in healthy human subjects. *J. Nucl. Med.* 58, 1464–1470. doi:10.2967/jnumed.116. 188474
- Schiffmann, S. N., Libert, F., Vassart, G., and Vanderhaeghen, J. J (1991). Distribution of adenosine A2 receptor mRNA in the human brain. *Neurosci. Lett.* 130, 177–181. doi:10.1016/0304-3940(91)90391-6
- Seale, T. W., Abla, K. A., Shamim, M. T., Carney, J. M., and Daly, J. W. (1993). 3,7-Dimethyl-1-propargylxanthine: a potent and selective *in vivo* antagonist of adenosine analogs. *Life Sci.* 43, 1671–1684. doi:10.1016/0024-3205(88)90478-x
- Sousa, J. B., and Diniz, C. (2017). The adenosinergic system as a therapeutic target in the vasculature: new ligands and challenges. *Molecules* 22, 752. doi:10.3390/ molecules22050752
- Stone-Elander, S., Thorell, J. O., Eriksson, L., Fredholm, B. B., and Ingvar, M. (1997). *In vivo* biodistribution of [N-11C-methyl]KF 17837 using 3-D-PET: evaluation as a ligand for the study of adenosine A(2A) receptors. *Nucl. Med. Biol.* 24, 187–191. doi:10.1016/S0969-8051(96)00216-8
- Svenningsson, P., Le Moine, C., Aubert, I., Burbaud, P., Fredholm, B. B., and Bloch, B. (1998). Cellular distribution of adenosine A2A receptor mRNA in the primate striatum. *J. Comp. Neurol.* 399, 229–240. doi:10.1002/(SICI)1096-9861(19980921)399:2<229::AID-CNE6>3.0
- Tang, Z., Ye, W., Chen, H., Kuang, X., Guo, J., Xiang, M., et al. (2019). Role of purines in regulation of metabolic reprogramming. *Purinergic Signal*. 15, 423–438. doi:10.1007/s11302-019-09676-z
- Tavares, A. A. S., Batis, J. C., Papin, C., Jennings, D., Alagille, D., Russell, D. S., et al. (2013). Kinetic modeling, test-retest, and dosimetry of 123I-MNI-420 in humans. J. Nucl. Med. 54, 1760–1767. doi:10.2967/jnumed.113.119933
- Todde, S., Moresco, R. M., Simonelli, P., Baraldi, P. G., Cacciari, B., Spalluto, G., et al. (2000). Design, radiosynthesis, and biodistribution of a new potent and selective ligand for *in vivo* imaging of the adenosine A2A receptor system using positron emission tomography. *J. Med. Chem.* 43, 4359–4362. doi:10.1021/ jm0009843

- Wang, W. F., Ishiwata, K., Nonaka, H., Ishii, S. I., Kiyosawa, M., Shimada, J. I., et al. (2000). Carbon-11-labeled KF21213: a highly selective ligand for mapping CNS adenosine A(2A) receptors with positron emission tomography. *Nucl. Med. Biol.* 27, 541–546. doi:10.1016/S0969-8051(00)00126-8
- Zhou, X., Boellaard, R., Ishiwata, K., Sakata, M., Dierckx, R. A. J. O., De Jong, J. R., et al. (2017a). *In vivo* evaluation of 11C-preladenant for PET imaging of adenosine A2A receptors in the conscious monkey. *J. Nucl. Med.* 58, 762–767. doi:10.2967/jnumed.116.182410
- Zhou, X., Elsinga, P. H., Khanapur, S., Dierckx, R. A. J. O., de Vries, E. F. J., and de Jong, J. R. (2017b). Radiation dosimetry of a novel adenosine A2A receptor radioligand [11C]preladenant based on PET/CT imaging and ex vivo biodistribution in rats. *Mol. Imag. Biol.* 19, 289–297. doi:10.1007/s11307-016-0992-3
- Zhou, X., Khanapur, S., De Jong, J. R., Willemsen, A. T. M., Dierckx, R. A. J. O., Elsinga, P. H., et al. (2017c). *In vivo* evaluation of [11C]preladenant positron emission tomography for quantification of adenosine A2A receptors in the rat brain. *J. Cerebr. Blood Flow Metabol.* 37, 577–589. doi:10.1177/0271678X16634714
- Zhou, X., Khanapur, S., Huizing, A. P., Zijlma, R., Schepers, M., Dierckx, R. A., et al. (2014). Synthesis and preclinical evaluation of 2-(2-furanyl)-7-[2-[4-[4-(2-[11C]methoxyethoxy)phenyl]-1-piperazinyl]ethyl]7H-pyrazolo[4,3-e]

[1,2,4]triazolo[1,5-c]pyrimidine-5-amine ([11C]Preladenant) as a PET tracer for the imaging of cerebral adenosine A2A receptors. *J. Med. Chem.* 57, 9204–9210. doi:10.1021/jm501065t

Zocchi, C., Ongini, E., Conti, A. M., Monopoli, A., and Dionisotti, S. P. (1996). The non-xanthine heterocyclic compound sch 58261 is a new potent and selective a2a adenosine receptor antagonist. *J. Pharmacol. Exp. Therapeut.* 276 (2), 398–404.

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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