



pISSN 2287-2728

eISSN 2287-285X

## Review

# Liver-lung axes in alcohol-related liver disease

Gavin E. Arteel

Division of Gastroenterology, Hepatology and Nutrition, Department of Medicine, Pittsburgh Liver Research Center, University of Pittsburgh, Pittsburgh, PA, USA

Alcohol-related liver disease (ALD) and alcohol-related susceptibility to acute lung injury are the leading causes of morbidity and mortality due to chronic alcohol abuse. Most commonly, alcohol-induced injury to both organs are evaluated independently, although they share many parallel mechanisms of injury. Moreover, recent studies indicate that there is a potential liver lung axis that may contribute to organ pathology. This mini-review explores established and potential mechanisms of organ-organ crosstalk in ALD and alcohol-related lung injury. (*Clin Mol Hepatol* 2020;26:670-676)

**Keywords:** Liver diseases; Alcoholic Liver disease; Acute lung injury; Respiratory distress syndrome, Adult

## ALCOHOL-RELATED LIVER AND LUNG DISEASE

The risk of alcohol-related liver disease (ALD) increases in a dose- and time-dependent manner with consumption of alcohol. Progression of ALD is well-characterized and is actually a spectrum of liver diseases, which ranges initially from simple steatosis, to inflammation and necrosis (steatohepatitis), to fibrosis and cirrhosis. The most effective therapy for ALD is orthotopic liver transplantation.<sup>1</sup> However, the usefulness of liver transplantation is limited, owing to a donor organ shortage, as well as by ethical issues concerning the treatment of individuals that have inveterate alcohol dependence. In the absence of a "cure" for ALD, the major clinical focus is to treat the sequelae of a failing liver (e.g., ascites, portal hypertension, and hepatorenal syndrome).<sup>1</sup> Although the successful treatment of these secondary effects pro-

longs the life of ALD patients, this therapy is only palliative. Furthermore, since underlying cirrhosis greatly increases the risk of developing hepatocellular carcinoma (HCC),<sup>2</sup> success in maintaining 'stable cirrhotics' may translate into an increase in the incidence of HCC. Indeed, HCC incidence is increasing in the USA and in Europe.<sup>3</sup>

Alcohol abuse is known to increase the risk for lung injury. In contrast to the liver, most studies do not support a direct pathogenic effect of ethanol on the lungs. Instead, it is hypothesized that alcohol consumption enhances the risks of injury caused by other 'hits.' For example, alcohol increases the risk for the development of lung infection.<sup>4,5</sup> This increased risk is mediated by physical factors that increase the risk of inoculation, including aspiration of gastric contents and/or microbes from the upper respiratory track (i.e., oropharyngeal flora), as well as decreased mu-

### Abbreviations:

ALD, alcohol-related liver disease; ALI, acute lung injury; ARDS, acute respiratory distress syndrome; AUD, alcohol use disorder; BALF, bronchoalveolar lavage fluid; CNS, central nervous system; DAMPs, damage-associated molecular patterns; EVs, extracellular vesicles; HCC, hepatocellular carcinoma; HMGB1, high motility group box-1; LPS, lipopolysaccharide; MTDs, mitochondrial damage-associated molecular patterns; MV, microvesicles; PAMPs, pathogen-associated molecular patterns; SIRS, systemic inflammatory response syndrome; TLR, toll-like receptor; TNFR1, tumor necrosis factor- $\alpha$  receptor; TNF $\alpha$ , tumor necrosis factor- $\alpha$

### Corresponding author : Gavin E. Arteel

Division of Gastroenterology, Hepatology and Nutrition, Department of Medicine, Pittsburgh Liver Research Center, University of Pittsburgh, BST1 W1143, 200 Lothrop Street, Pittsburgh, PA 15213, USA  
Tel: +1-412-648-4187, Fax: +1-412-648-4055  
E-mail: [gearteel@pitt.edu](mailto:gearteel@pitt.edu)  
<https://orcid.org/0000-0002-2253-5984>

**Editor:** Jung-Hwan Yu, Inha University School of Medicine, Korea

**Received :** Jul. 16, 2020 / **Revised :** Aug. 25, 2020 / **Accepted :** Aug. 25, 2020

cous-facilitated clearance of bacterial pathogens from the upper airway.<sup>6</sup> Moreover, alcohol ingestion disrupts the normal beating motion of the cilia that is important in the clearance mechanisms aimed to help remove pathogens from the airways.<sup>7</sup> These abnormalities, together with documented impairments in pulmonary host defenses, likely explain the increased infection rate observed in individuals that abuse alcohol.<sup>8</sup> Alcohol consumption is also associated with worse outcomes after pulmonary infection. Specifically, individuals diagnosed with an alcohol use disorder (AUD) are more susceptible to the development of the acute respiratory distress syndrome (ARDS) in response to a pulmonary or systemic infection.<sup>9</sup> ARDS is the most severe form of acute lung injury (ALI) with an incidence close to 200,000 cases per year in the USA and with a mortality rate of around 40%.<sup>10</sup> ARDS occurs 3.7 times more often in people who meet the diagnostic criteria for AUD. Lastly, it was recently shown that alcohol exposure to rodents using the acute-on-chronic binge model does cause subtle functional changes to the lung,<sup>11</sup> bringing into question the assumption that alcohol abuse does not directly damage the lungs.

## LIVER-LUNG INTERACTIONS IN ALD

End-stage ALD is well-recognized as a systemic disorder. The idea of the liver-lung axis in the setting of chronic alcohol exposure is based on clinical data demonstrating that patients with a diagnosed AUD have increased incidence of and mortality from ARDS.<sup>9,12,13</sup> Furthermore, in ARDS patients with hepatic failure, mortality increases to almost 100%.<sup>9</sup> The finding that most individuals with an AUD have at least subclinical ALD further supports coexistence of liver disease in alcohol-abusing patients with ARDS.<sup>14</sup> Pulmonary injury induced by lipopolysaccharide (LPS) can be altered by mediators released from the liver (e.g., tumor necrosis factor- $\alpha$  [TNF $\alpha$ ]). Indeed, it was demonstrated in rats that extrathoracic LPS-induced lung damage required perfusion with the liver.<sup>15</sup>

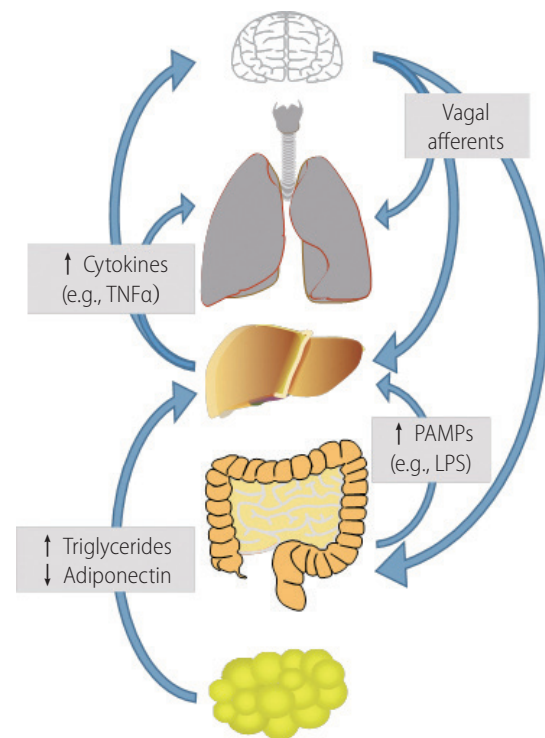
## POTENTIAL MECHANISMS OF INTER-ORGAN COMMUNICATION BETWEEN THE LIVER AND THE LUNGS

Although end organ diseases are often studied in isolation. Cross-talk between organs is not uncommon in the maintenance of homeostasis, as well as in mounting a coordinated response to

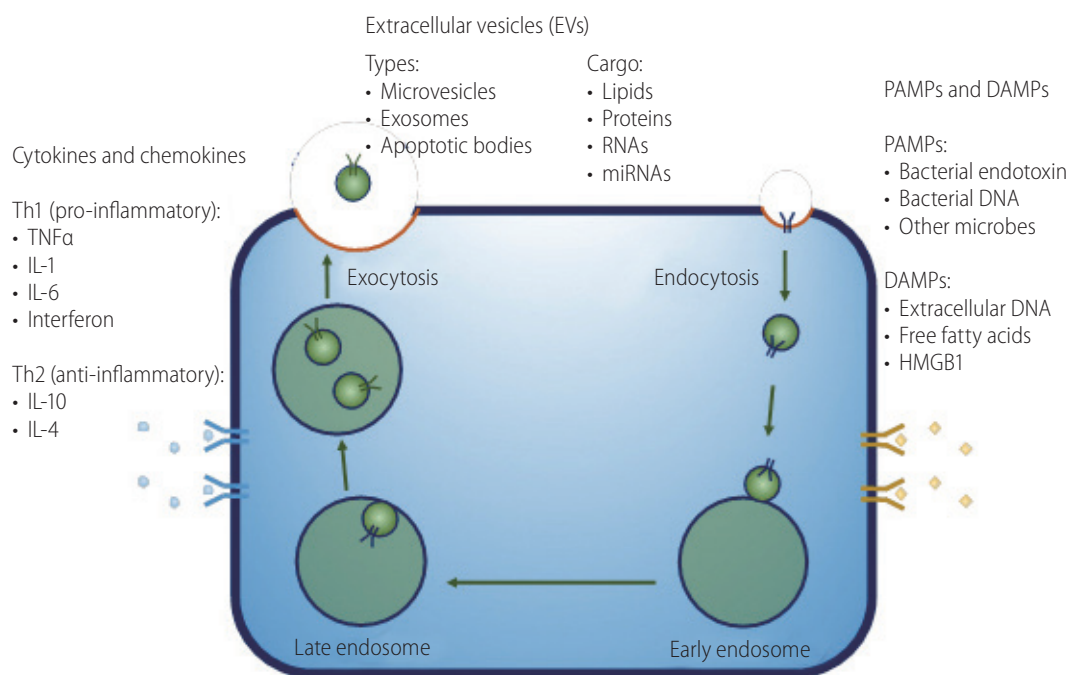
dyshomeostasis. As mentioned above, the existence of a liver-lung axis, at least unidirectional, has been established experimentally. Below, potential mechanisms that mediate organ-organ crosstalk are discussed (Fig. 1, 2).

## Cytokines and chemokines

In addition to TNF $\alpha$  (see above), a wide variety of hepatic cell types both produce and respond to cytokines, including inflammatory cells, such as monocytes, macrophages, and neutrophils, as well as parenchymal cells. Cytokines can be primarily classified into two groups: "proinflammatory" T helper 1 and "anti-inflammatory" T helper 2. Homeostasis mediated by these cytokines ensures appropriate immune and inflammatory responses, with minimal normal tissue damage. These mediators also facilitate a coordinated response to insults and injuries that stem well beyond the primary target organ.



**Figure 1.** The liver at the center of organ-organ axes. There are several physiologic and pathophysiologic mechanisms by which inter-organ communication can be mediated, such as via nutrients, hormones and hepatokines, afferent and efferent innervation, release of extracellular vesicles, cytokines and PAMPs and DAMPs. Modified from Poole et al.<sup>57</sup> TNF $\alpha$ , tumor necrosis factor- $\alpha$ ; PAMPs, pathogen-associated molecular patterns; LPS, lipopolysaccharide; DAMPs, damage-associated molecular patterns.



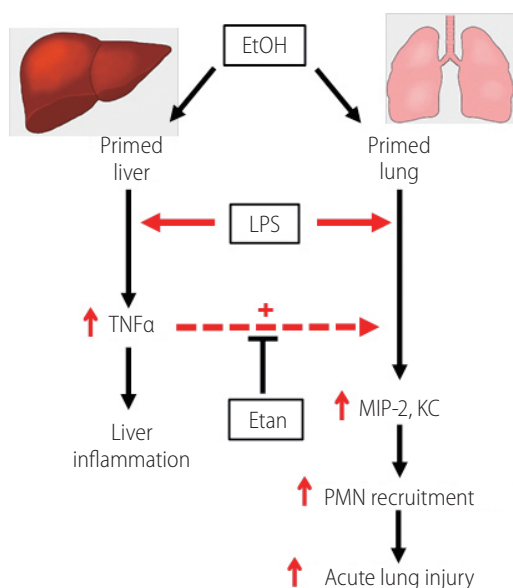
**Figure 2.** Specific mechanisms of communication in response to stress. Injured cells can release several signals to mediate communication between them and distal tissues. The profiles of both Th1 (i.e., proinflammatory) and Th2 (i.e., anti-inflammatory or profibrotic) cytokines is altered during hepatic injury. Additionally, the liver can respond to PAMPs (e.g., bacterial LPS) from other organs, such as the gastrointestinal tract, and/or respond or release DAMPs (e.g., HMGB1) that serve as ‘danger signals’ to pattern recognition receptors in other organs. Modified from Poole et al.<sup>57</sup> Th1, T helper 1; TNF $\alpha$ , tumor necrosis factor- $\alpha$ ; IL, interleukin; Th2, T helper 2; PAMPs, pathogen-associated molecular patterns; DAMPs, damage-associated molecular patterns; HMGB1, high motility group box-1; LPS, lipopolysaccharide.

Dysregulated cytokine production has been implicated in the development and progression of ALD.<sup>16</sup> Kupffer cells, the resident hepatic macrophages, play a key role in monitoring, clearing and mediating responses to gut-derived toxins, such as bacterial LPS. Alcohol consumption has long been known to increase gut permeability, leading to increased LPS in the portal circulation.<sup>17,18</sup> LPS and other bacterial toxins interact with Kupffer cells via toll-like receptors (TLRs) including TLR4. Activation of the TLR4 signaling pathway leads to increased transcription of several pro-inflammatory cytokines. It is therefore unsurprising that patients with ALD have increased levels of several circulating cytokines, including TNF $\alpha$ .<sup>19</sup> Other cytokines also likely play key roles, but TNF $\alpha$  is likely the most well studied.

TNF $\alpha$  is widely recognized as having a potential role in organ-organ communication in alcohol-induced organ injury. TNF $\alpha$  is suspected to be a common mechanism of alcohol-induced pathology not only in the liver,<sup>20,21</sup> but also in other organs, such as the lungs and the brain. In the lung, chronic alcohol pre-exposure enhanced endotoxemia-induced ALI, which was prevented by blocking systemic TNF $\alpha$  with etanercept.<sup>22</sup> In the brain, alcohol expo-

sure enhances the increase in TNF $\alpha$  levels in caused by LPS, which is also prevented by deleting the canonical TNF $\alpha$  receptor (TNFR1).<sup>23</sup> The source of this systemic TNF $\alpha$  ultimately remains unknown, although liver is clearly a likely key player.

TNF $\alpha$  has also been potentially implicated in alcohol-enhanced ALI. Lung damage was more severe in mice that were exposed to chronic alcohol subsequently injected with intraperitoneal LPS.<sup>22</sup> The differential effects on cytokine expression in systemic circulation and locally in the lung (i.e., bronchoalveolar lavage fluid [BALF]) were examined. Animals pre-exposed to ethanol diet had significantly elevated levels of plasma TNF $\alpha$  after LPS injection compared to animals fed a control diet. In the BALF, however, ethanol pre-exposed animals had elevated levels of the TNF $\alpha$ -responsive chemokines, macrophage inflammatory protein-2 and KC. This elevated chemokine expression also correlated with increased pulmonary neutrophil recruitment. Interestingly, blocking systemic TNF $\alpha$  using a TNF $\alpha$ -inhibiting antibody, etanercept, significantly attenuated the alcohol-enhanced pulmonary chemokine expression, and ultimately, alcohol-enhanced lung injury and inflammation after LPS (Fig. 3). While the liver is not the sole source



**Figure 3.** Potential mechanism of crosstalk between the liver and the lungs in alcohol-induced organ injury. Ethanol preexposure primes both the liver and lung to enhanced injury and inflammation caused by LPS. In the work presented here, ethanol preexposure significantly enhanced LPS induced TNF $\alpha$  mRNA expression in the liver but not the lung. Interestingly, ethanol preexposure enhanced LPS induced increases in the TNF $\alpha$  responsive genes MIP-2 and KC in the lung. Modified from Poole et al.<sup>57</sup> LPS, lipopolysaccharide; TNF $\alpha$ , tumor necrosis factor- $\alpha$ ; MIP-2, macrophage inflammatory protein-2.

of systemic TNF $\alpha$  in this experimental setting, other studies have demonstrated that ablation of Kupffer cells before LPS injection decreased systemic TNF $\alpha$  levels by almost 90%, indicating that these cells are in fact a predominate source of plasma TNF $\alpha$  in experimental endotoxemia.<sup>24</sup>

### Extracellular vesicles (EVs)

EVs, a term that includes microvesicles (MV), exosomes, and apoptotic bodies, are an emerging mechanism of organ-organ communication in many diseases, including ALD.<sup>25</sup> EVs can carry a diverse cargo, including lipids, proteins, RNAs, and miRNAs and are attractive therapeutic targets, due to both their potential mechanistic role in disease progression, as well as for being potential surrogate biomarkers. Over 10 years ago, it was proposed that MVs are (at least) surrogate biomarkers of advanced ALD.<sup>26</sup> Furthermore, alcohol exposure causes a release of hepatic EVs that contain a distinct miRNA profile.<sup>27</sup> It was also found that patients with ALD had elevated circulating EVs, and that these EVs also may carry a unique miRNA “barcode”.<sup>27</sup> EVs may not only

serve as surrogate biomarkers of ALD, but they may also play a mechanistic role in disease progression. In addition to mediating intra-organ communication between cells, EVs can also mediate inter-organ communication.<sup>28</sup> Indeed, it is now hypothesized that EVs contribute to the development and severity of ARDS.<sup>29</sup> Furthermore, remote preconditioning, such as the cardioprotective effect of hindlimb ischemia-reperfusion injury, is also hypothesized to be mediated by EVs (e.g.,<sup>30,31</sup>). The potential role of circulating EVs in mediating liver-lung communication in the context of ALD are not currently understood and would be an interesting target for further investigation.

### Pathogen-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs)

As mentioned above, the gut-derived toxin, LPS, has long been identified as playing a potential role in ALD. The primary receptor for LPS (TLR9) belongs to a family of pattern recognition receptors. Ligands for these receptors are grouped together as molecular “danger signals,” and include PAMPs and DAMPs. PAMPs include products of microbial (e.g., bacterial endotoxin or bacterial DNA), viruses, fungi, and parasites. DAMPs, on the other hand, are endogenous danger signals released from dead or dying cells; examples include extracellular DNA or RNA, free fatty acids, and high motility group box-1 (HMGB1), among others.<sup>32</sup> Signals derived by PAMPs/DAMPs can span multiple organs, and are thought to contribute to systemic inflammatory responses (e.g., systemic inflammatory response syndrome [SIRS]).<sup>33</sup> The innate immune system surveils qualitative and quantitative changes in the spectrum of PAMPs and DAMPs as a means to mount rapid and coordinated responses to any perceived threat that is driving those changes. Although this response is important for normal immune/inflammatory function, dysregulation of this response can lead to inappropriate inflammation and tissue damage. Ethanol consumption appears to broadly dysregulate the response of these receptors and enhances expression of several TLRs, as well as their responses to stimuli.<sup>34</sup>

The role of PAMPs in organ-organ interaction during is perhaps best understood in the context of the gut-liver axis. As early as 1893, Pavlov observed that low levels of intestinally-derived “toxins” are normally present in the portal blood and are cleared by the liver.<sup>35</sup> More than half a century later, several studies suggested that experimental fibrosis caused by choline deficiency was dependent on products derived from intestinal bacteria. For example, Rutenburg and colleagues showed that non-absorbable

antibiotics protected against diet-induced cirrhosis in rats.<sup>36</sup> The causal role of bacterial PAMPs in liver injury was not confined to models of diet-induced cirrhosis, but was further established in studies using acute hepatotoxins such as carbon tetrachloride.<sup>37,38</sup> High systemic levels of the PAMP, LPS, are found in both acute and chronic liver diseases.<sup>39,40</sup> Aside from the liver, GI-derived PAMPs during alcohol exposure can potentially affect other organs, including the lungs.

One of the best-understood DAMPs, or “alarmins,” is HMGB1. HMGB1 is a constitutively-expressed nuclear protein that is released from necrotic cells. A recent study demonstrated that HMGB1 translocation from the nucleus to the cytosol correlated with disease severity in liver biopsies from ALD patients.<sup>41</sup> Additionally, knocking out HMGB1 in hepatocytes protects mice against alcohol-induced liver injury. HMGB1 signaling has also been shown to be elevated in alcohol-induced injury in the brain<sup>42,43</sup> and pancreas.<sup>44</sup> However, the role of HMGB1 and other DAMPs in organ-organ crosstalk in the setting of ALD has been largely unexplored.

Although less characterized in the setting of ALD, the concept that circulating DAMPs may be involved in organ-organ crosstalk in other disease states is clearly established. For example, trauma elevates circulating mitochondrial DAMPs (MTDs), such as mitochondrial DNA, and is hypothesized to contribute to SIRS under that condition.<sup>45</sup> Human neutrophils exposed to MTDs have increased expression of the chemokine interleukin-8. In the same study, it was demonstrated that MTDs derived from rat liver cause lung injury in recipient rats; this injury was characterized by vascular leak, pulmonary edema, neutrophil infiltration, and accumulation of inflammatory cytokines in the alveolar space. As mentioned above, SIRS has been established as a significant risk factor for mortality from alcohol-related hepatitis.<sup>46</sup> Therefore, it is not unlikely that circulating DAMPs may be involved.

### Other potential mechanisms

There are also less direct means by which the liver can influence other organs via axes (Fig. 2). For example, the liver plays an absolutely critical role in supplying fuel to other organs.<sup>47</sup> Therefore, alterations in the flux of carbohydrates and lipids through the liver can indirectly impact distal organs, as their energy sensing mechanisms respond to these changes. Nutrient levels in the liver also directly mediate responsive changes in the central nervous system (CNS) via glucose sensing afferent neurons in the liver and other organs; these sensors are hypothesized to mediate rapid central

responses to short-term energy status alterations.

There is intricate crosstalk between these metabolic systems that is controlled by a complex interplay of nuclear receptors, intracellular signaling pathways and transcription factors. Hormones and other endocrine mediators play a key role in regulating these responses. The liver is well known to respond to several endocrine hormones, including insulin, glucagon, thyroid hormones and cortisol. The liver also produce several hormones that can mediate several extrahepatic effects, such insulin-like growth factor, angiotensinogen and thrombopoietin. Furthermore, it has become increasingly clear that the liver produces several endocrine-like “hepatokines” that play key roles in regulating extrahepatic responses (e.g., FGF21).<sup>48,49</sup> The net effect of these interactions is to generate an organ that rapidly responds to endocrine signals, as well as rapidly produces endocrine signals in response to stimuli.

The liver also has several afferent neurons that mediate and coordinate its responses with extrahepatic targets, especially the CNS. This circuitry plays a key role in metabolic homeostasis, stress responses, and inflammation.<sup>50-52</sup> There is evidence suggesting that ethanol and the sympathetic nervous system interact with and partially mimic each other. For example, acute alcohol intoxication increases plasma levels of adrenaline and noradrenaline.<sup>53</sup> Furthermore, ethanol causes a hypermetabolic state in liver that is abolished by adrenalectomy, hepatic vagotomy or by adrenoceptor blockade.<sup>54,55</sup> Repeated administration of ethanol also increases plasma catecholamines and gene expression of enzymes for catecholamine synthesis in the adrenal medulla.<sup>56</sup> Although it is understood that such networks can mediate coordination between organs in response to stress or dyshomeostasis in general, the specific impact of these interactions in the context of organ-organ crosstalk in ALD is incompletely understood.

### SUMMARY AND CONCLUSIONS

In conclusion, while multiple organ failure is a hallmark of decompensated, end-stage alcoholic liver disease, there is an increasing appreciation for communication between organs during earlier stages of the disease. Organs can communicate with one another via several potential mechanisms, including EVs, cytokines and chemokines, PAMPs and DAMPs, and the nervous system. The liver is proposed to communicate with other organs, such as the gut, brain, lung, and adipose tissue using these mechanisms, as well as others. Understanding the mechanisms by which organs communicate during the inflammatory injury phase



of ALD may allow for the development of targeted therapeutics to protect one or all of these systems from alcohol-mediated toxicities.

## Conflicts of Interest

The authors have no conflicts to disclose.

## REFERENCES

1. Bergheim I, McClain CJ, Arteel GE. Treatment of alcoholic liver disease. *Dig Dis* 2005;23:275-284.
2. La Vecchia C, Negri E, D'Avanzo B, Boyle P, Franceschi S. Medical history and primary liver cancer. *Cancer Res* 1990;50:6274-6277.
3. Venook AP, Papandreou C, Furuse J, de Guevara LL. The incidence and epidemiology of hepatocellular carcinoma: a global and regional perspective. *Oncologist* 2010;15 Suppl 4:5-13.
4. Jong GM, Hsiue TR, Chen CR, Chang HY, Chen CW. Rapidly fatal outcome of bacteremic *Klebsiella pneumoniae* pneumonia in alcoholics. *Chest* 1995;107:214-217.
5. Perlino CA, Rimland D. Alcoholism, leukopenia, and pneumococcal sepsis. *Am Rev Respir Dis* 1985;132:757-760.
6. Kershaw CD, Guidot DM. Alcoholic lung disease. *Alcohol Res Health* 2008;31:66-75.
7. Wyatt TA, Gentry-Nielsen MJ, Pavlik JA, Sisson JH. Desensitization of PKA-stimulated ciliary beat frequency in an ethanol-fed rat model of cigarette smoke exposure. *Alcohol Clin Exp Res* 2004;28:998-1004.
8. Zhang P, Bagby GJ, Xie M, Stoltz DA, Summer WR, Nelson S. Acute ethanol intoxication inhibits neutrophil beta2-integrin expression in rats during endotoxemia. *Alcohol Clin Exp Res* 1998;22:135-141.
9. Moss M, Bucher B, Moore FA, Moore EE, Parsons PE. The role of chronic alcohol abuse in the development of acute respiratory distress syndrome in adults. *JAMA* 1996;275:50-54.
10. ARDS Definition Task Force, Ranieri VM, Rubenfeld GD, Thompson BT, Ferguson ND, Caldwell E, et al. Acute respiratory distress syndrome: the Berlin Definition. *JAMA* 2012;307:2526-2533.
11. Poole LG, Beier JI, Torres-Gonzales E, Schlueter CF, Hudson SV, Artis A, et al. Chronic + binge alcohol exposure promotes inflammation and alters airway mechanics in the lung. *Alcohol* 2019;80:53-63.
12. Afshar M, Smith GS, Terrin ML, Barrett M, Lissauer ME, Mansoor S, et al. Blood alcohol content, injury severity, and adult respiratory distress syndrome. *J Trauma Acute Care Surg* 2014;76:1447-1455.
13. Moss M, Parsons PE, Steinberg KP, Hudson LD, Guidot DM, Burnham EL, et al. Chronic alcohol abuse is associated with an increased incidence of acute respiratory distress syndrome and severity of multiple organ dysfunction in patients with septic shock. *Crit Care Med* 2003;31:869-877.
14. Seitz HK, Bataller R, Cortez-Pinto H, Gao B, Gual A, Lackner C, et al. Alcoholic liver disease. *Nat Rev Dis Primers* 2018;4:16.
15. Siore AM, Parker RE, Stecenko AA, Cuppels C, McKean M, Christman BW, et al. Endotoxin-induced acute lung injury requires interaction with the liver. *Am J Physiol Lung Cell Mol Physiol* 2005;289:L769-L776.
16. McClain CJ, Barve S, Deaciuc I, Kugelmass M, Hill D. Cytokines in alcoholic liver disease. *Semin Liver Dis* 1999;19:205-219.
17. Bode C, Kugler V, Bode JC. Endotoxemia in patients with alcoholic and non-alcoholic cirrhosis and in subjects with no evidence of chronic liver disease following acute alcohol excess. *J Hepatol* 1987;4:8-14.
18. Nolan JP. The role of endotoxin in liver injury. *Gastroenterology* 1975;69:1346-1356.
19. Khoruts A, Stahnke L, McClain CJ, Logan G, Allen JI. Circulating tumor necrosis factor, interleukin-1 and interleukin-6 concentrations in chronic alcoholic patients. *Hepatology* 1991;13:267-276.
20. Jimuro Y, Gallucci RM, Luster MI, Kono H, Thurman RG. Antibodies to tumor necrosis factor alpha attenuate hepatic necrosis and inflammation caused by chronic exposure to ethanol in the rat. *Hepatology* 1997;26:1530-1537.
21. Yin M, Wheeler MD, Kono H, Bradford BU, Gallucci RM, Luster MI, et al. Essential role of tumor necrosis factor alpha in alcohol-induced liver injury in mice. *Gastroenterology* 1999;117:942-952.
22. Massey VL, Poole LG, Siow DL, Torres E, Warner NL, Schmidt RH, et al. Chronic alcohol exposure enhances lipopolysaccharide-induced lung injury in mice: potential role of systemic tumor necrosis factor-alpha. *Alcohol Clin Exp Res* 2015;39:1978-1988.
23. Qin L, Wu X, Block ML, Liu Y, Breese GR, Hong JS, et al. Systemic LPS causes chronic neuroinflammation and progressive neurodegeneration. *Glia* 2007;55:453-462.
24. Bautista AP, Skrepnik N, Niesman MR, Bagby GJ. Elimination of macrophages by liposome-encapsulated dichloromethylene diphosphonate suppresses the endotoxin-induced priming of Kupffer cells. *J Leukoc Biol* 1994;55:321-327.
25. Maji S, Matsuda A, Yan IK, Parasramka M, Patel T. Extracellular vesicles in liver diseases. *Am J Physiol Gastrointest Liver Physiol* 2017;312:G194-G200.
26. Ogasawara F, Fusegawa H, Haruki Y, Shiraishi K, Watanabe N, Matsuzaki S. Platelet activation in patients with alcoholic liver disease. *Tokai J Exp Clin Med* 2005;30:41-48.
27. Eguchi A, Lazaro RG, Wang J, Kim J, Povero D, Williams B, et al. Extracellular vesicles released by hepatocytes from gastric infusion model of alcoholic liver disease contain a microRNA barcode that can be detected in blood. *Hepatology* 2017;65:475-490.
28. Yoshioka Y, Katsuda T, Ochiya T. Circulating microRNAs as hormones: intercellular and inter-organ conveyors of epigenetic information.

- mation? *Exp Suppl* 2015;106:255-267.
29. Mahida RY, Matsumoto S, Matthay MA. Extracellular vesicles: a new frontier for research in acute respiratory distress syndrome. *Am J Respir Cell Mol Biol* 2020;63:15-24.
  30. Ma F, Liu H, Shen Y, Zhang Y, Pan S. Platelet-derived microvesicles are involved in cardio-protective effects of remote preconditioning. *Int J Clin Exp Pathol* 2015;8:10832-10839.
  31. Giricz Z, Varga ZV, Baranyai T, Sipos P, Pálóczi K, Kittel Á, et al. Cardioprotection by remote ischemic preconditioning of the rat heart is mediated by extracellular vesicles. *J Mol Cell Cardiol* 2014;68:75-78.
  32. Tilg H, Moschen AR, Szabo G. Interleukin-1 and inflammasomes in alcoholic liver disease/acute alcoholic hepatitis and nonalcoholic fatty liver disease/nonalcoholic steatohepatitis. *Hepatology* 2016;64:955-965.
  33. Hirsiger S, Simmen HP, Werner CM, Wanner GA, Rittirsch D. Danger signals activating the immune response after trauma. *Mediators Inflamm* 2012;2012:315941.
  34. Gustot T, Lemmers A, Moreno C, Nagy N, Quertinmont E, Nicaise C, et al. Differential liver sensitization to toll-like receptor pathways in mice with alcoholic fatty liver. *Hepatology* 2006;43:989-1000.
  35. Pavlov M. The anti-toxic function of the liver. *Lancet* 1893;2:1092.
  36. Rutenburg AM, Sonnenblick E, Koven I, Aprahamian HA, Reiner L, Fine J. The role of intestinal bacteria in the development of dietary cirrhosis in rats. *J Exp Med* 1957;106:1-14.
  37. Nolan JP, Ali MV. Endotoxin and the liver. II. Effect of tolerance on carbon tetrachloride-induced injury. *J Med* 1973;4:28-38.
  38. Nolan JP, Leibowitz AI. Endotoxin and the liver. III. Modification of acute carbon tetrachloride injury by polymyxin b--an antiendotoxin. *Gastroenterology* 1978;75:445-449.
  39. Wilkinson SP, Arroyo V, Gazzard BG, Moodie H, Williams R. Relation of renal impairment and haemorrhagic diathesis to endotoxaemia in fulminant hepatic failure. *Lancet* 1974;1:521-524.
  40. Tarao K, Moroi T, Nagakura Y, Ikeuchi T, Suyama T, Endo O, et al. Relationship between endotoxaemia and protein concentration of ascites in cirrhotic patients. *Gut* 1979;20:205-210.
  41. Ge X, Antoine DJ, Lu Y, Arriazu E, Leung TM, Klepper AL, et al. High mobility group box-1 (HMGB1) participates in the pathogenesis of alcoholic liver disease (ALD). *J Biol Chem* 2014;289:22672-22691.
  42. Coleman LG Jr, Zou J, Crews FT. Microglial-derived miRNA let-7 and HMGB1 contribute to ethanol-induced neurotoxicity via TLR7. *J Neuroinflammation* 2017;14:22.
  43. Zou JY, Crews FT. Release of neuronal HMGB1 by ethanol through decreased HDAC activity activates brain neuroimmune signaling. *PLoS One* 2014;9:e87915.
  44. Ren Z, Wang X, Xu M, Yang F, Frank JA, Ke ZJ, et al. Binge ethanol exposure causes endoplasmic reticulum stress, oxidative stress and tissue injury in the pancreas. *Oncotarget* 2016;7:54303-54316.
  45. Zhang Q, Raoof M, Chen Y, Sumi Y, Sursal T, Junger W, et al. Circulating mitochondrial DAMPs cause inflammatory responses to injury. *Nature* 2010;464:104-107.
  46. Michelena J, Altamirano J, Abrales JG, Affò S, Morales-Ibanez O, Sancho-Bru P, et al. Systemic inflammatory response and serum lipopolysaccharide levels predict multiple organ failure and death in alcoholic hepatitis. *Hepatology* 2015;62:762-772.
  47. Rui L. Energy metabolism in the liver. *Compr Physiol* 2014;4:177-197.
  48. Oh KJ, Lee DS, Kim WK, Han BS, Lee SC, Bae KH. Metabolic adaptation in obesity and type II diabetes: myokines, adipokines and hepatokines. *Int J Mol Sci* 2016;18:8.
  49. Lebensztejn DM, Flisiak-Jackiewicz M, Białokoz-Kalinowska I, Bobrus-Chociej A, Kowalska I. Hepatokines and non-alcoholic fatty liver disease. *Acta Biochim Pol* 2016;63:459-467.
  50. Yamada T, Oka Y, Katagiri H. Inter-organ metabolic communication involved in energy homeostasis: potential therapeutic targets for obesity and metabolic syndrome. *Pharmacol Ther* 2008;117:188-198.
  51. Ferro JM, Oliveira S. Neurologic manifestations of gastrointestinal and liver diseases. *Curr Neurol Neurosci Rep* 2014;14:487.
  52. de la Monte SM. Insulin resistance and neurodegeneration: progress towards the development of new therapeutics for Alzheimer's disease. *Drugs* 2017;77:47-65.
  53. Klingman GI, Goodall M. Urinary epinephrine and levarterenol excretion during acute sublethal alcohol intoxication in dogs. *J Pharmacol Exp Ther* 1957;121:313-318.
  54. Bravo IR, Acevedo CG, Gallardo V. Acute effects of ethanol on liver blood circulation in the anesthetized dog. *Alcohol Clin Exp Res* 1980;4:248-253.
  55. Yuki T, Bradford BU, Thurman RG. Role of hormones in the mechanism of the swift increase in alcohol metabolism in the rat. *Pharmacol Biochem Behav* 1980;13 Suppl 1:67-71.
  56. Patterson-Buckendahl P, Kubovcakova L, Krizanova O, Pohorecky LA, Kvetnansky R. Ethanol consumption increases rat stress hormones and adrenomedullary gene expression. *Alcohol* 2005;37:157-166.
  57. Poole LG, Dolin CE, Arteel GE. Organ-organ crosstalk in alcoholic liver disease. *Biomolecules* 2017;7:62.