



Misunderstandings Between Platelets and Neutrophils Build in Chronic Inflammation

Giuseppe A. Ramirez^{1,2}, Angelo A. Manfredi^{1,2} and Norma Maugeri^{1,2*}

¹ Vita-Salute San Raffaele University, Milan, Italy, ² Division of Immunology, Transplantation and Infectious Diseases, IRCCS Ospedale San Raffaele, Milan, Italy

Regulated hemostasis, inflammation and innate immunity entail extensive interactions between platelets and neutrophils. Under physiological conditions, vascular inflammation offers a template for the establishment of effective intravascular immunity, with platelets providing neutrophils with an array of signals that increase their activation threshold, thus limiting collateral damage to tissues and promoting termination of the inflammatory response. By contrast, persistent systemic inflammation as observed in immune-mediated diseases, such as systemic vasculitides, systemic sclerosis, systemic lupus erythematosus or rheumatoid arthritis is characterized by platelet and neutrophil reciprocal activation, which ultimately culminates in the generation of thrombo-inflammatory lesions, fostering vascular injury and organ damage. Here, we discuss recent evidence regarding the multifaceted aspects of platelet-neutrophil interactions from bone marrow precursors to shed microparticles. Moreover, we analyse shared and disease-specific events due to an aberrant deployment of these interactions in human diseases. To restore communications between the pillars of the immune-hemostatic continuum constitutes a fascinating challenge for the near future.

OPEN ACCESS

Edited by:

Craig N. Jenne, University of Calgary, Canada

Reviewed by:

Rostyslav Bilyy, Danylo Halytsky Lviv National Medical University, Ukraine Hadi Goubran Messiha, University of Saskatchewan, Canada

> *Correspondence: Norma Maugeri maugeri.norma@hsr.it

Specialty section:

This article was submitted to Molecular Innate Immunity, a section of the journal Frontiers in Immunology

Received: 04 August 2019 Accepted: 07 October 2019 Published: 22 October 2019

Citation:

Ramirez GA, Manfredi AA and Maugeri N (2019) Misunderstandings Between Platelets and Neutrophils Build in Chronic Inflammation. Front. Immunol. 10:2491. doi: 10.3389/fimmu.2019.02491 Keywords: platelets, neutrophil, inflammation, autoimmunity, systemic lupus erythematosus, systemic sclerosis, rheumatoid arthritis, vasculitis

INTRODUCTION

The Immune-Hemostatic Continuum

The circulatory system provides functional integration to tissues throughout the body and constitutes a dynamic platform for tasks, such as immune patrolling and defense against threats (1-3). Consistently, abnormalities in blood cells and cardiovascular manifestations are disproportionately represented in systemic autoimmune diseases, reflecting a network of interactions among circulating elements in the blood (4-7). Humoral moieties, such as complement, opsonins, components of the coagulation cascade and regulators of the vascular tone lie at the lowest level of complexity in this system and provide immediate, stereotyped responses to abnormal changes in the environment, such as volume loss, vascular injury or pathogen invasion, besides supporting more elaborate, long-term tasks performed by cells or subcellular elements (8–11).

Cellular membranes allow the segregation of selected information in compartments and modulate their subsequent effects on the environment by integrating multiple stimuli. Circulating membrane-endowed players in the immuno-hemostatic network encompass leukocytes, platelets and microparticles, with the endothelium as a fourth static counterpart (12, 13). Platelets and neutrophils play a crucial role in the maintenance of vascular and tissue integrity and interact extensively and productively.

1

Consistently, alterations in platelet-neutrophil cross-talk have dramatic long-range effects on vascular and immune homeostasis (14–16) and are growingly appreciated as targets for therapeutic intervention (17, 18).

PATHOPHYSIOLOGY OF INTERACTIONS BETWEEN PLATELETS AND NEUTROPHILS

Characters on the Stage: Platelets, Neutrophils, and Microparticles

Hemostasis and inflammation counterbalance the effect of injuring external stimuli. Selective regulation and polarization of these pathways enhance homeostatic responses at sites of tissue or vascular injury and minimize the detrimental effects to the host. Multiple mechanisms have developed, including variability in the lifespan of players involved in the immunehemostatic balance and availability of soluble or membranebound moieties. Emission of membrane-endowed subcellular particles fine-tunes cellular activation and converts locally concentrated high-intensity responses into a sum of smaller but widespread and reciprocally independent biological events. Generation of extracellular vesicles enables the extension of the total membrane area interacting with the environment as well as of the range of potential cellular targets. In addition, segregation of information in multiple signaling quanta discloses the possibility of independent interactions with distinct cellular counterparts according to the differential needs of target tissues. Platelets are anucleate cell fragments released by megakaryocytes. After a multi-stage process of cytoplasm compartmentalisation and concentration of bioactive compounds into granules taking place over the course of days, platelets are released as elongated precursors (proplatelets), which undergo multiple iterative fission events to reach their final size. Preplatelets are roundshaped precursors constituting a reversible intermediate stage in the transition from proplatelets to platelets (19). Small vessels of the bone marrow, spleen and lung might deliver signals that facilitate the final process of platelet maturation (20).

Increased platelet demand and/or consumption during acute systemic inflammation warrants adaptation of megakaryocytes. Inflammatory cytokines, such as IL6 promote megakaryocyte increase in ploidy and prompt thrombocytopoiesis through increased liver synthesis of thrombopoietin as part of the acute phase response (21). Platelets released under inflammatory stress are usually larger in volume, which correlates with an increased ischemic risk at a clinical level (22). Alternative sites of thrombocytopoiesis, such as the lung, might become activated under stress conditions in mice (23) and possibly in patients with lung cancer (24). In addition, stem-like megakaryocyte progenitors can be activated on demand during interferon-adriven inflammatory responses (25). Platelets survive for 7-10 days in circulation, where they surrogate the damaged endothelium during vascular injury, recognize and control invading pathogens and release stimuli to promote tissue repair (12).

Controlled exocytosis or integration of bioactive compounds into platelet membrane is crucial for these tasks. Platelets are endowed with three classes of granules: alpha-granules; dense granules and few lysosomes. Some authors also described "T granules" equipped with Toll-like receptor 9 as a potential fourth platelet compartment (26-28). Besides being providers of bioactive compounds through exocytosis, platelets also produce microparticles. Platelet-derived microparticles (PDµP) constitute a substantial fraction of circulating microparticles in humans under physiological conditions (29). PDµP can present with a variety of sizes, contents and functions (30-32) that range from facilitation of coagulation through tissue factor (TF) and phospholipid (phosphatidylserine) scaffolds (33, 34) to angiogenesis, tissue repair (35-37) and defensive responses (38). Modulation of neutrophil behavior through delivery of nucleic acids (RNA) or inflammatory signal intercellular transfer also occurs under inflammatory conditions (39-42). In addition, mitochondria-enriched PDµP modulate target cell metabolism (32). Megakaryocytes release microparticles as well, influencing bone marrow homeostasis and synergising with PDµP (32, 43, 44).

Neutrophils constitute the most abundant leukocyte population in the blood and are in charge of the early innate effector response to noxious stimuli (45). Neutrophils express a vast array of oxidative and proteolytic enzymes, which are preformed and stored in ready-to-use granules. Activated neutrophils express TF, promoting isolation of injured tissues through thrombosis (46, 47) and contributing to immunethrombosis upon interaction with platelets (2, 3). Neutrophils have a limited lifespan (45), which is mirrored by the timing of multiple acute or hyperacute clinical manifestations of infectious and immune-mediated diseases (48-50). Autophagy induction under inflammatory conditions might extend neutrophil survival, causing chronicization of tissue damage and facilitating autoimmunity (40, 51-55). In addition hematopoietic stem cells and myeloid progenitors respond to extreme inflammatory stimuli (thanks to the expression of innate germline encoded receptors, such as Toll-like receptors) causing massive granulopoiesis (56, 57).

Phagocytosis and digestion of invading pathogens constitutes the default-mode defensive task performed by neutrophils. However, frustrated microbial phagocytosis promotes the generation of extracellular traps (NETs) (58), i.e., the extracellular release of threads of decondensed chromatin and microbicidal moieties with or without loss of membrane integrity and cell vitality (suicidal vs. vital NET generation) (59). At least in cases leading to cell death, granule content is partially repurposed to facilitate chromatin remodeling and histone citrullination (60-63). Besides having a role in antimicrobial defense, NETs are also generated in response to unconventional stimuli, such as amorphous crystals, apoptotic bodies, cytokines, microparticles and changes in osmolarity (64, 65). Factors driving neutrophils to "choose" NET generation as opposed to phagocytosis are partially understood (66). Unsolicited formation and impaired clearance of NETs implies persistent exposure of self-antigens and inflammatory stimuli, which facilitates the development of autoimmunity (67–73).

Neutrophils also account for tissue and vascular damage in immune-mediated diseases including inflammatory bowel diseases, systemic lupus erythematosus (SLE) rheumatoid arthritis (RA) and vasculitides either directly or through NETinduced facilitation of thrombosis (2, 67, 68, 74-76). Neutrophils cooperate to regulate immune activation by secreting soluble pattern recognition receptors, such as pentraxin-3 (PTX3) or ficolins (3, 77, 78). Furthermore, they influence the activation and survival of other immunocompetent cells and tune the degree of systemic inflammation (45). Finally, neutrophils emit microparticles loaded with nucleic acids and/or digestive enzymes and armed with tissue factor (47, 79, 80). Neutrophilderived microparticles prime endothelial cells, macrophages and neutrophils themselves to inflammatory activation (80, 81) and possibly destabilize genomic integrity in target tissues preventing resolution of the immune response (82). Elevated levels of such microparticles, possibly interacting with NET constituents are detectable in blood of patients with immune-mediated diseases (83, 84).

Weaving the Plot: General Features of Platelet-Neutrophil Interactions

Platelets interact productively with multiple cells types (12), although neutrophils constitute preferential partners (62) (**Figure 1**). P-selectin expression on platelet surface (which interacts with P-selectin granulocyte ligand 1, PSGL1, constitutively expressed by neutrophils) initiates platelet-neutrophil interaction (85, 86) and occurs after vessel injury/endothelial activation (87), pathogen recognition (88), or aging (89). P-selectin-dependent platelet-neutrophil interaction recruits downstream integrin-dependent pathways and culminates in neutrophil activation and facilitated extravasation (90). Neutrophils interacting with platelets can either: (a) phagocytose them, quenching their thrombogenic and inflammatory potential (85); (b) progressing to the generation of NETs (88), an event influenced by the neutrophil metabolic state (66, 91, 92).

Vital neutrophil internalization (emperilopolesis) into megakaryocytes occurs in the bone marrow. Engulfed neutrophils provide megakaryocytes with activating stimuli (causing a rise in platelet production) and donate membrane segments causing enhanced phosphatidylserine expression by chimeric daughter platelets (93). It is tempting to speculate that emperilopolesis holds the key to understanding the mechanisms of the later interactions between neutrophils and platelets in the circulation.

Platelets and megakaryocytes communicate long-range with neutrophils through exocytosed mediators and microparticles with enhancing actions on neutrophil activation (94, 95). The prototypic alarmin/damage-associated molecular pattern HMGB1, either as a soluble moiety or loaded into microparticles, is a potent promoter of extended neutrophil survival and of NET generation (40, 51, 96–99). Cooperation and bidirectional exchange of lipid metabolites between neutrophils and platelets through microparticles maximize the synthesis of prostaglandins, such as the pro-coagulant and vasoconstrictor signal, thromboxane A2 (100, 101) and to the activation of the complement cascade, which in turn promotes neutrophil recruitment and activation (102).

An Anthology of Recurring *Topoi*: Shared Events Linked to Platelet and Neutrophil Biology

The interaction between platelets and neutrophils impacts on multiple stereotyped pathological manifestations (Table 1). A first hint can be found in laboratory tests, such as blood cell counts. In most cases, platelet and leukocyte numbers roughly correlate with systemic inflammation (147), either due to disease activity or infections. By contrast, reduced blood cell counts are a hallmark of a minority of autoimmune conditions, including SLE and overlap syndromes and Felty's syndrome, but might dominate during sepsis, affecting survival. No specific evidence is available about the reciprocal interactions between platelets and neutrophils during severe thrombocytopenia as an isolated phenomenon. This fact might also be due to difficulties in uncoupling variations in platelet counts and neutrophil responses from shared inciting stimuli underlying both events at an experimental level. Nonetheless, consistent evidence from sepsis, idiopathic thrombocytopenic purpura, thrombotic thrombocytopenic purpura and thrombocytopenia induced by heparin or other drugs suggests that neutrophils are intravascularly activated and tend to generate NETs in association with low platelet counts, possibly contributing to dysregulated hemostasis and adverse clinical outcomes (148-153). Interestingly, viral infections causing thrombocytopenia seem to associate with the expansion of low-density neutrophils (which are thought to have a higher propensity to form NETs), as observed in autoimmune diseases, such as SLE (154). Pancytopenia is a distinctive feature of hemophagocytic lymphohistiocytosis, a severe disorder developing either as a complication of multiple autoimmune diseases or as a standalone disorder (155). Variations in cell volume are thought to correlate with cytoskeletal remodeling and changes in cell function and metabolism (156-159). Mean platelet volume (MPV) is a variable provided in the context of blood count, significantly susceptible to the effects of multiple confounders at the analytical and preanalytical level. Altered MPV has been detected in SLE, RA, large- and small-vessel vasculitides, systemic sclerosis (SSc) and chronic spontaneous urticaria (103, 104, 117, 118, 123, 129, 135, 136, 160). However, disease activity in these settings has been associated with high and low MPV values, preventing a straightforward translation of this laboratory tool into clinical practice (161). Gasparyan et al. (137) and Scherlinger et al. (97) offer a possible syncretistic perspective: high platelet volume might reflect chronic low-grade platelet activation (156), whereas low MPV can be consequent to more severe inflammation, causing extensive shedding of platelet microparticles (32). Neutrophils showing increased cellular volume and consequent lower granular density have long been recognized as a hallmark of systemic immune-mediated diseases (159), although they are also detectable in sepsis (162) and cancer (163). This cell population is characterized by enhanced ability to extravasate,



FIGURE 1 Platelet-neutrophil crosstalk. A complex network of interactions connects platelet (PLT) to neutrophil (N) biology. Megakaryocytes (MK) are able to interact with neutrophils residing in the bone marrow and engulf them, preserving their vitality (a phenomenon called emperilopolesis). Neutrophils may eventually escape megakaryocyte engulfment after donating membrane components to the host cell. This event causes enhanced platelet production and release of chimeric platelets. Activated platelets and neutrophils can further interact in the circulating blood, either directly through cell-cell contact and/or through the exchange of soluble compounds or microparticles. Engagement of platelets by neutrophils through the P-selectin/PSGL-1 axis, and later on, integrin-mediated bonds can lead to platelet-phagocytosis, resulting in neutrophil exhaustion. Alternatively, failed platelet clearance can promote neutrophil activation (heralded by expression of surface markers, such as tissue factor and activation of integrins – red spikes in the figure) and facilitate neutrophil extravasation through the endothelial wall (E). Activated neutrophils can also enter autophagy programmes (green circular arrow), extending their survival, and progress toward the formation of neutrophil extracellular traps (NET). Platelet-derived microparticles (NDµP) constitute and additional channel for platelet-neutrophil interchanges and are thought to have a role in lipid metabolism.

extended survival and by a tendency to form NETs (68, 158, 164). By contrast, consistent with the model of inverse association between NETosis and phagocytosis (**Figure 1**), low density neutrophils are less able to engulf substrates (159).

Patients with inflammatory diseases of the blood vessels (124, 130, 131), connective tissue diseases and chronic arthritides have higher risks of ischemic events (105, 106, 113, 135, 138, 139, 165-167). Accelerated atherosclerosis is a common finding in SLE (7, 106, 168-170), RA (7, 139, 165, 171), or systemic vasculitides (170). Inflammation-related atherosclerotic mechanisms are only partially understood (172). NETs are major determinants of endothelial dysfunction and have been detected in atherosclerotic lesions (173). Platelets are also involved in the early phases of atherosclerosis, where they facilitate leukocyte egress toward the subendothelial vascular layers (174, 175). Antiphospholipid antibodies (aPL) constitute a major independent risk factor in the antiphospholipid syndrome (APS), a condition characterized by arterial or venous thrombosis and/or pregnancy complications occurring as a standalone disorder or secondary to SLE and other autoimmune diseases (176). aPL promote HMGB1-related response in platelets and monocytes (177) and, in cooperation with platelet Toll-like receptor 4, induce NETosis (178), possibly contributing to immunothrombosis (76). In addition, they might impair microparticle scavenging by glycoprotein I, increasing the likelihood of pro-coagulant platelet-neutrophil activation (179). Besides atherosclerotic lesions, NETs have been detected into coronary thrombi (96, 180).

Aberrant coagulation is detectable in immune mediated diseases (34, 107, 108, 119). Altered coagulation cascade and increased cardiovascular risk are common in asthmatic patients (181, 182), while patients with chronic spontaneous urticaria show aberrant thrombin generation but suffer no excess prevalence of cardiovascular disease (125, 141–146).

Thrombotic microangiopathy, consisting in diffuse deposition of thrombi along small vessels due to widespread endothelial activation with hemolysis and platelet consumption, might complicate SLE, SSc, antiphospholipid syndrome and other immune-mediated diseases (183–185). Under intravascular hemolytic conditions, free hemoglobin favors platelet activation. In turn, activated platelet boost neutrophil activation, possibly further feeding the inflammatory cascade (186). In addition, sera from patients with thrombotic microangiopathies fail to degrade NETs, which, in turn, can trigger thrombosis (76, 187).

The lung is a major inflammatory target (188). Neutrophils and platelets undergo unique pathophysiological interaction with the lung vasculature, a reservoir for neutrophils that can thus easily respond to airborne infectious or sterile inflammatory stimuli (189). Unleashed neutrophil effector functions contribute to acute lung injury/acute respiratory distress syndrome (ARDS) during sepsis or vasculitis (50), to the long-term effects on bronchial tissue of chronic inflammation in cystic fibrosis (190, 191) and to interstitial fibrosis in SSc (40, 192). NET generation has a central role in neutrophilic lung injury (193, 194). The lung vasculature influences platelets since lung microvessels are (a) a site of maturation of platelet precursors from the bone marrow (20); (b) a niche for human and mouse megakaryocyte homing (23, 195); (c) a site of detoxification of platelet mediators,

	Increased ischemic risk	Abnormal thrombin generation	Thrombotic microangiopathy	Lung involvement	Leukopenia	Thrombocytopenia	Altered platelet volume	Low complement	Vasculitis	Urticaria	Vascular remodeling	Fibrosis	References
SLE	+	+	-/+	+	+	+	+	+	-/+	-/+	I	I	(34, 103–112)
APS	+	+	-/+	+	I	-/+	Ċ	-/+	-/+	I	I	I	(113-116)
SSc	+	-/+	-/+	+	I	I	+	-/+	I	I	+	+	(117–122)
AN	+	+	I	+	I	I	+	I	+	-/+	I	I	(123–128)
N	+	ć	I	I	I	I	+	I	+	I	+	I	(126, 129–134)
٩	+	+	I	-/+	I	I	+	I	-/+	I	I	I	(119, 135–140)
NSU SSU	I	+	I	I	I	I	+	-/+	-/+	+	I	I	(141–146)

such as serotonin (196). Conversely platelets contribute to persistent vasoconstriction (potentially leading to pulmonary hypertension), interstitial fibrosis (197), and NET generation in acute lung injury (198).

ABNORMAL PLATELET-NEUTROPHIL INTERACTIONS IN SELECTED DISEASE SETTINGS

Systemic Vasculitides

Systemic vasculitides encompass a very large set of immunemediated diseases characterized by vascular injury and downstream organ ischemia as the core pathophysiological event (199). They can be roughly classified into two main subsets, large- and small-vessel vasculitides, based on (prominent) sites of vascular involvement. From a pathophysiological point of view, vascular remodeling with immune infiltration, disassembly of the physiological extracellular architecture and aberrant proliferation of macrophages, fibroblasts and endothelial cells predominate in large-vessel vasculitides, resulting in stroke-like acute failure of large portions of tissues or entire organ due to vessel occlusion. By contrast, small-vessel vasculitis are characterized by necrosis associated or not to granulomatous inflammation and increased thrombotic risk (200). Altered cross-talk among platelet, neutrophils and microparticles might contribute to vascular injury (201).

Neutrophils cause vascular damage in small-vessel vasculitis, possibly reflected by the accumulation of leukocyte cellular debris in peri-vasculitic lesions (leukocytoclasia). In anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitides (AAV), neutrophil mediated vascular injury is part of a vicious circle linking exposure of myeloperoxidase (MPO) and/or proteinase-3 from granules to neutrophil surface or into the setting of NETs (202) to the development of ANCA, which in turn activate neutrophils and the vascular endothelium (3, 67). In patients with AAV neutrophil responses couple with platelet activation (126). Consistently, patients with active AAV show increased plasmatic levels of P-selectin (127) and HMGB1 (203, 204). Elevation of HMGB1 has also been detected in other vasculitic settings, such as IgA-vasculitis (formerly Henoch-Schonlein's purpura) and Kawasaki disease (205).

In large vessel-vasculitides, activation of platelets may contribute to promote vascular remodeling through the release of signals, such as VEGF (206). Platelets in large-vessel vasculitides show signs of activation (129, 201), and in giant cell arteritis are significantly increased in number during active disease (147) and form hetero-aggregates with leukocytes, possibly contributing to exacerbate ischemic risk (132, 201). This evidence provides a rationale for the use of aspirin in primary prevention (207) and for the employment of platelets as diagnostic surrogates (126). Results in the literature (208) and reports on a potential cyclooxygenase-independent mechanism for aspirin in giant cell arteritis (209) suggest however that the role of platelets should be interpreted with caution.

Little is known on platelet-leukocyte interactions in large vessel vasculitides, although relative depletion of neutrophil

granule content has been reported in giant cell arteritis in association with platelet activation (126, 132). Notably, items of small-vessel (peri)vasculitis have been reported in large series of temporal artery biopsies from patients with giant cell arteritis (210) and might be secondary to stereotyped events resembling those observed in small-vessel vasculitides, possibly targeting the *vasa vasorum* and entailing aberrant platelet-neutrophil cross-talk.

Systemic Lupus Erythematosus

SLE is a multi-organ autoimmune disease with a wide spectrum of clinical manifestations and pathogenic mechanisms (211). Failure of clearance mechanisms and/or exposure of cell death debris in an inflammatory setting promotes autoimmunity and subsequent tissue damage (212). Hematological manifestations constitute a hallmark of SLE and are detected in >80% of patients (213). Cytopenia is the most frequent modality of presentation and affects either red blood cells, platelets and leukocytes. Bone marrow abnormalities are frequent, although no clear correlation can be established with disease activity (214-217). Accordingly, primary bone marrow failure is a rare cause of cytopenia (218), with most relevant mechanisms (besides drugs) being inflammation-induced iron deficiency and cytolysis. Neutropenia occurs in up to one third of SLE cases, in most cases due to antibodies (219), which is not apparently associated to infectious risk (109). Thrombocytopenia is also common in patients with SLE (220). Megakaryocyte number is generally increased during disease activity, reflecting extensive platelet production (214, 220). Patients with SLE and thrombocytopenia have an increased risk of a severe disease course and of mortality in large cohort studies (221).

Cardiovascular manifestations are frequent in patients with SLE and a cause of morbidity and mortality (222). Accelerated atherosclerosis, aPL and dysfunctional coagulation likely converge to determine this risk (106). Despite low absolute platelet counts, patients with SLE frequently show extensive platelet activation (223–227). Higher levels of P-selectin are detectable in urines from patients with lupus nephritis (228). Platelets also contribute to mesangial remodeling and renal vascular damage (229, 230).

Endothelial derived microparticles constitute the most abundant microparticle subset in patients with SLE and correlate with endothelial dysfunction and interferon- α signature (231, 232). However, PD μ P also accumulate during active SLE (233, 234) and might impact on inflammation and hemostasis (34, 234). PD μ P facilitate coagulation by providing phosphatidylserine scaffolds and intravascularly expressed TF. In addition, they promote neutrophil activation and NET generation being reservoirs of HMGB1 (96) and CD40L (234). Finally PD μ P synergise with NETs as inducers of anti-nuclear immunity by constituting a source of mitochondria, which behave as potent damage-associated molecular patterns due to their bacterial origin (235).

Mechanistically, platelet activation in SLE might depend on circulating immunocomplexes, which are abundant in SLE patients biological fluids and are recognized on platelet surface by $Fc\gamma RIIA$ and Toll-like receptor 4,7 (236). PD μ P themselves could take part in immunocomplexes, enforcing an inflammatory/immunogenic self-sustaining loop (235). Ensuing complement activation in turn amplifies and propagates neutrophil and platelet activation (102, 231, 234).

Systemic Sclerosis

SSc is a systemic autoimmune disease, characterized by unrelenting inflammation with a wound repair response consisting in mesenchymal extracellular matrix deposition leading to fibrosis, and by microvascular dysfunction and aberrant neoangiogenesis (120, 237, 238). Platelets and aberrant platelet-neutrophil interactions play a role in SSc (239). Possibly in response to microvascular damage, platelets of patients with SSc are constitutively activated and express signals driving neutrophil interaction (240, 241). P-selectin dependent cell-cell interactions seem to be relatively less represented in SSc, due to the lower leukocyte expression of PSGL-1 (242). Neutrophils have a pericellular distribution of granules and of their content, causing enhanced degradation of fibrinogen by exposed neutrophil proteases and eventually impairing fibrinogen dependent interactions between neutrophil CD11b/CD18 (also known as Mac-1 or aMB2 integrin) and platelet glycoprotein IIbIIIa (40, 96). Indeed, platelet-neutrophil heterotypic aggregates are less frequently detected in SSc compared to other inflammatory conditions (40, 96, 242).

Activated platelets in SSc contribute to impaired vascular tone [due to altered arachidonic acid metabolism (197, 239)] and to fibrosis. In fact, platelets release multiple fibrogenic mediators, such as transforming growth factor beta, plateletderived growth factor, CXCL4 (also known as platelet factor 4), beta-thromboglobulin, serotonin and HMGB1 (27, 97, 242-244). NETs promote fibroblast differentiation and function and might synergise with platelet in supporting fibrosis (192), also in light of the abundance of NET byproducts in the blood of patients with SSc (40). Synergistic NET/platelet-induced fibrosis is expected to be particularly significant in lung tissue, where neutrophil and platelets are abundant (189). Consistently, PDµP (retrieved from the plasma of patients with SSc) induce neutrophil granule mobilization and autophagy, culminating in extended neutrophil survival and generation of NETs through a HMGB1-dependent mechanism. Furthermore, neutrophils stimulated by SSc platelet microparticles migrate in murine lungs, associate with interstitial endothelial damage and promote lung fibrosis (40).

Rheumatoid Arthritis

Rheumatoid arthritis is a relatively frequent autoimmune disease characterized by prominent involvement of the synovial joints. Although extra-skeletal manifestations are relatively less frequent compared to other immune-mediated diseases, patients with RA show an increased ischemic risk, pointing to the existence of a core pathophysiological event linking inflammatory manifestations to vascular dysfunction (245).

Neutrophils undergoing NET generation might provide autoantigens in RA. Patients in fact frequently develop antibodies against citrullinated peptides (ACPA). Citrullination occurs thanks to the activity of deiminating enzymes, such as protein-arginine deiminase 4 (PAD4), abundantly expressed

in neutrophils (246). Citrullinated histones constitute a fundamental component of chromatin threads within NETs (73, 247). Platelets might also contribute to ACPA formation due to their expression of vimentin, a preferential target of citrullination (248). ACPA sustain joint inflammation by perpetuating macrophage activation within the synovia, eventually causing chronically elevated levels of tumor necrosis factor alpha (TNFa), which in turn promotes synovial proliferation, bone reabsorption, neoangiogenesis and enhanced synovial infiltration through activated endothelium (249). Enhanced expression of TF on activated platelets and neutrophils, which coexist in the synovial fluid of inflamed joints in patients with RA, provides an interesting hint on potential mechanisms involved in RA-associated enhanced ischemic risk. Platelets and leukocytes from patients with RA are activated due increased plasmatic concentration of TNFa (140). Indeed, platelet respond to TNFa thanks to the expression of TNF receptors 1 and 2 (250) resulting in increased P-selectin expression, platelet degranulation, phosphatidylserine up-regulation and TF expression. TNFα-activated platelets prompt thrombin generation and activation of leukocytes due to P-selectin. Consistently, platelet and leukocyte activation is reduced in patients treated with anti-TNF agents (140), who also face relatively lower rates of cardiovascular events in the long-term (251, 252).

Platelets can be activated by collagen through the megakaryocyte lineage-specific glycoprotein VI and thus prompted to generate microparticles. Boilard and colleagues (253) showed that, following this mechanism, high concentrations of PD μ P (possibly shuttled by engaging leukocytes) are detectable in synovial fluid of patients with RA and are required for arthritis development in a murine model. PD μ P contain significant amounts of interleukin 1 α and β , promoting synoviocyte proliferation, and of IL8, enhancing neutrophil recruitment and ensuring maintenance of inflammation (253).

CONCLUSION

Platelets and neutrophils are major determinants of the immune-hemostatic continuum and extensively interact based on cell-cell contact and/or exchange of soluble signals and microparticles to synergise in contrasting the noxious effects of endogenous or environmental stimuli toward vessel and tissue integrity and to promote physiological tissue renewal and homeostasis. These events, part of a set of simple, innate, but evolutionarily preserved stereotyped responses, are disproportionately active and self-sustained in patients with immune-mediated diseases, such as systemic vasculitides, SLE, SSc, RA and possibly allergic disorders and might account for the development of either some inflammatory manifestations and of cardiovascular complications. Patients with immunemediated diseases consistently show signs of platelet (and/or $PD\mu P$) activation, possibly prompting either the formation of heterotypic aggregates with neutrophils (as in giant cell arteritis) or neutrophil activation toward enhanced survival and eventually NET generation (as observed in small-vessel vasculitis, SLE, RA and to a higher extent SSc). Diagnostic and therapeutic strategies currently employed in the setting of autoimmune diseases to prevent disease progression and the occurrence of secondary complications are generally not targeted on these pathogenic mechanisms, suggesting the existence of a largely unexplored window of opportunity to improve survival and quality of life for patients by dampening sustained neutrophil-platelet interactions.

AUTHOR CONTRIBUTIONS

GR and NM collected the literature data, drafted, and revised the manuscript. AM revised the manuscript and

REFERENCES

- Keragala CB, Draxler DF, McQuilten ZK, Medcalf RL. Haemostasis and innate immunity–a complementary relationship: a review of the intricate relationship between coagulation and complement pathways. *Br J Haematol.* (2018) 180:782–98. doi: 10.1111/bjh.15062
- Engelmann B, Massberg S. Thrombosis as an intravascular effector of innate immunity. Nat Rev Immunol. (2013) 13:34–45. doi: 10.1038/nri3345
- Ramirez GA, Rovere-Querini P, Sabbadini MG, Manfredi AA. Parietal and intravascular innate mechanisms of vascular inflammation. *Arthritis Res Ther.* (2015) 17:16. doi: 10.1186/s13075-015-0528-2
- Mason JC, Libby P. Cardiovascular disease in patients with chronic inflammation: mechanisms underlying premature cardiovascular events in rheumatologic conditions. *Eur Heart J.* (2015) 36:482–9. doi: 10.1093/eurheartj/ehu403
- Fernandez-Gutierrez B, Perrotti PP, Gisbert JP, Domenech E, Fernandez-Nebro A, Canete JD, et al. Cardiovascular disease in immune-mediated inflammatory diseases: a cross-sectional analysis of 6 cohorts. *Medicine (Baltimore)*. (2017) 96:e7308. doi: 10.1097/MD.00000000000 07308
- Libby P, Nahrendorf M, Swirski FK. Leukocytes link local and systemic inflammation in ischemic cardiovascular disease: an expanded "Cardiovascular Continuum". J Am Coll Cardiol. (2016) 67:1091–103. doi: 10.1016/j.jacc.2015.12.048
- Symmons DP, Gabriel SE. Epidemiology of CVD in rheumatic disease, with a focus on RA and SLE. Nat Rev Rheumatol. (2011) 7:399–408. doi: 10.1038/nrrheum.2011.75
- Manfredi AA, Rovere-Querini P, Bottazzi B, Garlanda C, Mantovani A. Pentraxins, humoral innate immunity and tissue injury. *Curr Opin Immunol.* (2008) 20:538–44. doi: 10.1016/j.coi.2008.05.004
- Fischetti F, Tedesco F. Cross-talk between the complement system and endothelial cells in physiologic conditions and in vascular diseases. *Autoimmunity*. (2006) 39:417–28. doi: 10.1080/089169306007 39712
- de Bont CM, Boelens WC, Pruijn GJM. NETosis, complement, and coagulation: a triangular relationship. *Cell Mol Immunol.* (2019) 16:19–27. doi: 10.1038/s41423-018-0024-0
- 11. Cicardi M, Zuraw BL. Angioedema due to bradykinin dysregulation. J Allergy Clin Immunol Pract. (2018) 6:1132-41. doi: 10.1016/j.jaip.2018.04.022
- 12. Semple JW, Italiano JE Jr, Freedman J. Platelets and the immune continuum. *Nat Rev Immunol.* (2011) 11:264–74. doi: 10.1038/nri2956
- Arraud N, Linares R, Tan S, Gounou C, Pasquet JM, Mornet S, et al. Extracellular vesicles from blood plasma: determination of their morphology, size, phenotype and concentration. *J Thromb Haemost.* (2014) 12:614–27. doi: 10.1111/jth.12554
- Manfredi AA, Rovere-Querini P, Maugeri N. Dangerous connections: neutrophils and the phagocytic clearance of activated platelets. *Curr Opin Hematol.* (2010) 17:3–8. doi: 10.1097/MOH.0b013e3283324f97

provided critical analysis of intellectual content. All authors approved the final version of the manuscript and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work will appropriately be investigated and resolved.

FUNDING

This work of the authors was supported by grants from the Associazione Italiana Ricerca sul Cancro (AIRC IG 2017-2035) and the Ministero della Salute (RF-2013-02358715).

- Totani L, Evangelista V. Platelet-leukocyte interactions in cardiovascular disease and beyond. *Arterioscler Thromb Vasc Biol.* (2010) 30:2357–61. doi: 10.1161/ATVBAHA.110.207480
- McGregor L, Martin J, McGregor JL. Platelet-leukocyte aggregates and derived microparticles in inflammation, vascular remodelling and thrombosis. *Front Biosci.* (2006) 11:830–7. doi: 10.2741/1840
- Burnouf T, Burnouf PA, Wu YW, Chuang EY, Lu LS, Goubran H. Circulatory-cell-mediated nanotherapeutic approaches in disease targeting. *Drug Discov Today*. (2018) 23:934–43. doi: 10.1016/j.drudis.2017.08.012
- Baldini M, Manfredi AA, Maugeri N. Targeting platelet-neutrophil interactions in giant-cell arteritis. *Curr Pharma Des.* (2014) 20:567–74. doi: 10.2174/138161282004140213144840
- Thon JN, Montalvo A, Patel-Hett S, Devine MT, Richardson JL, Ehrlicher A, et al. Cytoskeletal mechanics of proplatelet maturation and platelet release. J Cell Biol. (2010) 191:861–74. doi: 10.1083/jcb.201006102
- Machlus KR, Italiano JE Jr. The incredible journey: from megakaryocyte development to platelet formation. J Cell Biol. (2013) 201:785–96. doi: 10.1083/jcb.201304054
- Kaushansky K. The molecular mechanisms that control thrombopoiesis. J Clin Invest. (2005) 115:3339–47. doi: 10.1172/JCI26674
- Slavka G, Perkmann T, Haslacher H, Greisenegger S, Marsik C, Wagner OF, et al. Mean platelet volume may represent a predictive parameter for overall vascular mortality and ischemic heart disease. *Arterioscler Thromb Vasc Biol.* (2011) 31:1215–8. doi: 10.1161/ATVBAHA.110.2 21788
- Lefrancais E, Ortiz-Munoz G, Caudrillier A, Mallavia B, Liu F, Sayah DM, et al. The lung is a site of platelet biogenesis and a reservoir for haematopoietic progenitors. *Nature.* (2017) 544:105–9. doi: 10.1038/nature21706
- Dejima H, Nakanishi H, Kuroda H, Yoshimura M, Sakakura N, Ueda N, et al. Detection of abundant megakaryocytes in pulmonary artery blood in lung cancer patients using a microfluidic platform. *Lung Cancer*. (2018) 125:128–35. doi: 10.1016/j.lungcan.2018.09.011
- Haas S, Hansson J, Klimmeck D, Loeffler D, Velten L, Uckelmann H, et al. Inflammation-Induced emergency megakaryopoiesis driven by hematopoietic stem cell-like megakaryocyte progenitors. *Cell Stem Cell*. (2015) 17:422–34. doi: 10.1016/j.stem.2015.07.007
- Sharda A, Flaumenhaft R. The life cycle of platelet granules. *F1000Res.* (2018) 7:236. doi: 10.12688/f1000research.13283.1
- 27. Nurden AT. Platelets, inflammation and tissue regeneration. *Thromb Haemost.* (2011) 105:S13–33. doi: 10.1160/THS10-11-0720
- Flaumenhaft R. Molecular basis of platelet granule secretion. Arterioscler Thromb Vasc Biol. (2003) 23:1152–60. doi: 10.1161/01.ATV.0000075965.88456.48
- Scherlinger M, Sisirak V, Richez C, Lazaro E, Duffau P, Blanco P. New insights on platelets and platelet-derived microparticles in systemic lupus erythematosus. *Curr Rheumatol Rep.* (2017) 19:48. doi: 10.1007/s11926-017-0678-0

- Pienimaeki-Roemer A, Kuhlmann K, Bottcher A, Konovalova T, Black A, Orso E, et al. Lipidomic and proteomic characterization of platelet extracellular vesicle subfractions from senescent platelets. *Transfusion*. (2015) 55:507–21. doi: 10.1111/trf.12874
- Dean WL, Lee MJ, Cummins TD, Schultz DJ, Powell DW. Proteomic and functional characterisation of platelet microparticle size classes. *Thromb Haemost.* (2009) 102:711–8. doi: 10.1160/TH09-04-243
- Boilard E, Duchez AC, Brisson A. The diversity of platelet microparticles. *Curr Opin Hematol.* (2015) 22:437–44. doi: 10.1097/MOH.00000000000166
- Key NS, Mackman N. Tissue factor and its measurement in whole blood, plasma, and microparticles. Semin Thromb Hemost. (2010) 36:865–75. doi: 10.1055/s-0030-1267040
- Pereira J, Alfaro G, Goycoolea M, Quiroga T, Ocqueteau M, Massardo L, et al. Circulating platelet-derived microparticles in systemic lupus erythematosus. Association with increased thrombin generation and procoagulant state. *Thromb Haemost.* (2006) 95:94–9. doi: 10.1160/TH05-05-0310
- Varon D, Hayon Y, Dashevsky O, Shai E. Involvement of platelet derived microparticles in tumor metastasis and tissue regeneration. *Thromb Res.* (2012) 130:S98–9. doi: 10.1016/j.thromres.2012.08.289
- Brill A, Dashevsky O, Rivo J, Gozal Y, Varon D. Plateletderived microparticles induce angiogenesis and stimulate postischemic revascularization. *Cardiovasc Res.* (2005) 67:30–8. doi: 10.1016/j.cardiores.2005.04.007
- 37. Gaetani E, Del Zompo F, Marcantoni M, Gatto I, Giarretta I, Porfidia A, et al. Microparticles produced by activated platelets carry a potent and functionally active angiogenic signal in subjects with Crohn's disease. *Int J Mol Sci.* (2018) 19:2921. doi: 10.3390/ijms19102921
- Sung PS, Huang TF, Hsieh SL. Extracellular vesicles from CLEC2-activated platelets enhance dengue virus-induced lethality via CLEC5A/TLR2. *Nat Commun.* (2019) 10:2402. doi: 10.1038/s41467-019-10360-4
- Laffont B, Corduan A, Rousseau M, Duchez AC, Lee CH, Boilard E, et al. Platelet microparticles reprogram macrophage gene expression and function. *Thromb Haemost.* (2016) 115:311–23. doi: 10.1160/th15-05-0389
- Maugeri N, Capobianco A, Rovere-Querini P, Ramirez GA, Tombetti E, Valle PD, et al. Platelet microparticles sustain autophagy-associated activation of neutrophils in systemic sclerosis. *Sci Transl Med.* (2018) 10:eaao3089. doi: 10.1126/scitranslmed.aao3089
- Provost P. The clinical significance of platelet microparticleassociated microRNAs. *Clin Chem Lab Med.* (2017) 55:657–66. doi: 10.1515/cclm-2016-0895
- Duchez AC, Boudreau LH, Naika GS, Bollinger J, Belleannee C, Cloutier N, et al. Platelet microparticles are internalized in neutrophils via the concerted activity of 12-lipoxygenase and secreted phospholipase A2-IIA. *Proc Natl Acad Sci USA*. (2015) 112:E3564–73. doi: 10.1073/pnas.1507905112
- Cunin P, Nigrovic PA. Megakaryocytes as immune cells. J Leukoc Biol. (2019) 105:1111–21. doi: 10.1002/JLB.MR0718-261RR
- 44. Jiang J, Kao CY, Papoutsakis ET. How do megakaryocytic microparticles target and deliver cargo to alter the fate of hematopoietic stem cells? *J Control Release.* (2017) 247:1–18. doi: 10.1016/j.jconrel.2016.12.021
- Mantovani A, Cassatella MA, Costantini C, Jaillon S. Neutrophils in the activation and regulation of innate and adaptive immunity. *Nat Rev Immunol.* (2011) 11:519–31. doi: 10.1038/nri3024
- Maugeri N, Brambilla M, Camera M, Carbone A, Tremoli E, Donati MB, et al. Human polymorphonuclear leukocytes produce and express functional tissue factor upon stimulation. *J Thromb Haemost.* (2006) 4:1323–30. doi: 10.1111/j.1538-7836.2006.01968.x
- 47. Kambas K, Chrysanthopoulou A, Vassilopoulos D, Apostolidou E, Skendros P, Girod A, et al. Tissue factor expression in neutrophil extracellular traps and neutrophil derived microparticles in antineutrophil cytoplasmic antibody associated vasculitis may promote thromboinflammation and the thrombophilic state associated with the disease. *Ann Rheum Dis.* (2014) 73:1854–63. doi: 10.1136/annrheumdis-2013-203430
- Schett G, Schauer C, Hoffmann M, Herrmann M. Why does the gout attack stop? A roadmap for the immune pathogenesis of gout. *RMD Open.* (2015) 1:e000046. doi: 10.1136/rmdopen-2015-000046
- 49. Hu Q, Shi H, Zeng T, Liu H, Su Y, Cheng X, et al. Increased neutrophil extracellular traps activate NLRP3 and inflammatory

macrophages in adult-onset Still's disease. Arthritis Res Ther. (2019) 21:9. doi: 10.1186/s13075-018-1800-z

- 50. Rebetz J, Semple JW, Kapur R. The pathogenic involvement of neutrophils in acute respiratory distress syndrome and transfusion-related acute lung injury. *Transf Med Hemother*. (2018) 45:290–8. doi: 10.1159/000492950
- Manfredi AA, Rovere-Querini P, D'Angelo A, Maugeri N. Low molecular weight heparins prevent the induction of autophagy of activated neutrophils and the formation of neutrophil extracellular traps. *Pharmacol Res.* (2017) 123:146–56. doi: 10.1016/j.phrs.2016.08.008
- Mitroulis I, Kambas K, Chrysanthopoulou A, Skendros P, Apostolidou E, Kourtzelis I, et al. Neutrophil extracellular trap formation is associated with IL-1beta and autophagy-related signaling in gout. *PLoS ONE.* (2011) 6:e29318. doi: 10.1371/journal.pone.00 29318
- Pan Q, Gao C, Chen Y, Feng Y, Liu WJ, Liu HF. Update on the role of autophagy in systemic lupus erythematosus: a novel therapeutic target. *Biomed Pharmacother*. (2015) 71:190–3. doi: 10.1016/j.biopha.2015.02.017
- Remijsen Q, Vanden Berghe T, Wirawan E, Asselbergh B, Parthoens E, De Rycke R, et al. Neutrophil extracellular trap cell death requires both autophagy and superoxide generation. *Cell Res.* (2011) 21:290–304. doi: 10.1038/cr.2010.150
- 55. Tang S, Zhang Y, Yin S, Gao X, Shi W, Wang Y, et al. Neutrophil extracellular trap formation is associated with autophagy-related signaling in ANCA-associated vasculitis. *Clin Exp Immunol.* (2015) 180:408–18. doi: 10.1111/cei.12589
- Manz MG, Boettcher S. Emergency granulopoiesis. Nat Rev Immunol. (2014) 14:302–14. doi: 10.1038/nri3660
- Chavakis T, Mitroulis I, Hajishengallis G. Hematopoietic progenitor cells as integrative hubs for adaptation to and fine-tuning of inflammation. *Nat Immunol.* (2019) 20:802–11. doi: 10.1038/s41590-019-0402-5
- Wartha F, Beiter K, Normark S, Henriques-Normark B. Neutrophil extracellular traps: casting the NET over pathogenesis. *Curr Opin Microbiol.* (2007) 10:52–6. doi: 10.1016/j.mib.2006.12.005
- Boeltz S, Amini P, Anders HJ, Andrade F, Bilyy R, Chatfield S, et al. To NET or not to NET:current opinions and state of the science regarding the formation of neutrophil extracellular traps. *Cell Death Differ*. (2019) 26:395–408. doi: 10.1038/s41418-018-0261-x
- Yipp BG, Kubes P. NETosis: how vital is it? *Blood.* (2013) 122:2784–94. doi: 10.1182/blood-2013-04-457671
- Zhao W, Fogg DK, Kaplan MJ. A novel image-based quantitative method for the characterization of NETosis. *J Immunol Methods*. (2015) 423:104–10. doi: 10.1016/j.jim.2015.04.027
- 62. Maugeri N, Rovere-Querini P, Manfredi AA. Disruption of a regulatory network consisting of neutrophils and platelets fosters persisting inflammation in rheumatic diseases. *Front Immunol.* (2016) 7:182. doi: 10.3389/fimmu.2016.00182
- Metzler KD, Fuchs TA, Nauseef WM, Reumaux D, Roesler J, Schulze I, et al. Myeloperoxidase is required for neutrophil extracellular trap formation: implications for innate immunity. *Blood.* (2011) 117:953–9. doi: 10.1182/blood-2010-06-290171
- Ramirez GA, Manfredi AA, Rovere-Querini P, Maugeri N. Bet on NETs! Or on how to translate basic science into clinical practice. *Front Immunol.* (2016) 7:417. doi: 10.3389/fimmu.2016.00417
- 65. Podolska MJ, Mahajan A, Knopf Hahn Boeltz J, J, Munoz L, et al. Autoimmune, rheumatic, chronic S. inflammatory diseases: neutrophil extracellular traps on parade. 10.1080/08916934.2018.15 Autoimmunity. (2018)51:281-7. doi: 19804
- Manfredi AA, Ramirez GA, Rovere-Querini P, Maugeri N. The neutrophil's choice: phagocytose vs make neutrophil extracellular traps. *Front Immunol.* (2018) 9:288. doi: 10.3389/fimmu.2018.00288
- 67. Sangaletti S, Tripodo C, Chiodoni C, Guarnotta C, Cappetti B, Casalini P, et al. Neutrophil extracellular traps mediate transfer of cytoplasmic neutrophil antigens to myeloid dendritic cells toward ANCA induction and associated autoimmunity. *Blood.* (2012) 120:3007–18. doi: 10.1182/blood-2012-03-416156
- 68. Villanueva E, Yalavarthi S, Berthier CC, Hodgin JB, Khandpur R, Lin AM, et al. Netting neutrophils induce endothelial damage, infiltrate tissues, and

expose immunostimulatory molecules in systemic lupus erythematosus. J Immunol. (2011) 187:538–52. doi: 10.4049/jimmunol.1100450

- Hakkim A, Furnrohr BG, Amann K, Laube B, Abed UA, Brinkmann V, et al. Impairment of neutrophil extracellular trap degradation is associated with lupus nephritis. *Proc Natl Acad Sci USA*. (2010) 107:9813–8. doi: 10.1073/pnas.0909927107
- Chapman EA, Lyon M, Simpson D, Mason D, Beynon RJ, Moots RJ, et al. Caught in a trap? Proteomic analysis of neutrophil extracellular traps in rheumatoid arthritis and systemic lupus erythematosus. *Front Immunol.* (2019) 10:423. doi: 10.3389/fimmu.2019.00423
- Bruschi M, Bonanni A, Petretto A, Vaglio A, Pratesi F, Santucci L, et al. Neutrophil extracellular traps (NETs) profiles in patients with incident SLE and lupus nephritis. *J Rheumatol.* (2019). doi: 10.3899/jrheum.181232. [Epub ahead of print].
- Pratesi F, Dioni I, Tommasi C, Alcaro MC, Paolini I, Barbetti F, et al. Antibodies from patients with rheumatoid arthritis target citrullinated histone 4 contained in neutrophils extracellular traps. *Ann Rheum Dis.* (2014) 73:1414–22. doi: 10.1136/annrheumdis-2012-2 02765
- 73. Sur Chowdhury C, Giaglis S, Walker UA, Buser A, Hahn S, Hasler P. Enhanced neutrophil extracellular trap generation in rheumatoid arthritis: analysis of underlying signal transduction pathways and potential diagnostic utility. *Arthritis Res Ther.* (2014) 16:R122. doi: 10.1186/ar4579
- Wright HL, Moots RJ, Edwards SW. The multifactorial role of neutrophils in rheumatoid arthritis. *Nat Rev Rheumatol.* (2014) 10:593–601. doi: 10.1038/nrrheum.2014.80
- Nakazawa D, Masuda S, Tomaru U, Ishizu A. Pathogenesis and therapeutic interventions for ANCA-associated vasculitis. *Nat Rev Rheumatol.* (2019) 15:91–101. doi: 10.1038/s41584-018-0145-y
- Jimenez-Alcazar M, Rangaswamy C, Panda R, Bitterling J, Simsek YJ, Long AT, et al. Host DNases prevent vascular occlusion by neutrophil extracellular traps. *Science*. (2017) 358:1202–6. doi: 10.1126/science.aam8897
- 77. Jaillon S, Peri G, Delneste Y, Fremaux I, Doni A, Moalli F, et al. The humoral pattern recognition receptor PTX3 is stored in neutrophil granules and localizes in extracellular traps. *J Exp Med.* (2007) 204:793–804. doi: 10.1084/jem.20061301
- Jaillon S, Ponzetta A, Magrini E, Barajon I, Barbagallo M, Garlanda C, et al. Fluid phase recognition molecules in neutrophildependent immune responses. *Semin Immunol.* (2016) 28:109–18. doi: 10.1016/j.smim.2016.03.005
- Polgar J, Matuskova J, Wagner DD. The P-selectin, tissue factor, coagulation triad. J Thromb Haemost. (2005) 3:1590–6. doi: 10.1111/j.1538-7836.2005.01373.x
- Moraes JA, Frony AC, Barcellos-de-Souza P, Menezes da Cunha M, Brasil Barbosa Calcia T, Benjamim CF, et al. Downregulation of microparticle release and pro-inflammatory properties of activated human polymorphonuclear neutrophils by LMW fucoidan. *J Innate Immun*. (2019) 11:330–46. doi: 10.1159/000494220
- Mesri M, Altieri DC. Endothelial cell activation by leukocyte microparticles. *J Immunol.* (1998) 161:4382–7.
- Butin-Israeli V, Bui TM, Wiesolek HL, Mascarenhas L, Lee JJ, Mehl LC, et al. Neutrophil-induced genomic instability impedes resolution of inflammation and wound healing. J Clin Invest. (2019) 129:712–26. doi: 10.1172/JCI122085
- 83. Sellam J, Proulle V, Jungel A, Ittah M, Miceli Richard C, Gottenberg JE, et al. Increased levels of circulating microparticles in primary Sjogren's syndrome, systemic lupus erythematosus and rheumatoid arthritis and relation with disease activity. *Arthritis Res Ther.* (2009) 11:R156. doi: 10.1186/ar2833
- 84. Wang Y, Luo L, Braun OO, Westman J, Madhi R, Herwald H, et al. Neutrophil extracellular trap-microparticle complexes enhance thrombin generation via the intrinsic pathway of coagulation in mice. *Sci Rep.* (2018) 8:4020. doi: 10.1038/s41598-018-22156-5
- Maugeri N, Rovere-Querini P, Evangelista V, Covino C, Capobianco A, Bertilaccio MT, et al. Neutrophils phagocytose activated platelets *in vivo*: a phosphatidylserine, P-selectin, and {beta}2 integrin-dependent cell clearance program. *Blood*. (2009) 113:5254–65. doi: 10.1182/blood-2008-09-180794
- Maugeri N, Evangelista V, Celardo A, Dell'Elba G, Martelli N, Piccardoni P, et al. Polymorphonuclear leukocyte-platelet interaction: role of P-selectin in thromboxane B2 and leukotriene C4 cooperative

synthesis. Thromb Haemost. (1994) 72:450-6. doi: 10.1055/s-0038-16 48888

- Page C, Pitchford S. Neutrophil and platelet complexes and their relevance to neutrophil recruitment and activation. *Int Immunopharmacol.* (2013) 17:1176–84. doi: 10.1016/j.intimp.2013.06.004
- Clark SR, Ma AC, Tavener SA, McDonald B, Goodarzi Z, Kelly MM, et al. Platelet TLR4 activates neutrophil extracellular traps to ensnare bacteria in septic blood. *Nat Med.* (2007) 13:463–9. doi: 10.1038/nm1565
- Brown SB, Clarke MC, Magowan L, Sanderson H, Savill J. Constitutive death of platelets leading to scavenger receptor-mediated phagocytosis. A caspaseindependent cell clearance program. *J Biol Chem.* (2000) 275:5987–96. doi: 10.1074/jbc.275.8.5987
- Kornerup KN, Salmon GP, Pitchford SC, Liu WL, Page CP. Circulating platelet-neutrophil complexes are important for subsequent neutrophil activation and migration. J Appl Physiol (1985). (2010) 109:758–67. doi: 10.1152/japplphysiol.01086.2009
- Han CZ, Ravichandran KS. Metabolic connections during apoptotic cell engulfment. Cell. (2011) 147:1442–5. doi: 10.1016/j.cell.2011.12.006
- Zhang S, Bories G, Lantz C, Emmons R, Becker A, Liu E, et al. Immunometabolism of phagocytes and relationships to cardiac repair. *Front Cardiovasc Med.* (2019) 6:42. doi: 10.3389/fcvm.2019.00042
- Cunin P, Bouslama R, Machlus KR, Martinez-Bonet M, Lee PY, Wactor A, et al. Megakaryocyte emperipolesis mediates membrane transfer from intracytoplasmic neutrophils to platelets. *Elife*. (2019) 8:e44031. doi: 10.7554/eLife.44031
- Kuravi SJ, Harrison P, Rainger GE, Nash GB. Ability of platelet-derived extracellular vesicles to promote neutrophil-endothelial cell interactions. *Inflammation*. (2019) 42:290–305. doi: 10.1007/s10753-018-0893-5
- Jy W, Mao WW, Horstman L, Tao J, Ahn YS. Platelet microparticles bind, activate and aggregate neutrophils *in vitro*. *Blood Cells Mol Dis*. (1995) 21:217–31; discussion 31a. doi: 10.1006/bcmd.1995.0025
- Maugeri N, Campana L, Gavina M, Covino C, De Metrio M, Panciroli C, et al. Activated platelets present high mobility group box 1 to neutrophils, inducing autophagy and promoting the extrusion of neutrophil extracellular traps. J Thromb Haemost. (2014) 12:2074–88. doi: 10.1111/jth.12710
- Scherlinger M, Guillotin V, Truchetet ME, Contin-Bordes C, Sisirak V, Duffau P, et al. Systemic lupus erythematosus and systemic sclerosis: all roads lead to platelets. *Autoimmun Rev.* (2018) 17:625–35. doi: 10.1016/j.autrev.2018.01.012
- Zhou H, Deng M, Liu Y, Yang C, Hoffman R, Zhou J, et al. Platelet HMGB1 is required for efficient bacterial clearance in intraabdominal bacterial sepsis in mice. *Blood Adv.* (2018) 2:638–48. doi: 10.1182/bloodadvances.2017011817
- 99. Kim SW, Lee H, Lee HK, Kim ID, Lee JK. Neutrophil extracellular trap induced by HMGB1 exacerbates damages in the ischemic brain. *Acta Neuropathol Commun.* (2019) 7:94. doi: 10.1186/s40478-019-0747-x
- 100. Rossaint J, Kuhne K, Skupski J, Van Aken H, Looney MR, Hidalgo A, et al. Directed transport of neutrophil-derived extracellular vesicles enables platelet-mediated innate immune response. *Nat Commun.* (2016) 7:13464. doi: 10.1038/ncomms13464
- 101. Maugeri N, Evangelista V, Piccardoni P, Dell'Elba G, Celardo A, de Gaetano G, et al. Transcellular metabolism of arachidonic acid: increased platelet thromboxane generation in the presence of activated polymorphonuclear leukocytes. *Blood.* (1992) 80:447–51. doi: 10.1016/0049-3848(92)9 0486-T
- 102. Eriksson O, Mohlin C, Nilsson B, Ekdahl KN. The human platelet as an innate immune cell: interactions between activated platelets and the complement system. *Front Immunol.* (2019) 10:1590. doi: 10.3389/fimmu.2019.01590
- 103. Bai M, Xing L, Feng J, Cui C, Huang L, Liang G. Mean platelet volume could reflect disease activity of adult patients with systemic lupus erythematosus. *Clin Lab.* (2016) 62:1317–22. doi: 10.7754/Clin.Lab.2015.151134
- 104. Yavuz S, Ece A. Mean platelet volume as an indicator of disease activity in juvenile SLE. *Clin Rheumatol.* (2014) 33:637–41. doi: 10.1007/s10067-014-2540-3
- 105. Fernandez-Nebro A, Rua-Figueroa I, Lopez-Longo FJ, Galindo-Izquierdo M, Calvo-Alen J, Olive-Marques A, et al. Cardiovascular events in systemic lupus erythematosus: a nationwide study in Spain

from the RELESSER registry. *Medicine (Baltimore)*. (2015) 94:e1183. doi: 10.1097/MD.0000000001183

- Ramirez GA, Efthymiou M, Isenberg DA, Cohen H. Under crossfire: thromboembolic risk in systemic lupus erythematosus. *Rheumatology (Oxf)*. (2019) 58:940–52. doi: 10.1093/rheumatology/key307
- Mehta BM, Kiani AN, Chen C, Jani J, Kickler TS, Petri M. Endogenous thrombin potential in the assessment of hypercoagulability in systemic lupus erythematosus. *Am J Hematol.* (2010) 85:83–5. doi: 10.1002/ajh.21566
- Kern A, Barabas E, Balog A, Burcsar S, Kiszelak M, Vasarhelyi B. Characterization of the thrombin generation profile in systemic lupus erythematosus. *Physiol Int.* (2017) 104:35–41. doi: 10.1556/2060.104.2017.1.5
- 109. Carli L, Tani C, Vagnani S, Signorini V, Mosca M. Leukopenia, lymphopenia, and neutropenia in systemic lupus erythematosus: prevalence and clinical impact-a systematic literature review. *Semin Arthritis Rheum.* (2015) 45:190–4. doi: 10.1016/j.semarthrit.2015.05.009
- 110. Ferriani MP, Silva MF, Pereira RM, Terreri MT, Saad Magalhaes C, Bonfa E, et al. Chronic spontaneous urticaria: a survey of 852 cases of childhood-onset systemic lupus erythematosus. *Int Arch Allergy Immunol.* (2015) 167:186–92. doi: 10.1159/000438723
- 111. Kolkhir P, Pogorelov D, Olisova O, Maurer M. Comorbidity and pathogenic links of chronic spontaneous urticaria and systemic lupus erythematosus-a systematic review. *Clin Exp Allergy.* (2016) 46:275–87. doi: 10.1111/cea.12673
- 112. Lood C, Tyden H, Gullstrand B, Nielsen CT, Heegaard NH, Linge P, et al. Decreased platelet size is associated with platelet activation and anti-phospholipid syndrome in systemic lupus erythematosus. *Rheumatology*. (2017) 56:408–16. doi: 10.1093/rheumatology/kex216
- 113. Ruiz-Irastorza G, Crowther M, Branch W, Khamashta MA. Antiphospholipid syndrome. *Lancet.* (2010) 376:1498–509. doi: 10.1016/S0140-6736(10)60709-X
- 114. Cervera R, Serrano R, Pons-Estel GJ, Ceberio-Hualde L, Shoenfeld Y, de Ramon E, et al. Morbidity and mortality in the antiphospholipid syndrome during a 10-year period: a multicentre prospective study of 1000 patients. *Ann Rheum Dis.* (2015) 74:1011–8. doi: 10.1136/annrheumdis-2013-204838
- 115. Arachchillage DR, Efthymiou M, Mackie IJ, Lawrie AS, Machin SJ, Cohen H. Anti-protein C antibodies are associated with resistance to endogenous protein C activation and a severe thrombotic phenotype in antiphospholipid syndrome. J Thromb Haemost. (2014) 12:1801–9. doi: 10.1111/jth.12722
- 116. Pierangeli SS, Girardi G, Vega-Ostertag M, Liu X, Espinola RG, Salmon J. Requirement of activation of complement C3 and C5 for antiphospholipid antibody-mediated thrombophilia. *Arthritis Rheum.* (2005) 52:2120–4. doi: 10.1002/art.21157
- 117. Soydinc S, Turkbeyler IH, Pehlivan Y, Soylu G, Goktepe MF, Bilici M, et al. Mean platelet volume seems to be a valuable marker in patients with systemic sclerosis. *Inflammation*. (2014) 37:100–6. doi: 10.1007/s10753-013-9716-x
- Maeda M, Kachi H, Mori S. Ultrastructural observation of platelets from patients with progressive systemic sclerosis (PSS). *J Dermatol.* (1998) 25:222– 30. doi: 10.1111/j.1346-8138.1998.tb02385.x
- 119. Solfietti L, Binello GB, Stella S, Bazzan M, Salierno M, Roccatello D. Thrombin generation assay: interactions between chronic inflammation and haemostasis in patients with autoimmune diseases. *Clin Exp Rheumatol.* (2016) 34:925–8.
- 120. Nicolosi PA, Tombetti E, Maugeri N, Rovere-Querini P, Brunelli S, Manfredi AA. Vascular remodelling and mesenchymal transition in systemic sclerosis. *Stem Cells Int.* (2016) 2016:4636859. doi: 10.1155/2016/4636859
- 121. Denton CP, Khanna D. Systemic sclerosis. Lancet. (2017) 390:1685-99. doi: 10.1016/S0140-6736(17)30933-9
- 122. Ames PR, Lupoli S, Alves J, Atsumi T, Edwards C, Iannaccone L, et al. The coagulation/fibrinolysis balance in systemic sclerosis: evidence for a haematological stress syndrome. Br J Rheumatol. (1997) 36:1045–50. doi: 10.1093/rheumatology/36.10.1045
- 123. Kim HJ, Jung SM, Song JJ, Park YB, Lee SW. Mean platelet volume can estimate the current vasculitis activity of microscopic polyangiitis. *Rheumatol Int.* (2018) 38:1095–101. doi: 10.1007/s00296-018-4011-7
- 124. Merkel PA, Lo GH, Holbrook JT, Tibbs AK, Allen NB, Davis JC, Jr, et al. Brief communication: high incidence of venous thrombotic events among patients with Wegener granulomatosis: the Wegener's Clinical Occurrence

of Thrombosis (WeCLOT) Study. Ann Intern Med. (2005) 142:620-6. doi: 10.7326/0003-4819-142-8-200505030-00011

- 125. Maino A, Rossio R, Cugno M, Marzano AV, Tedeschi A. Hypereosinophilic syndrome, Churg-Strauss syndrome and parasitic diseases: possible links between eosinophilia and thrombosis. *Curr Vasc Pharmacol.* (2012) 10:670– 5. doi: 10.2174/157016112801784594
- 126. Maugeri N, Rovere-Querini P, Evangelista V, Godino C, Demetrio M, Baldini M, et al. An intense and short-lasting burst of neutrophil activation differentiates early acute myocardial infarction from systemic inflammatory syndromes. *PLoS ONE*. (2012) 7:e39484. doi: 10.1371/journal.pone.0039484
- 127. Tomasson G, Lavalley M, Tanriverdi K, Finkielman JD, Davis JC Jr, Hoffman GS, et al. Relationship between markers of platelet activation and inflammation with disease activity in Wegener's granulomatosis. J Rheumatol. (2011) 38:1048–54. doi: 10.3899/jrheum.100735
- 128. Ma TT, Huang YM, Wang C, Zhao MH, Chen M. Coagulation and fibrinolysis index profile in patients with ANCA-associated vasculitis. *PLoS ONE.* (2014) 9:e97843. doi: 10.1371/journal.pone.0097843
- Peng YF, Guo J, Deng YB. The role of mean platelet volume in patients with Takayasu arteritis. Ann Clin Biochem. (2017) 54:273–8. doi: 10.1177/0004563216658312
- Weyand CM, Goronzy JJ. Medium- and large-vessel vasculitis. N Engl J Med. (2003) 349:160–9. doi: 10.1056/NEJMra022694
- Tombetti E, Mason JC. Takayasu arteritis: recent advances and hopes for the future. *Rheumatology*. (2019) 17:238–47. doi: 10.1111/1756-185X.12309
- 132. Maugeri N, Baldini M, Rovere-Querini P, Maseri A, Sabbadini MG, Manfredi AA. Leukocyte and platelet activation in patients with giant cell arteritis and polymyalgia rheumatica: a clue to thromboembolic risks? *Autoimmunity.* (2009) 42:386–8. doi: 10.1080/089169309028 32629
- 133. Akkececi NS, Cetin GY, Gogebakan H, Acipayam C. The C-reactive protein/albumin ratio and complete blood count parameters as indicators of disease activity in patients with Takayasu Arteritis. *Med Sci Monit.* (2019) 25:1401. doi: 10.12659/MSM.912495
- Kaiser M, Weyand CM, Bjornsson J, Goronzy JJ. Platelet-derived growth factor, intimal hyperplasia, and ischemic complications in giant cell arteritis. *Arthritis Rheum.* (1998) 41:623–33.
- 135. Maden M, Pamuk GE, Pamuk ON. Development of atherosclerotic cardiovascular mortality in gouty arthritis and rheumatoid arthritis patients: are they associated with mean platelet volume and neutrophil-lymphocyte ratio? A comparative study. Arch Rheumatol. (2017) 32:39–45. doi: 10.5606/ArchRheumatol.20 17.6033
- 136. Yazici S, Yazici M, Erer B, Erer B, Calik Y, Ozhan H, et al. The platelet indices in patients with rheumatoid arthritis: mean platelet volume reflects disease activity. *Platelets*. (2010) 21:122–5. doi: 10.3109/09537100903474373
- 137. Gasparyan AY, Ayvazyan L, Mikhailidis DP, Kitas GD. Mean platelet volume: a link between thrombosis and inflammation? *Curr Pharma Des.* (2011) 17:47–58. doi: 10.2174/1381612117950 49804
- Gasparyan AY, Stavropoulos-Kalinoglou A, Mikhailidis DP, Douglas KM, Kitas GD. Platelet function in rheumatoid arthritis: arthritic and cardiovascular implications. *Rheumatol Int.* (2011) 31:153–64. doi: 10.1007/s00296-010-1446-x
- Lauper K, Gabay C. Cardiovascular risk in patients with rheumatoid arthritis. Semin Immunopathol. (2017) 39:447–59. doi: 10.1007/s00281-017-0632-2
- 140. Manfredi AA, Baldini M, Camera M, Baldissera E, Brambilla M, Peretti G, et al. Anti-TNFalpha agents curb platelet activation in patients with rheumatoid arthritis. Ann Rheum Dis. (2016) 75:1511–20. doi: 10.1136/annrheumdis-2015-2 08442
- 141. Chandrashekar L, Rajappa M, Sundar I, Munisamy M, Ananthanarayanan PH, Thappa DM, et al. Platelet activation in chronic urticaria and its correlation with disease severity. *Platelets*. (2014) 25:162–5. doi: 10.3109/09537104.2013.786822
- 142. Sakurai Y, Morioke S, Takeda T, Takahagi S, Hide M, Shima M. Increased thrombin generation potential in patients with chronic spontaneous urticaria. *Allergol Int.* (2015) 64:96–8. doi: 10.1016/j.alit.2014.07.006

- Takeda T, Sakurai Y, Takahagi S, Kato J, Yoshida K, Yoshioka A, et al. Increase of coagulation potential in chronic spontaneous urticaria. *Allergy*. (2011) 66:428–33. doi: 10.1111/j.1398-9995.2010.02506.x
- 144. Tedeschi A, Kolkhir P, Asero R, Pogorelov D, Olisova O, Kochergin N, et al. Chronic urticaria and coagulation: pathophysiological and clinical aspects. *Allergy.* (2014) 69:683–91. doi: 10.1111/all.12389
- 145. Yanase Y, Takahagi S, Hide M. Chronic spontaneous urticaria and the extrinsic coagulation system. *Allergol Int.* (2018) 67:191–4. doi: 10.1016/j.alit.2017.09.003
- 146. Egeberg A, Kofoed K, Gislason GH, Vestergaard C, Thyssen JP. Cardiovascular risk is not Increased in patients with chronic urticaria: a retrospective population-based cohort study. *Acta Derm Venereol.* (2017) 97:261–2. doi: 10.2340/00015555-2516
- 147. Chan FLY, Lester S, Whittle SL, Hill CL. The utility of ESR, CRP and platelets in the diagnosis of GCA. BMC Rheumatol. (2019) 3:14. doi: 10.1186/s41927-019-0061-z
- 148. Claushuis TA, van Vught LA, Scicluna BP, Wiewel MA, Klein Klouwenberg PM, Hoogendijk AJ, et al. Thrombocytopenia is associated with a dysregulated host response in critically ill sepsis patients. *Blood.* (2016) 127:3062–72. doi: 10.1182/blood-2015-11-680744
- 149. Hillgruber C, Poppelmann B, Weishaupt C, Steingraber AK, Wessel F, Berdel WE, et al. Blocking neutrophil diapedesis prevents hemorrhage during thrombocytopenia. J Exp Med. (2015) 212:1255–66. doi: 10.1084/jem.20142076
- Alzubiedi S, Saleh MI. Predictors of severe thrombocytopenia secondary to peginterferon Alfa-2a treatment in subjects with hepatitis C virus infection. *Am J Ther.* (2017) 24:e670–5. doi: 10.1097/MJT.00000000000356
- 151. Gollomp K, Kim M, Johnston I, Hayes V, Welsh J, Arepally GM, et al. Neutrophil accumulation and NET release contribute to thrombosis in HIT. *JCI Insight*. (2018) 3:99445. doi: 10.1172/jci.insight.99445
- 152. Leffler J, Prohaszka Z, Mikes B, Sinkovits G, Ciacma K, Farkas P, et al. Decreased neutrophil extracellular trap degradation in Shiga toxinassociated haemolytic uraemic syndrome. *J Innate Immun.* (2017) 9:12–21. doi: 10.1159/000450609
- 153. Perdomo J, Leung HHL, Ahmadi Z, Yan F, Chong JJH, Passam FH, et al. Neutrophil activation and NETosis are the major drivers of thrombosis in heparin-induced thrombocytopenia. *Nat Commun.* (2019) 10:1322. doi: 10.1038/s41467-019-09160-7
- 154. Li Y, Wang H, Peng C, Li H, Jie S. Low-density neutrophils in severe fever with thrombocytopenia syndrome (SFTS) display decreased function to phagocytose SFTS virus and enhanced capacity to synthesize cytokines. *Int J Clin Exp Pathol.* (2017) 4:4069–78.
- 155. Esteban YM, de Jong JLO, Tesher MS. An overview of hemophagocytic lymphohistiocytosis. *Pediatr Ann.* (2017) 46:e309–13. doi: 10.3928/19382359-20170717-01
- 156. Thompson CB, Eaton KA, Princiotta SM, Rushin CA, Valeri CR. Size dependent platelet subpopulations: relationship of platelet volume to ultrastructure, enzymatic activity, and function. Br J Haematol. (1982) 50:509–19. doi: 10.1111/j.1365-2141.1982.tb01947.x
- 157. Thompson CB, Scher I, Schaefer ME, Lindsten T, Finkelman FD, Mond JJ. Size-dependent B lymphocyte subpopulations: relationship of cell volume to surface phenotype, cell cycle, proliferative response, and requirements for antibody production to TNP-Ficoll and TNP-BA. J Immunol. (1984) 133:2333–42.
- 158. Denny MF, Yalavarthi S, Zhao W, Thacker SG, Anderson M, Sandy AR, et al. A distinct subset of proinflammatory neutrophils isolated from patients with systemic lupus erythematosus induces vascular damage and synthesizes type I IFNs. J Immunol. (2010) 184:3284–97. doi: 10.4049/jimmunol.0902199
- Hacbarth E, Kajdacsy-Balla A. Low density neutrophils in patients with systemic lupus erythematosus, rheumatoid arthritis, and acute rheumatic fever. Arthritis Rheum. (1986) 29:1334–42. doi: 10.1002/art.1780291105
- 160. Vena GA, Cassano N, Marzano AV, Asero R. The role of platelets in chronic urticaria. Int Arch Allergy Immunol. (2016) 169:71–9. doi: 10.1159/000444085
- 161. Korniluk A, Koper-Lenkiewicz OM, Kaminska J, Kemona H, Dymicka-Piekarska V. Mean platelet volume (MPV): new perspectives for an old marker in the course and prognosis of inflammatory conditions. *Mediat Inflamm.* (2019) 2019:9213074. doi: 10.1155/2019/9213074

- 162. Park DH, Park K, Park J, Park HH, Chae H, Lim J, et al. Screening of sepsis using leukocyte cell population data from the Coulter automatic blood cell analyzer DxH800. *Int J Lab Hematol.* (2011) 33:391–9. doi: 10.1111/j.1751-553X.2011.01298.x
- 163. Sagiv JY, Michaeli J, Assi S, Mishalian I, Kisos H, Levy L, et al. Phenotypic diversity and plasticity in circulating neutrophil subpopulations in cancer. *Cell Rep.* (2015) 10:562–73. doi: 10.1016/j.celrep.2014.12.039
- 164. Worthen GS, Henson PM, Rosengren S, Downey GP, Hyde DM. Neutrophils increase volume during migration *in vivo* and *in vitro*. *Am J Respir Cell Mol Biol.* (1994) 10:1–7. doi: 10.1165/ajrcmb.10.1.82 92373
- 165. Chung CP, Giles JT, Kronmal RA, Post WS, Gelber AC, Petri M, et al. Progression of coronary artery atherosclerosis in rheumatoid arthritis: comparison with participants from the Multi-Ethnic Study of Atherosclerosis. Arthritis Res Ther. (2013) 15:R134. doi: 10.1186/ar4314
- 166. Ruiz-Irastorza G, Egurbide MV, Ugalde J, Aguirre C. High impact of antiphospholipid syndrome on irreversible organ damage and survival of patients with systemic lupus erythematosus. *Arch Intern Med.* (2004) 164:77– 82. doi: 10.1001/archinte.164.1.77
- Palatinus A, Adams M. Thrombosis in systemic lupus erythematosus. Semin Thromb Hemost. (2009) 35:621–9. doi: 10.1055/s-0029-1242716
- 168. Kay SD, Poulsen MK, Diederichsen AC, Voss A. Coronary, carotid, and lower-extremity atherosclerosis and their interrelationship in danish patients with systemic lupus erythematosus. J Rheumatol. (2016) 43:315–22. doi: 10.3899/jrheum.150488
- 169. Roman MJ, Crow MK, Lockshin MD, Devereux RB, Paget SA, Sammaritano L, et al. Rate and determinants of progression of atherosclerosis in systemic lupus erythematosus. *Arthritis Rheum.* (2007) 56:3412–9. doi: 10.1002/art.22924
- 170. Seyahi E, Ugurlu S, Cumali R, Balci H, Seyahi N, Yurdakul S, et al. Atherosclerosis in Takayasu arteritis. Ann Rheum Dis. (2006) 65:1202–7. doi: 10.1136/ard.2005.047498
- 171. Salmon JE, Roman MJ. Subclinical atherosclerosis in rheumatoid arthritis and systemic lupus erythematosus. Am J Med. (2008) 121:S3–8. doi: 10.1016/j.amjmed.2008.06.010
- 172. Bruce IN. 'Not only ... but also': factors that contribute to accelerated atherosclerosis and premature coronary heart disease in systemic lupus erythematosus. *Rheumatology*. (2005) 44:1492–502. doi: 10.1093/rheumatology/kei142
- 173. Qi H, Yang S, Zhang L. Neutrophil extracellular traps and endothelial dysfunction in atherosclerosis and thrombosis. *Front Immunol.* (2017) 8:928. doi: 10.3389/fimmu.2017.00928
- 174. Mezger M, Nording H, Sauter R, Graf T, Heim C, von Bubnoff N, et al. Platelets and immune responses during thromboinflammation. *Front Immunol.* (2019) 10:1731. doi: 10.3389/fimmu.2019.01731
- 175. Massberg S, Brand K, Gruner S, Page S, Muller E, Muller I, et al. A critical role of platelet adhesion in the initiation of atherosclerotic lesion formation. *J Exp Med.* (2002) 196:887–96. doi: 10.1084/jem.20012044
- 176. Bertolaccini ML, Sanna G. Recent advances in understanding antiphospholipid syndrome. *F1000Res.* (2016) 5:2908. doi: 10.12688/f1000research.9717.1
- 177. Manganelli V, Truglia S, Capozzi A, Alessandri C, Riitano G, Spinelli FR, et al. Alarmin HMGB1 and soluble RAGE as new tools to evaluate the risk stratification in patients with the antiphospholipid syndrome. *Front Immunol.* (2019) 10:460. doi: 10.3389/fimmu.2019. 00460
- 178. Yalavarthi S, Gould TJ, Rao AN, Mazza LF, Morris AE, Nunez-Alvarez C, et al. Release of neutrophil extracellular traps by neutrophils stimulated with antiphospholipid antibodies: a newly identified mechanism of thrombosis in the antiphospholipid syndrome. *Arthritis Rheumatol.* (2015) 67:2990–3003. doi: 10.1002/art.39247
- 179. de Groot PG, Meijers JC. beta(2)-Glycoprotein I: evolution, structure and function. J Thromb Haemost. (2011) 9:1275–84. doi: 10.1111/j.1538-7836.2011.04327.x
- 180. Stakos DA, Kambas K, Konstantinidis T, Mitroulis I, Apostolidou E, Arelaki S, et al. Expression of functional tissue factor by neutrophil extracellular traps in culprit artery of acute myocardial infarction. *Eur Heart J.* (2015) 36:1405–14. doi: 10.1093/eurheartj/ehv007

- Bazan-Socha S, Mastalerz L, Cybulska A, Zareba L, Kremers R, Zabczyk M, et al. Asthma is associated with enhanced thrombin formation and impaired fibrinolysis. *Clin Exp Allergy*. (2016) 46:932–44. doi: 10.1111/cea.12734
- 182. Tattersall MC, Guo M, Korcarz CE, Gepner AD, Kaufman JD, Liu KJ, et al. Asthma predicts cardiovascular disease events: the multi-ethnic study of atherosclerosis. *Arterioscler Thromb Vasc Biol.* (2015) 35:1520–5. doi: 10.1161/ATVBAHA.115.305452
- 183. Liszewski MK, Atkinson JP. Too much of a good thing at the site of tissue injury: the instructive example of the complement system predisposing to thrombotic microangiopathy. *Hematol Am Soc Hematol Educ Program.* (2011) 2011:9–14. doi: 10.1182/asheducation-2011.1.9
- Ruggenenti P, Noris M, Remuzzi G. Thrombotic microangiopathy, hemolytic uremic syndrome, and thrombotic thrombocytopenic purpura. *Kidney Int.* (2001) 60:831–46. doi: 10.1046/j.1523-1755.2001.060003831.x
- 185. Song D, Wu LH, Wang FM, Yang XW, Zhu D, Chen M, et al. The spectrum of renal thrombotic microangiopathy in lupus nephritis. *Arthritis Res Ther.* (2013) 15:R12. doi: 10.1186/ar4142
- 186. Bhasym A, Annarapu GK, Saha S, Shrimali N, Gupta S, Seth T, et al. Neutrophils develop rapid proinflammatory response after engulfing Hbactivated platelets under intravascular hemolysis. *Clin Exp Immunol.* (2019) 197:131–40. doi: 10.1111/cei.13310
- 187. Jimenez-Alcazar M, Napirei M, Panda R, Kohler EC, Kremer Hovinga JA, Mannherz HG, et al. Impaired DNase1-mediated degradation of neutrophil extracellular traps is associated with acute thrombotic microangiopathies. J Thromb Haemost. (2015) 13:732–42. doi: 10.1111/jth.12796
- Doyle TJ, Dellaripa PF. Lung manifestations in the rheumatic diseases. Chest. (2017) 152:1283–95. doi: 10.1016/j.chest.2017.05.015
- Aulakh GK. Neutrophils in the lung: "the first responders". Cell Tissue Res. (2018) 371:577–88. doi: 10.1007/s00441-017-2748-z
- 190. Yang C, Chilvers M, Montgomery M, Nolan SJ. Dornase alfa for cystic fibrosis. *Cochrane Database Syst Rev.* (2016) 4:CD001127. doi: 10.1002/14651858.CD001127.pub3
- 191. Yoo DG, Winn M, Pang L, Moskowitz SM, Malech HL, Leto TL, et al. Release of cystic fibrosis airway inflammatory markers from Pseudomonas aeruginosa-stimulated human neutrophils involves NADPH oxidase-dependent extracellular DNA trap formation. *J Immunol.* (2014) 192:4728–38. doi: 10.4049/jimmunol.1301589
- 192. Chrysanthopoulou A, Mitroulis I, Apostolidou E, Arelaki S, Mikroulis D, Konstantinidis T, et al. Neutrophil extracellular traps promote differentiation and function of fibroblasts. *J Pathol.* (2014) 233:294–307. doi: 10.1002/path.4359
- 193. Liu S, Su X, Pan P, Zhang L, Hu Y, Tan H, et al. Neutrophil extracellular traps are indirectly triggered by lipopolysaccharide and contribute to acute lung injury. *Sci Rep.* (2016) 6:37252. doi: 10.1038/srep37252
- 194. Gregoire M, Uhel F, Lesouhaitier M, Gacouin A, Guirriec M, Mourcin F, et al. Impaired efferocytosis and neutrophil extracellular trap clearance by macrophages in ARDS. *Eur Respir J.* (2018) 52:1702590. doi: 10.1183/13993003.02590-2017
- 195. Kosaki G. In vivo platelet production from mature megakaryocytes: does platelet release occur via proplatelets? Int J Hematol. (2005) 81:208–19. doi: 10.1532/IJH97.04177
- 196. Yamamoto Y, Hasegawa H, Inoue F, Ikeda K, Ichiyama A. Serotonin in the lung. Demonstration of a close correlation to blood platelet. *Agents Actions*. (1986) 18:351–8. doi: 10.1007/BF01964996
- 197. Pattanaik D, Brown M, Postlethwaite BC, Postlethwaite AE. Pathogenesis of systemic sclerosis. *Front Immunol.* (2015) 6:272. doi: 10.3389/fimmu.2015.00272
- 198. Caudrillier A, Kessenbrock K, Gilliss BM, Nguyen JX, Marques MB, Monestier M, et al. Platelets induce neutrophil extracellular traps in transfusion-related acute lung injury. J Clin Invest. (2012) 122:2661–71. doi: 10.1172/JCI61303
- 199. Jennette JC, Falk RJ, Bacon PA, Basu N, Cid MC, Ferrario F, et al. 2012 Revised International Chapel Hill consensus conference nomenclature of vasculitides. *Arthritis Rheum*. (2013) 65:1–11. doi: 10.1002/art.37715
- 200. Ramirez GA, Maugeri N, Sabbadini MG, Rovere-Querini P, Manfredi AA. Intravascular immunity as a key to systemic vasculitis: a work in progress, gaining momentum. *Clin Exp Immunol.* (2014) 175:150–66. doi: 10.1111/cei.12223

- Maugeri N, Baldini M, Ramirez GA, Rovere-Querini P, Manfredi AA. Platelet-leukocyte deregulated interactions foster sterile inflammation and tissue damage in immune-mediated vessel diseases. *Thromb Res.* (2012) 129:267–73. doi: 10.1016/j.thromres.2011.12.001
- Kessenbrock K, Krumbholz M, Schonermarck U, Back W, Gross WL, Werb Z, et al. Netting neutrophils in autoimmune small-vessel vasculitis. *Nat Med.* (2009) 15:623–5. doi: 10.1038/nm.1959
- 203. Bruchfeld A, Wendt M, Bratt J, Qureshi AR, Chavan S, Tracey KJ, et al. Highmobility group box-1 protein (HMGB1) is increased in antineutrophilic cytoplasmatic antibody (ANCA)-associated vasculitis with renal manifestations. *Mol Med.* (2011) 17:29–35. doi: 10.2119/molmed.2010.00132
- 204. Wibisono D, Csernok E, Lamprecht P, Holle JU, Gross WL, Moosig F. Serum HMGB1 levels are increased in active Wegener's granulomatosis and differentiate between active forms of ANCA-associated vasculitis. *Ann Rheum Dis.* (2010) 69:1888–9. doi: 10.1136/ard.2009.119172
- 205. Wang C, de Souza AW, Westra J, Bijl M, Chen M, Zhao MH, et al. Emerging role of high mobility group box 1 in ANCA-associated vasculitis. *Autoimmun Rev.* (2015) 14:1057–65. doi: 10.1016/j.autrev.2015.07.010
- 206. Baldini M, Maugeri N, Ramirez GA, Giacomassi C, Castiglioni A, Prieto-Gonzalez S, et al. Selective up-regulation of the soluble pattern-recognition receptor pentraxin 3 and of vascular endothelial growth factor in giant cell arteritis: relevance for recent optic nerve ischemia. *Arthritis Rheum.* (2012) 64:854–65. doi: 10.1002/art.34515
- 207. Nesher G, Berkun Y, Mates M, Baras M, Rubinow A, Sonnenblick M. Lowdose aspirin and prevention of cranial ischemic complications in giant cell arteritis. *Arthritis Rheum*. (2004) 50:1332–7. doi: 10.1002/art.20171
- Berger CT, Wolbers M, Meyer P, Daikeler T, Hess C. High incidence of severe ischaemic complications in patients with giant cell arteritis irrespective of platelet count and size, and platelet inhibition. *Rheumatology.* (2009) 48:258–61. doi: 10.1093/rheumatology/ken480
- Weyand CM, Kaiser M, Yang H, Younge B, Goronzy JJ. Therapeutic effects of acetylsalicylic acid in giant cell arteritis. *Arthritis Rheum.* (2002) 46:457–66. doi: 10.1002/art.10071
- 210. Cavazza A, Muratore F, Boiardi L, Restuccia G, Pipitone N, Pazzola G, et al. Inflamed temporal artery: histologic findings in 354 biopsies, with clinical correlations. Am J Surg Pathol. (2014) 38:1360–70. doi: 10.1097/PAS.0000000000 00244
- Bengtsson AA, Ronnblom L. Systemic lupus erythematosus: still a challenge for physicians. J Intern Med. (2017) 281:52–64. doi: 10.1111/joim.12529
- 212. Tsokos GC. Systemic lupus erythematosus. N Engl J Med. (2011) 365:2110–21. doi: 10.1056/NEJMra1100359
- 213. Aleem A, Al Arfaj AS, khalil N, Alarfaj H. Haematological abnormalities in systemic lupus erythematosus. *Acta Reumatol Port.* (2014) 39:236–41.
- 214. Barrera-Vargas A, Campos-Guzmán J, Govea-Peláez S, García-Ramos A, Demichelis-Gómez R, Bourlon C, et al. *THU0260 Systemic Lupus Erythematosus and Cytopenias: The Key Findings in Bone Marrow*. Madrid: BMJ Publishing Group Ltd (2019).
- 215. Pereira RM, Velloso ER, Menezes Y, Gualandro S, Vassalo J, Yoshinari NH. Bone marrow findings in systemic lupus erythematosus patients with peripheral cytopenias. *Clin Rheumatol.* (1998) 17:219–22. doi: 10.1007/BF01451051
- 216. Martinez-Banos D, Crispin JC, Lazo-Langner A, Sanchez-Guerrero J. Moderate and severe neutropenia in patients with systemic lupus erythematosus. *Rheumatology*. (2006) 45:994–8. doi: 10.1093/rheumatology/kel016
- Hepburn AL, Narat S, Mason JC. The management of peripheral blood cytopenias in systemic lupus erythematosus. *Rheumatology*. (2010) 49:2243– 54. doi: 10.1093/rheumatology/keq269
- Del Porto F, Tatarelli C, Di Napoli A, Proietta M. Systemic lupus erythematosus and myelofibrosis: a case report and revision of literature. *Leuk Res Rep.* (2018) 9:58–64. doi: 10.1016/j.lrr.2018.04.004
- Velo-Garcia A, Castro SG, Isenberg DA. The diagnosis and management of the haematologic manifestations of lupus. J Autoimmun. (2016) 74:139–60. doi: 10.1016/j.jaut.2016.07.001
- 220. Ziakas PD, Giannouli S, Zintzaras E, Tzioufas AG, Voulgarelis M. Lupus thrombocytopenia: clinical implications and prognostic significance. Ann Rheum Dis. (2005) 64:1366–9. doi: 10.1136/ard.2004.033100

- 221. Fernandez M, Alarcon GS, Apte M, Andrade RM, Vila LM, Reveille JD, et al. Systemic lupus erythematosus in a multiethnic US cohort: XLIII. The significance of thrombocytopenia as a prognostic factor. *Arthritis Rheum.* (2007) 56:614–21. doi: 10.1002/art.22376
- 222. Liu Y, Kaplan MJ. Cardiovascular disease in systemic lupus erythematosus: an update. *Curr Opin Rheumatol.* (2018) 30:441–8. doi: 10.1097/BOR.00000000000528
- 223. Joseph JE, Harrison P, Mackie IJ, Isenberg DA, Machin SJ. Increased circulating platelet-leucocyte complexes and platelet activation in patients with antiphospholipid syndrome, systemic lupus erythematosus and rheumatoid arthritis. *Br J Haematol.* (2001) 115:451–9. doi: 10.1046/j.1365-2141.2001.03101.x
- Nagahama M, Nomura S, Ozaki Y, Yoshimura C, Kagawa H, Fukuhara S. Platelet activation markers and soluble adhesion molecules in patients with systemic lupus erythematosus. *Autoimmunity.* (2001) 33:85–94. doi: 10.3109/08916930108995993
- 225. Duffau P, Seneschal J, Nicco C, Richez C, Lazaro E, Douchet I, et al. Platelet CD154 potentiates interferon-alpha secretion by plasmacytoid dendritic cells in systemic lupus erythematosus. *Sci Transl Med.* (2010) 2:47ra63. doi: 10.1126/scitranslmed.3001001
- 226. Tanaka A, Ito T, Kibata K, Inagaki-Katashiba N, Amuro H, Nishizawa T, et al. Serum high-mobility group box 1 is correlated with interferon- α and may predict disease activity in patients with systemic lupus erythematosus. *Lupus.* (2019) 28:1120–7. doi: 10.1177/09612033198 62865
- 227. Lood C, Tyden H, Gullstrand B, Jonsen A, Kallberg E, Morgelin M, et al. Platelet-derived S100A8/A9 and cardiovascular disease in systemic lupus erythematosus. *Arthritis Rheumatol.* (2016) 68:1970–80. doi: 10.1002/art.39656
- 228. Wu T, Xie C, Wang HW, Zhou XJ, Schwartz N, Calixto S, et al. Elevated urinary VCAM-1, P-selectin, soluble TNF receptor-1, and CXC chemokine ligand 16 in multiple murine lupus strains and human lupus nephritis. J Immunol. (2007) 179:7166–75. doi: 10.4049/jimmunol.179.10.7166
- 229. Delmas Y, Viallard JF, Solanilla A, Villeneuve J, Pasquet JM, Belloc F, et al. Activation of mesangial cells by platelets in systemic lupus erythematosus via a CD154-dependent induction of CD40. *Kidney Int.* (2005) 68:2068–78. doi: 10.1111/j.1523-1755.2005.00663.x
- 230. Schwarzenberger C, Sradnick J, Lerea KM, Goligorsky MS, Nieswandt B, Hugo CP, et al. Platelets are relevant mediators of renal injury induced by primary endothelial lesions. *Am J Physiol Renal Physiol.* (2015) 308:F1238– 46. doi: 10.1152/ajprenal.00535.2014
- 231. Tyden H, Lood C, Gullstrand B, Nielsen CT, Heegaard NHH, Kahn R, et al. Endothelial dysfunction is associated with activation of the type I interferon system and platelets in patients with systemic lupus erythematosus. *RMD Open.* (2017) 3:e000508. doi: 10.1136/rmdopen-2017-000508
- 232. Ostergaard O, Nielsen CT, Tanassi JT, Iversen LV, Jacobsen S, Heegaard NHH. Distinct proteome pathology of circulating microparticles in systemic lupus erythematosus. *Clin Proteomics*. (2017) 14:23. doi: 10.1186/s12014-017-9159-8
- 233. McCarthy EM, Moreno-Martinez D, Wilkinson FL, McHugh NJ, Bruce IN, Pauling JD, et al. Microparticle subpopulations are potential markers of disease progression and vascular dysfunction across a spectrum of connective tissue disease. *BBA Clin.* (2017) 7:16–22. doi: 10.1016/j.bbacli.2016.11.003
- 234. Mobarrez F, Vikerfors A, Gustafsson JT, Gunnarsson I, Zickert A, Larsson A, et al. Microparticles in the blood of patients with systemic lupus erythematosus (SLE): phenotypic characterization and clinical associations. *Sci Rep.* (2016) 6:36025. doi: 10.1038/srep36025
- 235. Mobarrez F, Fuzzi E, Gunnarsson I, Larsson A, Eketjall S, Pisetsky DS, et al. Microparticles in the blood of patients with SLE: size, content of mitochondria and role in circulating immune complexes. J Autoimmun. (2019) 102:142–9. doi: 10.1016/j.jaut.2019.05.003
- Linge P, Fortin PR, Lood C, Bengtsson AA, Boilard E. The non-haemostatic role of platelets in systemic lupus erythematosus. *Nat Rev Rheumatol.* (2018) 14:195–213. doi: 10.1038/nrrheum.2018.38
- Denton CP, Khanna D. Systemic sclerosis. Lancet. (2017) 390:1685–99. doi: 10.1093/med/9780199642489.003.0121_update_001

- Kuwana M, Okazaki Y, Yasuoka H, Kawakami Y, Ikeda Y. Defective vasculogenesis in systemic sclerosis. *Lancet.* (2004) 364:603–10. doi: 10.1016/S0140-6736(04)16853-0
- Ramirez GA, Franchini S, Rovere-Querini P, Sabbadini MG, Manfredi AA, Maugeri N. The role of platelets in the pathogenesis of systemic sclerosis. *Front Immunol.* (2012) 3:160. doi: 10.3389/fimmu.2012.00160
- Agache I, Radoi M, Duca L. Platelet activation in patients with systemic scleroderma-pattern and significance. *Roman J Intern Med.* (2007) 45:183– 91.
- 241. Yoshizaki A, Komura K, Iwata Y, Ogawa F, Hara T, Muroi E, et al. Clinical significance of serum HMGB-1 and sRAGE levels in systemic sclerosis: association with disease severity. *J Clin Immunol.* (2009) 29:180–9. doi: 10.1007/s10875-008-9252-x
- 242. Maugeri N, Rovere-Querini P, Baldini M, Baldissera E, Sabbadini MG, Bianchi ME, et al. Oxidative stress elicits platelet/leukocyte inflammatory interactions via HMGB1: a candidate for microvessel injury in sytemic sclerosis. *Antioxid Redox Signal.* (2014) 20:1060–74. doi: 10.1089/ars.2013.5298
- Dees C, Akhmetshina A, Zerr P, Reich N, Palumbo K, Horn A, et al. Plateletderived serotonin links vascular disease and tissue fibrosis. *J Exp Med.* (2011) 208:961–72. doi: 10.1084/jem.20101629
- 244. Maugeri N, Franchini S, Campana L, Baldini M, Ramirez GA, Sabbadini MG, et al. Circulating platelets as a source of the damageassociated molecular pattern HMGB1 in patients with systemic sclerosis. *Autoimmunity*. (2012) 45:584–7. doi: 10.3109/08916934.2012.7 19946
- 245. Smolen JS, Aletaha D, McInnes IB. Rheumatoid arthritis. Lancet. (2016) 388:2023-38. doi: 10.1016/S0140-6736(16)30173-8
- 246. Kusunoki Y, Nakazawa D, Shida H, Hattanda F, Miyoshi A, Masuda S, et al. Peptidylarginine deiminase inhibitor suppresses neutrophil extracellular trap formation and MPO-ANCA production. *Front Immunol.* (2016) 7:227. doi: 10.3389/fimmu.2016.00227
- 247. Khandpur R, Carmona-Rivera C, Vivekanandan-Giri A, Gizinski A, Yalavarthi S, Knight JS, et al. NETs are a source of citrullinated autoantigens and stimulate inflammatory responses in rheumatoid arthritis. *Sci Transl Med.* (2013) 5:178ra40. doi: 10.1126/scitranslmed.3005580
- Habets KL, Huizinga TW, Toes RE. Platelets and autoimmunity. Eur J Clin Invest. (2013) 43:746–57. doi: 10.1111/eci.12101
- 249. Farrugia M, Baron B. The role of TNF-alpha in rheumatoid arthritis: a focus on regulatory T cells. *J Clin Transl Res.* (2016) 2:84–90. doi: 10.18053/jctres.02.201603.005
- Pignatelli P, De Biase L, Lenti L, Tocci G, Brunelli A, Cangemi R, et al. Tumor necrosis factor-alpha as trigger of platelet activation in patients with heart failure. *Blood.* (2005) 106:1992–4. doi: 10.1182/blood-2005-03-1247
- 251. Greenberg JD, Kremer JM, Curtis JR, Hochberg MC, Reed G, Tsao P, et al. Tumour necrosis factor antagonist use and associated risk reduction of cardiovascular events among patients with rheumatoid arthritis. *Ann Rheum Dis.* (2011) 70:576–82. doi: 10.1136/ard.2010.129916
- 252. Westlake SL, Colebatch AN, Baird J, Curzen N, Kiely P, Quinn M, et al. Tumour necrosis factor antagonists and the risk of cardiovascular disease in patients with rheumatoid arthritis: a systematic literature review. *Rheumatology*. (2011) 50:518–31. doi: 10.1093/rheumatology/keq316
- 253. Boilard E, Nigrovic PA, Larabee K, Watts GF, Coblyn JS, Weinblatt ME, et al. Platelets amplify inflammation in arthritis via collagendependent microparticle production. *Science*. (2010) 327:580–3. doi: 10.1126/science.1181928

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2019 Ramirez, Manfredi and Maugeri. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.