

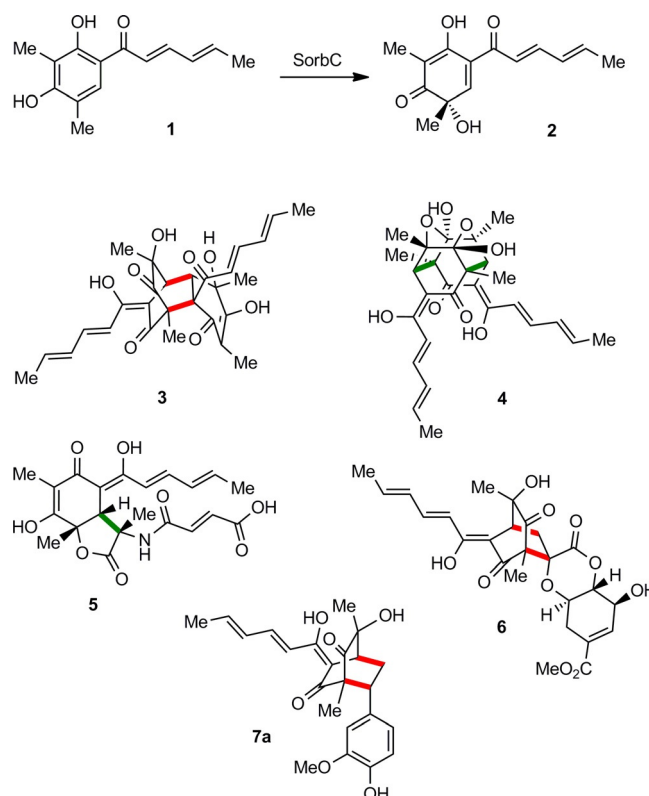
Chemoenzymatic Total Synthesis of Sorbicatechol Structural Analogues and Evaluation of Their Antiviral Potential

Anna Sib⁺,^[a] Tobias M. Milzarek⁺,^[a, b] Alexander Herrmann,^[c] Lila Oubraham,^[d]
Jonas I. Müller,^[b] Andreas Pichlmair,^[d] Ruth Brack-Werner,^[c] and Tobias A. M. Gulder*^[a, b]

Sorbicillinoids are fungal polyketides characterized by highly complex and diverse molecular structures, with considerable stereochemical intricacy combined with a high degree of oxygenation. Many sorbicillinoids possess promising biological activities. An interesting member of this natural product family is sorbicatechol A, which is reported to have antiviral activity, particularly against influenza A virus (H1N1). Through a straightforward, one-pot chemoenzymatic approach with recently developed oxidoreductase SorbC, the characteristic bicyclo[2.2.2]octane core of sorbicatechol is structurally diversified by variation of its natural 2-methoxyphenol substituent. This facilitates the preparation of a focused library of structural analogues bearing substituted aromatic systems, alkanes, heterocycles, and ethers. Fast access to this structural diversity provides an opportunity to explore the antiviral potential of the sorbicatechol family.

Sorbicillinoids are a large polyketide natural product family consisting of more than 50 members.^[1] They can be isolated

from a diverse set of marine and terrestrial fungi and can be categorized into four different groups, according to their molecular structures: monomeric, dimeric, trimeric, and further functionalized sorbicillinoids. Biosynthetically, all sorbicillinoids are derived from stereo- and regioselective oxidative dearomatization of sorbicillin (1) to the highly reactive sorbicillinol (2) by the oxidoreductase SorbC (Scheme 1).^[2] The di-/trimeric sorbicillinoids result from subsequent di-/trimerization of 2 with 1 or 2 by Michael addition or Diels–Alder cycloaddition due to the inherent respective reactivities of 2, leading to beautiful molecular architectures, such as bisorbicillinol (3, resulting from dimerization through a Diels–Alder reaction)^[3] or trichodimerol (4, resulting from double Michael addition/ketalization;^[4] Scheme 1). Further functionalized sorbicillinoids are likewise formed by these transformations, but instead of a simple dime-



Scheme 1. Top: Oxidative dearomatization of 1 to 2. Bottom: Examples of dimeric sorbicillinoid natural products 3 and 4, and of further functionalized congeners formed by Michael addition (green bonds), for example, sorbicillactone A (5), or Diels–Alder cycloaddition chemistry (red bonds), for example, spirosorbicillinol A (6) and sorbicatechol A (7a).

[a] A. Sib,⁺ T. M. Milzarek,⁺ Prof. Dr. T. A. M. Gulder
Biosystems Chemistry, Department of Chemistry and
Center for Integrated Protein Science Munich (CIPSM)
Technical University of Munich
Lichtenbergstrasse 4, 85748 Garching (Germany)
E-mail: tobias.gulder@ch.tum.de

[b] T. M. Milzarek,⁺ J. I. Müller, Prof. Dr. T. A. M. Gulder
Chair of Technical Biochemistry, Technical University of Dresden
Bergstrasse 66, 01069 Dresden (Germany)
E-mail: tobias.gulder@tu-dresden.de

[c] A. Herrmann, Prof. Dr. R. Brack-Werner
Helmholtz Zentrum München
German Research Center for Environmental Health (GmbH)
Institute of Virology
Ingolstädter Landstrasse 1, 85764 Neuherberg (Germany)

[d] Dr. L. Oubraham, Prof. Dr. A. Pichlmair
Immunopathology of Virus Infections Laboratory
Institute of Virology, Technical University of Munich
Schneckenburgerstrasse 8, 81675 Munich (Germany)

[*] These authors contributed equally to this work.

Supporting information and the ORCID identification numbers for the authors of this article can be found under <https://doi.org/10.1002/cbic.201900472>.

© 2019 The Authors. Published by Wiley-VCH Verlag GmbH & Co. KGaA. This is an open access article under the terms of the Creative Commons Attribution Non-Commercial NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

This article is part of the young researchers' issue ChemBioTalents. To view the complete issue, visit <http://chembiochem.org/chembiotalents>

zation, these transformations involve non-sorbicillinoid nucleophiles or dienophiles. Structurally and biomedically interesting examples of this class of sorbicillinoids include **5**,^[5] **6**,^[6] and **7a** (Scheme 1).^[7]

The biological activities of the functionalized sorbicillinoids are as diverse as their structures. Compound **5** is active against murine leukemic lymphoblast cell line L5178y, with an IC_{50} of 2.2 mg mL^{-1} , and protects human T cells from HIV-1 over a concentration range of 0.3 and 3.0 mg mL^{-1} .^[5] Compound **6** shows 2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging activity,^[6] this property is also typical for dimeric sorbicillinoids, such as **3**.^[3] Although currently far less studied, compared with the above-mentioned compounds, compound **7a** was reported to exhibit promising antiviral activity against H1N1.^[7] With an IC_{50} value of $85 \text{ }\mu\text{M}$, compound **7a** shows identical potential to that of the antiviral drug ribavirin (IC_{50} of $84 \text{ }\mu\text{M}$).

Because the structural variability of any given type of natural sorbicillinoid core is rather limited and the isolation of these compounds from the complex metabolic matrix of the fungal producer strains is tedious, synthetic strategies are required to facilitate biomedical studies on these fascinating molecules. Although a number of elegant total synthetic routes towards dimeric sorbicillinoids, such as **3** and **4**, do exist, most of these are not stereoselective and/or rather lengthy.^[8] In the case of the sorbicatchol core structure, only a single total synthetic approach exists. This led to the synthesis of *ent*-rezishanone C, with an ethoxy substituent instead of the 2-methoxyphenol substituent in **7a**. Overall, the synthetic route contains >20 individual steps, with a combined yield of less than 3%;^[9] this clearly provides evidence of the need for improved synthetic strategies towards the sorbicillinoids. To this end, we have recently developed chemoenzymatic approaches for the efficient one-pot synthesis of dimeric sorbicillinoids, such as **3** and **4**, utilizing synthetically readily available **1**, which is oxidatively dearomatized by employing SorbC to give **2** enantioselectively. Compound **2** can be dimerized in a controlled manner, depending on the organic cosolvent employed during the biocatalytic reaction.^[10] This methodology was further extended to

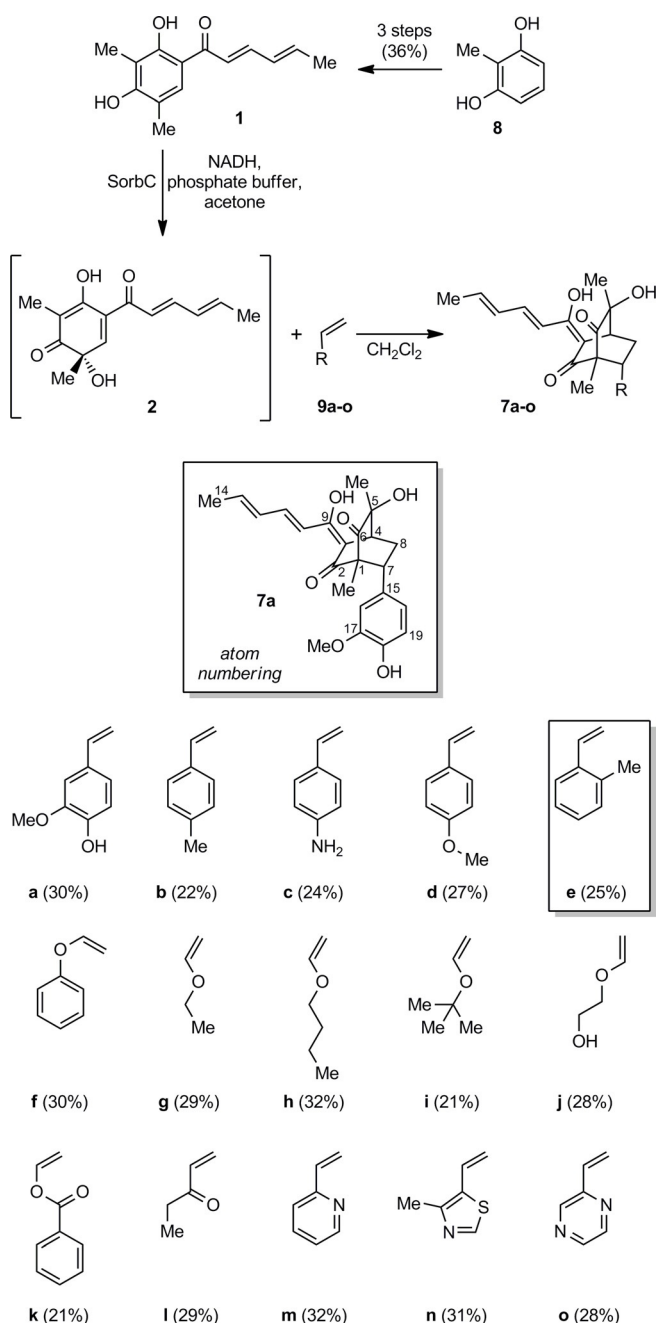
enable functionalization of **2** with external nucleophiles, for example, leading to the first stereoselective total synthesis of epoxysorbicillinol, and dienophiles, yielding the first synthetic access to rezishanones and **7a**.^[11] We herein present the application of this chemoenzymatic toolkit, featuring a facile intramolecular Diels–Alder reaction, for the straightforward preparation of a focused library of sorbicatchol-type structural analogues to enable the evaluation of their antiviral potential.

The required synthetic starting material, **1**, for the biocatalytic reaction was prepared in three-steps starting from 2-methylresorcinol, following our published procedure.^[11] In a regio- and stereoselective oxidative dearomatization reaction with SorbC, compound **1** was transformed into **2**, which was the reactive precursor for all sorbicatchol derivatives. This oxidative dearomatization reaction proceeds with perfect stereocontrol, exclusively delivering the desired product, (*S*)-**2**.^[10] Quenching of the reaction solution containing **2** with a diverse set of dienophiles readily delivered the target compounds in yields ranging from 21 to 32% (Scheme 2), irrespective of the substitution of the ene function. This is in the range of typical product yields for all sorbicillinoids following this chemoenzymatic strategy, and can be explained by unreacted starting material **1** and the formation of dimeric side products, particularly **3**, which cannot be fully suppressed due to the intrinsically high reactivity of **2**. Besides the synthesis of **7a**, sets of styrenes substituted at the aromatic portion (**9a–e**), vinyl ethers (**9f–k**), ethyl vinyl ketone (**9l**), and heteroaromatic building blocks (**9m–o**) were employed to give the respective sorbicatchol analogues **7a–o**. All compounds were isolated by means of semipreparative HPLC to give pure and stereochemically defined material (Figure S33 in the Supporting Information).

In the original isolation paper, the absolute configuration of the biologically more active *endo* Diels–Alder product, **7a**, and its likewise isolated, minor *exo* congener was thoroughly investigated through a combination of DFT calculations, 2D NOESY measurements, and analysis of experimental versus calculated circular dichroism (CD) spectra; thus allowing for an unambiguous assignment of the absolute configuration of both compounds.^[7] A comparison of the NMR spectroscopy data of compound **7a** produced by our approach with the data reported for the respective *endo* and *exo* products clearly revealed that our material corresponded to the *endo* product. This can most convincingly be deduced from chemical shift analysis of the ^{13}C NMR spectroscopy data, most importantly at C2 (**7a**: $\delta = 198.0$ ppm, reported for *endo*: $\delta = 197.9$ ppm, reported for *exo*: $\delta = 199.7$ ppm), C3 ($\delta = 112.2, 112.0, 110.5$ ppm), C7 ($\delta = 47.8, 47.8, 49.9$ ppm), C8 ($\delta = 31.5, 31.4, 30.2$ ppm), C16 ($\delta = 110.4, 110.2, 111.2$ ppm), and C5-methyl ($\delta = 24.4, 24.3, 25.2$ ppm), as well as from differences in ^1H NMR chemical shifts, particularly at H8 ($\delta = 1.84$ and $3.05, 1.84$ and $3.05, 2.25$ and 2.57 ppm), H16 ($\delta = 6.44, 6.45, 6.71$ ppm), and H20 ($\delta = 24.4, 24.3, 25.2$ ppm). In addition, NOESY interactions between the protons of C1-methyl at the sorbicillin-derived core structure with H16 and C17-OMe can only be seen in the *exo* analogue,^[7] and are consequently also absent in our product **7a** (Figure S4); this is in agreement with the above *endo* assignment. Most importantly, the large chemical shift difference of the geminal

Tobias A. M. Gulder holds degrees in chemistry from the University of Würzburg (Diploma 2004, PhD 2008). After postdoctoral training at the Scripps Institution of Oceanography (2008–2010), he started his independent work at the University of Bonn (2011–2014). In 2014, he was appointed Professor of Biosystems Chemistry at the Department of Chemistry and the Center for Integrated Protein Science Munich, Technical University of Munich. He then became Chair of Technical Biochemistry at the Technical University of Dresden. Since 2018, he has been Chair of the Editorial Board of *Natural Product Reports*. He is interested in the structure, biosynthesis, and synthesis of bacterial natural products.





Scheme 2. Synthesis of **7a** and 14 structural analogues (**7b–o**). Yields for each analogue are given in parentheses. NAD^+ : nicotinamide adenine dinucleotide.

protons attached to C8, which are typical for the *endo* product **7a** (difference 1.21 ppm), compared with the *exo* product (0.32), can also be found in all other compounds presented herein (**7b**: 1.15; **7c**: 1.23, **7d**: 1.16, **7e**: 1.24, **7f**: 1.22, **7g**: 1.10, **7h**: 1.09, **7i**: 1.17, **7j**: 1.12, **7k**: 1.42, **7l**: 1.46, **7m**: 1.53, **7n**: 1.45, **7o**: 0.89); thus corroborating that all products are *endo*. The regioselectivity of the Diels–Alder cycloadditions can generally be explained by the directing effect of the C1-methyl group (see numbering of **7a** in Scheme 2, *ortho* rule).

Because of the reported antiviral activity of **7a** against H1N1, a set of the prepared sorbicatechol analogues, **7a–o**,

were evaluated for their antiviral activity against influenza A virus (IAV). Initial tests were performed by using the *Gaussia* luciferase reporter virus.^[12] The abundance of *Gaussia* luciferase activity that accumulates in the supernatant of cells infected with this virus can be used as a proxy for virus replication. A resazurine conversion assay was also performed to assess the cytotoxic effect of the compounds on the cells.

Unfortunately, the tested compounds exhibited cytotoxicity to the viral host cells at concentrations below the observation of antiviral effects. This effect thus not only made it impossible to assess any antiviral effects against IAV, but also raised the question of whether the initially reported antiviral activity of **7a**^[7] was a true antiviral effect or rather the result of the cytotoxicity of the compound to the host cells. To further broaden our antiviral screening, we thus decided to expand from only testing *Orthomyxoviridae* (IAV) to *Retroviridae* by including tests against HIV-1. This system was also chosen due to the significant need for finding compounds active against HIV. Despite the fact that current anti-HIV drugs can minimize virus replication and thereby prevent the outbreak of AIDS, a series of severe problems remain, including the rapid emergence of resistant viruses, high virus variability, high costs, and adverse side effects.^[13] As a test system for anti-HIV activity, the EASY-HIT assay^[14] was performed; this is based on the reporter cell line LCS-RIC, which contains a stably integrated fluorescent reporter gene. Upon HIV infection, these cells express the fluorescent marker dsRed, which can be directly connected to the anti-HIV activity of a compound when reduced during treatment.^[14] We used the resazurin conversion assay, which is based on the enzymatic reduction of resazurin,^[15] to check for any signs of negative effects on the vitality and metabolism of the LCS-RIC cells that could appear during the course of the EASY-HIT assay. Of the 15 sorbicatechol analogues evaluated for anti-HIV properties, 6 compounds showed activities beyond their respective cytotoxicity values, as determined by the viability assay.^[15] Interestingly, all active compounds were either derived from a styrene (**9a–e**) or equipped with an aromatic portion in close proximity to the ene functionality in the substrate, as in phenyl vinyl ether **9f**. No other compounds exhibited a measurable activity, including all additional ethers and all heteroaromatic systems, as well as the benzoylated derivative **7k**, with only one additional keto function between the alkene and aromatic moiety, when compared with **7f**. These results strongly indicate that an aromatic portion in close proximity to the bicyclo[2.2.2]octane core structure of the sorbicatechols is required for activity. The selectivity indices (equaling the $\text{CC}_{50}/\text{IC}_{50}$) for the active compounds are mostly in the range of 1.37 to 1.72 (Table 1). In this range, cytotoxic effects on the host cells might indeed influence the antiviral activity values. For the *ortho*-methyl substituted analogue, **7e**, however, a significant increase in activity can be observed, along with a slight decrease of cytotoxicity. Overall, this leads to a selectivity index of 3.49 for **7e**. Interestingly, a simple repositioning of the methyl substituent from the *para* (as in **7b**) to the *ortho* position led to a twofold increase in activity.

In summary, we have synthesized 15 sorbicatechol derivatives **7a–o** by application of a chemoenzymatic synthesis of **2**

Table 1. Anti-HIV activity given as IC₅₀ values and cytotoxicity determined as CC₅₀ in the viability assay.^[15] The selectivity index is calculated from CC₅₀/IC₅₀. Emtricitabine (2',3'-dideoxy-5-fluoro-3'-thiacytidine, FTC) was used as the HIV inhibition control.

Compound	Anti-HIV activity IC ₅₀ [μM]	Viability assay CC ₅₀ [μM]	Selectivity index (CC ₅₀ /IC ₅₀)
7a	65.9 ± 4.68	102*	1.55
7b	68.8 ± 6.97	105.2*	1.53
7c	75.9 ± 7.37	125.2*	1.65
7d	62.7 ± 10.08	108.4*	1.72
7e	32.2 ± 2.52	112.3*	3.49
7f	76.6 ± 4.28	105.2*	1.37
FTC	0.7 ± 0.22	> 100.0	> 140

followed by quenching with a diverse set of dienophiles. Evaluation of their activity against IAV has revealed their cytotoxicity against the host cell system, rather than a true antiviral effect. Screening of the compound library against HIV-1 showed that aromatic substitution, as the bicyclo[2.2.2]octane substructure, was required to obtain antiviral activity, although it remained weak. The strongest activity was obtained with **7e**, with an IC₅₀ value of approximately 32 μM and a selectivity index of 3.49. The activity is thus about 50 times lower than that of the commercial virostatic drug emtricitabine (FTC), with a >40-fold decrease in selectivity index. In contrast to previous reports, our findings did not reveal antiviral activity of the sorbicatechols over a range that would be promising for further antiviral lead optimization.

Experimental Section

Experimental details: The synthesis of sorbicatechol derivatives followed a two-phase procedure, which is described here for the production of the most active derivative, **7e**, as an example. First, compound **1** was dissolved in acetone and added to phosphate buffer (50 mM, pH 8) with the enzyme SorbC. The reaction started upon the addition of NADH and was incubated for 6 h at room temperature. Second, product **2** was extracted with organic solvent and *o*-methylstyrene (**9e**) was added. After slow evaporation of the organic solvent under reduced pressure, to increase the concentration of the dienophile slowly over time, the crude product was purified by means of preparative HPLC.

Acknowledgements

We thank the groups of Prof. Dr. S. Sieber (Chair of Organic Chemistry II, TUM) for measuring HRMS data and of Prof. Dr. Tanja Gulder (Biomimetic Catalysis, TUM) for providing some of the chemical starting materials. T.M.M. thanks the Stiftung der Deutschen Wirtschaft (SDW) for his scholarship. We are very grateful to the DFG for generous financial support of this work

(GU 1233/1-1 and the Center for Integrated Protein Science Munich CIPSM).

Conflict of Interest

The authors declare no conflict of interest.

Keywords: antiviral agents · biocatalysis · natural products · sorbicillinoids · total synthesis

- [1] A. M. Harned, K. A. Volp, *Nat. Prod. Rep.* **2011**, *28*, 1790–1810.
- [2] A. Al-Fahad, A. Abood, K. M. Fisch, A. Osipow, J. Davison, M. Avramovic, C. P. Butts, J. Piel, T. J. Simpson, R. J. Cox, *Chem. Sci.* **2014**, *5*, 523–527.
- [3] N. Abe, T. Murata, A. Hirota, *Biosci. Biotechnol. Biochem.* **1998**, *62*, 661–666.
- [4] R. Andrade, W. A. Ayer, P. P. Mebe, *Can. J. Chem.* **1992**, *70*, 2526–2536.
- [5] a) G. Bringmann, G. Lang, T. A. M. Gulder, H. Tsuruta, J. Mühlbacher, K. Maksimenka, S. Steffens, K. Schaumann, R. Stöhr, J. Wiese, J. F. Imhoff, S. Perovic-Ottstadt, O. Boreiko, W. E. G. Müller, *Tetrahedron* **2005**, *61*, 7252–7265; b) G. Bringmann, T. A. M. Gulder, G. Lang, S. Schmitt, R. Stöhr, J. Wiese, K. Nagel, J. F. Imhoff, *Mar. Drugs* **2007**, *5*, 23–30.
- [6] K. Washida, N. Abe, Y. Sugiyama, A. Hirota, *Biosci. Biotechnol. Biochem.* **2009**, *73*, 1355–1361.
- [7] J. Peng, X. Zhang, L. Du, W. Wang, T. Zhu, Q. Gu, D. Li, *J. Nat. Prod.* **2014**, *77*, 424–428.
- [8] a) K. C. Nicolaou, R. Jautelat, G. Vassilikogiannakis, P. S. Baran, K. B. Simonsen, *Chem. Eur. J.* **1999**, *5*, 3651–3665; b) K. C. Nicolaou, G. Vassilikogiannakis, K. B. Simonsen, P. S. Baran, Y.-L. Zhong, V. P. Vidali, E. N. Pit-sinos, E. A. Couladouros, *J. Am. Chem. Soc.* **2000**, *122*, 3071–3079; c) R. Hong, Y. Cheng, L. Deng, *Angew. Chem. Int. Ed.* **2005**, *44*, 3478–3481; *Angew. Chem.* **2005**, *117*, 3544–3547; d) D. Barnes-Seeman, E. J. Corey, *Org. Lett.* **1999**, *1*, 1503–1504.
- [9] Q. Yan, M. G. Banwell, M. L. Coote, R. Lee, A. C. Willis, *Chem. Asian J.* **2017**, *12*, 1480–1484.
- [10] A. Sib, T. A. M. Gulder, *Angew. Chem. Int. Ed.* **2017**, *56*, 12888–12891; *Angew. Chem.* **2017**, *129*, 13068–13071.
- [11] A. Sib, T. A. M. Gulder, *Angew. Chem. Int. Ed.* **2018**, *57*, 14650–14653; *Angew. Chem.* **2018**, *130*, 14861–14864.
- [12] N. Eckert, F. Wrensch, S. Gärtner, N. Palanisamy, U. Goedecke, N. Jäger, S. Pöhlmann, M. Winkler, *PLoS One* **2014**, *9*, e97695.
- [13] A. A. Waheed, G. Tachedjian, *Curr. Top. Med. Chem.* **2016**, *16*, 1343–1349.
- [14] S. Kremb, M. Helfer, W. Heller, D. Hoffmann, H. Wolff, A. Kleinschmidt, S. Cepok, B. Hemmer, J. Durner, R. Brack-Werner, *Antimicrob. Agents Chemother.* **2010**, *54*, 5257–5268.
- [15] T. L. Riss, R. A. Moravec, A. L. Niles, S. Duellman, H. A. Benink, T. J. Worzella, L. Minor in *Assay Guidance Manual, Vol. 1* (Eds: G. S. Sittampalam, N. P. Coussens, K. Brimacombe, A. Grossman, M. Arkin, D. Auld, C. Austin, J. Baell, B. Bejcek, J. M. M. Caaverio, T. D. Y. Chung, J. L. Dahlin, V. Devanaryan, T. L. Foley, M. Glicksman, M. D. Hall, J. V. Haas, J. Ingles, P. W. Iversen, S. D. Kahl, S. C. Kales, M. Lal-Nag, Z. Li, J. McGee, O. McManus, T. Riss, O. J. Trask, J. R. Weidner, M. J. Wildey, M. Xia, X. Xu), Eli Lilly & Company and the National Center for Advancing Translational Sciences, Bethesda, **2004**, pp. 305–335.

Manuscript received: July 31, 2019

Accepted manuscript online: August 25, 2019

Version of record online: November 19, 2019