

# Generating Phenotypic Diversity in a Fungal Biocatalyst to Investigate Alcohol Stress Tolerance Encountered during Microbial Cellulosic Biofuel Production

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## Abstract

Consolidated bioprocessing (CBP) of lignocellulosic biomass offers an alternative route to renewable energy. The crop pathogen *Fusarium oxysporum* is a promising fungal biocatalyst because of its broad host range and innate ability to co-saccharify and ferment lignocellulose to bioethanol. A major challenge for cellulolytic CBP-enabling microbes is alcohol inhibition. This research tested the hypothesis that *Agrobacterium tumefaciens*-mediated transformation (ATMT) could be exploited as a tool to generate phenotypic diversity in *F. oxysporum* to investigate alcohol stress tolerance encountered during CBP. A random mutagenesis library of gene disruption transformants ( $n=1,563$ ) was constructed and screened for alcohol tolerance in order to isolate alcohol sensitive or tolerant phenotypes. Following three rounds of screening, exposure of select transformants to 6% ethanol and 0.75% *n*-butanol resulted respectively in increased ( $\geq 11.74\%$ ) and decreased ( $\leq 43.01\%$ ) growth compared to the wild-type (WT). Principal component analysis (PCA) quantified the level of phenotypic diversity across the population of genetically transformed individuals and isolated candidate strains for analysis. Characterisation of one strain, Tr. 259, ascertained a reduced growth phenotype under alcohol stress relative to WT and indicated the disruption of a coding region homologous to a putative sugar transporter (FOXG\_09625). Quantitative PCR (RT-PCR) showed FOXG\_09625 was differentially expressed in Tr. 259 compared to WT during alcohol-induced stress ( $P<0.05$ ). Phylogenetic analysis of putative sugar transporters suggests diverse functional roles in *F. oxysporum* and other filamentous fungi compared to yeast for which sugar transporters form part of a relatively conserved family. This study has confirmed the potential of ATMT coupled with a phenotypic screening program to select for genetic variation induced in response to alcohol stress. This research represents a first step in the investigation of alcohol tolerance in *F. oxysporum* and has resulted in the identification of several novel strains, which will be of benefit to future biofuel research.

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## Introduction

Lignocellulosic biomass is an abundant feedstock and attractive source of sugars for biofuel production. Large-scale utilisation is however challenged by the general lack of low-cost technologies that can overcome cellulose recalcitrance [1]. One potential route to eco-friendly sustainable energy production is the consolidated bioprocessing (CBP) of biomass into biofuels [2].

Several yeasts (e.g. *Saccharomyces cerevisiae*, *Pichia stipitis*, *Candida shehatae*, *Pachysolen tannophilus*) and bacteria (e.g. *Escherichia coli*, *Klebsiella oxytoca*, *Zymomonas*

*mobilis*) have been engineered for CBP ethanol production however their capacity to secrete saccharification and fermentation enzymes at sufficient yield remains an obstacle [3–7]. In contrast, filamentous fungi including *Trichoderma* sp., *Neurospora* sp., *Aspergillus* sp., *Monilinia* sp., *Rhizopus* sp., *Mucor* sp., *Paecilomyces* sp. and *Fusarium* sp., possess a large repertoire of lignocellulolytic enzymes due to their co-evolution with plants, and can convert released plant-derived sugars into ethanol [7–10]. In particular, the broad host range phytopathogen *Fusarium oxysporum* [11] can degrade and produce ethanol from various cellulosic substrates (e.g. untreated and pre-treated straw [12,13], brewer's spent grain

[14], potato waste [15]). Previous work [13] identified *F. oxysporum* strain 11C as a promising microbial biocatalyst capable of producing high bioethanol yields from delignified wheat straw.

At present no organism can ferment cellulosic materials to ethanol at rates and titres necessary to achieve economic feasibility [5]. CBP requires robust microbes able to: (i) degrade lignocellulosic materials in the absence of supplementary exogenous enzymes, (ii) utilise at high efficiency both hexose and pentose sugars including their conversion into high-titer ethanol and (iii) tolerate toxic compounds notably the primary alcohol product formed during fermentation [16]. Whereas the endogenous ability of *F. oxysporum* to tolerate inhibitory compounds encountered during CBP including lignocellulosic hydrolytates (carboxylic acids, phenolic compounds, furan derivatives) [17] and the fermentation by-product acetic acid [18] has been reported, a knowledge deficit exists in regards to *F. oxysporum*'s capability to tolerate ethanol.

Ethanol stress affects cell growth and viability in addition to productivity [19,20]. While other compounds compatible as biofuels such as *n*-butanol offer advantages over ethanol (e.g. high energy content and miscibility with gasoline), production is limited by the sensitivity of native producers such as *Clostridium* sp. to the alcohol [21]. This has led to the investigation of alternative butanol production hosts [22-24], prompting the investigation of *n*-butanol (hereafter referred to as butanol) tolerance in addition to ethanol in this study. For the cost-effective and commercially viable production of biofuels, product yields must exceed native microbial tolerance levels [25] therefore engineering of stress tolerant strains is critical to achieve high survival and production rates [25,26].

Several strategies have been investigated to improve alcohol tolerance in yeast and bacteria, including gene over-expression or random knockout libraries, genome shuffling and transcriptional or translational engineering [27]. For filamentous fungi ATMT has established itself as a valuable and powerful tool for genetic studies, greatly facilitating the identification and analysis of genes involved in complex biological processes such as host pathogenicity [28] and, as we hypothesise here, in regards to alcohol tolerance. As the gram-negative soil bacterium *A. tumefaciens* facilitates the stable transfer and integration of the tumour inducing (Ti) plasmid segment (known as T- DNA) into the targeted host cell [29], ATMT holds significant advantages over more traditional methods of gene disruption. For example, *A. tumefaciens*' broad host range, its potential to transform a range of starting materials and its propensity for low copy number insertion events [30]. ATMT also provides an ability to 'tag' the disrupted gene(s) thereby facilitating their identification via genome walking. In the case of *F. oxysporum* this is further assisted by the recent availability of the organism's genome sequence [31] (<http://www.broadinstitute.org>).

In this study, we hypothesised that ATMT could be exploited as a tool to generate significant degrees of phenotypic diversity in *F. oxysporum* strain 11C in response to alcohol stress. To this end, a modified ATMT protocol was established to generate a library of random gene disruption transformants, which underwent a three tier physiological evaluation. As a

result, the initial population of *F. oxysporum* transformants was reduced from 1,563 to 29 individuals, which were subjected to ethanol tolerance screening and cross-resistance to butanol. The level of recorded phenotypic variation confirmed the occurrence of ATMT-derived genotypes with several *F. oxysporum* transformants identified for future analysis. To the best of our knowledge this is the first report detailing the use of random insertional mutagenesis to investigate alcohol tolerance in a fungal CBP agent.

## Materials and Methods

### Origin and maintenance of fungi

*Fusarium oxysporum* strain 11C originated from Irish soils as previously described [13]. For ATMT, *F. oxysporum* was sub-cultured onto potato dextrose agar (PDA; Difco, UK) plates, incubated at 25°C for 5 days prior to producing fungal conidial inoculum in Mung bean broth [32].

### Plasmid and ATMT transformation

The binary vector used for fungal gene disruption was pSK1019, which is equipped with the *hph* antibiotic resistance marker gene under the control of an *Aspergillus nidulans* *TrpC* promoter and was donated by Professor Seogchan Kang (The Pennsylvania State University, USA). *Agrobacterium tumefaciens* strain AGL-1 was transformed with the binary vector pSK1019 [33] and ATMT of *F. oxysporum* strain 11C was optimised based on a previously published protocol [34] (see Methods S1). Three independent ATMTs of *F. oxysporum* strain 11C were conducted.

### Molecular analysis of transformants

For PCR analysis, fungal genomic DNA was extracted from mycelia using a modified method [35] (see Methods S1). DNA was quantified using a Qubit™ Quant-iT assay (Invitrogen, U.S) and stored at -80°C. PCR was used to detect the presence/absence of the *hph* transgene in DNA extracts from putative transformants (see Methods S1). For Southern blot analysis, genomic DNA was extracted [36] and quantified using a NanoDrop® ND-1000 Spectrophotometer (NanoDrop Technologies, USA). Southern blot analysis was used to determine plasmid copy number in transformants (see Methods S1).

### Primary alcohol tolerance screen

A total of 1,563 putatively transformed hygromycin-resistant colonies were isolated into microplates (96-well) containing 200 µl minimal medium (supplemented with hygromycinB, 60 µg ml<sup>-1</sup>, Sigma, UK) per well [37] which were then sealed and incubated at 25°C for 3 days and subsequently used as inoculum for primary screening of alcohol tolerance. For the primary screen (first of three), 96-well plates were prepared containing 140µl minimal medium [37] per well supplemented with six treatments either hygromycinB (60 µg ml<sup>-1</sup>), no ethanol or 0.5, 6.0 or 10% vv<sup>-1</sup> ethyl alcohol (Sigma, UK) plus or minus hygromycinB (60 µg ml<sup>-1</sup>; Sigma, UK). To target different response levels, low (0.5% vv<sup>-1</sup> ethanol), medium (6% vv<sup>-1</sup>

ethanol) and high (10%  $vv^{-1}$ ) ethanol concentrations were then selected. Wells were inoculated with 10  $\mu$ l of fungal material (one well per treatment per transformant/wild type fungus; plate layout is shown in Figure S1A). Plates were sealed with a lid and parafilm and incubated at 25°C with fungal growth ( $OD_{600nm}$ ) measured after both 48 and 96 hours using a spectrophotometer (Safire<sup>2</sup>, Tecan, Austria). Putative transformants with a  $\geq 2$ -fold higher  $OD_{600nm}$  in 10% ( $vv^{-1}$ ) ethanol relative to wild type fungus 11C were purified prior to secondary screening. This primary screening of the transformant collection was conducted once followed by the selection of putative transformants for purification and graduation to the secondary screening (see Methods S1).

### Secondary alcohol tolerance screen

Conidial suspensions of the *F. oxysporum* 11C wild type strain and transformants of interest (20  $\mu$ l from the from monoconidial 20% ( $vv^{-1}$ ) -80°C glycerol stock) were inoculated into 220  $\mu$ l minimal medium in microtiter (96-well) plate wells and incubated at 25°C for 24 hours. This served as inoculum for the secondary screen. For secondary screening; 100  $\mu$ l of fungal culture was transferred into microtiter plate wells containing 100  $\mu$ l minimal medium [37] supplemented with either 0, 6, 7, 8, 9 or 10%  $vv^{-1}$  ethanol (single well per treatment per transformant/wild type; see Figure S1B for plate layout). Plates were sealed with a lid and parafilm and incubated at 25°C for 96 hours after which fungal growth ( $OD_{600nm}$ ) was recorded. Data was normalised relative to the control (wild-type strain 11C). All datasets (0-10  $vv^{-1}$  ethanol) were used for selecting transformants of interest. From the secondary screen, *F. oxysporum* strain 11C putative transformants demonstrating either ethanol sensitive or ethanol tolerant phenotypes relative to the wild-type strain 11C control, at concentrations ranging between 6-10% ( $vv^{-1}$ ) were selected for tertiary screening. This experiment was conducted once.

### Tertiary alcohol tolerance screen

In the tertiary screen, tolerance to both ethanol and butanol was assessed, with the goal of examining the potential for cross-correlation between ethanol and butanol tolerance. Conidial inoculum for the tertiary assessment was generated by individually inoculating each transformant into Mung bean broth [32] (100 ml conical flask) with three fungal plugs and incubating at 25°C and 150 rpm for 5 days. Inoculum was harvested by centrifugation and washed twice with sterile distilled water and resuspended in minimal medium [37] to a concentration of  $10^6$   $ml^{-1}$  conidia. Microtiter (96-well) plates were prepared with wells containing 100  $\mu$ l minimal medium supplemented with either no alcohol, ethanol [2, 4, 6, 8 or 10% ( $vv^{-1}$ )] or butanol (0.25, 0.5, 0.75, 1 and 1.25% ( $vv^{-1}$ )) with a single treatment tested per 96-well plate (see Figure S1C for plate layout). Each plate was inoculated with 100  $\mu$ l fungal conidia ( $10^6$   $ml^{-1}$ ) (eight wells per treatment per wild type/transformant strain). Plates were sealed with a lid and parafilm and incubated at 25°C for 7 days. Fungal growth was measured using a spectrophotometer  $OD_{600nm}$  (Spectra Max 340 PC 96-well Plate Reader, Molecular Devices, USA), and percentage increase or decrease in growth relative to wild-type

control was determined. For data analysis see Methods S1. This experimental screening was repeated three times.

### Phenotypic analysis of *F. oxysporum* 11C and Tr. 259

A series of phenotypic assays of *F. oxysporum* 11C and Tr. 259 were conducted to investigate the effect of increasing alcohol concentration on growth, temporal analysis of alcohol tolerance, the effect of alcohol tolerance stress on spore germination and biomass production (see Methods S1). All experiments were conducted either twice or three times. For data analysis see Methods S1.

### Genome walking and sequence analysis

Genome walking was conducted on Tr. 259 to characterise the degree of genetic disruption underpinning the transformant's observed phenotype (see Methods S1 & Table S1).

### RT-PCR of putative hexose transporter (FOXG\_09625) under alcohol stress

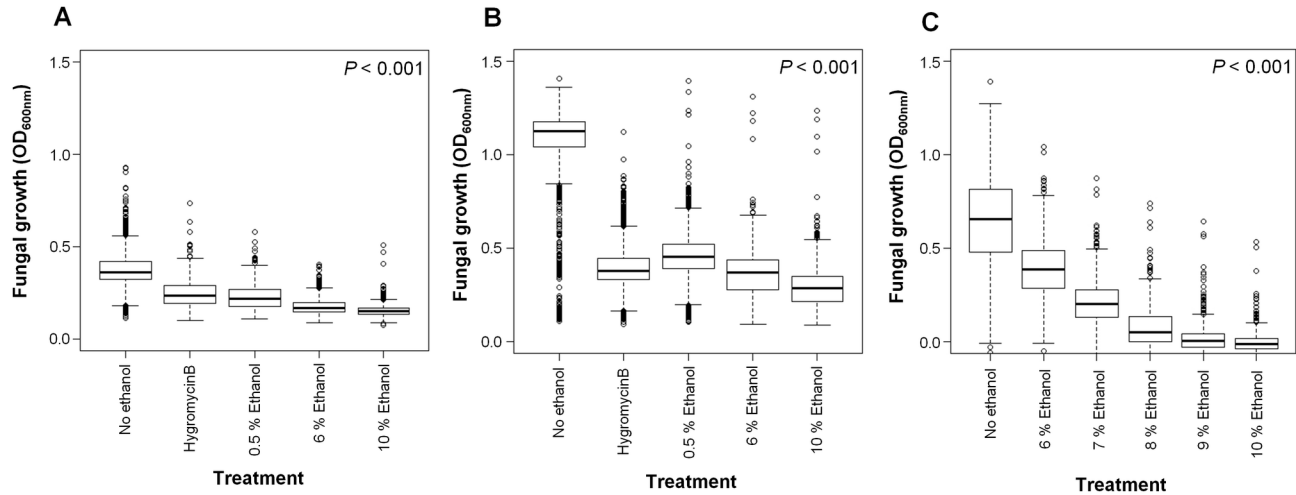
An experiment was designed to investigate the temporal accumulation of a putative hexose transporter under alcohol stress (Figure S2) (see Methods S1 & Table S1). Each experiment was conducted either three times (no alcohol, ethanol) or twice (butanol) with three replicates per strain per treatment per time point. Real-time quantification of the target and housekeeping gene respectively were performed in separate reactions. The threshold cycle ( $C_T$ ) values obtained from RT-PCR were used to calculate the accumulation of the target gene (relative mRNA accumulation) relative to  $\beta$ -tubulin transcript by the  $2^{-\Delta\Delta C_T}$  method, where  $\Delta\Delta C_T = (C_T, \text{target gene} - C_T, \beta\text{-tubulin})$  [38]. Results were based on the average obtained for two replica RT-PCR reactions per sample. For data analysis see Methods S1.

### Phylogenetic analysis of putative sugar transporters in *F. oxysporum*

A phylogram of putative *F. oxysporum* strain 4287 sugar transporters was constructed using amino acid sequences from transcripts showing  $\geq 30\%$  homology to FOXG\_09625 (See Methods S1).

### Data analysis

The primary/secondary screen dataset was analysed using the Boxplot function in R (R v2.15.2 R Development Core Team, 2012) and a Box and Whisker plot was used to depict the distribution of transformant response across various treatments. The primary/secondary screen dataset was non-normally distributed as determined within Minitab (Minitab release © 16, 2011 Minitab Inc.). The significance between treatments at each time point (48 and 96 hours respectively) or ethanol treatments was analysed within the Statistical Package for the Social Sciences (SPSS 18, SPSS Inc.) using Kruskal-Wallis H-test for non-parametrical data. The significance between treatments and transformants amongst the 29 individuals selected from secondary screening was determined using one-way ANOVA as the data could be transformed to fit



**Figure 1. Primary and second – tier screening of putative *Fusarium oxysporum* strain 11C transformants.** Box and Whisker plot depicting the distribution of putative *Fusarium oxysporum* strain 11C transformants ( $n = 1,563$ ) screened for altered ethanol tolerance during primary screening at 48 hours (A) and 96 hours (B) and transformants ( $n = 402$ ) screened for altered ethanol tolerance during secondary screening (C). Fungi were grown at 25°C in minimal medium [37] with five different treatments; no ethanol control; hygromycinB 60  $\mu\text{g ml}^{-1}$  control; 0.5% ( $\text{v}v^{-1}$ ) ethanol; 6% ( $\text{v}v^{-1}$ ) ethanol; 10% ( $\text{v}v^{-1}$ ) ethanol (Primary-tier) (A, B) or six treatments; no alcohol control; 6% ( $\text{v}v^{-1}$ ) ethanol; 7% ( $\text{v}v^{-1}$ ) ethanol; 8% ( $\text{v}v^{-1}$ ) ethanol; 9% ( $\text{v}v^{-1}$ ) ethanol; 10% ( $\text{v}v^{-1}$ ) ethanol (Second-tier) (C). Fungal growth ( $\text{OD}_{600\text{nm}}$ ) was measured after 96 hours using a spectrophotometer (Safire<sup>2</sup>, Tecan, Austria).

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a normal distribution using Johnson transformation within Minitab. For tertiary screen analysis see Methods S1.

## Results

### ATMT library construction for primary alcohol screening

A transformation platform was established for *F. oxysporum* strain 11C to generate a library of gene disruption transformants ( $n = 1,563$ ) (Figure S3). The library was screened using a high throughput alcohol screen based on microtiter (96-well) plates to discriminate against low to high ethanol (0.5–10% ( $\text{v}v^{-1}$ )) response levels measured across 48 and 96 hours (Figure 1A & 1B & Figure S3). Transformant selection for second – tier screening, as previously described [Methods], was based on recordings at 96 hours for which greater phenotypic variation was observed compared to 48 hours. An established threshold [Methods] led to the isolation of 182 putative transformants, which when purified (4 monoconidial cultures produced per putative transformant) yielded 443 viable monoconidial-derived cultures for PCR analysis and secondary alcohol screening.

### PCR analysis and secondary alcohol screening

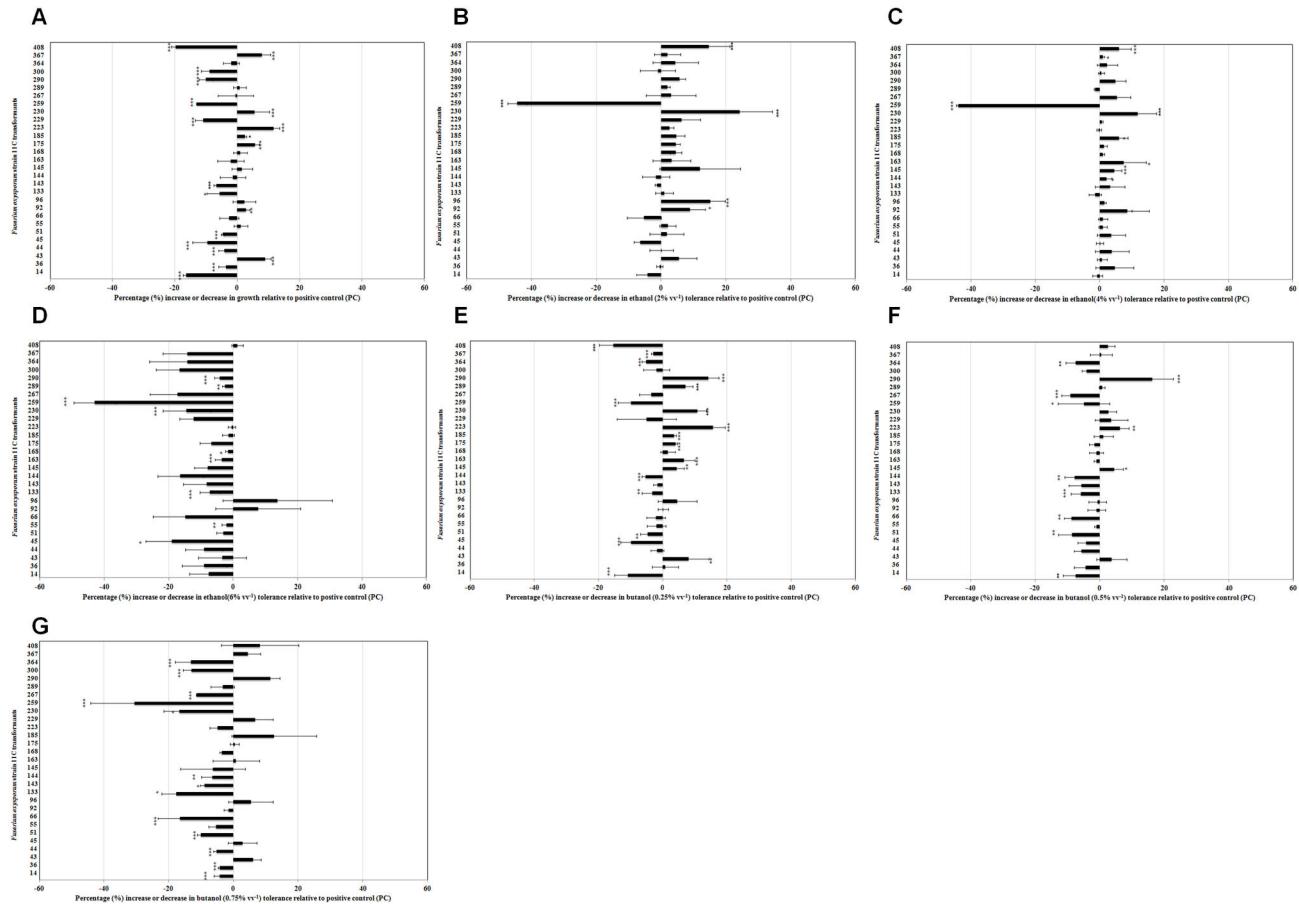
Over 90% (402/443) of the monoconidial lines generated from primary transformants were PCR-positive for the *hph* transgene (Figure S4A). On re-screening the ethanol tolerance of this population, a significant difference between treatments was observed between the derived monoconidial lines ( $P < 0.001$ ) (Figure 1C). A total of 29 PCR-verified transformants

indicating hypo- or hyper-ethanol growth phenotypes were selected for tertiary screening.

### Tertiary alcohol screening and transformant selection

Tertiary screening was used to assess tolerance of the 29 selected transformants to ethanol and cross-tolerance to butanol (Figure 2 & Table 1). A significant level of phenotypic diversity was recorded between strains across all treatments ( $P < 0.001$ ) (Figure 2). At 2 and 4% ( $\text{v}v^{-1}$ ) ethanol the highest level of hyper- or hypo-tolerance relative to parent strain 11C was observed for Tr. 230 and Tr. 259 (Figure 2B & 2C). At the highest ethanol concentration tested (6%  $\text{v}v^{-1}$ ), Tr. 259 continued to demonstrate a substantial hypo-tolerance to ethanol relative to the parental strain ( $-43.01\% \pm 6.40$ ) (Figure 2D). A strong positive correlation was noted between ethanol and butanol treatments respectively ( $r = 0.414$ ;  $n = 6$ ,  $P = 0.01$ ). Similar to results recorded for ethanol, at the highest butanol concentration (0.75%  $\text{v}v^{-1}$ ) Tr. 259 ( $-30.61\% \pm 13.49$ ) showed the highest hypo-butanol phenotype and Tr. 185 ( $12.57\% \pm 13.16$ ) the most hyper-butanol phenotype relative to strain 11C (Figure 2G).

Southern blot analysis of a random sub-set of nine transformants using an *hph* probe indicated an average transgene copy number of 1.55 (Figure S4B). PCA analysis (Figure 3) highlighted the degree of phenotypic diversity among transformants and isolated Tr. 259 as a candidate strain of interest for further analysis. Southern analysis of this transformant suggests a single insertion event (data not shown). From the presented evidence, nine transformants were identified as candidate strains for future studies showing



**Figure 2. Tertiary screening of *Fusarium oxysporum* strain 11C transformants (n=29).** Fungi were grown in minimal medium [37] supplemented with; no alcohol (A); 2% (v<sup>-1</sup>) ethanol (ethyl alcohol; Sigma, UK) (B), 4% (v<sup>-1</sup>) ethanol (C), 6% (v<sup>-1</sup>) ethanol (D) or 0.25% (v<sup>-1</sup>) butanol (*n*-butanol; Sigma, UK) (E), 0.5% (v<sup>-1</sup>) butanol (F) or 0.75% (v<sup>-1</sup>) butanol (G) and incubated at 25°C for 7 days. Percentage increase or decrease in fungal growth relative to the positive control (wild-type strain 11C) was determined. Results represent the mean of three independent experiments and bars indicate SEM. Transformants significantly different from the positive control (wild-type strain 11C) are highlighted with an asterisk (level of significance: <0.050 \*, <0.01\*\*, <0.001\*\*\*). doi: 10.1371/journal.pone.0077501.g002

significantly ( $P<0.05$ ) altered tolerance relative to the positive control (wild type strain 11C) to ethanol (respectively Tr. 14, 51,133,144 and 364), butanol (respectively Tr. 92, 230 and 408) or significantly ( $P<0.05$ ) for both alcohols, Tr. 259.

**Phenotypic analysis of Tr. 259**

The effect of alcohol stress on temporal growth, spore germination and biomass production was investigated to confirm the hypo-tolerant phenotype of Tr. 259 observed during tertiary screening. The effect of increasing alcohol concentration on strain growth was firstly assessed (Figure S5). Growth of both Tr. 259 and 11C decreased with increasing ethanol ( $r\geq-0.967$ ,  $n=5$ ,  $P\leq0.02$ ) and butanol ( $r\geq-0.929$ ,  $n=8$ ,  $P\leq0.01$ ) concentration whereas 11C showed significantly better growth than Tr. 259 up to 6% (v<sup>-1</sup>) ethanol and 1% (v<sup>-1</sup>) butanol ( $P<0.05$ ) (Figure S5). At 4% (v<sup>-1</sup>) ethanol and 0.75% (v<sup>-1</sup>) butanol a substantial difference in growth ( $\geq 42\%$ ) was

recorded between Tr. 259 and 11C prompting the selection of these concentrations for further studies (Figure S5).

In the absence of alcohol, both Tr. 259 and 11C exhibited similar mycelial growth and spore germination rates (Figure 4A & Figure S6D). In contrast, 11C achieved better mycelial growth with either 4% ethanol or 0.75% butanol present in the medium, as compared to Tr. 259 (Figure 4B & 4C). Both 4% ethanol and 0.75% butanol severely impacted spore germination, however 11C achieved significantly greater spore germination than Tr. 259 ( $P\leq0.01$ ) (Figure S6B-C & S6E-F).

**Genomic analysis of Tr. 259**

Genome walking was conducted to determine the potential genomic region associated with Tr. 259’s hypo-tolerant phenotype (Figure S7). This resulted in the cloning of a 1031nt sequence external to the T-DNA right border (RB) (sequence analysis confirmed the amplicon was flanked by the universal

**Table 1.** Summary of tertiary screening of *Fusarium oxysporum* strain 11C transformants to investigate the level of altered alcohol tolerance across a selected sub-population of individuals ( $n=29$ ).

Treat (vv <sup>-1</sup> ) <sup>a</sup>	Population (%) <sup>b</sup>	Individuals (N) <sup>c</sup>	Hyper (N) <sup>d</sup>	Hypo (N) <sup>e</sup>
Control 0	66	19	7	12
Ethanol 2	17	5	4	1
Ethanol 4	31	9	8	1
Ethanol 6	31	9	0	9
Butanol 0.25	62	18	9	9
Butanol 0.5	38	11	3	8
Butanol 0.75	45	13	0	13

a. Positive control (no alcohol) or alcohol (ethanol or butanol) tested

b. Percentage (%) population tested ( $n=29$ ) showing hypo-hyper- phenotype relative to wild-type strain 11C ( $P<0.05$ )

c. Total number of individuals showing hypo-hyper- phenotype relative to wild-type strain 11C ( $P<0.05$ )

d. Total number of hyper-altered phenotypes ( $P<0.05$ )

e. Total number of hypo-altered phenotypes ( $P<0.05$ )

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and RB specific primer as expected; Figure S8A). Further analysis did not result in the recovery of full-length T-DNA in the identified coding region, indicating partial T-DNA integration. *In silico* analysis of the recovered coding sequence showed the highest homology in the ORF of a putative sugar transporter gene FOXG\_09625 in *F. oxysporum* f.sp.*lycospersici* (strain 4287) at both the protein (68% identity) and nucleotide (83% identity) level respectively (Figure S8B & S8C).

FOXG\_09625 showed homology (percent identity  $\geq 30\%$ ) with 30 different proteins all containing MFS and sugar transporter domains respectively, encoded within the genome of *F. oxysporum* f.sp.*lycospersici* (strain 4287) (FGCD). Five of these proteins were annotated as hexose transporters (FOXG\_02491, FOXG\_09722, FOXG\_05876, FOXG\_13263, FOXG\_014666) including FOXG\_09625, and the remaining as hypothetical proteins. Comparison of these proteins showed considerable sequence diversity indicating potentially diverse roles of these proteins in *F. oxysporum* (Figure S9A). In total, the genome of *F. oxysporum* f.sp. *lycospersici* (strain 4287) encodes 13 proteins annotated as hexose transporters (Figure S9B).

RT-PCR analysis was used to analyse FOXG\_09625 transcript levels in strain 11C compared to Tr. 259, following short-term (0.5-12 hours) cultivation of germinated spores (pre-cultured for 12 hours) in the absence or presence of alcohol (Figure 5). In the absence of alcohol, basal expression was low in both 11C and Tr. 259 ( $P>0.05$ ). (Figure 5A). Reduced transcript levels were recorded for both strains under ethanol stress compared to normal conditions from 0.5 to 12 hours with a significant difference between strains at 2 hours ( $P<0.01$ ) (Figure 5B). In the presence of butanol, a significant difference between Tr. 259 and 11C was recorded for all time points tested ( $P<0.001$ ) with transcript levels highest at 0.5 hours (Figure 5C).

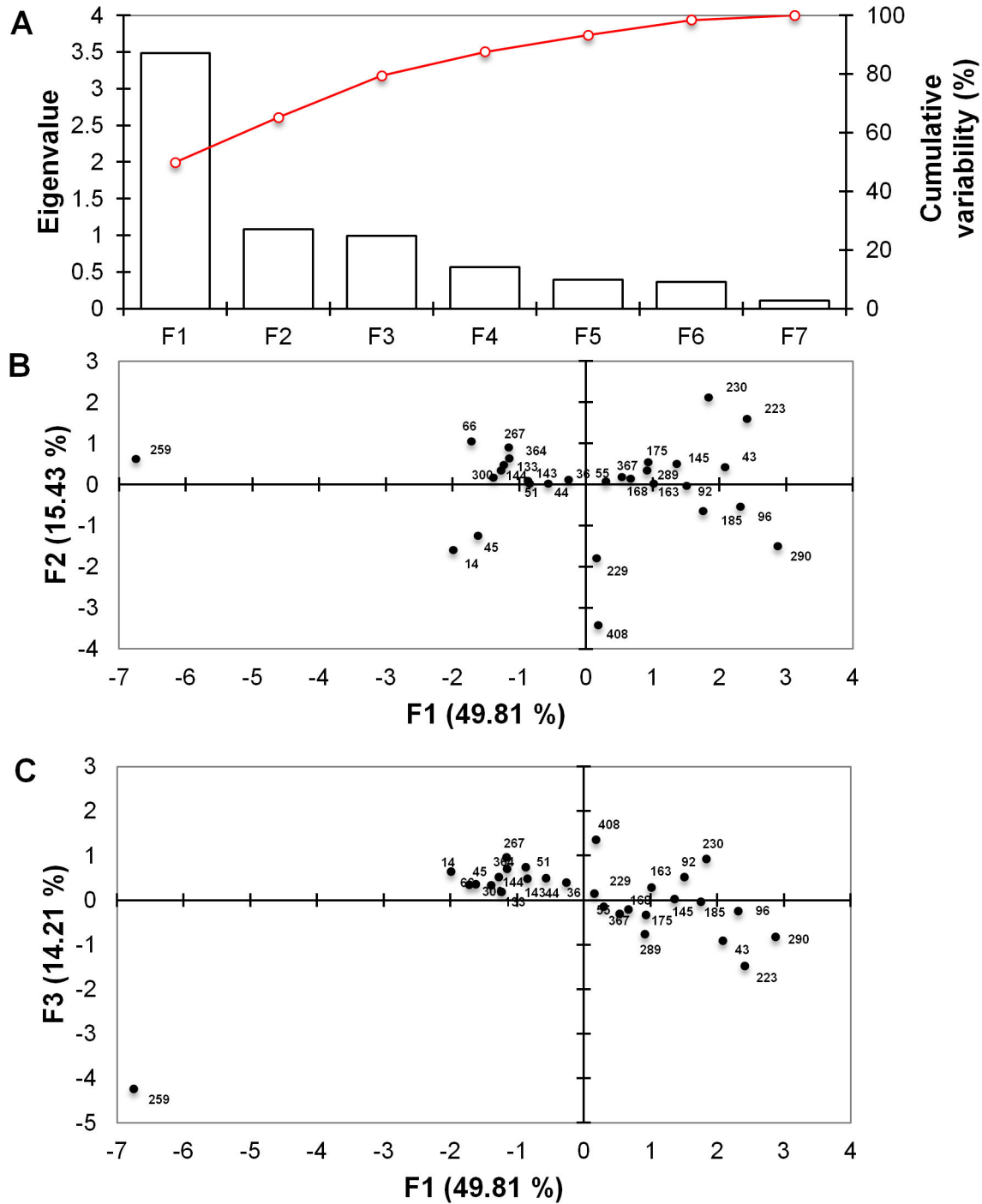
## Discussion

This study has shown that ATMT is an effective tool to generate phenotypic diversity in response to alcohol stress within *F. oxysporum* strain 11C. Although the generation of fungal insertion libraries is well documented, the integration of a three-tier alcohol screen into an ATMT platform is novel, and led to the generation and isolation of phenotypically diverse transformants from the library, indicating genotype-dependent variation and a functional ATMT system.

Ethanol tolerance of strain 11C ( $>6\%$  (vv<sup>-1</sup>) ethanol) was high compared to *Candida albicans*, which like *F. oxysporum* can ferment glucose to ethanol but can only tolerate 1% (vv<sup>-1</sup>) ethanol stress [39]. In contrast, yeast can tolerate high ethanol concentrations ( $>15\%$  (vv<sup>-1</sup>)), however at lower concentrations (4-6% (vv<sup>-1</sup>)) a 50% reduction in yeast's growth has been reported [40]. Unsurprisingly, strain 11C could not tolerate equivalent butanol concentrations as a barrier between 1-2% (vv<sup>-1</sup>) exists for most microbes with the exception of some *Lactobacillus* strains (3% (vv<sup>-1</sup>)) [41] and *Bacillus subtilis* (5% (vv<sup>-1</sup>)) [42].

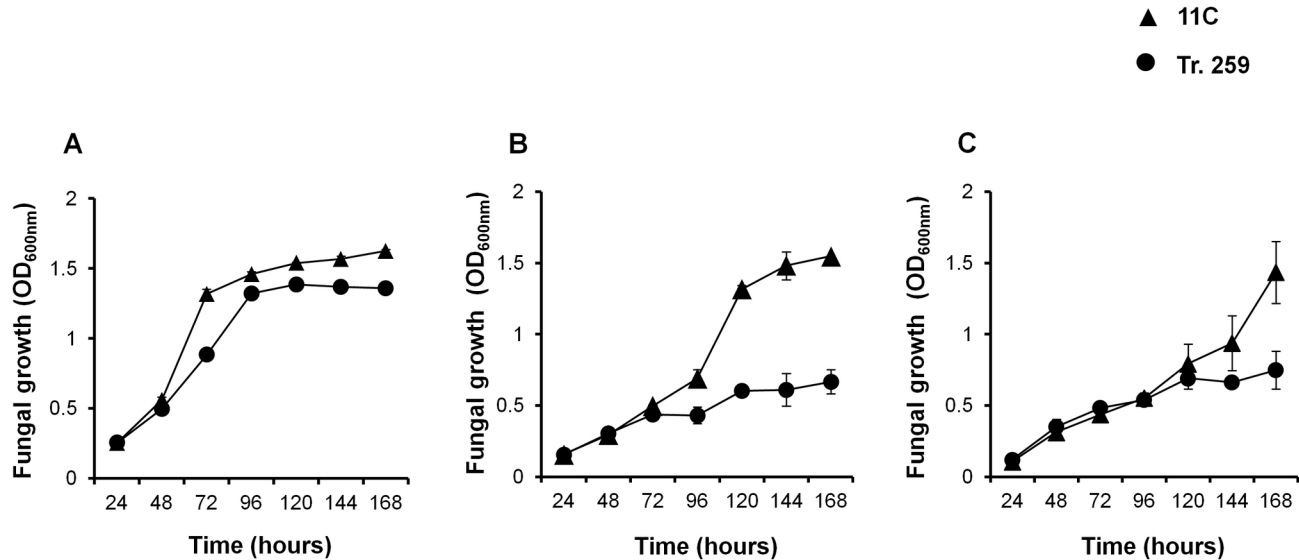
Large-scale screening of filamentous fungi can be challenging as manipulating fungal material (e.g. mycelium) can interfere with pipette-aided transfer between plates [43]. Fungal growth rates or hyphal growth patterns can vary greatly among isolates with some strains growing poorly in low volume space-limited microplates. Regardless, microplate-based assays facilitate the screening of large populations: a significant advantage over the labour-intensive regime of individual shake flasks or tubes [43]. Screening of *F. oxysporum* ATMT-derived transformants for alcohol tolerance has not yet been described. In other ascomycetous fungi notably the rice blast fungus *Magnaporthe grisea*, large-scale ATMT mutagenesis coupled with phenotypic plate-based screens has been used in pathogenicity related studies [44,45].

The three-tier screening system was designed to specifically discriminate either alcohol sensitive or tolerant phenotypes. Integration of the primary screen into the ATMT protocol provided a course but effective method to instigate a logistically feasible sorting of the library, thereby avoiding the laborious and time-consuming task of purifying thousands of putative transformants prior to screening. Non-purified, putative transformants underwent primary screening in contrast to the secondary screen, which used single-spored *hph*-PCR positive transformants. It cannot be ruled out that false negatives were lost to the analysis during these two stages. These primary rounds of screening were based on single reads to accelerate the phenotyping process and isolate transformants of interest for further analysis using an intensive confirmatory tertiary screen. This latter screen highlighted transformant Tr. 259 which underwent phenotypic and genomic analysis to confirm its hypo-alcohol tolerant phenotype and the ATMT-based disruption of a coding region showing homology to a putative sugar transporter (FOXG\_09625) in *F. oxysporum* f.sp. *lycospersici* (strain 4287). Southern hybridisation suggested a single insertion site within Tr. 259 with genome walking indicating partial T-DNA integration but it cannot be ruled out



**Figure 3. Principal component analysis (PCA) of tertiary screen.** Principal component analysis (PCA) was used to reduce the dimension of the tertiary dataset comprising of several interrelated variables (no alcohol, ethanol and butanol treatments on all 29 transformants) whilst retaining the variation present within the dataset. The dataset was transformed into variables (F1-F7) through component analysis using XLSTAT (Addinsoft) v2013.1 software. Scree plot of PCA analysis with the first three Eigenvalues (F1-F3) corresponding to the highest percentage of variance (**A**) Observation plot (built on Scree plot) in the factor space according to the first and second Principal components F1 and F2 (**B**) Observation plot (built on Scree plot) in the factor space according to the first and second Principal components F1 and F3 (**C**).

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**Figure 4. Temporal analysis of alcohol tolerance in *F. oxysporum*.** Fungal conidial inoculum was produced in Mung bean broth [32] and resuspended in minimal medium [37] at a concentration of  $10^6$  ml<sup>-1</sup>. A volume 100  $\mu$ l conidia ( $10^6$  ml<sup>-1</sup>) was added to microtiter (96-well; Sarstedt, Germany) plates with either no alcohol (A), ethanol (Ethyl absolute, Sigma, UK) at a concentration of 4% (v/v<sup>-1</sup>) (B) or butanol (*n*-Butanol Butyl alcohol, Sigma, UK) at a concentration of 0.75% (v/v<sup>-1</sup>) (C). Fungi were maintained at 25°C for 168 hours and growth (OD<sub>600nm</sub>) was measured every 24 hours. Fungal growth (OD<sub>600nm</sub>) was measured using a spectrophotometer (Spectra Max 340 PC 96-well plate reader, Molecular Devices, USA). Bars indicate SEM. Growth of Tr. 259 significantly different from 11C is highlighted with an asterisk (level of significance: \* $<0.05$ , \*\*  $<0.01$ ).

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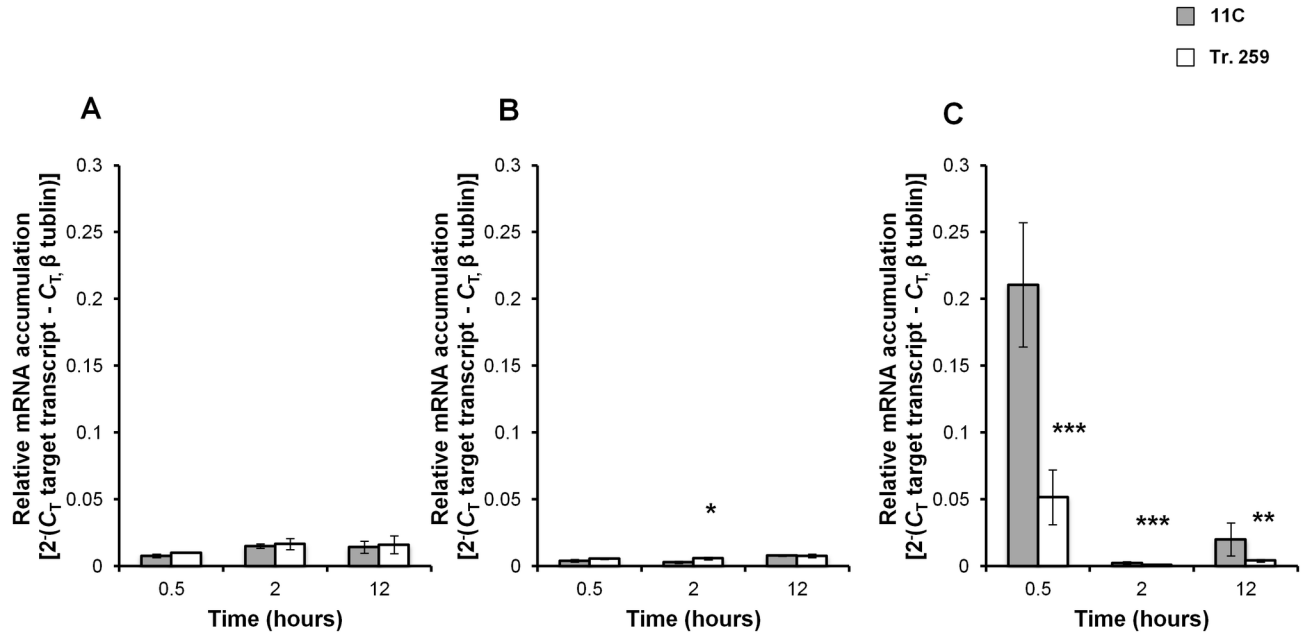
that additional T-DNA insertion events have also occurred through the genome of Tr. 259.

Eukaryotic transporters are best characterised in the yeast 20-member family of major hexose transporter genes (HXT1-4, HXT6-7, HXT8-17) which includes two sensor-coding genes (SNF3, RGT2) and the GAL2 gene [46]. In contrast to yeasts (*S. cerevisiae*, *Pichia stipitis*, *Schizosaccharomyces pombe*, *Kluyveromyces lactis*), few sugar transporters have been identified and characterised in filamentous fungi [47,48]. The genome of *F. oxysporum* f.sp. *lycospersici* (strain 4287) has at least 13 putative hexose transporters including FOXG\_09625, similar to *Aspergillus nidulans* reported to have 17 [47]. RT-PCR analysis indicated FOXG\_09625 was differentially expressed in Tr. 259 compared to WT during alcohol-induced stress and it would be interesting to investigate by Western blotting whether FOXG\_09625 is also differentially expressed at the protein level. In yeast, the closest protein homologue of FOXG\_09625 is HXT13, which is induced in the presence of non-fermentable carbon sources (e.g. alcohols), low sugar levels and repressed under high sugar concentrations [49]. It was hypothesised that FOXG\_09625 would be poorly expressed under normal conditions and highly expressed in the presence of ethanol or butanol. Whilst this hypothesis was not observed under ethanol stress, results recorded for butanol were as expected. For yeast, the association of hexose transport and glycolysis associated genes with higher expression levels under 4% (v/v<sup>-1</sup>) ethanol stress led to the proposal that the cell enters a pseudo-starvation state during

ethanol stress, whereby sugar-based nutrients are no longer accessible to the cell despite being present in the culture medium [50,51]. In this study, the highest FOXG\_09625 transcript levels were recorded under butanol stress (0.5 hours) suggesting that cells may have immediately entered a pseudo-starvation state in response to the increased toxicity of butanol, possibly inhibiting sugar up-take. In yeast, increased expression of hexose transporters and glycolysis genes improve both rapid production and consumption of ethanol [52].

A previous study [53] identified two sugar transporters overexpressed in strain 11C during CBP. In the analysis of additional transformants we would expect to identify an overlap with the SSH library for genes holding dual function i.e. relating to both production and tolerance. However, ethanol stress studies to date have identified genes belonging to diverse functional categories hence the identification of novel genes could also be anticipated. Interestingly, FOXG\_09625 showed 29% protein homology to the SSH – identified hexose transporter and no homology to the high affinity glucose transporter indicating potentially different roles for all three genes in either tolerance or production. Yet, a recent study [54] has indicated FOXG\_09625 is associated (directly/indirectly) with ethanol production since mutants overexpressing a hexose transporter resulted in increased ethanol yield coupled with compensatory changes in the expression of other transporters notably the up-regulation of FOXG\_09625. Investigations, including overexpression/knockout studies of FOXG\_09625, while not possible due to a limitation in





**Figure 5. RT-PCR analysis of putative hexose transporter under alcohol stress.** Temporal analysis of putative hexose transporter transcript (FOXG\_09625) during shake flask growth of *Fusarium oxysporum* strain 11C and transformant Tr. 259 under normal conditions (A), ethanol stress (B) and butanol stress (C). *F. oxysporum* (2ml 10<sup>6</sup> ml<sup>-1</sup>) was aerobically cultured in Erlenmeyer flasks (100 ml) in 48 ml of minimal medium [37] for 12 hours shaking at 150 rpm at 25°C. At 12 hours, flasks were amended with 2ml of water, ethanol or butanol to a final concentration of 0%, 4% and 0.75% (v/v<sup>-1</sup>), respectively. RT-PCR was conducted on RNA samples harvested at 0.5, 2, 12 hours respectively post – inoculation. Putative HXT accumulation was quantified relative to the housekeeping gene β-tubulin (FOXG\_06228) [38]. Results are based on three experiments (no alcohol, ethanol) or two experiments (butanol), each with three replicates per time point per strain tested. RT-PCR was conducted twice on each experiment. Bars indicate SEM. Tr. 259 FOXG\_09625 mRNA accumulation significantly different from strain 11C is highlighted with an asterisk (level of significance: \* < 0.05, \*\* ≤ 0.01, \*\*\* < 0.001).

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resources for this study will be required to fully elucidate the complexity of this gene interaction with alcohol and determine the degree to which over-expression/silencing of FOXG\_09625 could improve consolidated bioprocessing yield.

Filamentous fungal transporters appear to hold diverse functional roles notably in plant-fungi symbiosis [55-57] pathogenicity [58,59], and ethanol production and tolerance [48,50,60-62]. In regards to tolerance, transcriptional studies have identified alcohol responsive genes associated with sugar transport [50,63,64]. This is not surprising since the rate of sugar transport is limited by accumulation of alcohol in the fermentation broth, which can become a significant stress during fermentation resulting in a 'survival versus production' conundrum for the producing microbe [65]. While major advances have been made investigating alcohol tolerance in model systems (yeast, *E. coli*), a knowledge deficit exists for filamentous fungi [66-68]. A better understanding of the molecular basis of alcohol stress and tolerance in such fungi is important if enhanced tolerance and production is to be achieved. Previous work [54] coupled with this study point towards sugar transporters as key targets for improving fungal-enabled CBP.

In conclusion, the work completed in this study represents a first step in the investigation of alcohol tolerance in *F. oxysporum* via ATMT, which has led to the generation of a collection of transformants, which are now available to the research community for future studies. This is all the more relevant in light of the recent advances and application of RNA sequencing to elucidate complex biological processes at the transcript level in an unlimited array of organisms [69]. On the basis of the results presented here, ATMT can be exploited as a tool to generate diverse alcohol tolerant phenotypes in the CBP agent *F. oxysporum*.

## Supporting Information

**Figure S1. Microplate (96-well; Sarstedt, Germany) layout for three-tier screening.** *Fusarium oxysporum* wild-type strain 11C (WT) and putative transformants were inoculated into columns (1-12) and screened across five treatments: T1; no ethanol, T2; hygryomycinB (60 μg ml<sup>-1</sup>; Sigma, UK), T3; 0.5% (v/v<sup>-1</sup>) ethanol (ethyl alcohol; Sigma, UK), T4; 6% (v/v<sup>-1</sup>) ethanol (ethyl alcohol; Sigma, UK), T5; 10% (v/v<sup>-1</sup>) ethanol with treatments positioned in rows (A-F) (Primary-tier) (A) or into rows (A-H) and then transferred and

inoculated into corresponding rows in a second microplate and screened across six treatments: T1; no ethanol, T2; 6% (v/v<sup>-1</sup>) ethanol (ethyl alcohol; Sigma, UK), T3; 7% (v/v<sup>-1</sup>) ethanol T4; 8% (v/v<sup>-1</sup>) ethanol, T5; 9% (v/v<sup>-1</sup>) ethanol, T6; 10% (v/v<sup>-1</sup>) ethanol (Second-tier) (B) or into columns (1-12) with eight replicates per strain tested (column A-H) and screened across a single treatment including media control (MC) per plate: either no alcohol, 2% (v/v<sup>-1</sup>) ethanol (ethyl alcohol; Sigma, UK), 2% (v/v<sup>-1</sup>) ethanol, 4% (v/v<sup>-1</sup>) ethanol, 6% (v/v<sup>-1</sup>) ethanol or 0.25% (v/v<sup>-1</sup>) butanol, 0.5% (v/v<sup>-1</sup>) butanol and 0.75% (v/v<sup>-1</sup>) butanol (C). Grey shading indicates microplate well not used during experiment. (TIF)

**Figure S2. Experimental design flow chart to investigate alcohol-induced stress in *Fusarium oxysporum* strain 11C and transformant Tr. 259.**

(TIF)

**Figure S3. Generating and screening *Fusarium oxysporum* strain 11C transformants.** Inoculation of *Fusarium oxysporum* strain 11C into Mung bean broth [32] for 5 days at 25°C and collection of conidia by filtration through sterile cheesecloth, washing twice with sterile distilled water and adjusting spore concentration to 10<sup>4</sup> conidia per ml (A) Inoculation of *Agrobacterium tumefaciens* strain AGL-1 to minimal medium (MM) [34] supplemented with Kanamycin (50 µg ml<sup>-1</sup>) for 2 days at 28°C (OD<sub>600nm</sub> 0.4-0.6) followed by dilution to OD<sub>600nm</sub> of 0.15 in induction medium (IM) [34] supplemented with 200 µM acetosyringone (AS) and incubation for 6 hours at 28°C (B) Co-cultivation of an equal volume of bacterial and fungal cells for 30 minutes at 28°C in liquid co-cultivation medium [34] followed by spreading 100 µl mix onto a UV-sterilised cellulose filter membrane placed on solid co-cultivation medium on large Petri plates (150x20mm) for 2 days at 25°C where if transformation is successful, T-DNA is transferred from AGL-1 to *F. oxysporum* strain 11C and randomly integration into the fungal genome results (C) Transfer of cellulose filter membrane to modified selection medium (SM) on large Petri plates (150x20mm) supplemented with 60 hygromycinB for 7-9 days at 25°C followed by isolation of hygromycinB (60 µg ml<sup>-1</sup>) resistant putatively transformed colonies into microtiter (96-well) plates with minimal medium [37] supplemented with hygromycinB (60 µg ml<sup>-1</sup>) for 3 days at 25°C (D) Transfer of putative transformants to microtiter (96-well) plate with minimal medium supplemented with ethanol and butanol selection for primary alcohol tolerance screening followed by purification and PCR prior to two additional rounds of screening (2° and 3°) and selection of candidates for future analysis (E).

(TIF)

**Figure S4. Molecular analysis of *Fusarium oxysporum* strain 11C transformants.** PCR was used to determine if fungal transformant genomic DNA (gDNA) extracts contained *hph* gene present in pSK1019. Lanes 2-6 represent gDNA from transformants, 7; PC – positive control (plasmid DNA pSK1019), 8; NTC – no template control (water), 9; WT - gDNA from wild-type strain 11C. Arrow indicates PCR product size

(576 bp) based on 100bp ladder (NEB, UK) (Lane 1) (A). Southern blot analysis was used to confirm transgene integration in *Eco*R1 digested fungal gDNA and to determine copy number, using a 741nt fragment of the hygromycin phosphotransferase gene (*hph*) as a probe. Lanes 1-11 represent DNA digested with *Eco*R1; 1, gDNA from wild type fungal strain 11C, 2-10, gDNA from strain 11C transformants Tr. 144, Tr. 175, Tr. 230, Tr. 51, Tr. 43, Tr. 133, Tr. 145, Tr. 55 and Tr. 168, 11, plasmid DNA from pSK1019. Red dots indicate bands and arrows indicate molecular size (Kb) based on 1 Kb DNA ladder (Solis BioDyne, Estonia) (B). (TIF)

**Figure S5. Effect of increasing alcohol concentration on *Fusarium oxysporum* growth.** Fungal conidial inoculum was produced in Mung bean broth [32] and resuspended in minimal medium [37] at a concentration of 10<sup>6</sup> ml<sup>-1</sup>. A volume 100 µl conidia (10<sup>6</sup> ml<sup>-1</sup>) was added to microtiter (96-well; Sarstedt, Germany) plates supplemented with either ethanol (Ethyl Absolute, Sigma, UK) (A) or butanol (*n*-Butanol Butyl alcohol, Sigma, UK) (B). Fungi were maintained at 25°C for 7 days. Fungal growth (OD<sub>600nm</sub>) was measured using a spectrophotometer (Spectra Max 340 PC 96-well plate reader, Molecular Devices, USA). Bars indicate SEM. Growth of Tr. 259 significantly different from 11C is highlighted with an asterisk (level of significance: \* < 0.05, \*\* < 0.01).

(TIF)

**Figure S6. Effect of alcohol stress on spore germination and biomass production in *F. oxysporum*.** Spore germination of *F. oxysporum* strain 11C and transformant Tr. 259 in the presence of no alcohol (A) 4% (v/v<sup>-1</sup>) ethanol (B) and 0.75% (v/v<sup>-1</sup>) butanol (C); fungal conidial inoculum was produced in Mung bean broth [32] and resuspended in minimal medium [37] at a concentration of 10<sup>6</sup> ml<sup>-1</sup>. Fungi were maintained at 25°C for 24 hours shaking at 150 rpm and percentage germination was measured every 2 hours. Bars indicate SEM. Biomass production of *F. oxysporum* strain 11C and transformant Tr. 259 post – treatment with either; no alcohol (D) 4% (v/v<sup>-1</sup>) ethanol (E) or 0.75% (v/v<sup>-1</sup>) butanol (F). Minimal medium (48 ml) was inoculated with fungal conidia (2ml of 10<sup>6</sup> ml<sup>-1</sup>) and flasks were incubated at 25°C, 150 rpm. After 12 hours, flasks were amended with 2ml of water, ethanol or butanol to a final concentration of 0, 4 and 0.75% (v/v<sup>-1</sup>), respectively. Fungal material was collected at 72, 120 and 168 hours post treatment. Bars indicate SEM. Spore germination or biomass production of Tr. 259 significantly different from 11C is highlighted with an asterisk (level of significance: \* < 0.05, \*\* < 0.01).

(TIF)

**Figure S7. Genome walking in *F. oxysporum* 11C transformant Tr. 259.** Genome walking analysis using APA™ Gold Genome Walking Kit (Bio S&T, Canada) to walk the right border (RB) of pSK1019. Lanes represent: 1 - 4; degenerate random tagging (DRT) primers A, B, C and D respectively with target specific primer (GSP) for RB of pSK1019 amplified with 500 ng of fungal genomic DNA from *Fusarium oxysporum*

strain 11C transformant Tr. 259. Arrow indicates PCR fragment sizes (bp) on Hyperladder™ III (Bioline, UK) (Lane 5). Red dots indicate gel excised bands for subsequent cloning (A). Digestion of plasmid DNA from pGEMT® easy (Promega, UK) cloned PCR products to verify correct fragment insert size. Plasmid DNA (500 ng) was digested in 20 µl reaction and incubated at 37°C for 2 hours. Lanes represent: 1, Hyperladder™ III (Bioline, UK), 2 - 5, bands excised from Figure S7A highlighted by red dots; 6, blue colony (negative control) (B). PCR analysis of cloned PCR products to verify presence of target specific primer (GSP) used in genome walking prior to sequencing. Lanes represent: Lanes represent: 1, Hyperladder™ III (Bioline, UK), 2 - 5, bands excised from Figure S7A highlighted by red dots; 6, blue colony (negative control); 7, no plasmid DNA (negative control) (C). (TIF)

**Figure S8. *In silico* analysis of identified coding sequence obtained via genome walking in Tr. 259.** Sequence analysis of cloned PCR product obtained via genome walking within *Fusarium oxysporum* strain 11C transformant Tr. 259. Walking was from the right border (RB) of the transformation vector pSK1019 using the APA™ Gold Genome Walking Kit (Bio S&T, Canada), a universal kit primer (UAP-2) (red) and the gene-specific (GSP) primer (blue) (3'-5') (A). Identified coding sequence was analysed against the *F. oxysporum* f.sp.*lycospersici* (strain 4287) genome (FCGD; ([http://www.broadinstitute.org/annotation/genome/fusarium\\_group](http://www.broadinstitute.org/annotation/genome/fusarium_group))) using Blastn (B) and Blastx (C). (TIF)

**Figure S9. Phylogenetic analysis of putative sugar transporters in *Fusarium oxysporum*.** Homology (≥30%) of FOXG\_09625 to putative sugar transporters in *F. oxysporum* 4287 (A). Comparison of putative hexose transporters in *F. oxysporum* 4287 compared to *Saccharomyces cerevisiae* and putative sugar transporters showing (≥30%) homology to

FOXG\_09625 in *Aspergillus nidulans*, *Trichoderma reesei*, *Mucor circinelloides*, *Neurospora crassa* (B). Amino acid sequences were obtained from FCGD ([http://www.broadinstitute.org/annotation/genome/fusarium\\_group](http://www.broadinstitute.org/annotation/genome/fusarium_group)) SGD (<http://www.yeastgenome.org>) and NCBI (<http://www.ncbi.nlm.nih.gov>). All protein sequences showed sugar transporter domains and were aligned using ClustalW. A phylogram was constructed using the Neighbor-Joining method within Phylogeny.fr software. Numbers on internal nodes represent bootstrap values (100 re-sampling). Arrow indicates putative hexose transporter (FOXG\_09625) used in RT-PCR analysis. Asterisk indicates transcripts annotated as hexose transporters in *F. oxysporum* 4287 (FCGD). (TIF)

**Table S1. Primers used in this study for genome walking and real-time PCR.** (DOC)

**Methods S1. Expanded description of methods.** (DOCX)

**References S1.** (DOCX)

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## Author Contributions

Conceived and designed the experiments: RH FD EM. Performed the experiments: RH. Analyzed the data: RH FD EM. Contributed reagents/materials/analysis tools: RH FD EM. Wrote the manuscript: RH FD EM.

## References

1. Brodeur G, Yau E, Badal K, Collier J, Ramachandran KB et al. (2011) Chemical and Physicochemical Pretreatment of Lignocellulosic Biomass: A Review. *Enzyme Res* 2011: 787532. PubMed: 21687609
2. Bhatia L, Johri S, Ahmad R (2012) A economic and ecological perspective of ethanol production from renewable agro waste: a review. *AMB Express* 2: 65. doi:10.1186/2191-0855-2-65. PubMed: 23217124.
3. Elkins JG, Raman B, Keller M (2010) Engineered microbial systems for enhanced conversion of lignocellulosic biomass. *Curr Opin Biotechnol* 21(5): 657-662. doi:10.1016/j.copbio.2010.05.008. PubMed: 20579868.
4. La Grange DC, den Haan R, van Zyl WH (2010) Engineering cellulolytic ability into bioprocessing organisms. *Appl Microbiol Biotechnol* 87(4): 1195-1208. doi:10.1007/s00253-010-2660-x. PubMed: 20508932.
5. Sakuragi H, Kuroda K, Ueda M (2011) Molecular Breeding of Advanced Microorganisms for Biofuel Production. *J Biomed Biotechnol*. Volume 2011: 416931. PubMed: 21318120.
6. Amore A, Faraco V (2012) Potential of fungi as category I Consolidated BioProcessing organisms for cellulosic ethanol production. *Renewable Sustain Energ Rev* 16 (5): 3286-3301. doi:10.1016/j.rser.2012.02.050.
7. Olson DG, McBride JE, Shaw AJ, Lynd LR (2012). ecent Progress Consolidated Bioprocessing 12 (3): 396-405
8. Lübbehüsen TL, Nielsen J, McIntyre M (2004) Aerobic and anaerobic ethanol production by *Mucor circinelloides* during submerged growth. *Appl Microbiol Biotechnol* 63: 543-548. doi:10.1007/s00253-003-1394-4. PubMed: 12879305.
9. Dashtban M, Schraft H, Qin W (2009) Fungal Bioconversion of Lignocellulosic Residues; Opportunities & Perspectives. *Int J Bios Sci* 5(6): 578-595. PubMed: 19774110.
10. Fan Z, Wu W, Hildebrand A, Kasuga T, Zhang R et al. (2012) A Novel Biochemical Route for Fuels and Chemicals Production from Cellulosic Biomass. *PLOS ONE* 7 (2).
11. Lakshman DK, Pandey R, Kamo K, Bauchan G, Mitra A (2012) Genetic transformation of *Fusarium oxysporum* f.sp. gladioli with *Agrobacterium* to study pathogenesis in Gladiolus. *Eur J Plant Pathol* 133 (3): 729-738. doi:10.1007/s10658-012-9953-0.
12. Christakopoulos PK, Koullas DP, Kekos D, Koukios EG, Macris BJ (1991) Direct conversion of straw to ethanol by *Fusarium oxysporum*: effect of cellulose crystallinity. *Enzyme Microb Technol*. 13(3): 272-274. doi:10.1016/0141-0229(91)90141-V.
13. Ali SS, Khan M, Fagan B, Mullins E, Doohan FM (2012) Exploiting the inter-strain divergence of *Fusarium oxysporum* for microbial bioprocessing of lignocellulose to bioethanol. *AMB Express*. 2(1): 1-9. doi:10.1186/2191-0855-2-1. PubMed: 22214346.
14. Xiros C, Christakopoulos P (2009) Enhanced ethanol production from brewer's spent grain by a *Fusarium oxysporum* consolidated system. *Biotechnol Biofuels* 2(1): 4. doi:10.1186/1754-6834-2-4. PubMed: 19208239.
15. Hossain SM, Anantharaman N, Das M (2012) Bioethanol fermentation from untreated and pretreated lignocellulosic wheat straw using fungi *Fusarium oxysporum*. *Indian J Chem Technol* 19: 63-70.

16. Favaro L, Jooste T, Basaglia M, Rose SH, Saayman M et al. (2013) Designing industrial yeasts for the consolidated bioprocessing of starchy biomass to ethanol. *Bioengineered* 4(2): 97-10. doi:10.4161/bioe.22268. PubMed: 22989992.
17. Xiros C, Vafiadi M, Paschos T, Christakopoulos P (2011) Toxicity tolerance of *Fusarium oxysporum* towards inhibitory compounds formed during pretreatment of lignocellulosic materials. *J Chem Technol Biotechnol* 86(2): 223-230. doi:10.1002/jctb.2499.
18. Panagioutou G, Pachidou F, Petroutsos D, Olsson L, Christakopoulos P (2008) Fermentation characteristics of *Fusarium oxysporum* grown on acetate. *Bioresour Technol* 99(15): 7397-7401. doi:10.1016/j.biortech.2008.01.017. PubMed: 18304808.
19. Singh A, Kumar PKR (1991) *Fusarium oxysporum*: Status in Bioethanol Production. *Crit Rev Biotechnol* 11(2): 129-147. doi:10.3109/07388559109040619. PubMed: 1913845.
20. Brown SD, Guss AM, Karpinets TV, Parks JM, Smolin N et al. (2011) Mutant alcohol dehydrogenase leads to improved ethanol tolerance in *Clostridium thermocellum*. *Proc Natl Acad Sci USA*, 108: 2011. PubMed: 21825121.
21. Pfromm PH, Amanor-Boadu V, Nelson R, Vadlani P, Madl P (2010) Bio-butanol vs. bio-ethanol: A technical and economic assessment for corn and switchgrass fermented by yeast or *Clostridium acetobutylicum*. *Biomass Bioenerg* 34: 515-524. doi:10.1016/j.biombioe.2009.12.017.
22. Atsumi S, Cann AF, Connor MR, Shen CR, Smith KM et al. (2008) Metabolic engineering of *Escherichia coli* for 1-butanol production. *Metab Eng* 10(6): 305-311. doi:10.1016/j.ymben.2007.08.003. PubMed: 17942358.
23. Knoshaug E, Zhang M (2009) Butanol Tolerance in a Selection of Microorganisms. *Appl Biochem Biotechnol*. 153(1): 13-20.
24. Rühl J, Schmid A, Blank LM (2009) Selected *Pseudomonas putida* Strains Able to Grow in the Presence of High Butanol Concentrations. *Appl Environ Microbiol* 75(13): 4653-4656. doi:10.1128/AEM.00225-09. PubMed: 19411419.
25. Dunlop MJ, Dossani ZY, Szmídt HL, Chu HC, Lee TS et al. (2011) Engineering microbial biofuel tolerance and export using efflux pumps. *Mol Syst Biol* 10(7): 487. PubMed: 21556065.
26. Zingaro KA, Papoutsakis ET (2012) Toward a Semisynthetic Stress Response System To Engineer Microbial Solvent Tolerance. *mBio* 3(5): ([MedlinePgn:]) PubMed: 23033472.
27. Jia K, Zhang Y, Li Y (2010) Systematic engineering of microorganisms to improve alcohol tolerance. *Eng Life Sci* 10(5): 422-429. doi:10.1002/elsc.201000076.
28. Maruthachalam K, Klosterman SJ, Kang S, Hayes RJ, Subbarao KV (2011) Identification of pathogenicity-related genes in the vascular wilt fungus *Verticillium dahliae* by *Agrobacterium tumefaciens*-mediated T-DNA insertional mutagenesis. *Mol Biotechnol* 49(3): 209-221. doi:10.1007/s12033-011-9392-8. PubMed: 21424547.
29. de Groot MJ, Bundock P, Hooykaas PJ, Beijersbergen AG (1998) *Agrobacterium*-mediated transformation of filamentous fungi. *Nat Biotechnol* 16: 839-842. doi:10.1038/nbt0998-839. PubMed: 9743116.
30. Michielse CB, Hooykaas PJJ, van den Hondel Camij, Ram AFJ (2005) *Agrobacterium* -mediated transformation as a tool for functional genomics in fungi. *Curr Genet* 48(1): 1-17. doi:10.1007/s00294-005-0578-0. PubMed: 15889258.
31. Ma LJ, van der Does C, Borkovich KA, Coleman JJ, Daboussi MJ et al. (2010) Comparative genomics reveals mobile pathogenicity chromosomes in *Fusarium*. *Nature* 464(7287): 367-373. doi:10.1038/nature08850. PubMed: 20237561.
32. Brennan J, Leonard G, Cooke B, Doohan F (2005) Effect of temperature on head blight of wheat caused by *Fusarium culmorum* and *F. graminearum*. *Plant Pathol*. 54: 156-160. doi:10.1111/j.1365-3059.2005.01157.x.
33. Sambrook J, Russell DW (2011) *Molecular Cloning: A Laboratory Manual*. Cold Spring Harbor Laboratory Press.
34. Mullins ED, Chen X, Romaine P, Raina R, Geiser DM et al. (2001) *Agrobacterium*-Mediated Transformation of *Fusarium oxysporum*: An Efficient Tool for Insertional Mutagenesis and Gene Transfer. *Phytopathology*. 91(2): 173-180. doi:10.1094/PHYTO.2001.91.2.173. PubMed: 18944391.
35. Liu D, Coloe S, Baird R, John P (2000) Rapid Mini-Preparation of Fungal DNA for PCR. *J Clin Microbiol* 38(1): 471pp. PubMed: 10681211.
36. Edel V, Steinberg C, Gautheron N, Alabouvette C (2000) Ribosomal DNA-targeted oligonucleotide probe and PCR assay specific for *Fusarium oxysporum*. *Mycol Res*. 104: 518-526. doi:10.1017/S0953756299001896.
37. Mishra C, Keskar S, Rao M (1984) Production and Properties of Extracellular Endoxylanase from *Neurospora crassa*. *Appl Environ Microbiol* 48(1): 224-228. PubMed: 16346591.
38. Livak KJ, Schmittgen TD (2001) Analysis of relative gene expression data using real-time quantitative PCR and the 2<sup>-</sup>[Delta][Delta] CT method. *Methods* 25(4): 402-408. doi:10.1006/meth.2001.1262. PubMed: 11846609.
39. Zeuthen ML, Aniebo CM, Howard DH (1988) Ethanol Tolerance and the Induction of Stress Proteins by Ethanol in *Candida albicans*. *J Gen Microbiol* 134(5): 1375-1384. PubMed: 3058867.
40. Snowdon C, Schierholtz R, Poliszczuk P, Hughes S, van der Merwe G (2009) ETP1/YHL010c is a novel gene needed for the adaptation of *Saccharomyces cerevisiae* to ethanol. *FEMS\_Yeast\_Res* 9: 372-380. PubMed: 19416103.
41. Liu S, Qureshi N (2009) How microbes tolerate ethanol and butanol. *New Biotechnol* 26(3-4): 117-121. PubMed: 19577017.
42. Kataoka N, Tajima T, Kato J, Rachadech W, Vangnai AS (2011) Development of butanol-tolerant *Bacillus subtilis* strain GRSW2-B1 as a potential bioproduction host. *AMB Expr* 1(1): 10. doi:10.1186/2191-0855-1-10. PubMed: 21906347.
43. Bills GF, Platas G, Fillola A, Jiménez MR, Collado J et al. (2007) Enhancement of antibiotic and secondary metabolite detection from filamentous fungi by growth on nutritional arrays. *J Appl Microbiol*. 10(4): 1644-1658. PubMed: 18298532.
44. Betts MF, Tucker SL, Galadima N, Meng Y, Patel G et al. (2007) Development of a high throughput transformation system for insertional mutagenesis in *Magnaporthe oryzae*. *Fungal Genet Biol* 44(10): 1035-1049. doi:10.1016/j.fgb.2007.05.001. PubMed: 17600737.
45. Jeon J, Park SY, Chi MH, Choi J, Park J et al. (2007) Genome-wide functional analysis of pathogenicity genes in the rice blast fungus. *Nat Genet* 39(4): 561-565. doi:10.1038/ng2002. PubMed: 17353894.
46. Horák J (2013) Regulations of sugar transporters: insights from yeast. *Curr Genet* 59(1-2): 1-31. doi:10.1007/s00294-013-0388-8. PubMed: 23455612.
47. Wei H, Vienken K, Weber R, Bunting S, Requena N et al. (2003) A putative high affinity hexose transporter, *hxtA*, of *Aspergillus nidulans* is induced in vegetative hyphae upon starvation and in ascogenous hyphae during cleistothecium formation. *Fungal Genet Biol* 41: 148-156.
48. Fernandes S, Murray P (2010) Metabolic engineering for improved microbial pentose fermentation. *Bioeng Bugs*. 1(6): 424-428. doi:10.4161/bbug.1.6.12724. PubMed: 21468211.
49. Greatrix BW, van Vuuren HJ (2006) Expression of the HXT13, HXT15 and HXT17 genes in *Saccharomyces cerevisiae* and stabilization of the HXT1 gene transcript by sugar-induced osmotic stress. *Curr Genet* 49(4): 205-217. doi:10.1007/s00294-005-0046-x. PubMed: 16397765.
50. Chandler M, Stanley GA, Rogers P, Chambers P (2004) A genomic approach to defining the ethanol stress response in the yeast *Saccharomyces cerevisiae*. *Ann Microbiol* 54(4): 427-454.
51. Stanley D, Bandara A, Fraser S, Chambers PJ, Stanley GA (2010) The ethanol stress response and ethanol tolerance of *Saccharomyces cerevisiae*. *J Appl Microbiol* 109(1): 13-24. PubMed: 20070446.
52. Anderson MJ, Barker SL, Boone C, Measday V (2012) Identification of RCN1 and RSA3 as ethanol-tolerant genes in *Saccharomyces cerevisiae* using a high copy barcoded library. *FEMS Yeast Res* 12(1): 48-60. doi:10.1111/j.1567-1364.2011.00762.x. PubMed: 22093065.
53. Ali SS, Khan M, Mullins E, Doohan F (2013) Identification of *Fusarium oxysporum* Genes Associated with Lignocellulose Bioconversion Competency. *Bioenergy Research*.
54. Ali SS, Nugent B, Mullins E, Doohan FM (2013) Insights from the Fungus *Fusarium oxysporum* Point to High Affinity Glucose Transporters as Targets for Enhancing Ethanol Production from Lignocellulose. *PLOS ONE* 8(1): e54701. PubMed: 23382943.
55. López MF, Dietz S, Grunze N, Bloschies J, Weiß M et al. (2008) The sugar porter gene family of *Laccaria bicolor*: function in ectomycorrhizal symbiosis and soil-growing hyphae. *New Phytol* 180: 365-378. doi:10.1111/j.1469-8137.2008.02539.x. PubMed: 18627493.
56. Eckhardt N (2011) A Symbiotic Sugar Transporter in the Arbuscular Mycorrhizal Fungus *Glomus* sp. *Plant Cell* 23(10): 3561. doi:10.1105/tpc.111.231010. PubMed: 22003078.
57. Helber N, Wipfel K, Sauer N, Schaarschmidt S, Hause B et al. (2011) A Versatile Monosaccharide Transporter That Operates in the Arbuscular Mycorrhizal Fungus *Glomus* sp Is Crucial for the Symbiotic Relationship with Plants. *Plant Cell* 23: 3812-3823. doi:10.1105/tpc.111.089813. PubMed: 21972259.
58. Talbot NJ (2010) Living the Sweet Life: How Does a Plant Pathogenic Fungus Acquire Sugar from Plants? *PLOS Biol* 8(2): e1000308. PubMed: 20161721.

59. Wahl R, Wippel K, Goos S, Kamper J, Sauer N (2010) A Novel High-Affinity Sucrose Transporter Is Required for Virulence of the Plant Pathogen *Ustilago maydis*. *PLOS Biol* 8(2).
60. Olofsson K, Bertilsson M, Lidén G (2008) A short review on SSF – an interesting process option for ethanol production from lignocellulosic feedstocks. *Biotechnol Biofuels* 1: 7. doi:10.1186/1754-6834-1-7. PubMed: 18471273.
61. Patrascu E, Rapeanu G, Hopulele T (2009) Current approaches to efficient biotechnological production of ethanol. *Innovative Romanian Food Biotechnol* Vol. 4.
62. Xu Q, Arjun Singh A, Himmel ME (2009) Perspectives and new directions for the production of bioethanol using consolidated bioprocessing of lignocellulose. *Curr Opin Biotechnol* 20: 364-371. doi: 10.1016/j.copbio.2009.05.006. PubMed: 19520566.
63. Alexandre H, Ansanay-Galeote V, Dequin S, Blondin B (2001) Global gene expression during short-term ethanol stress in *Saccharomyces cerevisiae*. *FEBS Lett* 498(1): 98-103. doi:10.1016/S0014-5793(01)02503-0. PubMed: 11389906.
64. Reyes LH, Almario MP, Kao KC (2011) Genomic Library Screens for Genes Involved in n-Butanol Tolerance in *Escherichia coli*. *PLOS ONE* 6(3): e17678. PubMed: 21408113.
65. Yarris L (2011) Striking the Right Balance: JBEI Researchers Counteract Biofuel Toxicity in Microbes (Press Release). Available: <http://newscenter.lbl.gov/news-releases/2011/05/11/efflux-pumps-for-biofuels/>.
66. Takó M, Farkas E, Lung S, Krisch J, Vágvölgyi C et al. (2010) Identification of acid- and thermotolerant extracellular  $\beta$ -glucosidase activities in Zygomycetes fungi. *Acta Biol Hung* 61(1): 101-110. doi: 10.1556/ABiol.61.2010.1.10. PubMed: 20194103.
67. Asimwe T, Krause K, Schlunk I, Kothe E (2012) Modulation of ethanol stress tolerance by aldehyde dehydrogenase in the mycorrhizal fungus *Tricholoma vaccinum*. *Mycorrhiza* 22(6): 471-484. doi:10.1007/s00572-011-0424-9. PubMed: 22159964.
68. Vinche HM, Asachi R, Zamani A, Karimi K (2012) Ethanol and chitosan production from wheat hydrolysate by *Mucor hiemalis*. *J Chem Technol Biotechnol*, 87: 1222–1228. doi:10.1002/jctb.3822. PubMed: 23329859.
69. Oszolák F, Milos PM (2011) RNA sequencing: advances, challenges and opportunities. *Nat Rev Genet* 12(2): 87-98. doi:10.1038/nrg2934. PubMed: 21191423.