Title

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Reward and punishment contingency shifting reveals distinct roles for VTA dopamine and GABA neurons in
 behavioral flexibility

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10 Abstract

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In dynamic environments where stimuli predicting rewarding or aversive outcomes unexpectedly change, it is 12 critical to flexibly update behavior while preserving recollection of previous associations. Dopamine and GABA 13 neurons in the ventral tegmental area (VTA) are implicated in reward and punishment learning, yet little is 14 known about how each population adapts when the predicted outcome valence changes. We measured VTA 15 dopamine and GABA population activity while male and female rats learned to associate three discrete 16 auditory cues to three distinct outcomes: reward, punishment, or no outcome within the same session. After 17 18 learning, the reward and punishment cue-outcome contingencies were reversed, and subsequently re-19 reversed. As expected, the dopamine population rapidly adapted to learning and contingency reversals by increasing the response to appetitive stimuli and decreasing the response to aversive stimuli. In contrast, the 20 21 GABA population increased activity to all sensory events regardless of valence, including the neutral cue. 22 Reversing learned contingencies selectively influenced GABA responses to the reward-predictive cue, prolonging increased activity within and across sessions. The observed valence-specific dissociations in the 23 directionality and temporal progression of VTA dopamine and GABA calcium activity indicates that these 24 25 populations are independently recruited and serve distinct roles during appetitive and aversive associative learning and contingency reversal. 26 27

28 Introduction

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In order to adaptively traverse an environment, one must learn to discriminate between cues that predict 30 31 outcomes of different valences. The ventral tegmental area (VTA) is a central hub for acquiring associations as 32 it contains a heterogeneous assembly of cells that respond to environmental stimuli, predominantly characterized as either dopamine or y-aminobutyric acid (GABA) neurons¹⁻⁴. Extensive research has established 33 a fundamental role for VTA dopamine activity in processing rewarding stimuli⁵⁻¹⁵, although studies have 34 determined that VTA dopamine neurons also exhibit responses to aversive stimuli^{8,16-18}. Historically, 35 investigation of VTA GABA activity has primarily focused on local modulation of dopamine function¹⁹⁻²⁴. 36 However, GABA neurons in the VTA also exert effects independent of VTA dopamine neurons²⁵⁻²⁸, and respond 37 38 to appetitive and aversive stimuli in a manner that is distinct from dopaminergic activity^{8,29,30}. This suggests 39 that VTA dopamine and GABA populations are independently recruited during appetitive and aversive 40 situations.

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The ability to flexibly update behavior is essential for navigating dynamic environments where cues predicting 42 rewarding or aversive consequences can unexpectedly change. To examine the role of VTA neurons in updating 43 learned associations, our earlier work developed a flexible contingency learning (FCL) paradigm to assess the 44 initial acquisition and subsequent reversal of positive and negative associations experienced within the same 45 session^{31,32}. Single unit recordings determined that VTA neurons were predominantly excited by appetitive 46 predictive cues and inhibited by aversive predictive cues, with these signals dynamically updating following 47 contingency reversal³¹. Correlated activity between VTA neurons increased to the appetitive association but 48 decreased in response to the aversive association³², indicating that VTA dopamine and non-dopamine neurons 49

produce separable responses throughout the reversal of contingencies. The response pattern of the VTA GABA 50 population, as well as temporal differences between dopamine and GABA signaling across different phases of 51 52 learning and reversal, however, remain poorly understood.

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Here, we combined the FCL behavioral paradigm with fiber photometry recordings in male and female rats to 54 55 track calcium influx dynamics in VTA dopamine and GABA populations throughout initial learning, reversal, and 56 re-reversal of appetitive and aversive associations. We identified dissociable roles between VTA dopamine and 57 GABA responses across FCL: increases in dopamine population calcium activity occurred in response to appetitive association, whereas the GABA population calcium activity increased to all stimuli. Additionally, 58 59 reversing the contingences evoked a change in how GABA, but not dopamine, processed the new appetitive contingency. The GABA population activity increased towards the appetitive cue across reversal sessions, and 60 61 within-session increases in GABA activity were dynamic during both early and late sessions of the reversal 62 phase. These data collectively illustrate directional and temporal dissociations in VTA dopamine and GABA calcium activity to flexible reward and punishment contingencies. 63

65 Methods

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68 All procedures were approved by the Oregon Health & Science University Institutional Animal Care and Use Committee and were conducted in accordance with the National Institute of Health Guide for the Care and Use 69 70 of Laboratory Animals. Male and female Long-Evans rats were bred in house on a TH::Cre (n = 8; 5 female) or GAD::Cre (*n* = 9; 5 female) transgenic background. All rats were given *ad libitum* access to water and chow, 71 72 group-housed with littermates until aged to P60-65 when surgical procedures began, and maintained on a 12 h reverse light/dark cycle (lights off at 9:00 am) where all experiments were performed during the dark phase. 73

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75 Surgery

76 Viral infusion surgery. Rats were placed under isoflurane anesthesia and received unilateral or bilateral 77 infusions of AAV1-Syn-Flex-GCaMP6f-WPRE-SV40 (1×10¹³ vg/mL, Addgene) to allow for Cre-dependent 78 expression of GCaMP in the VTA (AP -5.5 mm, ML +/-0.6 mm from bregma, DV -7.5, -6.5 mm from dura). Each infusion was 250 nL, administered at a rate of 100 nL/min, with the most ventral infusion performed first. The 79 80 syringe was left in place for 10 min to allow virus to diffuse before slowly removing the needle. Animals were given 5 mg/kg of carprofen after surgery, after which they were single-housed. 81

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83 Fiber implant surgery. After a minimum of 3 weeks following viral infusion surgery, subjects were implanted with a 400 µm diameter optical fiber targeting the VTA (AP -5.5 mm, ML +/-0.6 mm from bregma with left/right 84 sides counterbalanced across animals, DV -7.3 mm from dura) along with 4 skull screws. Fibers were secured to 85 86 the skull using a light-curing dental cement (lvoclar Vivadent) followed by powder acrylic cement (Lang Dental). 87 Subjects were administered 5 mg/kg of carprofen after surgery and were given at least 1 week to recover 88 before behavioral testing began.

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Behavioral testing 90

After recovering from surgery, rats were placed and maintained on mild food restriction to target 90% free-91 feeding weight. Behavioral sessions were performed in conditioning chambers that included grid floors

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connected to a shock generator, a food trough, and three auditory stimulus generators (4.5 kHz tone, white 93

noise, and clicker; Coulbourn Instruments). The chamber floors were thoroughly cleansed with disinfectant, 94

- 95 and the walls and food port were cleaned with 70% ethanol solution between every subject. To familiarize rats with the chamber and food retrieval, rats underwent a single magazine training session in which 25 sucrose 96
- 97 pellets (45 mg; BioServ) were noncontingently delivered at a 90 ± 15 s variable interval.
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99 Flexible contingency learning (FCL) task. Rats were trained on a modified version of our previously developed FCL paradigm ^{31,32}. In this task, subjects underwent 24 Pavlovian conditioning sessions in which the termination 100 of a 5 s auditory cue [conditioned stimulus (CS); tone, white noise, or click] resulted in the delivery of an 101 appetitive (sucrose pellet), aversive (0.2 mA, 180 ms shock), or neutral (nothing) outcome [unconditioned 102 stimulus (US)]. All associations were presented in every session, and the CS-US pairings were counterbalanced 103 across subjects. Each session contained 25 appetitive trials, 25 aversive trials, and 25 neutral trials delivered in 104 105 a pseudorandom order, with a 45 ± 5 s intertrial interval between trials. After 8 sessions of initial training, the 106 appetitive and aversive associations reversed in that the cue previously associated with the sucrose pellet (CS_1) instead preceded the foot shock, and the cue previously associated with the foot shock (CS₂) instead preceded 107 the sucrose pellet. After 8 sessions of this reversal, rats underwent re-reversal where the appetitive and 108 aversive cue-outcome associations returned to the original assignments (as experienced during initial training) 109 for 8 final sessions. The neutral cue (CS-), which was associated with no outcome, did not change across 110 sessions. Conditioned responding was quantified as the change in the rate of head entries to the food port 111 during the 5 s CS relative to the 5 s preceding the CS delivery ³³. We also guantified the latency to initiate a 112 head entry during the CS, and the probability of initiating a head entry during the CS. For the post-outcome 113 114 analysis, we calculated the average number of head entries made during a 5 s post-US delivery time window.

116 Fiber photometry

Fiber photometry recordings for the detection of VTA dopamine or GABA population activity were performed 117 in all sessions using a system with optical components from Doric lenses, with LED modulation controlled by a 118 real-time processor from Tucker Davis Technologies (TDT; RZ5P). Rats were attached to a fiber optic cable in 119 which 465 nm (signal) and 405 nm (isosbestic control) LEDs were modulated at 211 and 330 Hz, respectively, to 120 the implanted cannula. The LED power was set for each animal to yield between 150–200 mV for each signal 121 ³⁴. Data was acquired with TDT Synapse software, and time stamps for the CS and US were collected via 5V TTL 122 signals from the behavioral chamber that interfaced with the TDT processor in order to align events with 123 calcium activity. 124

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126 Analyses of GCaMP signals were performed using custom Python scripts based on those previously described ³⁵⁻³⁷. The isosbestic control and signal channels were low pass filtered at 3 Hz using a butterworth filter to 127 reduce noise, and then the isosbestic control channel was fitted to the signal channel using a least squares 128 polynomial fit of degree 1. Data was then separated into epochs based on the start and end of a given trial. 129 The change in fluorescence (ΔF/F) was calculated by subtracting the fitted isosbestic control channel from the 130 signal channel before dividing by the fitted isosbestic control channel. The signal was then z-scored by 131 132 subtracting the mean Δ F/F from the Δ F/F signal, divided by the standard deviation of the Δ F/F signal. The zscore comparison window was the 5 s prior to the CS onset. To quantify signal changes in response to each CS, 133 the average z-score was calculated during the entire 5 s CS, as well as during the first 2 s and last 3 s of the CS. 134 To quantify responses to the US, the average z-score was calculated during the 3 s following US delivery, and 135 the peak US response was calculated by taking the maximum z-score during the 3 s following CS termination 136 137 relative to 0.5 s before US delivery. We additionally analyzed data in 5-trial bins in order to assess changes in 138 calcium activity between the beginning and end of a session. To determine whether within-session changes in calcium activity occurred during early or late training sessions, the resulting binned data was averaged across 139 the first 4 or last 4 sessions of each training phase. 140

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Fiber photometry permutation analysis. We used a permutation-based approach to compare changes in neural calcium activity as described previously^{35,38} using Python. For each subject and session, a z-score response to the appetitive, aversive, and neutral associations were separately calculated. For each comparison, a null distribution was generated by shuffling the data, randomly selecting the data into two groups, and calculating the mean difference between groups. This was performed 1000 times for each time point. A p-value was obtained by determining the percentage of times a value in the null distribution was greater than or equal to

the observed difference in the unshuffled data (two-tailed for all comparisons). To control for multiple
 comparisons we utilized a consecutive threshold approach based on the 3 Hz lowpass filter window^{35,38,39},
 where a p-value < 0.05 was required for 14 consecutive samples in order to be considered significant.

- 151
- 152 Data analysis and statistics

Detailed results of all statistical tests are found in the **Statistical Tables**. Aside from permutation tests, all statistical analyses were performed in GraphPad Prism 10. FCL behavioral responding, quantification of neural data, and binned trial data were analyzed using a 2-way mixed-effects model fit (restricted maximum likelihood method), repeated measures where appropriate, followed by *post hoc* Tukey's or Sidak's tests. The Geisser– Greenhouse correction was applied to address unequal variances between groups. Unpaired t-tests were used

- 158 to compare open field data between groups.
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- 160 Histology

After behavioral testing, rats were deeply anesthetized with chloral hydrate (400 mg/kg, i.p.), then 161 transcardially perfused with phosphate-buffered saline followed by 4% paraformaldehyde. Brains were 162 removed and postfixed for >24 h, then subsequently placed in 30% sucrose solution. Sections were cut at 40 163 microns on a cryostat (Leica Microsystems) and stored in phosphate-buffered saline (PBS) with 0.05% sodium 164 azide. Immunohistochemistry was performed to verify localization of GCaMP6f viral expression in VTA 165 166 dopamine or GABA neurons for fiber photometry experiments, or to verify GiDREADD/control mCherry and CAV-cre expression for chemogenetic experiments. Brain slices were first permeabilized in 3% bovine serum 167 albumin (BSA), 0.1% Triton X, and 1% Tween 80 in PBS + 0.05% sodium azide for 2 h at room temperature. 168 Sections were then incubated with the primary antibody mouse anti-GFP (1:500, Abcam) as well as either 169 rabbit anti-glutamate decarboxylase (GAD; 1:500, Abcam) or chicken anti-tyrosine hydroxylase (TH; 1:500, 170 Abcam) for fiber photometry experiments, diluted in PBS + Azide, 3% BSA + 0.15% Triton X for 24 h at 4°C. 171 172 Slices were then washed in PBS + azide, 3% BSA + 0.15% Triton X, three times for five minutes each. Brain sections were next incubated with the secondary antibodies donkey-anti-mouse Alexa-488 (1:1000, Abcam), 173 goat-anti-chicken Alexa-594 (1:1000, Abcam), and goat-anti-rabbit Alexa-594 (1:1000, Abcam), diluted in PBS + 174 175 Azide, 3% BSA + 0.15% Triton X for 2 h at room temperature. Sections were washed again as outlined above and mounted to slides with Vectashield anti-fade mounting medium (Vector Labs). Brain slices were imaged for 176 viral expression and fiber placement on a Zeiss Axio Observer microscope. 177

- 179 Results
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181 Behavioral responding differentiates between associative valence and adapts to contingency reversal 182

Rats were trained on a flexible contingency learning (FCL) paradigm that was modified from our previous 183 work^{31,32} to allow for the simultaneous acquisition of three distinct conditioned associations. In each session, 3 184 discrete auditory cues were used as conditioned stimuli (CS), each of which signaled the delivery of a different 185 186 unconditioned stimulus (US): sucrose pellet (appetitive outcome), mild foot shock (aversive outcome), or nothing (neutral outcome; Figure 1A, left). After 8 sessions of this initial learning phase, the appetitive and 187 aversive associations were reversed, where the CS previously paired with a reward (CS₁) instead preceded 188 shock delivery and the CS previously paired with a shock (CS₂) instead preceded reward delivery. Rats were 189 tested on the reversed contingencies for 8 sessions before the associations were re-reversed back to the initial 190 assignments for 8 final sessions. The neutral association (CS-) did not change across the 24 total sessions of FCL 191 (Figure 1A, right). Conditioned responding was quantified as the change in the rate of food port head entries 192 during the 5 s CS relative to the 5 s preceding the CS³³. Rats increased conditioned responding to the food port 193 in response to the initial appetitive CS₁, but not the aversive CS₂ or neutral CS-, throughout the initial learning 194 phase (two-way mixed-effects analysis; initial learning phase CS effect: $F_{(1.14, 18.16)} = 22.71$, p < 0.0001; session x 195 196 CS interaction effect: $F_{(2.60, 41.66)} = 11.81$, p < 0.0001; Figure 1B). When the learned appetitive and aversive

associations reversed, rats adapted their behavior by increasing conditioned responding to the newly appetitive CS₂, and decreased responding during the previously appetitive CS₁ that became aversive (reversal phase CS effect: $F_{(1.06, 16.98)} = 12.22$, p = 0.002; session x CS interaction effect: $F_{(1.75, 26.90)} = 11.97$, p = 0.0003; **Figure 1B**). After re-reversal in which CS-US pairings returned to the initial assignments, rats again updated conditioned responding in favor of the appetitive CS₁ (re-reversal phase CS effect: $F_{(1.13, 15.79)} = 18.99$, p =0.0004; session x CS interaction effect: $F_{(1.73, 23.05)} = 9.15$, p = 0.002; **Figure 1B**).

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Conditioned responding adapted when learned associations were reversed. To determine if the rate of 204 responding differed depending on whether the CS was novel or previously paired with an aversive outcome, 205 we compared conditioned responding to the appetitive association between all FCL phases. Rats acquired the 206 appetitive association at similar rates across initial learning, reversal, and re-reversal phases (Supplemental 207 Figure 1A). We also examined whether conditioned responding differed between the TH and GAD genotypes, 208 and found no effect of genotype on conditioned responding to the appetitive CS during either initial learning 209 or the reversal phase (Supplemental Figure 1B). While the aim of this study was not to examine sex 210 differences, we included both male and female rats and performed further analysis with sex as a factor. There 211 212 was no effect of sex on conditioned responding to the appetitive CS during either initial learning or the reversal of associations (Supplemental Figure 1C). Therefore, data were collapsed across genotype and sex for the 213 remainder of the analyses. 214

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Although subjects reduced rates of conditioned responding to the aversive and neutral CSs throughout FCL 216 (Figure 1B), they continued to explore the food port with low levels of approach probability (Figure 1C). During 217 the initial learning phase, the appetitive CS_1 produced the highest probability of approach and the aversive CS_2 218 produced the lowest, demonstrating the aversive properties of the shock-paired CS₂ (initial learning phase CS 219 effect: $F_{(1.61, 25.69)} = 51.20$, p < 0.0001; session x CS interaction effect: $F_{(5.33, 85.35)} = 10.25$, p < 0.0001; Figure 1C). 220 The probability of approach was modified to reflect the new contingencies during reversal (reversal phase CS 221 effect: $F_{(1.99, 31.87)} = 34.49$, p < 0.0001; session x CS interaction effect: $F_{(4.94, 75.79)} = 13.61$, p < 0.0001), as well as 222 the re-reversal phase (re-reversal phase CS effect: $F_{(1,89,26,44)} = 40.29$, p < 0.0001; session x CS interaction effect: 223 224 $F_{(4.24, 56.67)} = 7.14$, p < 0.0001; Figure 1C). When rats approached the food port during a CS, the appetitive CS₁ also elicited a quicker latency across the initial learning phase (initial learning phase CS effect: $F_{(1.81, 28.98)} = 0.54$, 225 p = 0.57; session x CS interaction effect: $F_{(6.85, 104.7)} = 2.26$, p = 0.04), and during the second reversal (re-reversal 226 CS effect: $F_{(1.95, 27.32)} = 4.95$, p = 0.02; Figure 1D). There was no difference in the latency to respond to the 227 appetitive, aversive, or neutral CSs in the reversal phase (Figure 1D). Post-outcome head entries increased 228 after the termination of the reward-paired CS₁ during initial learning (initial learning phase CS effect: F(1.36, 21.79) 229 = 81.44, p < 0.0001; session x CS interaction effect: $F_{(5.21, 83.38)}$ = 7.03, p < 0.0001; Figure 1E), reversal (reversal 230 phase CS effect: $F_{(1.16, 18.54)} = 61.81$, p < 0.0001; session x CS interaction effect: $F_{(4.39, 67.40)} = 1.60$, p = 0.18) and 231 re-reversal (re-reversal phase CS effect: $F_{(1.66, 23, 24)} = 41.51$, p < 0.0001; session x CS interaction effect: $F_{(4, 29, 57, 35)}$ 232 = 5.40, p = 0.0007; Figure 1E). These findings illustrate that rats learn to distinguish between simultaneously 233 acquired appetitive, aversive, and neutral cues, and adapt their behavior when contingencies update. 234

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- 236 Dissociable VTA dopamine and GABA responses dynamically adapt during learning and reversal of 237 contingencies
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- VTA dopamine and GABA neurons are implicated in associative learning, but their distinct roles in flexible updating of cue-outcome associations is not well understood. We used fiber photometry to measure cue- and outcome-elicited calcium responses in VTA dopamine and GABA neuron populations throughout FCL. To record from the VTA dopamine neuron population, TH::cre rats expressed GCaMP6f in TH+ cells in the VTA, and fiber placement was centered above the virus injection (**Figure 2A**). Differences in neural calcium activity between the first and last session of each FCL phase were assessed using a permutation-based approach^{35,38}. In the first session (session 1), VTA dopamine calcium activity increased in response to reward but not the appetitive CS₁

246 predicting reward. By the end of the initial learning phase (session 8), a large phasic increase in response to the appetitive predictive CS₁ developed (Figure 2B). The dopamine response to the aversive CS₂ or neutral CS- did 247 not significantly change across initial sessions (Figure 2B). By session 8, however, dopamine population calcium 248 activity was lower during the aversive CS₂ compared to the neutral CS- (Supplemental Figure 2A). When the 249 appetitive and aversive associations reversed (session 9), the dopamine population first displayed a prediction 250 error-like response: decreased activity to the unexpected shock and increased activity to the unexpected 251 252 reward (Figure 2C; Supplemental Figure 2B; Supplemental Figure 3A). By the end of the reversal phase (session 16), the dopamine population had adjusted to the new contingencies and displayed a similar response 253 profile to the contingencies as before the reversal (Figure 2C; Supplemental Figure 4A-B). During re-reversal, 254 the dopamine population followed a similar pattern by initially responding in a prediction error-like manner 255 (session 17) and adapted by the end of the phase (session 24; Figure 2D; Supplemental Figure 2C; 256 Supplemental Figure 3B; Supplemental Figure 4A-B). 257

To record from the VTA GABA neuron population, we used fiber photometry in GAD::cre rats expressing 258 GCaMP6f in GAD+ cells (Figure 3A). In session 1, the GABA population displayed a phasic increase in response 259 to all three CSs as well as both shock and reward USs (Figure 3B; Supplemental Figure 2D). Despite subjects 260 successfully learning the contingencies, by the end of the initial phase GABA activity in response to these 261 events, including the neutral CS, remained the same (Figure 1; Figure 3B; Supplemental Figure 1; 262 **Supplemental Figure 2D).** The only exception was that the GABA response to appetitive CS_1 increased after 263 learning (Figure 3B; Supplemental Figure 2D). When the appetitive and aversive associations reversed, the 264 GABA population also displayed a prediction error-like response in session 9 by decreasing activity to the 265 unexpected shock and increasing activity to the unexpected reward (Figure 3C; Supplemental Figure 2E; 266 Supplemental Figure 3C). By the end of reversal learning, GABA responses were nearly identical to the last 267 session of initial learning (Figure 3C; Supplemental Figure 4C-D). During the second reversal, the GABA 268 population again showed a prediction error-like response and adapted by the end of the re-reversal phase 269 (Figure 3D; Supplemental Figure 2F; Supplemental Figure 3D; Supplemental Figure 4C-D). The GABA response 270 to the neutral CS remained unchanged even after extensive training. 271

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When a CS was contingent on an outcome, neural calcium activity of both dopamine and GABA populations 273 dynamically changed between the first and last session of each phase in FCL (Figures 2-3). To quantify the 274 flexible progression of CS-evoked dopamine and GABA responses in different FCL phases, we averaged activity 275 during the full 5 s CS presentation, as well the early (first 2 s) and late (last 3 s) periods of CS presentation 276 (Figure 4A-D) over all sessions. When considering the full CS response in the initial learning phase, VTA 277 dopamine activity was largest to the appetitive CS_1 compared to the aversive and neutral CSs (initial learning 278 phase CS effect: $F_{(1.82, 12.79)} = 43.07$, p < 0.0001; session x CS interaction effect: $F_{(3.28, 21.52)} = 5.42$, p = 0.005; 279 Figure 4A). By the end of the initial learning phase, dopamine responses to the shock-predictive CS₂ were 280 lower than to the neutral CS-. This pattern of responding flexibly updated to the new contingencies during the 281 reversal phase (reversal phase CS effect: $F_{(1.19, 8.32)} = 21.34$, p = 0.001; session x CS interaction effect: $F_{(2.83, 17.96)} =$ 282 10.30, p = 0.0004) as well as during the re-reversal phase (re-reversal phase CS effect: $F_{(1.33, 9.33)} = 22.81$, p =283 0.0005; session x CS interaction effect: $F_{(2.15, 11.81)} = 10.61$, p = 0.002; Figure 4A). In contrast to VTA dopamine, 284 VTA GABA calcium activity ubiquitously increased to all CSs across the initial learning phase (initial learning 285 phase CS effect: $F_{(1.30, 10.37)} = 3.77$, p = 0.07; session x CS interaction effect: $F_{(2.68, 20.29)} = 2.15$, p = 0.13; Figure 286 4B). Reversing the learned contingencies, however, increased GABA activity to the new appetitive CS₂ 287 compared to the other CSs (reversal phase CS effect: $F_{(1.44, 11.50)} = 6.00$, p = 0.02; session x CS interaction effect: 288 $F_{(2.34, 16.71)} = 7.82$, p = 0.003; Figure 4B). This response pattern from the GABA population persisted into the re-289 reversal phase with an interaction effect between session and CS (re-reversal phase CS effect: $F_{(1.03, 7.23)} = 4.32$, 290 p = 0.07; session x CS interaction effect: $F_{(2.60, 13.20)} = 4.31$, p = 0.03; Figure 4B). 291 292

293 Regardless of task phase or predicted outcome valence, throughout FCL CS-evoked GABA activity was higher than dopamine activity (Supplemental Figure 5A). Whereas VTA dopamine calcium activity displayed a phasic 294 increase at CS onset that diminished towards baseline by the end of the CS (in the case of the appetitive 295 association), GABA calcium activity remained amplified throughout the CS (see Figures 2-3). We therefore 296 quantified differences between the early CS period (0-2 s after CS onset) and the late CS period (2-5 s after CS 297 onset; Figure 4C-D). In the initial learning phase, every CS produced a larger response during early CS 298 299 compared to late CS in both the dopamine population (initial learning phase CS₁ early v late CS effect: $F_{(1,7)}$ = 21.87, p = 0.002; CS₁ session x early v late CS interaction effect: $F_{(2.45, 15.78)} = 5.74$, p = 0.01; CS₂ early v late CS 300 effect: $F_{(1,7)} = 19.91$, p = 0.003; CS- early v late CS effect: $F_{(1,7)} = 10.04$, p = 0.02; Figure 4C) and the GABA 301 population (CS₁ early v late CS effect: $F_{(1, 8)}$ = 7.63, p = 0.02; CS₂ early v late CS effect: $F_{(1, 8)}$ = 9.71, p = 0.01; CS-302 early v late CS effect: $F_{(1, 8)}$ = 14.35, p = 0.005; Figure 4D). The early response to the appetitive and aversive CSs 303 continued in the dopamine population throughout reversal and re-reversal phases (reversal phase CS₁ early v 304 late CS effect: $F_{(1,7)} = 13.36$, p = 0.008; CS₁ session x early v late CS interaction effect: $F_{(2.49, 15.29)} = 5.00$, p = 0.02; 305 CS₂ early v late CS effect: $F_{(1,7)} = 8.56$, p = 0.02; CS₂ session x early v late CS interaction effect: $F_{(2.53, 15.53)} = 3.63$, 306 p = 0.04; re-reversal phase CS₁ session x early v late CS interaction effect: $F_{(1.69, 8.46)} = 7.82$, p = 0.01; CS₂ early v 307 late CS effect: $F_{(1,7)} = 9.07$, p = 0.02; Figure 4C). The reversal phase also increased reward-evoked US responses 308 in dopamine and GABA populations (Supplemental Figure 5B-E). However, reversing the contingencies 309 eliminated differences in GABA activity between early and late CS responses for the new appetitive CS₂ and 310 311 neutral CS-, and for all CSs during re-reversal (Figure 4D). Thus, the CS-evoked activity during the reversal phase was encoded differently by GABA, and not dopamine, populations. The GABA response to the appetitive 312 CS significantly increased during reversal, and differences between early and late CS-evoked activity were no 313 314 longer significant.

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316 Reversing learned associations increases within-session activity in the VTA GABA population

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Dopamine and GABA calcium activity changed over FCL sessions, reflecting learning by both populations across 318 days (Figures 2-4). To determine the temporal dynamics of this change, we parsed sessions into 5-trial bins and 319 320 compared full CS responses between the first bin and the last bin in each session (Figure 5A-B; Supplemental Figure 6; and equivalent analysis of behavioral responding in Supplemental Figure 7). In all FCL phases, both 321 dopamine and GABA populations exhibited within-session increases in CS-evoked calcium activity. However, 322 there was a stark difference in the temporal progression of within-session activity between dopamine and 323 GABA populations. In particular, CS-evoked dopamine activity increased to the appetitive association within 324 the first several sessions of each FCL phase, whereas CS-evoked GABA within-session activity increased to both 325 326 appetitive and aversive associations throughout the FCL phases (Supplemental Figure 6A,C). To quantify these differences, we averaged the binned activity between early (first 4) and late (last 4) sessions of each FCL phase 327 (Figure 5C). 328

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The temporal pattern of within-session responses to the appetitive CS were different between dopamine and 330 GABA populations. Dopamine calcium activity increased to the reward-predictive CS within only the early 331 sessions of every FCL phase (dopamine early sessions CS₁ trial effect: $F_{(1,7)} = 28.22$, p = 0.001; FCL phase x CS₁ 332 trial interaction effect: $F_{(1.96, 13.69)} = 9.95$, p = 0.002; CS₂ trial effect: $F_{(1, 7)} = 7.75$, p = 0.03; FCL phase x CS₂ trial 333 interaction effect: $F_{(1.35, 9.43)} = 4.78$, p < 0.05; Figure 5D). In contrast, the GABA population did not exhibit 334 within-session changes in activity to the appetitive association during the initial learning phase (Figure 5E). 335 Reversing the learned contingencies, however, elicited increased within-session GABA activity to the appetitive 336 association during both early and late sessions, an effect that persisted into the re-reversal phase (GABA early 337 sessions CS₁ trial effect: $F_{(1,8)} = 20.33$, p = 0.002; FCL phase x CS₁ trial interaction effect: $F_{(1.33, 9.28)} = 8.22$, p =338 0.01; CS₂ trial effect: $F_{(1, 8)} = 15.86$, p = 0.004; late sessions CS₁ trial effect: $F_{(1, 8)} = 14.37$, p = 0.005; CS₂ trial 339 effect: $F_{(1, 8)} = 29.74$, p = 0.0006; Figure 5E). Furthermore, only the GABA population exhibited increased 340 within-session activity to the aversive association, which occurred during late sessions of the initial learning 341

and reversal phases (Figure 5E). Changes in dopamine and GABA responses to the neutral CS-, as well as to the
 reward and shock USs, remained minimal within FCL sessions (Supplemental Figure 6). Within-session analysis
 of behavior indicates that conditioned responding increased to the appetitive association during both early
 and late sessions throughout FCL phases, implicating both dopamine and GABA within-session increased
 activity in modulating behavioral responding (Supplemental Figure 7). Thus while within-session signaling is
 modulated in both VTA dopamine and GABA populations, it is dissociable through distinct temporal and
 valence-specific response profiles.

350 Discussion

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A large number of studies have characterized the activity of VTA dopamine neurons during reward learning⁵⁻¹⁵, 352 whereas the role of GABA activity has been primarily limited to its modulation of dopamine function¹⁹⁻²⁴. 353 Some prior research, however, has shown that VTA GABA neurons elicit responses that are distinct from 354 dopamine neurons^{8,29,30,32}, suggesting that VTA GABA and dopamine populations may have independent roles 355 in encoding associative learning. Here, we compared VTA dopamine and GABA calcium activity using fiber 356 357 photometry during a flexible contingency learning paradigm that assessed initial learning and reversal of appetitive and aversive associations acquired simultaneously. The initial acquisition of cue-outcome 358 associations elicited responses from both dopamine and GABA VTA populations. Reversing the learned reward 359 and punishment contingencies, however, selectively influenced GABA population responses to the reward-360 predictive cue, by prolonging increased calcium activity both within and across sessions. These findings reveal 361 that the VTA GABA population is critically modulated when learned contingencies update, and further suggests 362 that dopamine and GABA neurons are independently recruited during learning of appetitive and aversive 363 associations. 364

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The ability to flexibly update responding when learned contingencies change has been examined across 366 species through reversal learning, latent inhibition, and counterconditioning paradigms⁴⁰⁻⁴². Reversal learning 367 typically involves switching between two appetitive outcomes, and latent inhibition comprises overriding a 368 neutral stimulus with an association that has valence. Counterconditioning paradigms, which employ reversals 369 between appetitive and aversive associations, examine each valence reversal independently^{43,44}. Our flexible 370 contingency learning task combines features of these paradigms to demonstrate that rats adapt to the valence 371 reversal of reward and punishment associations experienced concurrently^{31,32}. Our current findings extend 372 these observations to both male and female rats, and we demonstrate rats also re-reverse their behavior when 373 associations are returned to their initial contingencies. We did not identify sex differences in behavioral 374 responding, potentially due to the strain used in this study^{33,45}. Pre-exposure to cues associated with a neutral 375 outcome (such as in latent inhibition) or an aversive outcome (such as in counterconditioning) has been found 376 to dampen behavioral responding when the contingency is subsequently shifted to an appetitive outcome⁴¹⁻ 377 ^{43,46,47}. However, in our task where multiple cue-outcome associations are experienced in the same session, we 378 379 find that rats learn to associate the previously aversive predictive cue with a rewarding outcome at the same 380 rate as the initial acquisition of the appetitive association. This suggests that a more complex environment may facilitate behavioral flexibility. 381

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We observed distinct calcium influx responses between dopamine and GABA neurons in the VTA to appetitive, 383 aversive, and neutral associations. On a population level, VTA dopamine neuron calcium activity increased to 384 the appetitive association and decreased to the aversive association, with minimal response to the neutral cue. 385 This is consistent with the response profile for the majority of individual VTA dopamine neurons examined 386 previously^{5,8,10,11,16,17,31}. In contrast, VTA GABA population calcium activity increased in response to all salient 387 stimuli. Initially this increased response was uniform, but after additional exposure the appetitive cue evoked a 388 more pronounced increase in GABA activity relative to the aversive and neutral cues. Although changes in 389 calcium activity measured with fiber photometry do not necessarily reflect spiking activity⁴⁸ (but see⁴⁹), our 390

findings are consistent with previously measured changes in spike rate of VTA dopamine and GABA neurons^{8,31}. 391 Furthermore, this approach allowed us to observe responses to the shock outcome, which causes noise in 392 electrophysiological recordings. Both shock and reward outcomes as well as all predictive cues, including the 393 neutral cue regardless of the extensive exposure to this association, elicited increases in VTA GABA calcium 394 activity, implicating this population in salience signaling⁵⁰. When the contingencies were reversed, both VTA 395 dopamine and GABA calcium activity exhibited a reward prediction error-like response. The purpose of 396 397 prediction error responses in dopamine neurons have been well-studied as a teaching signal for adaptive learning^{5,6,8-10,51,52}. Limited prior research has also identified increased VTA GABA firing to the delivery of an 398 unexpected reward⁵³, which may function to convey outcome value to downstream targets such as the ventral 399 pallidum⁵⁴. 400

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While both dopamine and GABA population calcium activity increased to the appetitive association, activation 402 of these populations produces different effects on reward-based behavioral responding. Optogenetic studies 403 have shown that activating VTA dopamine neurons during either an appetitive cue or reward delivery enhances 404 cued reward seeking^{51,55}. In contrast, activation of the VTA GABA population prior to or during reward delivery 405 respectively decreases anticipatory conditioned responding and reward consumption^{22,53}. Nevertheless, fiber 406 photometry and optogenetic approaches used in vivo are unable to disentangle potential heterogeneity 407 between individual neurons. We appreciate that both dopamine and GABA cell groups in the VTA can display 408 diverse responses to stimuli that can depend on their anatomical location or projection targets^{3,18,30,56}. Our 409 observation that within-session changes in cue-evoked GABA activity occur throughout training suggests that 410 we likely recorded signals from both local and long-range projecting GABA neurons, which may individually be 411 412 critical at separate stages of training. Elevated local VTA GABA activity could serve as a salience or prediction signal to dopamine neurons during initial learning^{50,53}, while increased activity in long-range GABA projections 413 to cholinergic interneurons in the ventral striatum could facilitate cue discrimination during reversal sessions²⁵. 414 Understanding the multifaceted role of VTA GABAergic activity during associative learning will be essential for 415 future research. 416

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In conclusion, we identify distinct patterns of responses between VTA dopamine and GABA populations during appetitive and aversive associative learning and contingency reversal. A critical observation was that VTA GABA neuron population calcium activity is selectively amplified by contingency reversal. This finding supports a role for GABA neurons in behavioral flexibility. Previous research examining flexible behavior focus on how cortical, striatal, and amygdalar regions mediate the reversal of learned contingencies⁴⁰⁻⁴², which all send projections to the VTA^{29,57-60}. Therefore, it will be critical to determine the impact of the afferent projections on VTA dopamine and GABA signaling when learned contingencies are updated.

426 Author contributions

M.J.L. and B.M. designed research; M.J.L. performed research, analyzed data, and wrote the first draft of the
paper; M.J.L. and B.M. edited the paper; M.J.L. and B.M. wrote the paper.

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430 Competing interests

431 The authors report no conflicts of interest.

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442 Data Availability

The datasets presented in the current study are available from the corresponding author upon reasonable request.

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446 Figures and Figure legends



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Figure 1. Behavioral responding adapts to reversal of contingencies during FCL. A, Schematic for the FCL task. 448 Left, diagram of the three cue-outcome associations presented in every FCL session. Right, chart depicting the 449 450 initial, reversal, and re-reversal phases of FCL. B, Conditioned responding towards CSs initially associated with reward (blue), shock (red) or nothing (gray open circles). **C**, Probability of approaching the food port during CS 451 presentation. **D**, Latency to respond with a head entry into the food port during CS presentation. **E**, Number of 452 food port head entries following US delivery. Gray shading represents reversal period in which appetitive and 453 454 aversive associations were switched. Data are presented as mean +/- SEM. Asterisks represent main effect of CS; pound signs represent interaction effect with session. #, *p < 0.05; ##, **p < 0.01; ###, ***p < 0.001;455 ####,**** p < 0.0001. 456

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463 Figure 2. VTA dopamine signaling encodes reward association across FCL phases. A, Left, GCaMP6f was 464 expressed in dopamine neurons of TH::cre rats. Right, optic fibers were placed above viral expression in the VTA. B, Average VTA dopamine population calcium activity towards CSs associated with reward (teal, left), 465 466 shock (orange, middle) or nothing (gray, right) during the first (Session 1) and last (Session 8) sessions of the 467 initial learning phase. The first session of each phase is depicted in darker shades, the last session of each phase is depicted in lighter shades. Colored lines above each trace represent a significant difference between 468 469 the first and last session of the phase detected via permutation test. C, Average calcium activity towards CSs associated with shock (teal, left), reward (orange, middle) or nothing (gray, right) during the first (Session 9) 470 and last (Session 16) sessions of the reversal phase. D, Average calcium activity towards CSs associated with 471 472 reward (teal, left), shock (orange, middle) or nothing (gray, right) during the first (Session 17) and last (Session 24) sessions of the re-reversal phase. Data are presented as mean + SEM. Vertical scale bars indicate 1 z-score. 473 474 Horizontal scale bars indicate 1 second.

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Figure 3. VTA GABA population responds to all associations across FCL phases. A, Left, GCaMP6f was expressed 478 in GABA neurons of GAD::cre rats, and there was no co-expression with TH neurons (inset). Right, optic fibers 479 480 were placed above viral expression in the VTA. **B**, Average VTA GABA population calcium activity towards CSs 481 associated with reward (green, left), shock (yellow, middle) or nothing (gray, right) during the first (Session 1) 482 and last (Session 8) sessions of the initial learning phase. The first session of each phase is depicted in darker shades, the last session of each phase is depicted in lighter shades. Colored lines above each trace represent a 483 484 significant difference between the first and last session of the phase detected via permutation test. C, Average calcium activity towards CSs associated with shock (green, left), reward (yellow, middle) or nothing (purple, 485 gray) during the first (Session 9) and last (Session 16) sessions of the reversal phase. D, Average calcium activity 486 towards CSs associated with reward (green, left), shock (yellow, middle) or nothing (gray, right) during the first 487 (Session 17) and last (Session 24) sessions of the re-reversal phase. Data are presented as mean + SEM. Vertical 488 scale bars indicate 1 z-score. Horizontal scale bars indicate 1 second. 489

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Figure 4. Quantification of VTA dopamine and GABA responses evoked by full, early or late periods of CS presentation across FCL sessions. **A**,**B**, Dopamine (A) and GABA (B) average calcium activity during the full 5 s CS presentation. Large asterisks above the graph represent main effect of CS, pound signs represent interaction effect with session. Small asterisks within graph represent post-hoc comparisons between CSs: black asterisks compare CS₁ and CS₂; teal (A) and green (B) asterisks compare CS₁ and CS-; orange (A) and yellow (B) asterisks compare CS₂ and CS-. C,D, Dopamine (C) and GABA (D) average calcium activity during the early period (first 2 s; darker shades) or the late period (last 3 s; lighter shades) of CS presentation. Data are presented as mean +/-SEM. Asterisks represent main effect between early and late CS, pound signs represent interaction effect with session. #,*p < 0.05; ##,**p < 0.01; ###,*** p < 0.001; ####,**** p < 0.0001.



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Figure 5. Within-session changes in CS-evoked calcium activity in VTA dopamine and GABA populations differ between early and late sessions in each FCL phase. **A**, Representative trace of within-session changes in calcium activity during an early phase session. Example trace of VTA dopamine calcium activity during a single session (Session 2) to the appetitive CS₁ portioned into 5-trial bins. Dotted rectangles represent first and last 5trial bins from the session. **B**. Depresentative trace of within session changes in calcium activity during a late.

525 trial bins from the session. **B**, Representative trace of within-session changes in calcium activity during a late

526 527	phase session. Example trace of VTA GABA calcium activity during a single session (Session 14) to the appetitive CS ₂ portioned into 5-trial bins. C , Schematic depicting early (first 4 sessions; left) and late (last 4		
528	sessions; right) sessions of each FCL phase. D , Dopamine within-session calcium activity during the 5 s CS		
529	period for the appetitive and aversive associations in early (left) and late (right) sessions. Sessions were split		
530	into 5-trial bins, and bins were averaged across sessions. The color intensity represents the beginning of the		
531	session (Trials 1-5, dark shades) or the end of the session (Trials 21-25, light shades). E. GABA within-session		
532	calcium activity during the 5 s CS period for the appetitive and aversive associations in early (left) and late		
533	(right)) sessions. Data are presented as mean +/- SEM, with individual subject points included. Asterisks	
534	repres	sent post-hoc comparisons between trial bins for each CS. * $p < 0.05$; ** $p < 0.01$.	
535	-1		
536	Refer	ences	
537			
538	1	Margolis, E. B., Lock, H., Hjelmstad, G. O. & Fields, H. L. The ventral tegmental area revisited: is there an	
539		electrophysiological marker for dopaminergic neurons? J Physiol 577, 907-924 (2006).	
540		https://doi.org:10.1113/iphysiol.2006.117069	
541	2	Nair-Roberts, R. G. <i>et al.</i> Stereological estimates of dopaminergic. GABAergic and glutamatergic	
542	-	neurons in the ventral tegmental area, substantia nigra and retrorubral field in the rat. <i>Neuroscience</i>	
543		152 . 1024-1031 (2008), https://doi.org:10.1016/i.neuroscience.2008.01.046	
544	3	Morales M & Margolis F B Ventral tegmental area: cellular heterogeneity connectivity and	
545	5	behaviour Nat Rev Neurosci 18 73-85 (2017) https://doi.org/10.1038/prp.2016.165	
546	4	Margolis F B Toy B Himmels P Morales M & Fields H L Identification of rat ventral tegmental	
547	•	area GABAergic neurons <i>PLoS One</i> 7 e42365 (2012) https://doi.org/10.1371/journal.none.0042365	
548	5	Schultz W Davan P & Montague P B A neural substrate of prediction and reward Science 275 1593-	
549	5	1599 (1997) https://doi.org/10/1126/science 275/5306/1593	
550	6	Lak A Stauffer W R & Schultz W Donamine prediction error responses integrate subjective value	
551	U	from different reward dimensions. Proc Natl Acad Sci U S A 111 , 2343-2348 (2014).	
552		https://doi.org/10.1073/pnas.1321596111	
553	7	Tobler, P. N., Fiorillo, C. D. & Schultz, W. Adaptive coding of reward value by dopamine neurons. <i>Science</i>	
554		307 . 1642-1645 (2005). https://doi.org:10.1126/science.1105370	
555	8	Cohen, I. Y., Haesler, S., Vong, L., Lowell, B. B. & Uchida, N. Neuron-type-specific signals for reward and	
556	C	punishment in the ventral tegmental area. <i>Nature</i> 482 , 85-88 (2012).	
557		https://doi.org:10.1038/nature10754	
558	9	Fiorillo, C. D., Tobler, P. N. & Schultz, W. Discrete coding of reward probability and uncertainty by	
559	C	dopamine neurons. <i>Science</i> 299 , 1898-1902 (2003), https://doi.org:10.1126/science.1077349	
560	10	Pan, W. X., Schmidt, R., Wickens, J. R. & Hyland, B. I. Dopamine cells respond to predicted events during	
561		classical conditioning: evidence for eligibility traces in the reward-learning network. J Neurosci 25.	
562		6235-6242 (2005), https://doi.org:10.1523/INEUROSCI.1478-05.2005	
563	11	Mohebi, A. et al. Dissociable dopamine dynamics for learning and motivation. Nature 570 , 65-70	
564		(2019), https://doi.org:10.1038/s41586-019-1235-v	
565	12	Wise, R. A. Dopamine, learning and motivation. <i>Nat Rev Neurosci</i> 5 , 483-494 (2004).	
566		https://doi.org:10.1038/nrn1406	
567	13	Fields, H. L., Hielmstad, G. O., Margolis, E. B. & Nicola, S. M. Ventral tegmental area neurons in learned	
568		appetitive behavior and positive reinforcement. Annu Rev Neurosci 30 , 289-316 (2007).	
569		https://doi.org:10.1146/annurey.neuro.30.051606.094341	
570	14	van Zessen, R. <i>et al.</i> Cue and Reward Evoked Dopamine Activity Is Necessary for Maintaining Learned	
571		Paylovian Associations, J Neurosci 41 , 5004-5014 (2021), https://doi.org/10.1523/INFUROSCI 2744-	
572		20.2021	
573	15	Garr. E. <i>et al.</i> Mesostriatal dopamine is sensitive to changes in specific que-reward contingencies. Sci	
574	-	Adv 10, eadn4203 (2024). https://doi.org:10.1126/sciadv.adn4203	

575 576	16	Ungless, M. A., Magill, P. J. & Bolam, J. P. Uniform inhibition of dopamine neurons in the ventral tegmental area by aversive stimuli. <i>Science</i> 303 , 2040-2042 (2004).
577		https://doi.org:10.1126/science.1093360
578	17	Brischoux, F., Chakraborty, S., Brierley, D. I. & Ungless, M. A. Phasic excitation of dopamine neurons in
579		ventral VIA by noxious stimuli. <i>Proc Natl Acad Sci U S A</i> 106 , 4894-4899 (2009).
580	4.0	https://doi.org:10.10/3/pnas.081150/106
581	18	Matsumoto, M. & Hikosaka, O. Two types of dopamine neuron distinctly convey positive and negative
582		motivational signals. <i>Nature</i> 459 , 837-841 (2009). <u>https://doi.org:10.1038/nature08028</u>
583	19	Creed, M. C., Ntamati, N. R. & Ian, K. R. VIA GABA neurons modulate specific learning behaviors
584		through the control of dopamine and cholinergic systems. Front Behav Neurosci 8, 8 (2014).
585	20	https://doi.org:10.3389/fnben.2014.00008
586	20	Ian, K. R. <i>et al.</i> GABA neurons of the VIA drive conditioned place aversion. <i>Neuron</i> 73 , 1173-1183
587	24	(2012). <u>https://doi.org:10.1016/j.neuron.2012.02.015</u>
588	21	Bocklisch, C. et al. Cocaine disinhibits dopamine neurons by potentiation of GABA transmission in the
589		ventral tegmental area. Science 341 , 1521-1525 (2013). <u>https://doi.org:10.1126/science.1237059</u>
590	22	van Zessen, R., Phillips, J. L., Budygin, E. A. & Stuber, G. D. Activation of VTA GABA neurons disrupts
591		reward consumption. Neuron 73 , 1184-1194 (2012). https://doi.org:10.1016/j.neuron.2012.02.016
592	23	Dobi, A., Margolis, E. B., Wang, H. L., Harvey, B. K. & Morales, M. Glutamatergic and nonglutamatergic
593		neurons of the ventral tegmental area establish local synaptic contacts with dopaminergic and
594		nondopaminergic neurons. <i>J Neurosci</i> 30 , 218-229 (2010). <u>https://doi.org:10.1523/JNEUROSCI.3884-</u>
595	~ .	
596	24	Omelchenko, N. & Sesack, S. R. Ultrastructural analysis of local collaterals of rat ventral tegmental area
597		neurons: GABA phenotype and synapses onto dopamine and GABA cells. Synapse 63, 895-906 (2009).
598	25	https://doi.org:10.1002/syn.20668
599	25	Brown, M. I. et al. Ventral tegmental area GABA projections pause accumbal cholinergic interneurons
600	26	to enhance associative learning. <i>Nature</i> 492 , 452-456 (2012). <u>https://doi.org:10.1038/nature1165/</u>
601	26	Carr, D. B. & Sesack, S. R. GABA-containing neurons in the rat ventral tegmental area project to the
602 CO2		prefrontal cortex. Synapse 38 , 114-123 (2000). <u>https://doi.org:10.1002/1098-</u>
603		2396(200011)38:2<114::AID-SYN2>3.0.CU;2-R
604 605	27	Al-Hasani, R. <i>et al.</i> Ventral tegmental area GABAergic inhibition of cholinergic interneurons in the
605		ventral nucleus accumpens snell promotes reward reinforcement. <i>Nat Neurosci</i> 24 , 1414-1428 (2021).
606	20	https://doi.org:10.1038/s41593-021-00898-2
607	28	Van Bockstaele, E. J. & Pickel, V. M. GABA-containing neurons in the ventral tegmental area project to
608		the nucleus accumbens in rat brain. Brain Res 682 , 215-221 (1995). <u>https://doi.org:10.1016/0006-</u>
609	20	<u>8993(95)00334-m</u>
610 C11	29	Bouarab, C., Thompson, B. & Polter, A. M. VIA GABA Neurons at the Interface of Stress and Reward.
611	20	Front Neural Circuits 13 , 78 (2019). <u>https://doi.org:10.3389/thcir.2019.00078</u>
61Z	30	Eium, J. E. <i>et al.</i> Distinct dynamics and intrinsic properties in ventral tegmental area populations
C14		https://dei.org/10.1016/i.colrop.2024.114668
014 615	21	<u>Mups://doi.org.10.1016/j.ceirep.2024.114668</u>
615 615	31	Kim, Y. B., Matthews, M. & Mognaddam, B. Putative gamma-aminobutyric acid neurons in the ventral
010 617		learning. Fur / Neurosci 22 , 1564, 1572 (2010), https://doi.org/10.1111/j.1460.0568.2010.07271.v
617 617	22	learning. Eur J Neurosci 32 , 1564-1572 (2010). <u>https://doi.org:10.1111/j.1460-9568.2010.07371.x</u>
618	32	Kim, Y., Wood, J. & Wiognaddam, B. Coordinated activity of ventral tegmental neurons adapts to
670 673		appennive and aversive learning. PLOS OTE 7, e29766 (2012).
02U 601	22	Intps://uoi.org.tu.ts/t/journal.pone.0029766
622	55	Lenner, IVI. J., Dejeux, IVI. I. & Wanat, IVI. J. Sex Differences in Benavioral Responding and Dopamine
022		Release outing Paviovian Learning. <i>Evento</i> 9 (2022). <u>https://doi.org.10.1523/ENEURO.0050-22.2022</u>

Seiler, J. L. et al. Dopamine signaling in the dorsomedial striatum promotes compulsive behavior. Curr 623 34 Biol 32, 1175-1188 e1175 (2022). https://doi.org:10.1016/j.cub.2022.01.055 624 Jacobs, D. S., Allen, M. C., Park, J. & Moghaddam, B. Learning of probabilistic punishment as a model of 35 625 anxiety produces changes in action but not punisher encoding in the dmPFC and VTA. Elife 11 (2022). 626 https://doi.org:10.7554/eLife.78912 627 36 Jacobs, D. S., Bogachuk, A. P. & Moghaddam, B. Orbitofrontal and prelimbic cortices serve 628 629 complementary roles in adapting reward seeking to learned anxiety. Biol Psychiatry (2024). 630 https://doi.org:10.1016/j.biopsych.2024.02.1015 Jacobs, D. S. & Moghaddam, B. Prefrontal Cortex Representation of Learning of Punishment Probability 631 37 During Reward-Motivated Actions. J Neurosci 40, 5063-5077 (2020). 632 https://doi.org:10.1523/JNEUROSCI.0310-20.2020 633 Jean-Richard-Dit-Bressel, P., Clifford, C. W. G. & McNally, G. P. Analyzing Event-Related Transients: 38 634 Confidence Intervals, Permutation Tests, and Consecutive Thresholds. Front Mol Neurosci 13, 14 (2020). 635 https://doi.org:10.3389/fnmol.2020.00014 636 Pascoli, V. et al. Stochastic synaptic plasticity underlying compulsion in a model of addiction. Nature 39 637 564, 366-371 (2018). https://doi.org:10.1038/s41586-018-0789-4 638 Izquierdo, A., Brigman, J. L., Radke, A. K., Rudebeck, P. H. & Holmes, A. The neural basis of reversal 639 40 learning: An updated perspective. Neuroscience 345, 12-26 (2017). 640 641 https://doi.org:10.1016/j.neuroscience.2016.03.021 Keller, N. E., Hennings, A. C. & Dunsmoor, J. E. Behavioral and neural processes in counterconditioning: 642 41 Past and future directions. Behav Res Ther 125, 103532 (2020). 643 644 https://doi.org:10.1016/j.brat.2019.103532 Miller, D. B., Rassaby, M. M., Collins, K. A. & Milad, M. R. Behavioral and neural mechanisms of latent 645 42 inhibition. Learn Mem 29, 38-47 (2022). https://doi.org:10.1101/lm.053439.121 646 647 43 Peck, C. A. & Bouton, M. E. Context and performance in aversive-to-appetitive and appetitive-toaversive transfer. Learning and Motivation 21, 1-31 (1990). https://doi.org:10.1016/0023-648 9690(90)90002-6 649 650 44 Bouton, M. E. & Peck, C. A. Spontaneous recovery in cross-motivational transfer (counterconditioning). 651 Animal Learning & Behavior 20, 313–321 (1992). https://doi.org:10.3758/BF03197954 Rivera-Garcia, M. T., McCane, A. M., Chowdhury, T. G., Wallin-Miller, K. G. & Moghaddam, B. Sex and 652 45 strain differences in dynamic and static properties of the mesolimbic dopamine system. 653 Neuropsychopharmacology 45, 2079-2086 (2020). https://doi.org:10.1038/s41386-020-0765-1 654 Scavio, M. J., Jr. Classical-classical transfer: effects of prior aversive conditioning upon appetitive 655 46 conditioning in rabbits (Oryctolagus cuniculus). J Comp Physiol Psychol 86, 107-115 (1974). 656 https://doi.org:10.1037/h0035966 657 Krank, M. D. Asymmetrical effects of Pavlovian excitatory and inhibitory aversive transfer on Pavlovian 47 658 659 appetitive responding and acquisition. Learning and Motivation 16, 35-62 (1985). https://doi.org:10.1016/0023-9690(85)90003-7 660 48 Legaria, A. A. et al. Fiber photometry in striatum reflects primarily nonsomatic changes in calcium. Nat 661 662 Neurosci 25, 1124-1128 (2022). https://doi.org:10.1038/s41593-022-01152-z Fleming, W., Jewell, S., Engelhard, B., Witten, D. M. & Witten, I. B. Inferring spikes from calcium imaging 663 49 in dopamine neurons. PLoS One 16, e0252345 (2021). https://doi.org:10.1371/journal.pone.0252345 664 50 Wakabayashi, K. T. et al. Chemogenetic activation of ventral tegmental area GABA neurons, but not 665 mesoaccumbal GABA terminals, disrupts responding to reward-predictive cues. 666 Neuropsychopharmacology 44, 372-380 (2019). https://doi.org;10.1038/s41386-018-0097-6 667 668 51 Steinberg, E. E. et al. A causal link between prediction errors, dopamine neurons and learning. Nat Neurosci 16, 966-973 (2013). https://doi.org:10.1038/nn.3413 669

52	Glimcher, P. W. Understanding dopamine and reinforcement learning: the dopamine reward prediction
	error hypothesis. Proc Natl Acad Sci U S A 108 Suppl 3 , 15647-15654 (2011).
	https://doi.org:10.1073/pnas.1014269108
53	Eshel, N. et al. Arithmetic and local circuitry underlying dopamine prediction errors. Nature 525, 243-
	246 (2015). <u>https://doi.org:10.1038/nature14855</u>
54	Zhou, W. L. et al. Activity of a direct VTA to ventral pallidum GABA pathway encodes unconditioned
	reward value and sustains motivation for reward. Sci Adv 8, eabm5217 (2022).
	https://doi.org:10.1126/sciadv.abm5217
55	Sharpe, M. J. et al. Dopamine transients are sufficient and necessary for acquisition of model-based
	associations. Nat Neurosci 20, 735-742 (2017). <u>https://doi.org:10.1038/nn.4538</u>
56	Lammel, S., Lim, B. K. & Malenka, R. C. Reward and aversion in a heterogeneous midbrain dopamine
	system. Neuropharmacology 76 Pt B , 351-359 (2014).
	https://doi.org:10.1016/j.neuropharm.2013.03.019
57	Sesack, S. R. & Pickel, V. M. Prefrontal cortical efferents in the rat synapse on unlabeled neuronal
	targets of catecholamine terminals in the nucleus accumbens septi and on dopamine neurons in the
	ventral tegmental area. J Comp Neurol 320 , 145-160 (1992). <u>https://doi.org:10.1002/cne.903200202</u>
58	Lodge, D. J. The medial prefrontal and orbitofrontal cortices differentially regulate dopamine system
	function. Neuropsychopharmacology 36 , 1227-1236 (2011). <u>https://doi.org:10.1038/npp.2011.7</u>
59	Watabe-Uchida, M., Zhu, L., Ogawa, S. K., Vamanrao, A. & Uchida, N. Whole-brain mapping of direct
	inputs to midbrain dopamine neurons. Neuron 74, 858-873 (2012).
	https://doi.org:10.1016/j.neuron.2012.03.017
60	Xia, Y. et al. Nucleus accumbens medium spiny neurons target non-dopaminergic neurons in the ventral
	tegmental area. <i>J Neurosci</i> 31 , 7811-7816 (2011). <u>https://doi.org:10.1523/JNEUROSCI.1504-11.2011</u>
	52 53 54 55 56 57 58 59 60