

Review Article

Pattern Recognition Receptor-Mediated Chronic Inflammation in the Development and Progression of Obesity-Related Metabolic Diseases

Lili Yu ^{1,2}, Yanhua Li,^{1,2} Cancan Du,^{1,2} Weidong Zhao,³ Hanxiao Zhang,³ Yun Yang ¹,
Aiping Sun,^{1,2} Xiangfeng Song ^{1,2} and Zhiwei Feng ^{1,2}

¹School of Basic Medical Sciences, Xinxiang Medical University, Xinxiang 453003, China

²Institute of Precision Medicine, Xinxiang Medical University, Xinxiang 453003, China

³School of Laboratory Medicine, Xinxiang Medical University, Xinxiang 453003, China

Correspondence should be addressed to Lili Yu; merrys222@126.com and Zhiwei Feng; 123066@xxmu.edu.cn

Received 20 April 2019; Accepted 13 August 2019; Published 8 September 2019

Academic Editor: Olivia Osborn

Copyright © 2019 Lili Yu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Obesity-induced chronic inflammation is known to promote the development of many metabolic diseases, especially insulin resistance, type 2 diabetes mellitus, nonalcoholic fatty liver disease, and atherosclerosis. Pattern recognition receptor-mediated inflammation is an important determinant for the initiation and progression of these metabolic diseases. Here, we review the major features of the current understanding with respect to obesity-related chronic inflammation in metabolic tissues, focus on Toll-like receptors and nucleotide-binding oligomerization domain-like receptors with an emphasis on how these receptors determine metabolic disease progression, and provide a summary on the development and progress of PRR antagonists for therapeutic intervention.

1. Introduction

Dramatic rises in obesity and related disorders have occurred globally in just decades, sparing no age groups. The prevalence of obesity among adults has tremendously increased in the past four decades. Approximately 2 billion adults worldwide are overweight, and among them, more than half a billion are obese (World Health Organization, 2018). The development of obesity is closely associated with serious metabolic disorders, such as insulin resistance, type 2 diabetes mellitus (T2DM), hepatic steatosis, and cardiovascular disease, which cause significant morbidity and mortality worldwide [1]. These kinds of chronic diseases are an enormous burden for individuals, families, and societies, as the quality of life is ruined and treatment requires considerable financial supplies and health care resources. The mechanism underlying the pathogenesis of these diseases should be determined, and an effective approach for relieving and healing should be developed.

Accumulating evidence indicates that obesity is associated with chronic low-grade inflammation, which is the key point in the initiation and progression of obesity-related metabolic diseases, especially insulin resistance, T2DM, nonalcoholic fatty liver disease (NAFLD), and atherosclerosis [2–5]. Inflammation occurs due to obesity, and considerable studies have shown that it may play a decisive role in homeostasis [6, 7]. Thus, the influence of obesity-related inflammation in the initiation and regulation of these diseases is a matter of significance [8]. The immune and metabolic systems are closely integrated and complementary [9–11]. The innate immune system constructs the first line of defense to detect and sense the majority of components elicited by infection and endogenous molecules. Thus, excessive metabolic proteins and metabolites associated with obesity can be recognized by innate pattern recognition receptors (PRRs) [12].

Several PRR subfamilies, such as Toll-like receptors (TLRs), retinoic acid-inducible gene I-like receptors, nucleotide-binding oligomerization domain- (NOD-) like

receptors (NLRs), C-type lectin receptors, and DNA sensors have been identified [13]. PRRs recognize pathogen-associated molecular patterns (PAMPs) induced by gut microbiota and infection and danger-associated molecular patterns (DAMPs) caused by metabolic stress or tissue damage to activate innate immune responses and lead to the expression of diverse arrays of downstream signaling pathways [14, 15]. TLRs and NLRs are the two most characterized and described innate receptors in the progression of metabolic diseases, which induce downstream intracellular signaling cascades to produce inflammatory molecules such as cytokines, chemokines, and costimulatory molecules. TLRs predominantly recognize the extracellular or endosomal compartments, whereas NLRs sense invading intracellular pathogens and perturbations associated with stress or damage [16, 17].

Sustained activation and uncontrolled regulation of PRR-mediated innate immune responses can lead to chronic inflammation, which promote the development and progression of many chronic diseases. Genetic, biochemical, and clinical studies have indicated the close link between PRRs and the risk of many chronic diseases. This review summarizes and discusses the recent advancements in understanding the role of PRRs and their downstream signals in the pathogenesis of prevalent obesity-associated diseases.

2. Obesity-Induced Chronic Inflammation in Metabolic Tissues

Multiple PRRs have been implicated in the recognition of metabolic stress and initiation of inflammatory responses in various tissues, which contribute to the development of metabolic diseases [18, 19]. Metabolic syndrome-associated chronic inflammation is related to multiple organs and tissues, including adipose tissues, pancreas, liver, muscle, blood vessel, hypothalamus, and gastrointestinal tract (Figure 1).

2.1. Adipose Tissue. Adipose tissue inflammation is considered a crucial event that leads to metabolic disease. The first hint is the increased expression and production of tumor necrosis factor- (TNF-) α in adipose tissue of obese individuals and its direct role in obesity-induced insulin resistance [20]. However, TNF- α antagonism does not show a significant improvement on insulin sensitivity in patients with metabolic syndrome or T2DM [21]. Accumulated data have confirmed other upregulations of proinflammatory cytokines and chemokines in enlarged adipose tissue sites of obese individuals and those with T2DM, such as interleukin- (IL-) 1β , IL-6, and monocyte chemoattractant protein- (MCP-) 1. One study suggests that galectin-3-mediated inflammation induces insulin resistance [22].

Adipose tissues normally contain multiple immune cells that together maintain the integrity and metabolism of adipocytes [23]. Under normal conditions, immune cells in adipose tissues operate in an anti-inflammatory state and mainly contain M2-resident macrophages, CD4⁺ T helper (Th) 2, and CD4⁺ regulatory T (Treg) cells, which secrete anti-inflammatory IL-10 and Th2 cytokines. In obesity, adipose tissues are associated with a marked accumulation

of proinflammatory immune cells, with M1-polarized macrophages having the most abundant population. Meanwhile, the number of CD4⁺ Th1 cells and Th17 cells is increased, which secrete interferon- (IFN-) γ and IL-17 as major proinflammatory cytokines in obese adipose tissues, whereas the number of Treg and Th2 cells is decreased [24]. In addition, the infiltration of CD8⁺ T cells and neutrophils is assumed to promote the recruitment and activation of macrophages, and B cells are implicated in producing pathogenic antibodies and proinflammatory cytokines contributing to chronic low-grade inflammation in obesity [25].

However, the molecular events and precise triggers of obesity-associated inflammation in adipose tissue are still not fully clear. One potential mechanism is that underlying elevated lipid metabolites can trigger the activation of PRRs, including saturated long-chain fatty acids (SFAs), ceramides, and modified low-density lipoproteins (LDLs) [1]. PRRs can also be triggered by the production of other endogenous damage-associated signals during the development of metabolic diseases, such as high mobility group box 1 (HMGB1) and fetuin A [16].

2.2. Pancreas. Inflammation in pancreatic islets contributes to β -cell dysfunction and apoptosis, which results in reduced insulin secretion [26]. As in adipose tissue, macrophages infiltrate and accumulate in pancreatic islets of obese T2DM individuals and diet-induced obesity mice, which produce proinflammatory cytokines and chemokines, such as TNF- α , IL- 1β , and MCP1 [27]. The most well-studied mechanism by which islet macrophages cause β -cell dysfunction is through the secretion of IL- 1β , and interference with the IL-1 pathway has been demonstrated to relieve T2DM and restore β -cell function in humans [28, 29]. A recent study indicated that two islet-resident macrophage populations, namely, intra- and peri-islet macrophages, exist in mice pancreas, and obesity induces the local expansion of resident intraislet macrophages, independent of recruitment from circulating monocytes [30]. Functionally, intraislet macrophages impair β -cell function in a cell-cell contact-dependent manner [30]. PRRs have also been reported in the process of pancreas inflammation. Islet amyloid polypeptide (IAPP), a protein that forms amyloid deposits in the pancreas during T2DM, can activate nucleotide-binding and oligomerization domain (NACHT), leucine-rich repeat (LRR), and pyrin domain- (PYD-) containing protein 3 (NLRP3) in causing infiltrated macrophages, generating IL- 1β , and reducing β -cell mass [31].

2.3. Liver. Hepatic steatosis is a common feature in obesity that can result in NAFLD and ultimately progress to nonalcoholic steatohepatitis (NASH) and fibrosis. The inflammatory cytokines derived from immune cells in the liver can potentiate steatosis. Two major kinds of macrophages, namely, Kupffer cells (KCs) and recruited hepatic macrophages (RHMs), are found in the liver. KCs are resident macrophages and are important for hepatic inflammatory states [32]. During obesity development, steatotic hepatocytes and KCs release chemokines such as MCP1 to recruit circulating monocytes. RHMs are abundantly accumulated

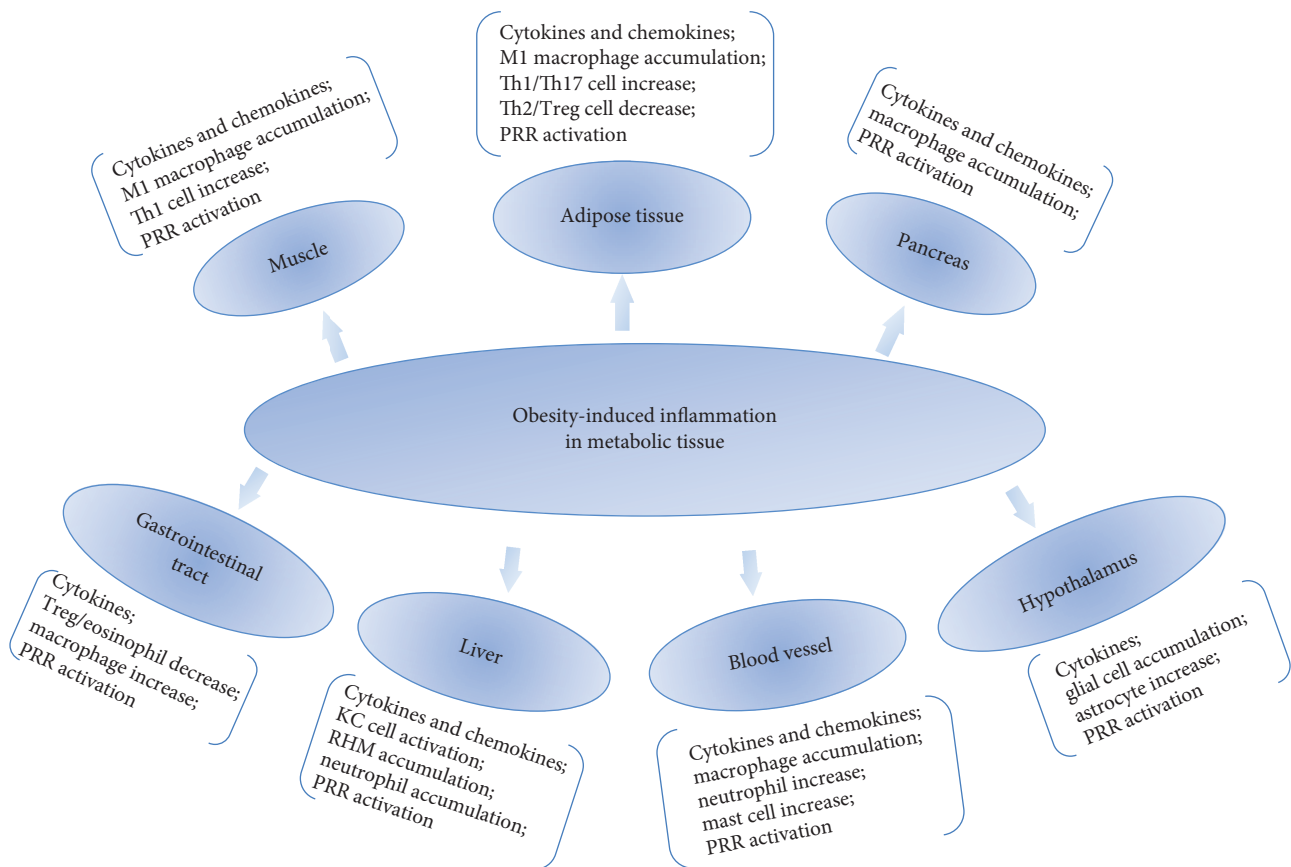


FIGURE 1: Obesity-induced chronic tissue inflammation states in metabolic tissues. Chronic tissue inflammation induces a range of effects on adipose tissue, muscle, liver, pancreas, gastrointestinal tract, blood vessel, and hypothalamus. These inflammatory changes include the secretion of cytokines and chemokines, infiltration of immune cells, and activation of PRRs, which are the key points in the initiation and progression of obesity-related metabolic diseases.

in the liver, and they are highly proinflammatory and characteristically express various cytokines, such as $\text{TNF-}\alpha$, $\text{IL-1}\beta$, and IL-6 . One study indicated that chemotactic eicosanoid LTB_4 promotes insulin resistance in obese mice by acting on macrophages, hepatocytes, and myocytes [33]. Similarly, neutrophils accumulate in the liver during obesity and release a similar set of cytokines and chemokines contributing to the overall inflammatory process. A combination of hepatic insults is proposed to drive hepatic steatosis pathogenesis and activate various innate immune pathways. TLRs and NLRs are widely expressed in multiple hepatic cells and KCs. Intrahepatic activation of PRR signaling by endogenous and pathogen-derived ligands plays a critical role in the development of hepatic steatosis and NAFLD [34].

2.4. Muscle. Skeletal muscle (SM) insulin resistance plays a key role in metabolic dysfunction. One possible etiologic factor that causes decreased muscle insulin sensitivity is inflammation. Numerous studies have demonstrated the increase of inflammatory markers in the SM of obese and diabetic individuals, which includes the activation of proinflammatory cytokines in muscle cells and circulating cytokines derived from other tissues. These muscle-derived mediators may also contribute to low-grade systemic inflammation with obesity. The first cytokine produced by the

muscle fiber is IL-6 , which is reported to accumulate in the bloodstream in response to muscle contraction and is shown to improve insulin sensitivity. However, long-term and chronic exposures to IL-6 during obesity can cause insulin resistance and inflammation in muscle tissues [35].

Similar to adipose tissue, resident macrophages are located in the epimysium and perimysium of SM. In obesity, many proinflammatory immune cells, including M1 macrophages and Th1 cells, infiltrate and accumulate in muscle tissues, which secrete proinflammatory cytokines, such as $\text{TNF-}\alpha$, $\text{IL-1}\beta$, and MCP1 [36]. This infiltration may be consecutive to chemokine release by resident macrophages and/or myocytes themselves in response to PRR-mediated recognition of PAMPs or DAMPs. Similar to adipose tissues, SM tissue is sensitive to lipopolysaccharide (LPS) and circulating SFAs, inducing the production of cytokines $\text{TNF-}\alpha$, IL-6 , and chemokines depending on PRRs [37].

2.5. Blood Vessel. Atherosclerosis is a chronic inflammatory disease of the blood vessels; it is a common cardiovascular disease associated with lipid accumulation and plaque formation in the blood vessel wall [23]. Atherosclerosis is promoted through obesity-associated hyperlipidemia and inflammation. At the initiation of atherosclerosis, circulating LDL particles are detained by subendothelial cells and oxidized

by oxygen radicals and enzymatic attacks. Then, the modified LDLs activate innate immune receptors and trigger inflammatory cascades to induce the expression of inflammatory cytokines, chemokines, and costimulatory molecules. Inflammation cytokines such as TNF- α , IL-1 β , and IL-18 are elevated in atherosclerosis and associated with disease severity. Monocytes, dendritic cells, and T cells are recruited into the intima by these chemokines. Many monocytes are differentiated into macrophages and eventually are transformed into foam cells as cholesterol accumulation. Neutrophils and mast cells are also recruited to atherosclerotic lesions, which produce proatherogenic mediators that contribute to lesion growth and disease progression [38]. Various PRRs are expressed in arterial lesions and activated by oxidized LDL particles, fibronectin, cholesterol crystals, and calcium crystals during atherosclerosis progression [39].

2.6. Hypothalamus. The hypothalamus plays an important role in the homeostatic regulation of body weight through energy intake and expenditures. Increased hypothalamic glial cell and/or astrocyte accumulation occurs during obesity [40]. Studies have confirmed that the expression of inflammatory cytokines is increased through PRRs in obesity, which is associated with hypothalamus insulin resistance. In particular, clinical and experimental studies have identified resistin as a key hormone linking obesity-induced hypothalamic inflammation and insulin resistance through the activation of TLR4 signaling pathways [41].

2.7. Gastrointestinal Tract. The development of obesity and related metabolic diseases is associated with altered gastrointestinal microbiota composition [42]. Microbiota can produce metabolites and byproducts to induce proinflammatory cytokines and modulate metabolism in the development of obesity, tissue inflammation, and metabolic dysfunction. Microbiota transplantation from obese humans into lean recipients leads to an increase in systemic insulin resistance [43]. Mice with high-fat-diet- (HFD-) induced obesity display altered immune cell populations in the intestinal lamina propria, including a reduced proportion of Treg cells and eosinophils and elevated macrophages [44]. Obesity could induce increased gastrointestinal permeability and endotoxemia, resulting in high levels of circulating metabolites and byproducts produced by microbiota [45]. Microbes and their products contain PAMPs, which are recognized by the host innate immune system through PRRs, including LPS, bacterial DNA, and peptidoglycan (PGN).

3. TLRs and Obesity-Related Metabolic Diseases

TLRs execute an irreplaceable role in controlling the inflammation of different metabolic organs in obesity-induced metabolic diseases. As widely expressed and highly conserved transmembrane receptors, TLRs are at the intersection of diet and metabolism and may therefore be important determinants of obesity and metabolic diseases [18, 46]. TLRs are type I transmembrane domain proteins with a tripartite structure: an amino- (N-) terminal extracellular domain as ligand recognition domains, a signal transmem-

brane spanning region, and a carboxyl- (C-) terminal cytoplasmic signaling domain [47]. Broadly, TLRs use two main adaptors, namely, myeloid differentiation primary response gene 88 (Myd88) and TIR-domain-containing adaptor-inducing IFN- β (TRIF), to mediate the downstream signaling pathways.

With the exception of TLR3, all TLRs require the engagement of Myd88 to induce the activation of NF- κ B, activator protein-1, mitogen-activated protein kinases (MAPKs), and the IFN regulatory factor- (IRF-) 7 pathway, which lead to the expression of cytokines and IFNs. In addition, TLR3 and TLR4 engage TRIF to trigger a specific signaling cascade that results in the activation of NF- κ B, MAPKs, and IRF3 to induce the production of proinflammatory cytokines and type I IFNs [48]. To date, at least 13 TLRs have been identified in humans and mice. Multiple TLRs, including TLR2, TLR3, TLR4, TLR5, TLR8, TLR9, and TLR10, have been indicated in the regulation of PAMPs from gut microbiota and DAMPs from metabolic stress and tissue damage, which initiate the inflammatory responses contributing to the development of obesity-related chronic diseases (Figure 2).

3.1. TLR2. TLR2 is located at the surface of the cell and forms dimers with other TLRs to recognize the largest range of ligands including free FAs (FFAs), lipopeptides, lipoteichoic acids, lipids, porins, lipoarabinomannan, zymosan, and lipomannan from bacteria and fungi [47]. Elevated plasma FFA levels can account for a large part of insulin resistance in obese patients with T2DM [49]. An in vitro study of human cells showed that TLR2 mediates FFA-induced proinflammatory responses in cultured adipocytes and monocytes/macrophages [50–52].

Human cross-sectional studies found that TLR2 activation and expression are enhanced in adipose tissue and peripheral monocytes of obese subjects compared with lean control [53]. Other studies showed that TLR2 expression is increased in patients with diabetes [54], and TLR2/TLR4 stimulation induces an enhanced inflammation in obese patients with atherosclerosis [55]. Although TLR2 is elevated in obese patients compared with healthy controls, it is unaltered in patients with limited liver disease compared with obesity-related NASH in two clinical studies [56, 57]. In addition, a clinical intervention showed that a one-week HFD leads to reduced TLR2 expression in young healthy men [58]. This discrepancy may be due to the short period of HFD intervention, and further clinical trials are required to confirm this finding.

TLR2 has been implicated in the pathogenesis of obesity and insulin resistance in dietary mouse models. The expression of TLR2 and TNF- α was increased in visceral adipose tissue by a HFD as a development of insulin resistance [59]. In a HFD-induced mouse model, macrophages infiltrate hypertrophic adipose tissue and are activated by FFAs via the TLR2 and JNK pathways [60]. In loss-of-function animal studies, TLR2-deficient mice fed with a HFD are associated with the suppression of adipocyte hypertrophy, decreased macrophage infiltration and inflammatory cytokine expression in adipose tissue, and reduced expression of inflammatory cytokines in the liver and pancreas [61–63]. In

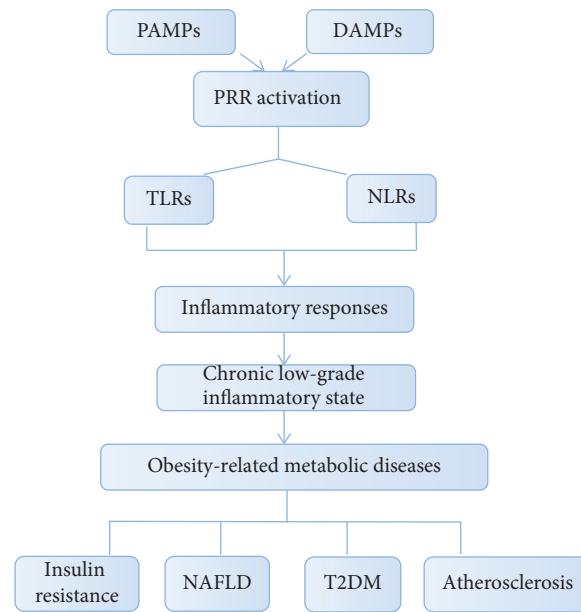


FIGURE 2: Overview of PRRs in linking inflammation. As important players of the innate immune system, PRRs sense DAMPs from metabolic stress and/or tissue damage and PAMPs from gut microbiota and/or infection. Activation of PRRs induces a series of inflammatory responses and systemically mediates a chronic, low-grade inflammation, which promotes the pathogenesis of obesity-related metabolic diseases, such as insulin resistance, NAFLD, T2DM, and atherosclerosis.

addition, TLR2-deficient mice are protected from HFD-induced adiposity, insulin resistance, impaired insulin secretion, and hypercholesterolemia [62, 64]. TLR2-deficient mice are also protected from choline-deficient L-amino-acid-defined diet-induced liver inflammation and diet-induced NASH [65]. Furthermore, TLR2 deficiency in apolipoprotein E- (ApoE-) deficient and LDL receptor- (LDLR-) deficient mice reduces atherosclerotic plaque formation in the progression of atherosclerosis [66, 67].

However, another study indicated that TLR2-deficient mice are protected from HFD-induced insulin resistance under germ-free conditions, whereas TLR2-knockout mice show a phenotype of metabolic syndrome under conventional conditions due to the specific role of gut microbiota and increased LPS absorption [68]. TLR2 deficiency in mice is conventionally associated with an increased abundance of *Firmicutes* and a decreased abundance of *Bacteroidetes* compared with controls [68]. By contrast, another group found that the impact of TLR deficiency on the composition of gut microbiota is minimal under homeostatic conditions [69]. In addition, TLR2-deficient mice are not protected from methionine- and choline-deficient diet- (MCDD-) induced steatohepatitis [70]. This condition is also related to gut microbiota. MCDD increases the amount LPS-producing bacteria, and increased LPS plasma levels could be due to the activation of TLR4 in TLR2-deficient mice. Further studies are needed to elucidate the effect between the TLR2 signaling pathway and gut microbiota.

In a therapeutic animal study, TLR2 suppression using an antisense oligonucleotide improves insulin sensitivity and signaling in muscle and white adipose tissue of mice fed with a HFD [71]. Blockade of TLR2 in intact aortas using an anti-TLR2 antibody attenuates an increase in vascular contraction

in rats with streptozotocin-induced diabetes [72]. Another small molecule, $C_{16}H_{15}NO_4$ (C29), was identified as a potential TLR2 inhibitor, which reduced TLR2-induced inflammation [73]. Taken together, although there are some uncertain results, TLR2 inhibition is beneficial to suppress sustained inflammation.

3.2. TLR3. TLR3 plays a crucial role in survival by recognizing various viral components, such as double-stranded RNA (dsRNA) and poly(I:C), which is a synthetic dsRNA polymer [74]. Upon recognition, TLR3 uses TRIF as an adaptor to induce the activation of IRF3 and increase the production of type I IFNs for increasing cellular antiviral defenses.

Studies of adipocytes show that TLR3 is present in human adipose-derived stem cells [75] and mature adipocytes [76]. Similarly, the expression of TLR3 is significantly higher than that of the nonadipocytes in the adipose tissue of mice [77]. In a human cross-sectional study, the expression of TLR3 was decreased in obese patients compared with nonobese subjects, and decreased TLR3 was observed in hyperplastic adipose tissue, blood, and inflamed adipocytes [78]. Although TLR3 is highly expressed in adipocytes, the contribution of TLR3 to insulin resistance is enigmatic. In an *in vitro* study of human cells, defects in TLR3 expression and latent endoribonuclease activation lead to decreased mitochondrial manganese superoxide dismutase expression and insulin resistance in muscle cells of obese individuals [79]. However, in a correlational study of humans, TLR3 expression in adipose tissue from 80 healthy donors did not correlate with BMI or insulin sensitivity (HOMA-IR) [76].

Few studies have indicated the importance of TLR3 in regulating glucose metabolic responses and insulin levels in

rodents. In loss-of-function animal models, although TLR3-deficient mice fed with a HFD developed obesity, they exhibited improved glucose tolerance and reduced liver steatosis compared with wild-type (WT) obese mice [80]. Conversely, another study indicated that TLR3-deficient mice with HFD-induced obesity exhibit increased glucose tolerance and serum insulin and decreased serum levels of very low-density lipoproteins and triglycerides (TG) [81]. However, another study showed that TLR3 deficiency in mice does not significantly influence HFD-induced obesity and system insulin sensitivity or inflammation [76]. The impact of TLR3 on metabolic regulation remains unclear. TLR3 appears to play a redundant role in obesity-induced inflammation and insulin resistance, which should be investigated.

3.3. TLR4. TLR4 is the most widely described PRR in obesity-induced chronic inflammation and development of metabolic diseases. TLR4 is broadly expressed in human adipocytes, hepatocytes, and monocytes [82–84]. TLR4 responds to ligands such as LPS and initiates its response by forming a complex with MD-2, leading to the activation of the Myd88 and TRIF pathways. Similar to TLR2, TLR4 signaling leads to the production of proinflammatory cytokines, including TNF- α , IL-1, and IL-6. Studies of human cells showed that SFAs, including lauric acid, palmitic acid, and stearic acid, activate TLR4 in adipocytes and macrophages in vitro [85]. HMGB1 has been shown to mediate TLR4 activation in human adipocytes, which are also elevated in obesity [82, 86].

Human cross-sectional studies showed that TLR4 expression is increased in adipose tissue and monocytes of obese or diabetic patients, which is correlated with the severity of insulin resistance [53, 87, 88]. Individuals with T2DM show increased cellular membrane levels of TLR4 in monocytes and higher serum concentrations of IL-1 β , IL-6, IL-8, and TNF- α compared with controls [53]. In another cross-sectional study, TLR4 mRNA levels were significantly increased in subjects with NASH compared with obese and NAFLD patients; this occurs within the setting of increased LPS and FAs [83]. A previous study also indicated that TLR4 is a new target for chemoprevention of hepatocellular carcinoma in obesity and steatohepatitis [89]. Human cross-sectional studies showed that TLR4 is involved in monocyte activation of patients with accelerated forms of atherosclerosis, and the upregulation of TLR4 on human monocyte subsets is closely associated with coronary plaque vulnerability in patients [90, 91].

In loss-of-function mouse models, TLR4-deficient mice are protected from diet-induced obesity due to the decrease in proinflammatory cytokine levels [92, 93]. TLR4-deficient mice fed with a HFD exhibit improved insulin resistance, reduced tissue inflammation, and decreased M1 macrophage infiltration [93–95]. Hepatocyte-specific TLR4 deficient mice are also protected from obesity and insulin resistance [96]. In this mouse model, inflammatory cell infiltration was suppressed in the liver and adipose tissue, which indicates that hepatocyte TLR4 signaling produces chemokines to recruit inflammatory cell infiltration. However, macrophage-specific TLR4-deficient mice are not protected from obesity

or insulin resistance induced by a HFD [96]. These results suggest that TLR4 expression in hepatocytes plays a decisive role in the initiation of inflammatory responses. In other mouse models, TLR4 deficiency in atherosclerosis-prone ApoE-deficient mice fed a HFD showed significant reduction of aortic plaque areas, plaque lipid content, macrophage infiltration, and circulating levels of proinflammatory cytokines [97]. However, in another study, TLR4 deficiency decreased atherosclerosis but did not protect against adipocyte hypertrophy and macrophage accumulation in obese LDLR-deficient mice [98]. TLR4 also plays a central role in NASH and liver fibrosis in NAFLD. Deficiency in TLR4 expression attenuates NASH and fibrosis in mice [99]. Moreover, TLR4 deficiency in LDLR-knockout mice prevents TG accumulation and enhances FA oxidation to alter the onset of NAFLD after a high-fat/high-cholesterol diet [100]. All these results suggest that TLR4 plays an irreplaceable role in regulating obesity-related metabolic diseases.

Enhanced TLR4 signaling is increased in muscle, visceral fat tissue, and liver under obesity, which is responsible for obesity inflammation, obesity-related insulin resistance, and liver diseases [101]. Increased expression of TLR4 enhances NF- κ B activation and subsequently increases the release of IL-6 and TNF- α . In particular, released FAs by hypertrophied adipocytes activate TLR4 signaling pathways to induce inflammatory cytokines and chemokines to induce cell dysfunction in adipose tissues [102, 103]. Circulating levels of FAs are significantly higher in obese compared with lean individuals, which results in increased TLR4 signaling in obesity and inflammation [101, 103]. In obesity, the interactions between adipocytes and macrophages aggravate adipose inflammation through TLR4 [104]. M1-type macrophages highly express TLR4 and infiltrate into the target metabolic organs of obesity. By contrast, M2-type macrophages, which show a low TLR4 expression, are dominant in lean subjects. In addition, circulating levels of LPS are moderately increased in patients with obesity, T2DM, metabolic syndrome, and NAFLD [105, 106]. Increased LPS in plasma could be due to increased intestinal permeability and enhanced absorption through TLR4 by HFD [107].

In therapeutic animal studies, TAK-242 (a TLR4 inhibitor) attenuates insulin resistance in muscle cells and the adverse neural effects of diet-induced obesity [108, 109]. In an acute experiment, rats that received a TLR4 inhibitor (TAK-242 or E5564) obtain partial protection against acute and chronic fat-induced insulin resistance [110]. Berberine may reduce insulin resistance, at least in part, by modulating the gut microbiota along with inhibiting LPS-TLR4 signaling in the liver [111].

3.4. TLR5. TLR5 recognizes flagellin, the primary structural component of flagella, and triggers inflammatory response through MyD88. TLR5 is expressed in the intestinal mucosa against metabolic diseases, which strongly influences the composition of the microbiota throughout life [48, 112]. In loss-of-function mouse models, TLR5-deficient mice show altered gut microbiota composition, low-grade inflammation, metabolic syndrome, and predisposition to the development of colitis [113]. In mice, epithelial TLR5-mediated

REG3 γ production is critical for the counterselection of colonizing flagella [112]. TLR5-deficient mice fed a HFD also exhibit exacerbated metabolic syndromes, including hyperlipidemia, insulin resistance, and increased adiposity [114]. In addition, transferring gut microbiota from TLR5-deficient mice to WT germ-free mice confers the features of metabolic syndrome to recipients [115]. Metabolic syndrome in TLR5-deficient mice is exacerbated by HFD, suggesting that TLR5 is responsible for the balance of microbiota.

3.5. TLR8. There is little information about TLR8 in obesity-mediated inflammation. One study indicated that TLR8 expression in adipose tissue of nondiabetic/T2DM obese individuals is higher than that of the overweight/lean population. The elevated levels of TLR8 in obesity/T2DM are consistent with the increase of inflammatory mediators and may represent an immune marker of metabolic inflammation [116].

3.6. TLR9. TLR9 recognizes viral and bacterial DNA (unmethylated CpG motifs) and activates signaling through MyD88. In addition, cell-free DNA from obesity-related adipocyte degeneration can activate TLR9 [117]. A clinical association study indicated that bacterial DNA translocation to the blood is associated with metabolic dysfunction and the onset of diabetes [118]. In humans, plasma ssDNA levels are significantly high in patients with computed tomography-determined visceral obesity and are associated with homeostasis model assessment of insulin resistance [117]. The mitochondrial DNA released from hepatocytes serves as a TLR9 ligand and promotes steatohepatitis. A clinical study indicated that the mtDNA copy number is 4.3- and 3.2-fold higher in nonalcoholic fatty liver and NASH patients than in healthy controls, respectively [119].

In a loss-of-function animal study, mitochondrial DNA released by adipocytes under obesity-induced excessive stress stimulates TLR9 in the immune cells and induces inflammation in adipose tissue [120]. HFD-fed TLR9-deficient mice demonstrated reduced macrophage accumulation and inflammation in adipose tissue and better insulin sensitivity compared with WT mice, whereas bone marrow reconstitution with WT bone marrow restored the attenuation of insulin resistance observed in HFD-fed TLR9-deficient mice [117]. TLR9-deficient mice were protected against steatohepatitis due to choline-amino acid depletion and a HFD [121]. In mice, NASH development in response to a HFD required TLR9 on lysozyme-expressing cells [122].

Administration of a TLR9 inhibitory oligonucleotide to fat-fed WT mice reduced the accumulation of macrophages in adipose tissue and improved insulin resistance [117]. In mice, a TLR7/9 antagonist (IRS954) was effective for improving hepatic steatosis and NAFLD [122]. TLR9 is a target of treatments for obesity and NAFLD.

3.7. TLR10. A recent study indicated that TLR10 impacts adipose tissue morphology in obesity. In humans, obese individuals with polymorphisms in the TLR10 gene displayed reduced macrophage infiltration in the adipose tissue accompanied by low leptin levels and high adiponectin levels in the

plasma [123]. Transgenic human TLR10 mice displayed reduced adipocyte size, adipose tissue weight, and lower plasma insulin levels compared with WT mice fed with a HFD [123]. More studies need to assess the value of TLR10 as a therapeutic target in obesity and metabolic syndrome.

4. NLRs and Metabolic Diseases

NLR-mediated inflammation caused by gut microbiota-derived PAMPs and metabolic stress-derived DAMPs during obesity induces metabolic disturbances and metabolic diseases (Figure 2). The NLR family contains 23 members [124]. A typical NLR is composed of three distinct common domains: the C-terminal LRR domain involved in the recognition of PAMPs and DAMPs; a centrally located NACHT that enables activation of a signaling complex via ATP-dependent oligomerization; and an N-terminal caspase recruitment domain (CARD) and PYD, which mediate downstream signaling [125].

4.1. NOD1/2. NOD1 and NOD2 are members of the NLR family. They are cytosolic PRRs that play a role in detecting intracellular microorganisms [126]. These receptors detect specific bacterial PGN motifs and initiate an inflammatory signaling cascade involved in host defense, which is a potential factor linking innate immunity and metabolic function [124]. NOD1 and NOD2 recognize their ligands through the LRR domain and NACHT domain oligomer by initiating downstream signaling to activate NF- κ B and phosphorylated MAPK, which mediate the expression of proinflammatory cytokines [124].

In a loss-of-function animal study, NOD1/2-double-deficient mice are protected from HFD-induced insulin intolerance, lipid accumulation, and inflammation in adipose tissue and liver [127]. In addition, PGN can cause acute systemic insulin resistance by activating NOD1, and NOD1-deficient mice have reduced bacterial translocation to metabolic tissues and improved insulin tolerance [128]. Moreover, NOD1 deficiency in ApoE-knockout mice showed reduced development of atherosclerotic lesions from the early stage and delayed progression of atherosclerosis [129]. However, one study showed that NOD2-deficient mice fed with a HFD have increased adipose tissue and liver inflammation and exacerbated insulin resistance [130]. This condition may be because NOD2-deficient mice have an impaired gut barrier function.

4.2. NLRP3 Inflammasome. As cytoplasmic complexes, inflammasomes recognize microbial and endogenous ligands and recruit procaspase-1 directly through interacting with CARD domains or indirectly through apoptosis-associated speck-like protein containing CARD (ASC), resulting in the formation of active caspase-1 and triggering the maturation of proinflammatory cytokines, such as IL-1 β and IL-18 [125]. The NLRP3 inflammasome is activated by PAMPs from microbial pathogens, such as cell wall proteoglycans, pore-forming toxins, and DNA/RNA, and by DAMPs, such as crystals, ATP, hyaluronan, β -amyloid, and monosodium urate crystals. Activation of the NLRP3 inflammasome

requires two steps: the first step is an NF- κ B-activating stimulus for cells to express pro-IL-1 β and optimal NLRP3 and the second step is the NLRP3 inflammasome that results in caspase-1 activation [131].

Several molecules have been identified as DAMPs involved in NLRP3 activation in HFD-induced obesity. SFAs, such as palmitate and ceramides, can stimulate NLRP3 inflammasome activation to induce inflammation and obesity-mediated T2DM [132]. As an inducer of NLRP3 activation, IAPP is also accumulated in the islet of T2DM patients [31]. In addition, endocannabinoids are found to induce NLRP3 inflammasome-dependent IL-1 β production by pancreatic infiltrating macrophages, resulting in pancreatic β -cell death [133]. Moreover, cholesterol crystals and oxidized LDL particles have been shown to activate the NLRP3 inflammasome, which leads to a vascular response during atherosclerosis progression [134, 135].

Human cross-sectional studies showed an increased secretion of IL-1 β , increased expression of NLRP3, increased number of adipose tissue macrophages, and decreased number of regulatory T cells in the visceral adipose tissue of metabolically unhealthy patients compared with metabolically healthy and lean participants [136]. T2DM subjects have significantly increased mRNA and protein expression of NLRP3, ASC, and proinflammatory cytokines in monocyte-derived macrophages compared with healthy controls [137]. Weight loss in obese patients with T2DM is associated with insulin sensitivity and reduction of NLRP3 and IL-1 β expression in adipose tissue [138]. Moreover, NLRP3 inflammasomes show high expression in the aorta of patients with atherosclerosis [139].

Animal studies provide a direct link among the NLRP3 inflammasome, chronic inflammation, and insulin resistance. A loss-of-function animal study showed that NLRP3-, ASC-, and caspase-1-deficient mice are protected from HFD-induced obesity and show improved glucose tolerance and insulin sensitivity [138, 140, 141]. NLRP3-deficient mice have reduced expression in visceral fat of markers of M1-type macrophage activation and an increase in markers of M2-type macrophage activation [138]. Gene silencing of NLRP3 suppresses atherosclerosis and stabilizes plaques in ApoE-deficient mice [142]. Liraglutide protects NAFLD via inhibiting NLRP3 inflammasome activation in a mouse model induced by HFD [143]. In addition, caspase-1 contributes to adipose tissue formation because mice lacking caspase-1 have reduced adipocyte size, reduced fat mass, increased adipogenic gene expression, and improved insulin sensitivity and gained less weight than WT controls [144]. In particular, IL-1 β was strongly linked to the pathogenesis of T2DM by promoting insulin resistance and causing β -cell functional impairment and apoptosis [131]. Blockade of IL-1 β inhibited atherosclerotic plaque formation in ApoE-deficient mice [145]. Similarly, elevated IL-18 levels have been shown to cause vascular inflammation and enhance the instability of atherosclerotic plaques, whereas IL-18-deficiency in ApoE-deficient mice results in reduced atherosclerotic lesion size [146].

A therapeutic animal study showed that MCC950, an NLRP3 selective inhibitor, improves NAFLD pathology

and fibrosis in obese diabetic mice and reduces liver fibrosis in MCD-fed mice [147]. A therapeutic human study found that glyburide, a small-molecule inhibitor used for T2DM treatment, can inhibit NLRP3-dependent IL-1 β production [148]. In addition, the IL-1 receptor antagonist (anakinra) or IL-1 β antagonism (gevokizumab, canakizumab, and LY2189102) can effectively control glucose levels and β -cell function in T2DM patients [18].

4.3. NLRP1 Inflammasome. NLRP1 does not require ASC to activate caspase-1, which recruits procaspase-1 by directly interacting with CARD domains to trigger the maturation of IL-1 β and IL-18. Toxoplasma infection and anthrax lethal toxin have been reported to activate NLRP1. NLRP1 functions in the context of metabolic stress to produce IL-18, preventing obesity and metabolic syndrome. NLRP1-deficient mice spontaneously develop a large increase in adipose tissue, which exacerbated by high-fat feeding and decreased IL-18 levels results in increased adiposity and resistance to diet-induced metabolic dysfunction [149].

4.4. NLRP6 Inflammasome. NLRP6 requires ASC to recruit procaspase-1 to activate caspase-1 and triggers the maturation of IL-1 β and IL-18. NLRP6 plays an important function in controlling the composition of the gut microbiota in metabolic diseases [150]. Colonic microbes trigger NLRP6-mediated secretion of IL-18, which induce the expression of antimicrobial peptides that control the colonic microbial community to regulate metabolic diseases [150]. NLRP6-deficient mice are highly resistant to infection with bacterial pathogens [151]. The above results reveal that NLRP6 is a negative regulator of inflammatory signaling.

5. Concluding Remarks

Given the findings mentioned above, whether PRRs are part of the pathogenesis of metabolic disorders or manifestations of other derangements that arise as disorder progresses need to be investigated. The nature of inflammatory responses to obesity is unique, and many organ systems are impacted by inflammation during obesity. In many patients, insulin resistance appears long before obesity; type 1 diabetes appears and persists in the absence of obesity, and its pathogenesis likely involves entirely different immunological disturbances [152]. In this article, we only focus on the roles of PRRs underlying obesity-caused inflammation in the pathogenesis of T2DM and other related metabolic diseases.

TLRs and NLRs have been demonstrated to sense various DAMPs derived from metabolic stress or tissue damage in obesity-associated metabolic disturbances. Obesity leads to various triggering events, such as endoplasmic reticulum stress, hypoxia, and lipotoxicity, which can initiate the activation of proinflammatory pathways within metabolic tissues, including adipose tissue, liver, pancreas, gastrointestinal tract, muscle, hypothalamus, and blood vessels [101]. These receptors play critical roles in bridging immune responses and metabolic homeostasis. Thus, the development of PRR inhibitors shows huge potential as a powerful therapeutic strategy [124, 153, 154]. In general, strategies utilized to

downmodulate the activation of TLRs/NLRs involve the use of antagonists, which block the binding of ligands or ligand complexes to the receptors.

At present, a number of TLR/NLR-related inhibitors have emerged [155, 156]. OPN-305, a TLR2-specific antibody, has completed clinical trials for myelodysplastic syndrome (NCT02363491 and NCT03337451). Eritoran (E5564), an antagonist of TLR4, has been terminated under clinical trials in patients with insulin resistance and T2DM for unknown reasons (NCT02321111 and NCT02267317). A small-molecule-specific inhibitor for TLR4, TAK-242 (Resatorvid), has been terminated under clinical trials in patients with sepsis-induced cardiovascular and respiratory failure due to safety and efficiency concerns (NCT00633477). A humanized monoclonal anti-TLR4 antibody (NI-0101) shows a therapeutic effect under phase-I clinical trials for LPS reaction [157]. The oligonucleotide IMO-3100 antagonizes TLR7/TLR9, and IMO-3100 has completed trials in patients with moderate to severe plaque psoriasis (NCT01622348). IMO-8400 inhibits TLR7/TLR8/TLR9 under phase-II clinical trials in patients with moderate to severe plaque psoriasis [158]. IMO-8400 has completed trials in adult patients with dermatomyositis (NCT02612857) and in patients with Waldenström's macroglobulinemia (NCT02363439). P2X7R antagonists that block the NLRP3 inflammasome are under phase-II clinical trials [159].

Although preclinical studies reported the promising inhibitory effects of some antagonistic ligands on TLR/NLR signaling, there are some challenges associated with these agents. The innate immune system is activated by PRRs and is a critical host "first responder" for exogenous pathogens and endogenous stresses. Chronic suppression may have the potential for extensive complications as implied in chronic metabolic disease. These agents mentioned above have been used in myelodysplastic syndrome, severe acute sepsis, and autoimmune diseases; only Eritoran has been administered for chronic metabolic disease. In fact, Eritoran is no longer in clinical development due to either clinical or nonclinical toxicity or lack of efficacy.

Additional studies are required to provide new insights into the therapeutic targeting of TLRs/NLRs, and the development of new screening methods and identification of drugs with good pharmacology is important. To date, the involvement of TLRs/NLRs in the development of inflammatory diseases continues to be unveiled, which indicates that the overactivation of these receptors causes disruption of homeostasis, increasing the risk for metabolic diseases. Thus, TLR/NLR-driven responses should be tightly regulated to prevent any detrimental effect from their aberrant activation.

Conflicts of Interest

The authors have declared that no competing interests exist.

Acknowledgments

This work was supported by grants from the National Natural Science Foundation of China (Grant Nos. 81500675 and

81671226) and the Natural Science Foundation of Henan Province of China (No. 162300410225).

References

- [1] A. R. Saltiel and J. M. Olefsky, "Inflammatory mechanisms linking obesity and metabolic disease," *The Journal of Clinical Investigation*, vol. 127, no. 1, pp. 1–4, 2017.
- [2] J. Lasselin and L. Capuron, "Chronic low-grade inflammation in metabolic disorders: relevance for behavioral symptoms," *Neuroimmunomodulation*, vol. 21, no. 2-3, pp. 95–101, 2014.
- [3] S. Crunkhorn, "Metabolic disorders: breaking the links between inflammation and diabetes," *Nature Reviews Drug Discovery*, vol. 12, no. 4, p. 261, 2013.
- [4] M. Navab, N. Gharavi, and A. D. Watson, "Inflammation and metabolic disorders," *Current Opinion in Clinical Nutrition and Metabolic Care*, vol. 11, no. 4, pp. 459–464, 2008.
- [5] G. S. Hotamisligil, "Inflammation and metabolic disorders," *Nature*, vol. 444, no. 7121, pp. 860–867, 2006.
- [6] G. S. Hotamisligil and E. Erbay, "Nutrient sensing and inflammation in metabolic diseases," *Nature Reviews Immunology*, vol. 8, no. 12, pp. 923–934, 2008.
- [7] J. I. Leon-Pedroza, L. A. Gonzalez-Tapia, E. del Olmo-Gil, D. Castellanos-Rodriguez, G. Escobedo, and A. Gonzalez-Chavez, "Low-grade systemic inflammation and the development of metabolic diseases: from the molecular evidence to the clinical practice," *Cirugia y Cirujanos*, vol. 83, no. 6, pp. 543–551, 2015.
- [8] M. F. Gregor and G. S. Hotamisligil, "Inflammatory mechanisms in obesity," *Annual Review of Immunology*, vol. 29, no. 1, pp. 415–445, 2011.
- [9] N. S. Kalupahana, N. Moustaid-Moussa, and K. J. Claycombe, "Immunity as a link between obesity and insulin resistance," *Molecular Aspects of Medicine*, vol. 33, no. 1, pp. 26–34, 2012.
- [10] O. Osborn and J. M. Olefsky, "The cellular and signaling networks linking the immune system and metabolism in disease," *Nature Medicine*, vol. 18, no. 3, pp. 363–374, 2012.
- [11] J. Wada and H. Makino, "Innate immunity in diabetes and diabetic nephropathy," *Nature Reviews Nephrology*, vol. 12, no. 1, pp. 13–26, 2016.
- [12] R. Ringseis, K. Eder, F. C. Mooren, and K. Kruger, "Metabolic signals and innate immune activation in obesity and exercise," *Exercise Immunology Review*, vol. 21, pp. 58–68, 2015.
- [13] X. Cao, "Self-regulation and cross-regulation of pattern-recognition receptor signalling in health and disease," *Nature Reviews Immunology*, vol. 16, no. 1, pp. 35–50, 2016.
- [14] J. J. Shin, E. K. Lee, T. J. Park, and W. Kim, "Damage-associated molecular patterns and their pathological relevance in diabetes mellitus," *Ageing Research Reviews*, vol. 24, pp. 66–76, 2015.
- [15] W. G. Land, "The role of damage-associated molecular patterns in human diseases: part I — promoting inflammation and immunity," *Sultan Qaboos University Medical Journal*, vol. 15, pp. e9–e21, 2015.
- [16] C. Jin and R. A. Flavell, "Innate sensors of pathogen and stress: linking inflammation to obesity," *The Journal of Allergy and Clinical Immunology*, vol. 132, no. 2, pp. 287–294, 2013.
- [17] B. K. Davis, H. Wen, and J. P.-Y. Ting, "The inflammasome NLRs in immunity, inflammation, and associated diseases," *Annual Review of Immunology*, vol. 29, no. 1, pp. 707–735, 2011.

- [18] N. Esser, S. Legrand-Poels, J. Piette, A. J. Scheen, and N. Paquot, "Inflammation as a link between obesity, metabolic syndrome and type 2 diabetes," *Diabetes Research and Clinical Practice*, vol. 105, no. 2, pp. 141–150, 2014.
- [19] C. Jin, J. Hena-Mejia, and R. A. Flavell, "Innate immune receptors: key regulators of metabolic disease progression," *Cell Metabolism*, vol. 17, no. 6, pp. 873–882, 2013.
- [20] G. S. Hotamisligil, P. Arner, J. F. Caro, R. L. Atkinson, and B. M. Spiegelman, "Increased adipose tissue expression of tumor necrosis factor- α in human obesity and insulin resistance," *The Journal of Clinical Investigation*, vol. 95, no. 5, pp. 2409–2415, 1995.
- [21] T. C. Wascher, J. H. N. Lindeman, H. Sourij, T. Kooistra, G. Pacini, and M. Roden, "Chronic TNF- α neutralization does not improve insulin resistance or endothelial function in "healthy" men with metabolic syndrome," *Molecular Medicine*, vol. 17, no. 3-4, pp. 189–193, 2011.
- [22] P. Li, S. Liu, M. Lu et al., "Hematopoietic-derived Galectin-3 causes cellular and systemic insulin resistance," *Cell*, vol. 167, no. 4, pp. 973–984.e12, 2016, e12.
- [23] Y. S. Lee, J. Wollam, and J. M. Olefsky, "An integrated view of immunometabolism," *Cell*, vol. 172, no. 1-2, pp. 22–40, 2018.
- [24] D. Mathis, "Immunological goings-on in visceral adipose tissue," *Cell Metabolism*, vol. 17, no. 6, pp. 851–859, 2013.
- [25] J. DeFuria, A. C. Belkina, M. Jagannathan-Bogdan et al., "B cells promote inflammation in obesity and type 2 diabetes through regulation of T-cell function and an inflammatory cytokine profile," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 13, pp. 5133–5138, 2013.
- [26] K. Eguchi and R. Nagai, "Islet inflammation in type 2 diabetes and physiology," *The Journal of Clinical Investigation*, vol. 127, no. 1, pp. 14–23, 2017.
- [27] K. Eguchi and I. Manabe, "Macrophages and islet inflammation in type 2 diabetes," *Diabetes, Obesity & Metabolism*, vol. 15, no. s3, pp. 152–158, 2013.
- [28] C. Cavelti-Weder, A. Babians-Brunner, C. Keller et al., "Effects of gevokizumab on glycemia and inflammatory markers in type 2 diabetes," *Diabetes Care*, vol. 35, no. 8, pp. 1654–1662, 2012.
- [29] A. Rissanen, C. P. Howard, J. Botha, and T. Thuren, "Effect of anti-IL-1 β antibody (canakinumab) on insulin secretion rates in impaired glucose tolerance or type 2 diabetes: results of a randomized, placebo-controlled trial," *Diabetes, Obesity and Metabolism*, vol. 14, no. 12, pp. 1088–1096, 2012.
- [30] W. Ying, Y. S. Lee, Y. Dong et al., "Expansion of islet-resident macrophages leads to inflammation affecting β cell proliferation and function in obesity," *Cell Metabolism*, vol. 29, no. 2, pp. 457–474.e5, 2019, e5.
- [31] S. L. Masters, A. Dunne, S. L. Subramanian et al., "Activation of the NLRP3 inflammasome by islet amyloid polypeptide provides a mechanism for enhanced IL-1 β in type 2 diabetes," *Nature Immunology*, vol. 11, no. 10, pp. 897–904, 2010.
- [32] A. T. Nguyen-Lefebvre and A. Horuzsko, "Kupffer cell metabolism and function," *Journal of Enzymology Metabolism*, vol. 1, 2015.
- [33] P. Li, D. Y. Oh, G. Bandyopadhyay et al., "LTB4 promotes insulin resistance in obese mice by acting on macrophages, hepatocytes and myocytes," *Nature Medicine*, vol. 21, no. 3, pp. 239–247, 2015.
- [34] J. A. Del Campo, P. Gallego, and L. Grande, "Role of inflammatory response in liver diseases: therapeutic strategies," *World Journal of Hepatology*, vol. 10, no. 1, pp. 1–7, 2018.
- [35] I. Nieto-Vazquez, S. Fernandez-Veledo, C. de Alvaro, and M. Lorenzo, "Dual role of interleukin-6 in regulating insulin sensitivity in murine skeletal muscle," *Diabetes*, vol. 57, no. 12, pp. 3211–3221, 2008.
- [36] I. M. Khan, X. Y. D. Perrard, G. Brunner et al., "Intermuscular and perimuscular fat expansion in obesity correlates with skeletal muscle T cell and macrophage infiltration and insulin resistance," *International Journal of Obesity*, vol. 39, no. 11, pp. 1607–1618, 2015.
- [37] M. A. McArdle, O. M. Finucane, R. M. Connaughton, A. M. McMorrow, and H. M. Roche, "Mechanisms of obesity-induced inflammation and insulin resistance: insights into the emerging role of nutritional strategies," *Frontiers in Endocrinology*, vol. 4, p. 52, 2013.
- [38] M. Nahrendorf and F. K. Swirski, "Immunology. Neutrophil-macrophage communication in inflammation and atherosclerosis," *Science*, vol. 349, no. 6245, pp. 237–238, 2015.
- [39] A. Gistera and G. K. Hansson, "The immunology of atherosclerosis," *Nature Reviews Nephrology*, vol. 13, no. 6, pp. 368–380, 2017.
- [40] A. Jais and J. C. Bruning, "Hypothalamic inflammation in obesity and metabolic disease," *The Journal of Clinical Investigation*, vol. 127, no. 1, pp. 24–32, 2017.
- [41] Y. Benomar and M. Taouis, "Molecular mechanisms underlying obesity-induced hypothalamic inflammation and insulin resistance: pivotal role of Resistin/TLR4 pathways," *Frontiers in Endocrinology*, vol. 10, p. 140, 2019.
- [42] C. Bleau, A. D. Karelis, D. H. St-Pierre, and L. Lamontagne, "Crosstalk between intestinal microbiota, adipose tissue and skeletal muscle as an early event in systemic low-grade inflammation and the development of obesity and diabetes," *Diabetes/Metabolism Research and Reviews*, vol. 31, no. 6, pp. 545–561, 2015.
- [43] C. A. Marotz and A. Zarrinpar, "Treating obesity and metabolic syndrome with fecal microbiota transplantation," *The Yale Journal of Biology and Medicine*, vol. 89, no. 3, pp. 383–388, 2016.
- [44] M. Spiljar, D. Merkler, and M. Trajkovski, "The immune system bridges the gut microbiota with systemic energy homeostasis: focus on TLRs, mucosal barrier, and SCFAs," *Frontiers in Immunology*, vol. 8, p. 1353, 2017.
- [45] D. A. Winer, H. Luck, S. Tsai, and S. Winer, "The intestinal immune system in obesity and insulin resistance," *Cell Metabolism*, vol. 23, no. 3, pp. 413–426, 2016.
- [46] I. Jialal, H. Kaur, and S. Devaraj, "Toll-like receptor status in obesity and metabolic syndrome: a translational perspective," *The Journal of Clinical Endocrinology and Metabolism*, vol. 99, no. 1, pp. 39–48, 2014.
- [47] L. Yu and Z. Feng, "The role of toll-like receptor signaling in the progression of heart failure," *Mediators of Inflammation*, vol. 2018, Article ID 9874109, 11 pages, 2018.
- [48] H. Hug, M. H. Mohajeri, and G. La Fata, "Toll-like receptors: regulators of the immune response in the human gut," *Nutrients*, vol. 10, no. 2, p. 203, 2018.
- [49] G. Boden, "Obesity, insulin resistance and free fatty acids," *Current Opinion in Endocrinology, Diabetes, and Obesity*, vol. 18, no. 2, pp. 139–143, 2011.

- [50] R. K. Murumalla, M. K. Gunasekaran, J. K. Padhan et al., "Fatty acids do not pay the toll: effect of SFA and PUFA on human adipose tissue and mature adipocytes inflammation," *Lipids in Health and Disease*, vol. 11, no. 1, p. 175, 2012.
- [51] S. Huang, J. M. Rutkowski, R. G. Snodgrass et al., "Saturated fatty acids activate TLR-mediated proinflammatory signaling pathways," *Journal of Lipid Research*, vol. 53, no. 9, pp. 2002–2013, 2012.
- [52] R. G. Snodgrass, S. Huang, I. W. Choi, J. C. Rutledge, and D. H. Hwang, "Inflammasome-mediated secretion of IL-1 β in human monocytes through TLR2 activation; modulation by dietary fatty acids," *The Journal of Immunology*, vol. 191, no. 8, pp. 4337–4347, 2013.
- [53] R. Ahmad, A. al-Mass, V. Atizado et al., "Elevated expression of the toll like receptors 2 and 4 in obese individuals: its significance for obesity-induced inflammation," *Journal of Inflammation*, vol. 9, no. 1, article 48, 2012.
- [54] M. Andrews, N. Soto, and M. Arredondo-Olguin, "Association between ferritin and hepcidin levels and inflammatory status in patients with type 2 diabetes mellitus and obesity," *Nutrition*, vol. 31, no. 1, pp. 51–57, 2015.
- [55] V. P. W. Scholtes, D. Versteeg, J. P. P. M. de Vries et al., "Toll-like receptor 2 and 4 stimulation elicits an enhanced inflammatory response in human obese patients with atherosclerosis," *Clinical Science*, vol. 121, no. 5, pp. 205–214, 2011.
- [56] M. T. Arias-Loste, P. Iruzubieta, Á. Puente et al., "Increased expression profile and functionality of TLR6 in peripheral blood mononuclear cells and hepatocytes of morbidly obese patients with non-alcoholic fatty liver disease," *International Journal of Molecular Sciences*, vol. 17, no. 11, p. 1878, 2016.
- [57] J. du Plessis, H. Korf, J. van Pelt et al., "Pro-inflammatory cytokines but not endotoxin-related parameters associate with disease severity in patients with NAFLD," *PLoS One*, vol. 11, no. 12, article e0166048, 2016.
- [58] Z. Wan, C. Durrer, D. Mah, S. Simtchouk, and J. P. Little, "One-week high-fat diet leads to reduced toll-like receptor 2 expression and function in young healthy men," *Nutrition Research*, vol. 34, no. 12, pp. 1045–1051, 2014.
- [59] K. Murakami, H. Bujo, H. Unoki, and Y. Saito, "High fat intake induces a population of adipocytes to co-express TLR2 and TNF α in mice with insulin resistance," *Biochemical and Biophysical Research Communications*, vol. 354, no. 3, pp. 727–734, 2007.
- [60] M. T. A. Nguyen, S. Favelukis, A. K. Nguyen et al., "A subpopulation of macrophages infiltrates hypertrophic adipose tissue and is activated by free fatty acids via Toll-like receptors 2 and 4 and JNK-dependent pathways," *Journal of Biological Chemistry*, vol. 282, no. 48, pp. 35279–35292, 2007.
- [61] L.-H. Kuo, P. J. Tsai, M. J. Jiang et al., "Toll-like receptor 2 deficiency improves insulin sensitivity and hepatic insulin signalling in the mouse," *Diabetologia*, vol. 54, no. 1, pp. 168–179, 2011.
- [62] J. A. Ehses, D. T. Meier, S. Wueest et al., "Toll-like receptor 2-deficient mice are protected from insulin resistance and beta cell dysfunction induced by a high-fat diet," *Diabetologia*, vol. 53, no. 8, pp. 1795–1806, 2010.
- [63] G. Betanzos-Cabrera, D. Estrada-Luna, H. Belefant-Miller, and J. C. Cancino-Diaz, "Mice fed with a high fat diet show a decrease in the expression of toll like receptor (TLR)2 and TLR6 mRNAs in adipose and hepatic tissues," *Nutrición Hospitalaria*, vol. 27, pp. 1196–1203, 2012.
- [64] J. E. Davis, D. R. Braucher, J. Walker-Daniels, and M. E. Spurlock, "Absence of Tlr2 protects against high-fat diet-induced inflammation and results in greater insulin-stimulated glucose transport in cultured adipocytes," *The Journal of Nutritional Biochemistry*, vol. 22, no. 2, pp. 136–141, 2011.
- [65] K. Miura, L. Yang, N. van Rooijen, D. A. Brenner, H. Ohnishi, and E. Seki, "Toll-like receptor 2 and palmitic acid cooperatively contribute to the development of nonalcoholic steatohepatitis through inflammasome activation in mice," *Hepatology*, vol. 57, no. 2, pp. 577–589, 2013.
- [66] X. Liu, T. Ukai, H. Yumoto et al., "Toll-like receptor 2 plays a critical role in the progression of atherosclerosis that is independent of dietary lipids," *Atherosclerosis*, vol. 196, no. 1, pp. 146–154, 2008.
- [67] A. E. Mullick, P. S. Tobias, and L. K. Curtiss, "Modulation of atherosclerosis in mice by Toll-like receptor 2," *The Journal of Clinical Investigation*, vol. 115, no. 11, pp. 3149–3156, 2005.
- [68] A. M. Caricilli, P. K. Picardi, L. L. de Abreu et al., "Gut microbiota is a key modulator of insulin resistance in TLR 2 knock-out mice," *PLoS Biology*, vol. 9, no. 12, article e1001212, 2011.
- [69] C. Ubeda, L. Lipuma, A. Gobourne et al., "Familial transmission rather than defective innate immunity shapes the distinct intestinal microbiota of TLR-deficient mice," *The Journal of Experimental Medicine*, vol. 209, no. 8, pp. 1445–1456, 2012.
- [70] C. A. Rivera, L. Gaskin, M. Allman et al., "Toll-like receptor-2 deficiency enhances non-alcoholic steatohepatitis," *BMC Gastroenterology*, vol. 10, no. 1, article 52, 2010.
- [71] A. M. Caricilli, P. H. Nascimento, J. R. Pauli et al., "Inhibition of toll-like receptor 2 expression improves insulin sensitivity and signaling in muscle and white adipose tissue of mice fed a high-fat diet," *Journal of Endocrinology*, vol. 199, no. 3, pp. 399–406, 2008.
- [72] L. Schmidt and M. A. Carrillo-Sepulveda, "Toll-like receptor 2 mediates vascular contraction and activates RhoA signaling in vascular smooth muscle cells from STZ-induced type 1 diabetic rats," *Pflügers Archiv*, vol. 467, no. 11, pp. 2361–2374, 2015.
- [73] P. Mistry, M. H. W. Laird, R. S. Schwarz et al., "Inhibition of TLR2 signaling by small molecule inhibitors targeting a pocket within the TLR2 TIR domain," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 112, no. 17, pp. 5455–5460, 2015.
- [74] C. J. Desmet and K. J. Ishii, "Nucleic acid sensing at the interface between innate and adaptive immunity in vaccination," *Nature Reviews Immunology*, vol. 12, no. 7, pp. 479–491, 2012.
- [75] L. Yu, Y. Xu, F. Wang, C. Yang, G. Liu, and X. Song, "Functional roles of pattern recognition receptors that recognize virus nucleic acids in human adipose-derived mesenchymal stem cells," *BioMed Research International*, vol. 2016, Article ID 9872138, 13 pages, 2016.
- [76] D. B. Ballak, E. J. P. van Asseldonk, J. A. van Diepen et al., "TLR-3 is present in human adipocytes, but its signalling is not required for obesity-induced inflammation in adipose tissue *in vivo*," *PLoS One*, vol. 10, no. 4, article e0123152, 2015.
- [77] L. Yu, K. Yan, P. Liu et al., "Pattern recognition receptor-initiated innate antiviral response in mouse adipose cells," *Immunology and Cell Biology*, vol. 92, no. 2, pp. 105–115, 2014.

- [78] J. Latorre, J. M. Moreno-Navarrete, M. Sabater et al., "Decreased TLR3 in hyperplastic adipose tissue, blood and inflamed adipocytes is related to metabolic inflammation," *Cellular Physiology and Biochemistry*, vol. 51, no. 3, pp. 1051–1068, 2018.
- [79] O. Fabre, C. Breuker, C. Amouzou et al., "Defects in TLR3 expression and RNase L activation lead to decreased MnSOD expression and insulin resistance in muscle cells of obese people," *Cell Death & Disease*, vol. 5, no. 3, article e1136, 2014.
- [80] L. H. Wu, C. C. Huang, S. Adhikarakunnathu et al., "Loss of toll-like receptor 3 function improves glucose tolerance and reduces liver steatosis in obese mice," *Metabolism*, vol. 61, no. 11, pp. 1633–1645, 2012.
- [81] D. Strodthoff, Z. Ma, T. Wirström et al., "Toll-like receptor 3 influences glucose homeostasis and β -cell insulin secretion," *Diabetes*, vol. 64, no. 10, pp. 3425–3438, 2015.
- [82] M. K. Gunasekaran, A.-L. Virama-Latchoumy, A.-C. Girard et al., "TLR4-dependant proinflammatory effects of HMGB1 on human adipocyte," *Adipocytes*, vol. 5, no. 4, pp. 384–388, 2016.
- [83] T. Sharifnia, J. Antoun, T. G. C. Verriere et al., "Hepatic TLR4 signaling in obese NAFLD," *American Journal of Physiology Gastrointestinal and Liver Physiology*, vol. 309, no. 4, pp. G270–G278, 2015.
- [84] E. F. Devezve, M. Renovato-Martins, K. Clement, C. Sautes-Fridman, I. Cremer, and C. Poitou, "Profiling of the three circulating monocyte subpopulations in human obesity," *Journal of Immunology*, vol. 194, no. 8, pp. 3917–3923, 2015.
- [85] K. Nishiyama, Y. Fujimoto, T. Takeuchi, and Y. T. Azuma, "Aggressive crosstalk between fatty acids and inflammation in macrophages and their influence on metabolic homeostasis," *Neurochemical Research*, vol. 43, no. 1, pp. 19–26, 2018.
- [86] M. K. Gunasekaran, W. Viranaicken, A. C. Girard et al., "Inflammation triggers high mobility group box 1 (HMGB1) secretion in adipose tissue, a potential link to obesity," *Cytokine*, vol. 64, no. 1, pp. 103–111, 2013.
- [87] I. Jialal, B. A. Huet, H. Kaur, A. Chien, and S. Devaraj, "Increased toll-like receptor activity in patients with metabolic syndrome," *Diabetes Care*, vol. 35, no. 4, pp. 900–904, 2012.
- [88] V. Catalán, J. Gómez-Ambrosi, A. Rodríguez et al., "Increased tenascin C and Toll-like receptor 4 levels in visceral adipose tissue as a link between inflammation and extracellular matrix remodeling in obesity," *The Journal of Clinical Endocrinology & Metabolism*, vol. 97, no. 10, pp. E1880–E1889, 2012.
- [89] J. Nguyen, J. Jiao, K. Smoot et al., "Toll-like receptor 4: a target for chemoprevention of hepatocellular carcinoma in obesity and steatohepatitis," *Oncotarget*, vol. 9, no. 50, pp. 29495–29507, 2018.
- [90] Y. Ozaki, T. Imanishi, S. Hosokawa et al., "Association of Toll-like receptor 4 on human monocyte subsets and vulnerability characteristics of coronary plaque as assessed by 64-slice multidetector computed tomography," *Circulation Journal*, vol. 81, no. 6, pp. 837–845, 2017.
- [91] H. Methe, J. O. Kim, S. Kofler, M. Weis, M. Nabauer, and J. Koglin, "Expansion of circulating Toll-like receptor 4-positive monocytes in patients with acute coronary syndrome," *Circulation*, vol. 111, no. 20, pp. 2654–2661, 2005.
- [92] A. K. Ghosh, M. O'Brien, T. Mau, and R. Yung, "Toll-like receptor 4 (TLR4) deficient mice are protected from adipose tissue inflammation in aging," *Aging*, vol. 9, no. 9, pp. 1971–1982, 2017.
- [93] N. Pierre, L. Deldicque, C. Barbe, D. Naslain, P. D. Cani, and M. Francaux, "Toll-like receptor 4 knockout mice are protected against endoplasmic reticulum stress induced by a high-fat diet," *PLoS One*, vol. 8, no. 5, article e65061, 2013.
- [94] H. Shi, M. V. Kokoeva, K. Inouye, I. Tzamelis, H. Yin, and J. S. Flier, "TLR4 links innate immunity and fatty acid-induced insulin resistance," *The Journal of Clinical Investigation*, vol. 116, no. 11, pp. 3015–3025, 2006.
- [95] J. S. Orr, M. J. Puglisi, K. L. J. Ellacott, C. N. Lumeng, D. H. Wasserman, and A. H. Hasty, "Toll-like receptor 4 deficiency promotes the alternative activation of adipose tissue macrophages," *Diabetes*, vol. 61, no. 11, pp. 2718–2727, 2012.
- [96] L. Jia, C. R. Vianna, M. Fukuda et al., "Hepatocyte Toll-like receptor 4 regulates obesity-induced inflammation and insulin resistance," *Nature Communications*, vol. 5, no. 1, article 3878, 2014.
- [97] K. S. Michelsen, M. H. Wong, P. K. Shah et al., "Lack of Toll-like receptor 4 or myeloid differentiation factor 88 reduces atherosclerosis and alters plaque phenotype in mice deficient in apolipoprotein E," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 29, pp. 10679–10684, 2004.
- [98] Y. Ding, S. Subramanian, V. N. Montes et al., "Toll-like receptor 4 deficiency decreases atherosclerosis but does not protect against inflammation in obese low-density lipoprotein receptor-deficient mice," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 32, no. 7, pp. 1596–1604, 2012.
- [99] T. Csak, A. Velayudham, I. Hritz et al., "Deficiency in myeloid differentiation factor-2 and toll-like receptor 4 expression attenuates nonalcoholic steatohepatitis and fibrosis in mice," *American Journal of Physiology Gastrointestinal and Liver Physiology*, vol. 300, no. 3, pp. G433–G441, 2011.
- [100] D. F. Ferreira, J. Fiamoncini, I. H. Prist, S. K. Ariga, H. P. de Souza, and T. M. de Lima, "Novel role of TLR4 in NAFLD development: modulation of metabolic enzymes expression," *Biochimica et Biophysica Acta (BBA) - Molecular and Cell Biology of Lipids*, vol. 1851, no. 10, pp. 1353–1359, 2015.
- [101] G. S. Hotamisligil, "Inflammation, metaflammation and immunometabolic disorders," *Nature*, vol. 542, no. 7640, pp. 177–185, 2017.
- [102] S. Kochumon, A. Wilson, B. Chandy et al., "Palmitate activates CCL4 expression in human monocytic cells via TLR4/MyD88 dependent activation of NF- κ B/MAPK/PI3K signaling systems," *Cellular Physiology and Biochemistry*, vol. 46, no. 3, pp. 953–964, 2018.
- [103] M. M. Rogero and P. C. Calder, "Obesity, inflammation, toll-like receptor 4 and fatty acids," *Nutrients*, vol. 10, no. 4, p. 432, 2018.
- [104] A. B. Engin, "Adipocyte-macrophage cross-talk in obesity," *Advances in Experimental Medicine and Biology*, vol. 960, pp. 327–343, 2017.
- [105] A. L. Harte, N. F. da Silva, S. J. Creely et al., "Elevated endotoxin levels in non-alcoholic fatty liver disease," *Journal of Inflammation*, vol. 7, no. 1, article 15, 2010.
- [106] P. D. Cani, J. Amar, M. A. Iglesias et al., "Metabolic endotoxemia initiates obesity and insulin resistance," *Diabetes*, vol. 56, no. 7, pp. 1761–1772, 2007.
- [107] K. A. Kim, W. Gu, I. A. Lee, E. H. Joh, and D. H. Kim, "High fat diet-induced gut microbiota exacerbates inflammation

- and obesity in mice via the TLR4 signaling pathway,” *PLoS One*, vol. 7, no. 10, article e47713, 2012.
- [108] V. A. Moser, M. F. Uchoa, and C. J. Pike, “TLR4 inhibitor TAK-242 attenuates the adverse neural effects of diet-induced obesity,” *Journal of Neuroinflammation*, vol. 15, no. 1, article 306, 2018.
- [109] S. E. Hussey, H. Liang, S. R. Costford et al., “TAK-242, a small-molecule inhibitor of Toll-like receptor 4 signalling, unveils similarities and differences in lipopolysaccharide- and lipid-induced inflammation and insulin resistance in muscle cells,” *Bioscience Reports*, vol. 33, no. 1, pp. 37–47, 2012.
- [110] N. Zhang, H. Liang, R. V. Farese, J. Li, N. Musi, and S. E. Hussey, “Pharmacological TLR4 inhibition protects against acute and chronic fat-induced insulin resistance in rats,” *PLoS One*, vol. 10, no. 7, article e0132575, 2015.
- [111] A. Pirillo and A. L. Catapano, “Berberine, a plant alkaloid with lipid- and glucose-lowering properties: from in vitro evidence to clinical studies,” *Atherosclerosis*, vol. 243, no. 2, pp. 449–461, 2015.
- [112] M. Fulde, F. Sommer, B. Chassaing et al., “Neonatal selection by Toll-like receptor 5 influences long-term gut microbiota composition,” *Nature*, vol. 560, no. 7719, pp. 489–493, 2018.
- [113] B. Chassaing, R. E. Ley, and A. T. Gewirtz, “Intestinal epithelial cell toll-like receptor 5 regulates the intestinal microbiota to prevent low-grade inflammation and metabolic syndrome in mice,” *Gastroenterology*, vol. 147, no. 6, pp. 1363–1377.e17, 2014, e17.
- [114] M. Vijay-Kumar, J. D. Aitken, F. A. Carvalho et al., “Metabolic syndrome and altered gut microbiota in mice lacking Toll-like receptor 5,” *Science*, vol. 328, no. 5975, pp. 228–231, 2010.
- [115] F. A. Carvalho, O. Koren, J. K. Goodrich et al., “Transient inability to manage proteobacteria promotes chronic gut inflammation in TLR5-deficient mice,” *Cell Host & Microbe*, vol. 12, no. 2, pp. 139–152, 2012.
- [116] R. Ahmad, S. Kochumon, R. Thomas, V. Atizado, and S. Sindhu, “Increased adipose tissue expression of TLR8 in obese individuals with or without type-2 diabetes: significance in metabolic inflammation,” *Journal of Inflammation*, vol. 13, no. 1, article 38, 2016.
- [117] S. Nishimoto, D. Fukuda, Y. Higashikuni et al., “Obesity-induced DNA released from adipocytes stimulates chronic adipose tissue inflammation and insulin resistance,” *Science Advances*, vol. 2, no. 3, article e1501332, 2016.
- [118] P. Bellot, J. C. García-Pagán, R. Francés et al., “Bacterial DNA translocation is associated with systemic circulatory abnormalities and intrahepatic endothelial dysfunction in patients with cirrhosis,” *Hepatology*, vol. 52, no. 6, pp. 2044–2052, 2010.
- [119] S. Kamfar, S. M. Alavian, M. Houshmand et al., “Liver mitochondrial DNA copy number and deletion levels may contribute to nonalcoholic fatty liver disease susceptibility,” *Hepatitis Monthly*, vol. 16, no. 12, article e40774, 2016.
- [120] C.-P. Hong, C. H. Yun, G. W. Lee, A. Park, Y. M. Kim, and M. H. Jang, “TLR9 regulates adipose tissue inflammation and obesity-related metabolic disorders,” *Obesity*, vol. 23, no. 11, pp. 2199–2206, 2015.
- [121] A. R. Mridha, F. Haczeyni, M. M. Yeh et al., “TLR9 is up-regulated in human and murine NASH: pivotal role in inflammatory recruitment and cell survival,” *Clinical Science*, vol. 131, no. 16, pp. 2145–2159, 2017.
- [122] I. Garcia-Martinez, N. Santoro, Y. Chen et al., “Hepatocyte mitochondrial DNA drives nonalcoholic steatohepatitis by activation of TLR9,” *The Journal of Clinical Investigation*, vol. 126, no. 3, pp. 859–864, 2016.
- [123] L. Boutens, A. M. Mirea, I. van den Munckhof et al., “A role for TLR10 in obesity and adipose tissue morphology,” *Cytokine*, vol. 108, pp. 205–212, 2018.
- [124] C. L. Feerick and D. P. McKernan, “Understanding the regulation of pattern recognition receptors in inflammatory diseases — a “Nod” in the right direction,” *Immunology*, vol. 150, no. 3, pp. 237–247, 2017.
- [125] G. Santoni, C. Cardinali, M. B. Morelli, M. Santoni, M. Nabissi, and C. Amantini, “Danger- and pathogen-associated molecular patterns recognition by pattern-recognition receptors and ion channels of the transient receptor potential family triggers the inflammasome activation in immune cells and sensory neurons,” *Journal of Neuroinflammation*, vol. 12, no. 1, p. 21, 2015.
- [126] R. Caruso, N. Warner, N. Inohara, and G. Nunez, “NOD1 and NOD2: signaling, host defense, and inflammatory disease,” *Immunity*, vol. 41, no. 6, pp. 898–908, 2014.
- [127] J. D. Schertzer, A. K. Tamrakar, J. G. Magalhaes et al., “NOD1 activators link innate immunity to insulin resistance,” *Diabetes*, vol. 60, no. 9, pp. 2206–2215, 2011.
- [128] K. L. Chan, T. H. Tam, P. Boroumand et al., “Circulating NOD1 activators and hematopoietic NOD1 contribute to metabolic inflammation and insulin resistance,” *Cell Reports*, vol. 18, no. 10, pp. 2415–2426, 2017.
- [129] S. Kanno, H. Nishio, T. Tanaka et al., “Activation of an innate immune receptor, Nod1, accelerates atherogenesis in *Apoe*^{-/-} mice,” *The Journal of Immunology*, vol. 194, no. 2, pp. 773–780, 2015.
- [130] E. Denou, K. Lolmède, L. Garidou et al., “Defective NOD2 peptidoglycan sensing promotes diet-induced inflammation, dysbiosis, and insulin resistance,” *EMBO Molecular Medicine*, vol. 7, no. 3, pp. 259–274, 2015.
- [131] D. Jiang, S. Chen, R. Sun, X. Zhang, and D. Wang, “The NLRP3 inflammasome: role in metabolic disorders and regulation by metabolic pathways,” *Cancer Letters*, vol. 419, pp. 8–19, 2018.
- [132] S. Legrand-Poels, N. Esser, L. L’Homme, A. Scheen, N. Paquot, and J. Piette, “Free fatty acids as modulators of the NLRP3 inflammasome in obesity/type 2 diabetes,” *Biochemical Pharmacology*, vol. 92, no. 1, pp. 131–141, 2014.
- [133] T. Jourdan, G. Godlewski, R. Cinar et al., “Activation of the Nlrp3 inflammasome in infiltrating macrophages by endocannabinoids mediates beta cell loss in type 2 diabetes,” *Nature Medicine*, vol. 19, no. 9, pp. 1132–1140, 2013.
- [134] J. F. Varghese, R. Patel, and U. C. S. Yadav, “Sterol regulatory element binding protein (SREBP) -1 mediates oxidized low-density lipoprotein (oxLDL) induced macrophage foam cell formation through NLRP3 inflammasome activation,” *Cellular Signalling*, vol. 53, pp. 316–326, 2019.
- [135] P. Duewell, H. Kono, K. J. Rayner et al., “NLRP3 inflammasomes are required for atherogenesis and activated by cholesterol crystals,” *Nature*, vol. 464, no. 7293, pp. 1357–1361, 2010.
- [136] N. Esser, L. L’homme, A. de Roover et al., “Obesity phenotype is related to NLRP3 inflammasome activity and immunological profile of visceral adipose tissue,” *Diabetologia*, vol. 56, no. 11, pp. 2487–2497, 2013.

- [137] H.-M. Lee, J. J. Kim, H. J. Kim, M. Shong, B. J. Ku, and E. K. Jo, "Upregulated NLRP3 inflammasome activation in patients with type 2 diabetes," *Diabetes*, vol. 62, no. 1, pp. 194–204, 2013.
- [138] B. Vandanmagsar, Y. H. Youm, A. Ravussin et al., "The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance," *Nature Medicine*, vol. 17, no. 2, pp. 179–188, 2011.
- [139] F. Zheng, S. Xing, Z. Gong, and Q. Xing, "NLRP3 inflammasomes show high expression in aorta of patients with atherosclerosis," *Heart, Lung & Circulation*, vol. 22, no. 9, pp. 746–750, 2013.
- [140] J. Rheinheimer, B. M. de Souza, N. S. Cardoso, A. C. Bauer, and D. Crispim, "Current role of the NLRP3 inflammasome on obesity and insulin resistance: a systematic review," *Metabolism*, vol. 74, pp. 1–9, 2017.
- [141] R. Stienstra, J. A. van Diepen, C. J. Tack et al., "Inflammasome is a central player in the induction of obesity and insulin resistance," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 108, no. 37, pp. 15324–15329, 2011.
- [142] F. Zheng, S. Xing, Z. Gong, W. Mu, and Q. Xing, "Silence of NLRP3 suppresses atherosclerosis and stabilizes plaques in apolipoprotein E-deficient mice," *Mediators of Inflammation*, vol. 2014, Article ID 507208, 8 pages, 2014.
- [143] W. Zhu, P. P. Feng, K. He, S. W. Li, and J. P. Gong, "Liraglutide protects non-alcoholic fatty liver disease via inhibiting NLRP3 inflammasome activation in a mouse model induced by high-fat diet," *Biochemical and Biophysical Research Communications*, vol. 505, no. 2, pp. 523–529, 2018.
- [144] R. Stienstra, L. A. B. Joosten, T. Koenen et al., "The inflammasome-mediated caspase-1 activation controls adipocyte differentiation and insulin sensitivity," *Cell Metabolism*, vol. 12, no. 6, pp. 593–605, 2010.
- [145] V. Bhaskar, J. Yin, A. M. Mirza et al., "Monoclonal antibodies targeting IL-1 beta reduce biomarkers of atherosclerosis *in vitro* and inhibit atherosclerotic plaque formation in apolipoprotein E-deficient mice," *Atherosclerosis*, vol. 216, no. 2, pp. 313–320, 2011.
- [146] L. Badimon, "Interleukin-18: a potent proinflammatory cytokine in atherosclerosis," *Cardiovascular Research*, vol. 96, no. 2, pp. 172–175, 2012, discussion 6–80.
- [147] A. R. Mridha, A. Wree, A. A. B. Robertson et al., "NLRP3 inflammasome blockade reduces liver inflammation and fibrosis in experimental NASH in mice," *Journal of Hepatology*, vol. 66, no. 5, pp. 1037–1046, 2017.
- [148] R. C. Coll, A. A. B. Robertson, J. J. Chae et al., "A small-molecule inhibitor of the NLRP3 inflammasome for the treatment of inflammatory diseases," *Nature Medicine*, vol. 21, no. 3, pp. 248–255, 2015.
- [149] A. J. Murphy, M. J. Kraakman, H. L. Kammoun et al., "IL-18 production from the NLRP1 inflammasome prevents obesity and metabolic syndrome," *Cell Metabolism*, vol. 23, no. 1, pp. 155–164, 2016.
- [150] T. Prochnicki and E. Latz, "Inflammasomes on the crossroads of innate immune recognition and metabolic control," *Cell Metabolism*, vol. 26, no. 1, pp. 71–93, 2017.
- [151] M. Levy, H. Shapiro, C. A. Thaiss, and E. Elinav, "NLRP6: a multifaceted innate immune sensor," *Trends in Immunology*, vol. 38, no. 4, pp. 248–260, 2017.
- [152] A. Al-Goblan, M. Al-Alfi, and M. Khan, "Mechanism linking diabetes mellitus and obesity," *Diabetes, Metabolic Syndrome and Obesity: Targets and Therapy*, vol. 7, pp. 587–591, 2014.
- [153] H. Guo, J. B. Callaway, and J. P. Ting, "Inflammasomes: mechanism of action, role in disease, and therapeutics," *Nature Medicine*, vol. 21, no. 7, pp. 677–687, 2015.
- [154] A. Achek, D. Yesudhas, and S. Choi, "Toll-like receptors: promising therapeutic targets for inflammatory diseases," *Archives of Pharmacological Research*, vol. 39, no. 8, pp. 1032–1049, 2016.
- [155] G. Lopez-Castejon and P. Pelegrin, "Current status of inflammasome blockers as anti-inflammatory drugs," *Expert Opinion on Investigational Drugs*, vol. 21, no. 7, pp. 995–1007, 2012.
- [156] W. Gao, Y. Xiong, Q. Li, and H. Yang, "Inhibition of Toll-like receptor signaling as a promising therapy for inflammatory diseases: a journey from molecular to nano therapeutics," *Frontiers in Physiology*, vol. 8, p. 508, 2017.
- [157] E. Monnet, G. Lapeyre, E. Poelgeest et al., "Evidence of NI-0101 pharmacological activity, an anti-TLR4 antibody, in a randomized phase I dose escalation study in healthy volunteers receiving LPS," *Clinical Pharmacology & Therapeutics*, vol. 101, no. 2, pp. 200–208, 2017.
- [158] D. M. W. Balak, M. B. A. van Doorn, R. D. Arbeit et al., "IMO-8400, a toll-like receptor 7, 8, and 9 antagonist, demonstrates clinical activity in a phase 2a, randomized, placebo-controlled trial in patients with moderate-to-severe plaque psoriasis," *Clinical Immunology*, vol. 174, pp. 63–72, 2017.
- [159] J. H. Park and Y. C. Kim, "P2X7 receptor antagonists: a patent review (2010–2015)," *Expert Opinion on Therapeutic Patents*, vol. 27, no. 3, pp. 257–267, 2017.