

# Diagnostic Yield of Oral Swab Testing by TB-LAMP for Diagnosis of Pulmonary Tuberculosis

This article was published in the following Dove Press journal:  
*Infection and Drug Resistance*

Yanhua Song,<sup>1\*</sup> Yifeng Ma,<sup>2,\*</sup>  
Rongmei Liu,<sup>1\*</sup> Yuanyuan Shang,<sup>3,\*</sup>  
Liping Ma,<sup>1</sup> Fengmin Huo,<sup>3</sup> Yunxu  
Li,<sup>3</sup> Wei Shu,<sup>4</sup> Yufeng Wang,<sup>5</sup>  
Mengqiu Gao,<sup>1</sup> Yu Pang<sup>3</sup>

<sup>1</sup>Department of Tuberculosis, Beijing Chest Hospital, Capital Medical University/Beijing Tuberculosis & Thoracic Tumor Research Institute, Beijing, People's Republic of China;

<sup>2</sup>Clinical Laboratory, Beijing Chest Hospital, Capital Medical University/Beijing Tuberculosis and Thoracic Tumor Research Institute, Beijing, People's Republic of China;

<sup>3</sup>Department of Bacteriology and Immunology, Beijing Key Laboratory on Drug-Resistant Tuberculosis Research, Beijing Chest Hospital, Capital Medical University/Beijing Tuberculosis & Thoracic Tumor Research Institute, Beijing, People's Republic of China; <sup>4</sup>Clinical Center on TB, Beijing Chest Hospital, Capital Medical University/Beijing Tuberculosis & Thoracic Tumor Research Institute, Beijing, People's Republic of China; <sup>5</sup>Department of Laboratory Quality Control, Innovation Alliance on Tuberculosis Diagnosis and Treatment (Beijing), Beijing, People's Republic of China

\*These authors contributed equally to this work

Correspondence: Mengqiu Gao  
Department of Tuberculosis, Beijing Chest Hospital, Capital Medical University/Beijing Tuberculosis & Thoracic Tumor Research Institute, No. 9, Beiguan Street, Tongzhou District, Beijing 101149, People's Republic of China  
Tel/Fax +86-10-8950 9322  
Email gaomqwdm@aliyun.com

Yu Pang  
Department of Bacteriology and Immunology, Beijing Key Laboratory on Drug-Resistant Tuberculosis Research, Beijing Chest Hospital, Capital Medical University/Beijing Tuberculosis & Thoracic Tumor Research Institute, No. 9, Beiguan Street, Tongzhou District, Beijing 101149, People's Republic of China  
Tel/Fax +86-10-8950 9322  
Email pangyupound@163.com

**Objective:** A prospective study was conducted to ascertain the accuracy of oral swab specimens collected in the early morning, spot and at night for detecting pulmonary tuberculosis (TB).

**Methods:** We prospectively enrolled patients with symptoms suggestive of pulmonary TB in Beijing Chest Hospital. An early morning sputum specimen was collected from each patient for GeneXpert MTB/RIF (Xpert) and mycobacterial culture. In addition, three oral swabs were collected for TB-LAMP testing.

**Results:** With the combined results of three oral swab specimens, the proportion of *Mycobacterium tuberculosis* (MTB)-positive cases achieved 40.6%, which was comparable to results for Xpert and MGIT ( $P=0.603$ ). Using Xpert plus MGIT as reference, the sensitivity of OS-LAMP on a single specimen ranged from 32.6% on the night oral swab to 50.0% on the morning swab. The combination of three oral swab specimens correctly identified 38 MTB-positive cases, indicating an overall sensitivity of 82.6%, which was significantly higher than that of a single oral swab specimen ( $P<0.001$ ,  $P=0.001$ ).

**Conclusion:** Oral swab can be used as an alternative specimen for diagnosis of pulmonary TB using TB-LAMP. Morning oral swab exhibits the highest sensitivity, and the inclusion of more specimens at different time points provides compensation in diagnostic sensitivity with single oral swab.

**Keywords:** diagnosis, oral swab, TB-LAMP, pulmonary tuberculosis, China

## Introduction

Tuberculosis (TB), caused by *Mycobacterium tuberculosis* (MTB) complex, poses a serious threat to public health, especially in developing countries.<sup>1,2</sup> In 2018, there were an estimated 10 million cases and 1.3 million deaths from tuberculosis worldwide.<sup>1</sup> Despite increases in TB notifications, a large gap occurs between the estimated number of incidents and the number of new cases reported (7.0 million).<sup>1</sup> More importantly, of the TB cases notified in 2018, only 55% were bacteriologically confirmed, and the remaining patients were diagnosed clinically.<sup>1</sup> Thus, intensified efforts are needed to improve access to early and accurate diagnosis that is essential for achieving the ultimate goal of TB elimination by 2035.<sup>3</sup>

A high-quality biological specimen is of great importance for facilitating the diagnosis of pulmonary TB.<sup>4</sup> Currently, the identification of active TB majorly relies on the examination of sputum specimens in individuals with symptoms suggestive of pulmonary TB.<sup>5</sup> However, a marked proportion of patients fail to produce good quality sputum specimens.<sup>4,6</sup> In the absence of quality sputum, multiple types of specimens have to be used as alternatives for TB diagnosis,

such as induced sputum and bronchoalveolar lavage fluid (BALF).<sup>7–9</sup> Induced sputum testing is considered a useful test in the diagnosis of sputum scarce patients, while its complex procedures become a barrier to gain acceptance from clinicians.<sup>10</sup> BALF is another promising alternative to the commonly used sputum specimen for detecting MTB in suspects.<sup>4</sup> The higher risks of nosocomial TB transmission and higher costs impedes clinical applications in resource-poor countries, where TB is endemic.<sup>10</sup> Therefore, a cheaper noninvasive method of providing quality specimens would be advantageous.

Oral swab testing has recently been reported to be a promising test in the diagnosis of bacteriologically confirmed TB patients.<sup>11,12</sup> Compared with collection of sputum specimens, oral swabbing is very easy to perform without production of aerosol. Although preliminary experiments yielded encouraging results for oral swabs,<sup>12</sup> the sample size was small, and the optimal sampling time point was undetermined. To address these concerns, a prospective study was conducted in the Beijing Chest Hospital to ascertain the accuracy of oral swab specimens collected in the early morning, spot and at night for detecting pulmonary TB.

## Materials and Methods

### Study Subjects

We prospectively enrolled patients aged >16 years with symptoms suggestive of pulmonary TB who were admitted to an inpatient TB unit in Beijing Chest hospital between November 2019 and April 2020. Patients meeting inclusion criteria had at least one symptom of TB and radiological abnormalities indicative of TB.<sup>13</sup> Symptoms of TB included a persistent cough of  $\geq 2$  weeks, unexplained fever for  $\geq 2$  weeks, weight loss, and night sweat. Demographic and clinical information was collected from electronic medical record.

### Collection of Sputum and Oral Swabs

Early morning sputum specimen was collected from each patient for GeneXpert MTB/RIF (Xpert) and mycobacterial culture. In addition, three oral swabs were collected as described previously.<sup>14</sup> Briefly, the clinicians firmly brushed the swab along the dorsum of the tongue seven-to-eight times. After swabbing, the swab was put into a 15-mL disposable tube containing 2 mL sterile normal saline.

## Routine Laboratory Examinations

Direct smear was conducted using light-emitting diode fluorescence microscopy for acid fast bacilli (AFB).<sup>15</sup> The positivity of smear was graded following national guidelines established by the Chinese Center for Diseases Control and Prevention. A volume of 1.0 mL of sputum was decontaminated with the N-acetyl-L-cysteine (NALC)-NaOH method for 15 min. After neutralization with sterile PBS (pH=7.0), the decontaminated specimen was centrifugated at 3000×g for 15 min then each pellet was resuspended in PBS. A volume of 0.5 mL of each suspension was inoculated into mycobacteria growth indicator tube (MGIT) supplemented with 0.8 mL of oleic acid-albumin-dextrose-catalase (OADC) along with PANTA. MGIT tubes were incubated into MGIT 960 instrument. The growth of bacteria was automatically recorded by the instrument. All positive cultures were confirmed as mycobacteria with Ziehl–Neelsen staining. The subsequent species identification was performed using a commercial Tibilias Rapid Test (Chuangxin, Hangzhou, China).

For Xpert testing, 1.0 mL of sputum was digested with 2.0 mL of sample reagent. After incubation at room temperature for 15 min, 2.0 mL of inactivated sputum sample was pipetted to a Xpert MTB/RIF cartridge. The cartridge was then loaded into the GeneXpert instrument. Results were automatically generated by the instrument within two hours.

### TB-LAMP

One milliliter of suspended swab sample was pipetted into a 1.5-mL centrifugation tube followed by centrifugation at 8000×g for three minutes. The supernatant was discarded, and the 60  $\mu$ L of pellet resuspension was used for TB-LAMP assay as described previously. The 60  $\mu$ L of resuspension was transferred to a heating tube containing extraction solution. Then the heating tube was incubated at 90°C for five minutes for inactivation and lysis of pathogens. Next the mixture in the heating tube was extruded into an adsorbent tube to remove the impurity. Thirty microliters of crude DNA solution was added into the reaction tube, and then loaded into the heating block at 67°C for 40 min. After amplification, the results of TB-LAMP were interpreted with the fluorescence detector.

### Statistical Analysis

The composite reference standard of mycobacterial culture and Xpert assay was used as the gold standard to assess

the diagnostic accuracy of TB-LAMP on oral swab samples. The chi-squared test was used to compare performance among the various laboratory methods. Nonparametric test was conducted to assess the difference in the values of test results between two groups. A Venn diagram was built using an online tool available at <https://bioinfogp.cnb.csic.es/tools/venny/index.html>. All statistical calculations were conducted using SPSS version 20.0 (IBM Corporation, Armonk, NY, USA). Differences were declared significant if *P*-values were less than 0.05.

## Results

### Participants

A total of 103 participants with symptoms suggestive of pulmonary TB were recruited in this study. Of these patients, two were excluded from the analysis as indicated in Figure 1. Ultimately, 101 were included (74.6% were male). The median age was 48.5 years (range: 17.0–88.0) (Table 1). Of the included patients 60.4% (61/101) were resident, and the most frequently observed comorbidity was diabetes (16/101, 15.8%) (Table 1). All participants in our study were HIV negative.

### Diagnostic Accuracy of TB-LAMP on Oral Swab Specimens

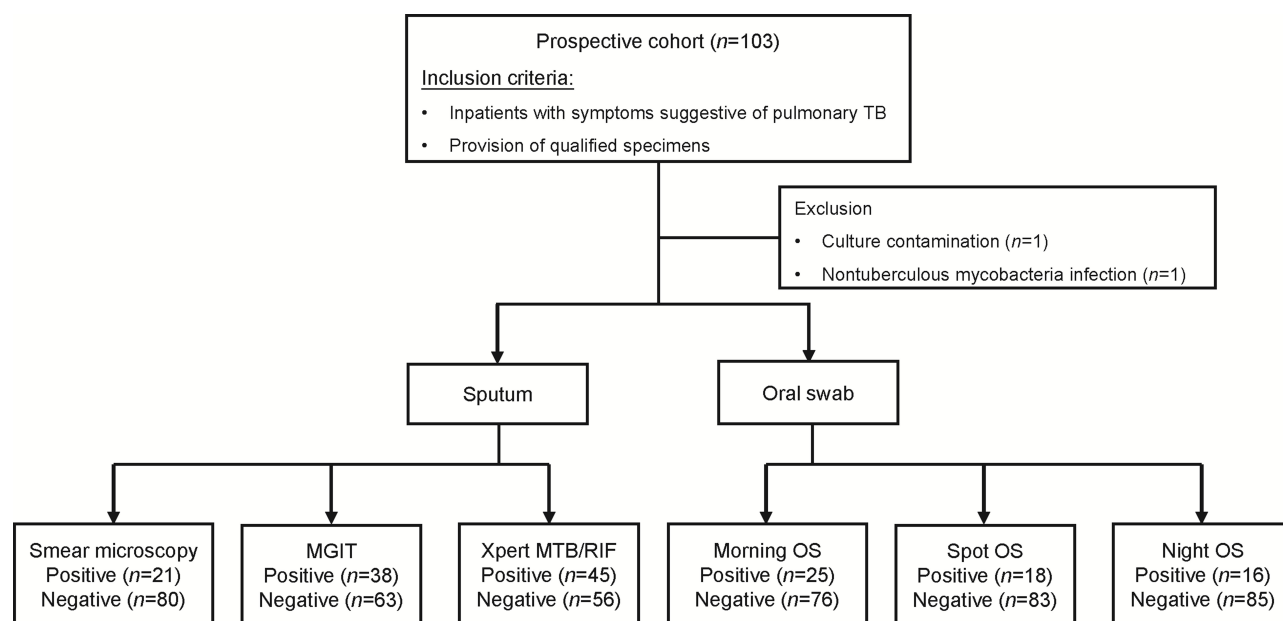
Among 101 specimens, Xpert and MGIT methods identified 45 and 38 MTB-positive cases, for detection rates of

**Table 1** Demographic and Clinical Characteristics of Individuals with Symptoms Suggestive of Pulmonary Tuberculosis

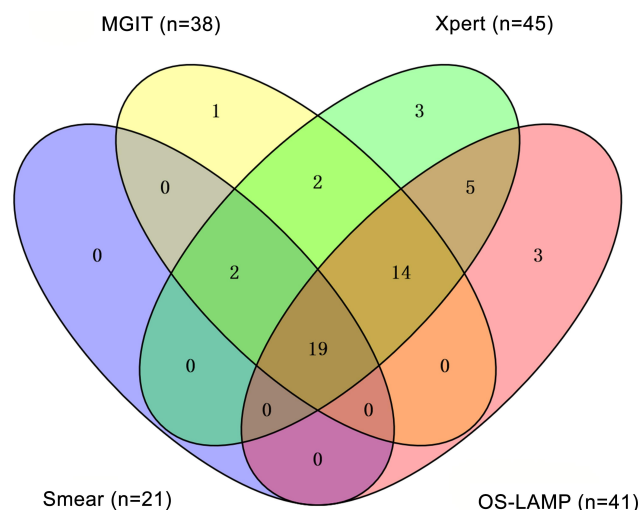
Characteristics	All Patients (n=101)
<b>Median age (range), years</b>	43.5 (17.0–88.0)
<b>Male sex, n (%)</b>	69 (74.6)
<b>Residence</b>	
Resident	61 (60.4)
Float	40 (39.6)
<b>Comorbidity/ies</b>	
Diabetes	16 (15.8)
Hypertension	10 (9.9)
Cardiac disease	8 (7.9)
Immunologic dysfunction	4 (4.0)
Kidney dysfunction	4 (4.0)
Hepatitis	3 (3.0)
Others	2 (2.0)

44.6% and 37.6%, respectively. The proportions of MTB-positive cases detected oral swab-TB-LAMP (OS-LAMP) were 24.8% (25/101), 15.8% (16/101) and 17.8% (18/101) for morning, night and spot specimens, respectively. With the combined results of three specimens, the proportion of MTB-positive cases achieved 40.6% (41/101), which was comparable to results for Xpert and MGIT (*P*=0.603).

The overlap between patients with a positive smear microscopy, Xpert, MGIT, or OS-LAMP is shown in Figure 2. Among 101 patients, there were 19 (18.8%)



**Figure 1** Participant enrollment.  
**Abbreviation:** OS, oral swab.



**Figure 2** Venn diagram showing the distribution of 101 patients for detection of MTB by using smear microscopy, MGIT, Xpert, and oral swab testing. Among 101 patients included, 41 were positive for MTB by oral swab testing, whereas, 45, 38, and 21 samples were positive by Xpert, MGIT and smear microscopy, respectively.

and 52 (51.5%) cases positive and negative for MTB, respectively by all four methods. Xpert detected MTB in total 45 cases either as alone or combined with the other three methods. Of 41 cases detected by OS-LAMP, three cases yielded negative Xpert and MGIT results, but were further diagnosed as confirmed laboratory TB patients from BALF specimens.

We further analyzed the performance of OS-LAMP for detection of MTB-positive cases using Xpert and MGIT results from sputum as gold standard. As summarized in Table 2, a total of 46 cases exhibited positive Xpert and/or MGIT results. The sensitivity of OS-LAMP on a single specimen ranged from 32.6% (15/46 95%CI: 19.1–46.2) in night oral swab to 50.0% (23/46, 95%CI: 35.6–64.4) in morning swab. The combination of three oral swab specimens correctly identified 38 MTB-positive cases, indicating an overall sensitivity of 82.6% (38/46, 95%CI: 71.7–

93.6), which was significantly higher than that of single oral swab specimen ( $P < 0.001$ ,  $P = 0.001$ ).

### Relationship of OS-LAMP Results with Bacterial Load

Of 45 cases with positive Xpert results, 38 had positive OS-LAMP results. The mean Ct values of LAMP positive group was  $21.5 \pm 1.0$  cycles, which was significantly lower than that of LAMP negative group ( $28.9 \pm 1.0$  cycles,  $P = 0.002$ ). Similarly, of 38 cases with positive MGIT results, the patients with negative OS-LAMP results ( $29.2 \pm 1.9$  days) had a longer time to detection of mycobacteria growth than the LAMP-positive group ( $16.8 \pm 1.1$  days,  $P < 0.001$ ) (Figure 3).

### Discussion

The diagnosis of TB is hampered by lack of sputum production or poor sample quality.<sup>16</sup> The use of alternative specimens is increasing in the diagnosis of this infectious disease. In this study, our data demonstrated that oral swab can be used as an alternative specimen for diagnosis of pulmonary TB using TB-LAMP. In a recent study from South Africa, two oral swab per patient exhibited a combined sensitivity of 92.8% relative to sputum Xpert assay by using a manual quantitative PCR targeting IS6110.<sup>11</sup> Along with the observed high sensitivity, the specificity of this assay was less than optimal due to sample contamination.<sup>11</sup> We hypothesize that this difference in sensitivity is likely because the manual PCR protocols were used in the latter study that resulted in remarkably high false positivity rate. Therefore the use of closed tube system in our analysis may better reflect the diagnostic yield of oral swab testing for pulmonary TB.

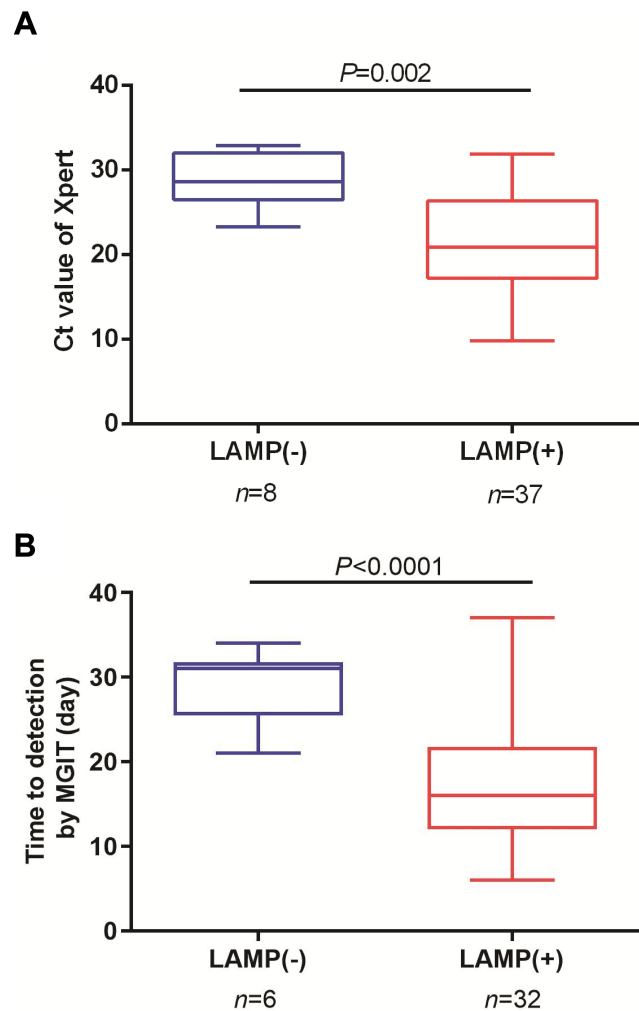
Another interesting finding of our study was the substantial lack of overlap in detection of MTB across three samples per patient. Although the morning oral swab

**Table 2** Detection of MTB with TB-LAMP from Various Oral Swab Samples

Specimen	n	TP	FP <sup>a</sup>	FN	TN	Sensitivity (95%CI)	Specificity (95%CI)	PPV (95%CI)	NPV (95%CI)
Morning	101	23	2	23	53	50.0 (35.6–64.4)	96.4 (91.4–100.0)	92.0 (81.4–100.0)	69.7 (59.4–80.1)
Night	101	15	1	31	54	32.6 (19.1–46.2)	98.2 (94.7–100.0)	93.8 (81.9–100.0)	63.5 (53.3–73.8)
Spot	101	17	1	29	54	37.0 (23.0–50.9)	98.2 (94.7–100.0)	94.4 (83.9–100.0)	65.1 (54.8–75.3)
Morning + Spot	101	31	3	15	52	67.4 (53.8–80.9)	94.5 (88.5–100.0)	91.2 (81.6–100.0)	77.6 (67.6–87.6)
Night + Spot	101	28	2	15	52	60.9 (46.8–75.0)	96.4 (91.4–100.0)	93.3 (84.4–100.0)	74.6 (64.5–84.8)
Morning + Night	101	32	2	14	53	69.6 (56.3–82.9)	96.4 (91.4–100.0)	94.1 (86.2–100.0)	79.1 (69.4–88.8)
All	101	38	3	8	52	82.6 (71.7–93.6)	94.5 (88.5–100.0)	92.7 (84.7–100.0)	86.7 (78.1–95.3)

**Note:** <sup>a</sup>FP is determined when Xpert and MGIT on sputum specimens are used as gold standard.

**Abbreviations:** TP, true positive; FP, false positive; FN, false negative; TN, true negative; PPV, positive predictive value; NPV, negative predictive value.



**Figure 3** Relationship of OS-LAMP results with bacterial load (A). Relationship of OS-LAMP results with bacterial load by Xpert; The Xpert-generated cycle threshold (Ct) value of Probe A was used as an indicator for bacterial load. (B) Relationship of OS-LAMP results with bacterial load by MGIT.

exhibited the highest sensitivity, OS-LAMP missed half of laboratory-confirmed pulmonary TB patients. In line with our data, the sensitivity of a single oral swab was 43% reported by Lima et al.<sup>17</sup> Considering that tubercle bacilli or its DNA are deposited on oral surfaces, this phenomenon suggests that the bacterial load in an oral swab specimen is significantly affected by daily oral activities such as eating, drinking and oral hygiene that decrease tubercle bacillus analytes on oral surface.<sup>11</sup> The morning oral swab should be collected prior to any oral activity to improve detection of MTB. In addition, the inclusion of more specimens at different times provides a practical alternative method for compensating the loss of diagnostic sensitivity with a single oral swab. Despite yielding the comparative performance as Xpert by the combination of three oral swab specimens, this raises concerns about

rising out-of-pocket costs for TB patients, especially patients in underdeveloped regions.<sup>18</sup> Further information is essential to inform the policy maker to formulate appropriate financial support strategy for incorporation OS-LAMP into the routine diagnostic algorithm.

As expected, we found that the false negative detection results by OS-LAMP were majorly associated with the low bacterial load in pulmonary TB patients, such as high C(t) values by Xpert and increased time to detection by MGIT. Notably, three patients who were Xpert negative and MGIT negative were identified as MTB-positive by OS-LAMP. Further analysis on BALF demonstrated that such patients were true TB cases. It is believed that the quality of sputum specimen has a great impact on diagnostic accuracy of MTB-positive TB patients. In China more than half of the samples were of insufficient quality.<sup>19</sup> Therefore the poor quality of sputum could be the main reason for missed detection of tubercle bacilli. This finding has several important implications: on the one hand, more effort should be made to collect good quality sputum samples that could guarantee the diagnostic accuracy; on the other hand, this highlights that oral swab testing could be used as an alternative to improve the detection of MTB for patients producing poor quality sputum.

BALF is considered as a promising alternative the commonly used sputum specimen for detecting MTB in patients with symptoms suggestive of TB;<sup>8</sup> however, it is worth mentioning that invasive diagnostic procedures hamper its implementation in resource-limited regions.<sup>8</sup> The easy-to-collect specimens are therefore a key priority for diagnostic development. If these criteria are applied, oral swab would be strongly favored over BALF for initial investigation of pulmonary TB, as the yield of OS-LAMP has been comparable to sputum Xpert.

We also acknowledged several obvious limitations to the present study. First, the major limitation was the single center recruitment of the small sample of individuals, which may weaken the significance of our conclusion. Further study is urgently needed to verify our data by enrollment of a large cohort of patients. Second, only TB-LAMP was used to detect tubercle bacilli in oral swab samples. Additional work is needed to adapt these samples for use on Xpert MTB/RIF Ultra, an automated diagnostic platform with higher sensitivity.<sup>20</sup> Third, as to the reference examinations, MGIT yielded fewer positive cases compared to Xpert. This phenomenon was mainly due to the high proportion of salivary sputum that limited the capability of MGIT culture in detecting



mycobacteria.<sup>19</sup> Fourth, the probability of true TB positives has a significant effect on the positive predictive value and negative predictive value. Considering that this study was conducted in the settings with high TB burden, the swab specimen showed acceptable performance. If prerequisite probability of tuberculosis was low, the swab specimen value may be lower than our observation. Fifth, the WHO raises target product profiles for pulmonary TB diagnostics that should meet for the performance and operational characteristics<sup>21</sup> Although the sensitivity and specificity of OS-LAMP achieved the minimal requirements, the unit price and laboratory infrastructure for maintaining functional LAMP further challenges its implementation in resource-poor countries. Finally, the feasibility for recovery of mycobacteria from oral swab was not assessed in our analysis. Despite these limitations, the results confirm that oral swab is sufficient to use as a promising alternative to sputum for diagnosis of pulmonary TB.

In conclusion, our data demonstrate that oral swab can be used as an alternative specimen for diagnosis of pulmonary TB using TB-LAMP. Morning oral swabs exhibit the highest sensitivity, and the inclusion of more specimens at different times provides compensation in diagnostic sensitivity with single oral swab. Future work is required to explore the utility of oral swabs for the diagnosis of TB with an automated diagnostic platform.

## Ethical Consideration

This study was conducted in accordance with the Declaration of Helsinki and approved by the Ethics Committee of Beijing Chest Hospital, Capital Medical University (No. 2019-86). The adults or the parents/legal guardians of patients under 18 years of age signed a written informed consent to agree with the anonymous use of clinical data.

## Consent for Publication

Not applicable.

## Funding

This work was supported by the Beijing Hospitals Authority' Ascent Plan (DFL20191601).

## Disclosure

The authors report no conflicts of interest in this work.

## References

1. World Health Organization. *Global Tuberculosis Report 2019*. WHO/HTM/TB/2019.13. Geneva: World Health Organization;2019.
2. Wang L, Zhang H, Ruan Y, et al. Tuberculosis prevalence in China, 1990–2010; a longitudinal analysis of national survey data. *Lancet*. 2014;383(9934):2057–2064. doi:10.1016/S0140-6736(13)62639-2
3. Floyd K, Glaziou P, Zumla A, Raviglione M. The global tuberculosis epidemic and progress in care, prevention, and research: an overview in year 3 of the End TB era. *Lancet Respir Med*. 2018;6(4):299–314. doi:10.1016/S2213-2600(18)30057-2
4. Theron G, Peter J, Meldau R, et al. Accuracy and impact of Xpert MTB/RIF for the diagnosis of smear-negative or sputum-scarce tuberculosis using bronchoalveolar lavage fluid. *Thorax*. 2013;68(11):1043–1051. doi:10.1136/thoraxjnl-2013-203485
5. Schoch OD, Rieder P, Tueller C, et al. Diagnostic yield of sputum, induced sputum, and bronchoscopy after radiologic tuberculosis screening. *Am J Respir Crit Care Med*. 2007;175(1):80–86. doi:10.1164/rccm.200608-1092OC
6. Sakundarno M, Nurjazuli N, Jati SP, et al. Insufficient quality of sputum submitted for tuberculosis diagnosis and associated factors, in Klaten district, Indonesia. *BMC Pulm Med*. 2009;9:16. doi:10.1186/1471-2466-9-16
7. Conde MB, Loivos AC, Rezende VM, et al. Yield of sputum induction in the diagnosis of pleural tuberculosis. *Am J Respir Crit Care Med*. 2003;167(5):723–725. doi:10.1164/rccm.2111019
8. Xu P, Tang P, Song H, et al. The incremental value of bronchoalveolar lavage for the diagnosis of pulmonary tuberculosis in a high-burden urban setting. *J Infect*. 2019;79(1):24–29. doi:10.1016/j.jinf.2019.05.009
9. Kordy F, Richardson SE, Stephens D, Lam R, Jamieson F, Kitai I. Utility of gastric aspirates for diagnosing tuberculosis in children in a low prevalence area: predictors of positive cultures and significance of non-tuberculous mycobacteria. *Pediatr Infect Dis J*. 2015;34(1):91–93. doi:10.1097/INF.0000000000000498
10. Menzies D. Sputum induction: simpler, cheaper, and safer—but is it better? *Am J Respir Crit Care Med*. 2003;167(5):676–677. doi:10.1164/rccm.2212008
11. Luabeya AK, Wood RC, Shenje J, et al. Noninvasive detection of tuberculosis by oral swab analysis. *J Clin Microbiol*. 2019;57(3).
12. Mesman AW, Calderon R, Soto M, et al. Mycobacterium tuberculosis detection from oral swabs with Xpert MTB/RIF ULTRA: a pilot study. *BMC Res Notes*. 2019;12(1):349. doi:10.1186/s13104-019-4385-y
13. Ou X, Li Q, Xia H, et al. Diagnostic accuracy of the PURE-LAMP test for pulmonary tuberculosis at the county-level laboratory in China. *PLoS One*. 2014;9(5):e94544. doi:10.1371/journal.pone.0094544
14. Wood RC, Luabeya AK, Weigel KM, et al. Detection of Mycobacterium tuberculosis DNA on the oral mucosa of tuberculosis patients. *Sci Rep*. 2015;5:8668. doi:10.1038/srep08668
15. Xia H, Song YY, Zhao B, et al. Multicentre evaluation of Ziehl-Neelsen and light-emitting diode fluorescence microscopy in China. *Int J Tuberc Lung Dis*. 2013;17(1):107–112. doi:10.5588/ijtld.12.0184
16. Ho J, Marks GB, Fox GJ. The impact of sputum quality on tuberculosis diagnosis: a systematic review. *Int J Tuberc Lung Dis*. 2015;19(5):537–544. doi:10.5588/ijtld.14.0798
17. Lima F, Santos AS, Oliveira RD, et al. Oral swab testing by Xpert (R) MTB/RIF Ultra for mass tuberculosis screening in prisons. *J Clin Tuberc Other Mycobact Dis*. 2020;19:100148. doi:10.1016/j.jctube.2020.100148
18. McAllister SM, Wiem Lestari B, Sullivan T, et al. Out-of-pocket costs for patients diagnosed with tuberculosis in different healthcare settings in Bandung, Indonesia. *Am J Trop Med Hyg*. 2020;103:1057–1064. doi:10.4269/ajtmh.19-0848

19. Shi J, Dong W, Ma Y, et al. GeneXpert MTB/RIF outperforms mycobacterial culture in detecting mycobacterium tuberculosis from salivary sputum. *Biomed Res Int.* 2018;(2018):1514381.
20. Dorman SE, Schumacher SG, Alland D, et al. Xpert MTB/RIF Ultra for detection of Mycobacterium tuberculosis and rifampicin resistance: a prospective multicentre diagnostic accuracy study. *Lancet Infect Dis.* 2018;18(1):76–84. doi:10.1016/S1473-3099(17)30691-6
21. *High-Priority Target Product Profiles for New Tuberculosis Diagnostics: Report of a Consensus Meeting.* Geneva: World Health Organization. WHO/HTM/TB/2014.18. 2014.

## Infection and Drug Resistance

Dovepress

### Publish your work in this journal

Infection and Drug Resistance is an international, peer-reviewed open-access journal that focuses on the optimal treatment of infection (bacterial, fungal and viral) and the development and institution of preventive strategies to minimize the development and spread of resistance. The journal is specifically concerned with the epidemiology of

antibiotic resistance and the mechanisms of resistance development and diffusion in both hospitals and the community. The manuscript management system is completely online and includes a very quick and fair peer-review system, which is all easy to use. Visit <http://www.dovepress.com/testimonials.php> to read real quotes from published authors.

Submit your manuscript here: <https://www.dovepress.com/infection-and-drug-resistance-journal>