**Results.** Thirty-seven of 70 (53%) of patients received vancomycin over 61 courses with a mean duration of 8 days; 14 (23%) of these courses were with neutropenic fever. No indication was documented by the provider for 21 (34%) vancomycin courses (Figure 1). Almost half of all courses given for neutropenic fever did not meet guideline indications (Figure 2). Adverse effects occurred in 19 (31%) of vancomycin courses, including 11 (18%) associated with acute kidney injury.

Vancomycin was associated with reduced relative abundance of organisms correlated with reduced risk of subsequent severe acute graft vs. host disease and *Clostridium scindens*, an organism protective against *C. difficile* infection (CDI) (Figure 3, in bold).

**Conclusion.** Indications for vancomycin were poorly documented and infrequently guideline based. Adverse events occurred in 1 in 3 courses of vancomycin. Vancomycin correlated with microbiome changes which have been associated with increased risk for aGVHD and CDI.



Disclosures. S. Pergam, Merck: Consultant, Consulting fee.

# 617. Characterization and Development of the Infant Gut Virome: A STORK Study

Andrea Granados, PhD<sup>1</sup>; Susanna K. Tan, MD<sup>2</sup>; Jerome Bouquet, PhD<sup>3</sup>; Charles Y. Chiu, MD, PhD<sup>1</sup>; Julie Parsonnet, MD, FIDSA<sup>4</sup> and Lauri Green, BSc<sup>3</sup>; <sup>1</sup>Department of Laboratory Medicine, University of California, San Francisco, San Francisco, California, <sup>2</sup>Medicine, Division of Infectious Diseases, Stanford University School of Medicine, Stanford, California, <sup>3</sup>University of California, San Francisco, San Francisco, California, <sup>4</sup>Department of Medicine, Division of Infectious Diseases and Geographic Medicine, Stanford University School of Medicine, Stanford, California

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**Background.** There is little known about the dynamics of the infant virome and how it relates to healthy growth and development. This study will establish the baseline gut virome and observe dynamic changes in a cohort of infants from birth to 3 years old. We hypothesize that changes in the gut virome will impact growth and immune development.

Methods. One hundred and twenty-eight infants were enrolled in the Stanford's Outcome Research in Kids (STORK) cohort prior to 36 weeks gestation. Stool samples were collected at an average of 90, 134, 162 days old/infant. Baseline data were collected at birth (height, weight, length, Apgar's score, antibiotic use) and health surveys were collected weekly. Stool samples (n = 477) were extracted using the EZ1 Viral Kit (Qiagen). Libraries were prepared using the Nextera XT kit (Illumina) and sequenced on an Illumina HiSeq 2500 on rapid mode (150/150 bp paired-end sequencing). Datasets were analyzed using SURPI; a bioinformatic pipeline for pathogen detection.

**Results.** A subset of the infants were tested (n = 27), 54% of which were male. The infants were 62% white, Hispanic, 26% white, non-Hispanic, 8% Asian, and 4% other. Seventy-five stool samples—sequenced at an average depth of 22 million reads—were analyzed from the 27 infants. Vertebrate viruses (42.8%) and phages (45.2%) represented the majority of the viral reads, while the other reads were invertebrate, plants or protozoa (12%). Virome abundance, richness, and diversity were 5.5e+04 species reads per million, 55.5 on the Chao Richness scale, and 1.45 on the Shannon Diversity Index respectively, with values increasing as the infants aged. The phage families most commonly identified were Myoviridae, Podoviridae, and Siphoviridae. There were seven different human viral families observed: Adenoviridae, Astroviridae, Caliciviridae, Parvoviridae, Picornaviridae, Reoviridae, and Anelloviridae. Five infants were documented to have cold symptoms within 7 days of sampling, they were found to have mastadenovirus C (1), mamastrovirus 1 (1), bocavirus (3). Three infants were documented to have caliciviruses (2) and adenovirus (1); however, no symptoms were reported.

*Conclusion.* This study will comprehensively characterize the development of the human virome and monitor its effect on growth and immune development. *Disclosures.* All authors: No reported disclosures.

#### 618. Do Clinical Factors Affect Microbial Engraftment After Fecal Microbiota Transplantation in Recurrent *Clostridium difficile* Infection?

<u>Shrish Budree</u>, MD<sup>1,2</sup>; Majdi Osman, MD MPH<sup>1</sup>; Pratik Panchal, MD<sup>3</sup>; Edina Shu, <u>BS<sup>4</sup></u>; Madeline Carrellas, BA<sup>4</sup>; Zain Kassam, MD MPH<sup>5</sup> and Jessica Allegretti, MD, MPH<sup>4</sup>; <sup>1</sup>OpenBiome, Somerville, Massachusetts, <sup>2</sup>Pediatrics, University of Cape Town, Cape Town, South Africa, <sup>3</sup>Clinical Research, OpenBiome, Somerville, Massachusetts, <sup>4</sup>Gastroenterology, Brigham and Women's Hospital, Boston, Massachusetts, <sup>5</sup>Finch Therapeutics, Somerville, Massachusetts

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**Background.** Fecal microbiota transplantation (FMT) is an effective treatment for recurrent *Clostridium difficile* infection (rCDI). Few studies have evaluated clinical factors associated with microbial engraftment. We describe microbial changes post-FMT and clinical factors impacting engraftment.

**Results.** A total of 12 patients received an FMT from 12 unique donors. The efficacy rate was 92%. Mean recipient age was 60 years (range: 33–87) with more females (7/12).

Recipients pre-FMT alpha diversity was significantly lower compare to donors (P = 0.04, Figure 1a). This difference dissipated post-FMT (P = 0.67). On  $\beta$ -diversity analysis, the recipients pre-FMT samples clustered separately from their post-FMT samples (P = 0.01, Figure 1b), with the post-FMT samples shifting closer to the donor samples. *Proteobacteria* was dominant in patients' pre-FMT samples and were substantially reduced post-FMT, combined with an expansion in *Bacteroidetes* (Figure 2).

On linear regression analysis, clinical factors (age, sex, previous recurrent CDI episodes, inflammatory bowel disease, proton pump inhibitor, immunosuppression, previous anti-CDI antibiotic courses, probiotics) were not significantly associated with engraftment outcomes.

**Conclusion.** There is a significant and durable shift in recipients' microbial profile to resemble their donor post-FMT. Recipients' pre-FMT clinical factors did not significantly affect microbial engraftment. Future metagenomic studies may help elucidate whether clinical factors impact engraftment.



Figure 1: (A) Alpha diversity. (B) Bray–Curtis principle component plot.





## 619. Intestinal Microbiome Changes Associated with Immune Status and *Clostridium difficile* Colonization in Hospitalized Children

Sindhu Mohandas, MD<sup>1</sup>; Vijaya L Soma, MD<sup>2</sup>; Tresa Ambooken, MD<sup>3</sup>; David Goldman, MD<sup>4</sup>; Dong-Ninh Tran, PhD<sup>5</sup>; George Weinstock, PhD<sup>5</sup>; Erica Sodergren, PhD<sup>5</sup> and Betsy C. Herold, MD, FIDSA, FPIDS<sup>6</sup>; <sup>1</sup>Children<sup>s</sup> Hospital of Wisconsin, Milwaukee, Wisconsin, <sup>2</sup>Division of Pediatric Infectious Disease, Children<sup>s</sup> Hospital at Montefiore, Bronx, New York, <sup>3</sup>Pediatrics, Bronx Lebanon hospital, Bronx, New York, <sup>4</sup>Children<sup>s</sup> Hospital at Montefiore, Bronx, New York, <sup>5</sup>Jackson Laboratory for Genomic Medicine, Farmington, Connecticut, <sup>6</sup>Department of Pediatrics and Microbiology-Immunology, Albert Einstein College of Medicine, Bronx, New York

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**Background.** The intestinal microbiome modulates local and systemic immune responses and may impact clinical outcomes. However, there are few studies in pediatric patients. We conducted a cross-sectional study of fecal microbiomes in hospitalized children on a single inpatient unit at Children's Hospital at Montefiore, Bronx, New York in 2016–2017 to test the hypothesis that "high-risk" children with chronic illnesses (cancer, transplant and sickle cell disease [SCD]) have decreased microbial diversity and higher rates of asymptomatic colonization with *C. difficile* compared with children hospitalized on the same ward but without similar risk factors.

*Methods.* Stool was collected within 72 hours of admission from patients who provided consent and assayed for *C. difficile* colonization by glutamate dehydrogenase (GDH); microbiome analysis was performed by 16S rRNA sequencing. Clinical and demographic data were obtained from the EMR.

**Results.** One hundred and six unique patients provided a sample for analysis. Sixtynine were categorized as high-risk, including 32 SCD patients. *C. difficile* colonization rates were 22% and 19% in the high-risk and low-risk groups, respectively, but highest in the subset of SCD patients on penicillin prophylaxis (33%). The high-risk group had a trend toward lower microbial diversity than controls, and SCD patients exhibited a diversity index greater than other high-risk patients. Antibiotic use in the last 3 months and PPI use were associated with decreased microbial diversity across the entire study population (P = 0.004, P = 0.007, respectively). Among children with SCD, those on penicillin prophylaxis had a trend toward decreased alpha diversity while folic acid was associated with increased diversity (P = 0.02). SCD patients had greater quantities of *Bacteroides* and *Parabacteroides* and fewer *Escherichia* and *Shigella* than the other cohorts.

**Conclusion.** SCD and penicillin prophylaxis might be risk factors for *C. difficile* colonization and intestinal dysbiosis. The implications of these findings require further, longitudinal study.

Disclosures. All authors: No reported disclosures.

#### 620. Oral β-Lactamase Therapies Prevent Microbiome Damage and Attenuate Antibiotic Resistance From IV and Oral Antibiotics in Large Animal Models of Antibiotic-Mediated Gut Dysbiosis

Sheila Connelly, PhD<sup>1</sup>; Christian Furlan-Freguia, PhD<sup>1</sup>; Brian Fanelli, BS<sup>2</sup>; Nur A. Hasan, PhD<sup>2</sup>; Rita R. Colwell, PhD<sup>2</sup> and Michael Kaleko, MD, PhD<sup>1</sup>; <sup>1</sup>Synthetic Biologics, Inc., Rockville, Maryland, <sup>2</sup>CosmosID, Inc., Rockville, Maryland

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**Background.** Antibiotics can damage the gut microbiome leading to overgrowth of pathogens and provide selective pressure for emergence of antibiotic resistance. SYN-004 (ribaxamase) is a clinical-stage  $\beta$ -lactamase formulated for oral delivery intended to degrade certain  $\beta$ -lactam antibiotics in the GI tract to preserve the gut microbiome. Ribaxamase was evaluated in a phase 2b clinical study that met its primary endpoint of

significantly reducing *C. difficile* infection in patients treated with IV ceftriaxone and demonstrated protection of the gut microbiome with reduced emergence of antibiotic resistance. Ribaxamase is intended for use with IV penicillins and cephalosporins, but does not degrade carbapenems. B-lactamase-mediated microbiome protection was expanded to include oral and carbapenem antibiotics.

Methods. For use with oral  $\beta$ -lactams, a ribaxamase formulation, SYN-007, was engineered for release in the lower small intestine, distal to the site of antibiotic absorption. For use with IV carbapenems, SYN-006, a novel metallo- $\beta$ -lactamase, was formulated for oral delivery. SYN-007 (10 mg, PO, TID) was evaluated in dogs treated with oral amoxicillin (40 mg/kg, PO, TID) for 5 days. SYN-006 (50 mg, PO, QID) was evaluated in pigs treated with ertapenem (30 mg/kg, IV, SID) for 4 days. Serum antibiotic levels were measured and fecal DNA whole-genome shotgun sequence analyses were performed.

**Results.** In dogs and pigs, systemic antibiotic levels were not significantly different ±SYN-007 or SYN-006. Fecal DNA metagenomics analyses demonstrated that oral amoxicillin and IV ertapenem resulted in significant changes to the gut microbiome. SYN-007 and SYN-006 attenuated microbiome damage and reduced emergence of antibiotic resistance.

**Conclusion.** Ribaxamase, SYN-007, and SYN-006 have the potential to protect the commensal gut microbiota from antibiotic-mediated collateral damage and to mitigate emergence and spread of antibiotic resistance, thereby broadening the utility of this prophylactic approach to include all classes of  $\beta$ -lactam antibiotics, delivered both systemically and orally. Antibiotic inactivation represents a new paradigm for preservation of the gut microbiome and reduction of antibiotic resistance.



Oral beta-lactamases do not affect systemic antibiotic levels and protect the gut microbiome from antibiotic-mediated damage. Serum was tested for amoxicillin or ertapenem using LCMSNAS analysis. Area under the curve was not significantly different between orale moxicilli amos vs. oral amoxicillin = VSN-007 (p=07.03) of for V trappenem and oral. V ertapenem + SYN-006 (p=07.42), indicating that the beta-lactamases did not affect serum antibiotic levels (eff panels). Focal DNA collected before and after antibiotic administration was subjected to metagenomics sequencing analyses. Shannon sphe diversity ratios (pertexament/post-treatment) were significantly different for onal amoxicillin alone vs. oral amoxicillin = SYN-007 (p=0.0007). Biot SYN-007 not SYN-007 (p=0.0003), and for 1) redicating minimal change in diversity of the gut microbiome pre and post antibiotic exposure (right panels). These data demonstrate tat or alb sta-tamases protectine gut microbiome pre and post antibiotic biod(levels).

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# **621. Treatment of Recurrent** *Clostridium difficile* Infection with SER-109 Increases the Concentration of Secondary Bile Acids in a Dose-Dependent Manner Matthew Henn, PhD<sup>1</sup>; Christopher Ford, PhD<sup>1</sup>; Edward O'Brien, PhD<sup>1</sup>; Jennifer Wortman, PhD<sup>1</sup>; Liyang Diao, PhD<sup>1</sup>; Christopher Desjardins, PhD<sup>1</sup>; Amelia Tomlinson, PhD<sup>1</sup>; Kevin Litcofsky, PhD<sup>1</sup>; Mark Wilcox, MD<sup>2</sup>; Anthony Buckley, PhD<sup>3</sup>; Patricia Bernardo, ScD<sup>1</sup>; Barbara McGovern, MD<sup>1</sup>; John G. Aunins, PhD<sup>1</sup>; David N. Cook, PhD<sup>1</sup> and Michele Trucksis, PhD, MD<sup>1</sup>; <sup>1</sup>Seres Therapeutics, Inc., Cambridge, Massachusetts, <sup>2</sup>Leeds Teaching Hospitals & University of Leeds, Leeds, UK

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**Background.** C. difficile recurs when dormant spores germinate in the dysbiotic gut, facilitated by an increase of 1° vs. 2° bile acids. SER-109, an ecology of bacterial spores purified from stool of healthy donors, is an investigational first-in-class microbiome therapeutic intended to facilitate microbiome restoration and reduce risk of recurrent *C. difficile* (rCDI). Rapid engraftment of spore-forming species is associated with (i) higher doses of SER-109 in our dose-ranging Phase 1b study (Ph1b) and (ii) reduced rCDI in our Phase 2 trial (Ph2). We explored whether higher doses of SER-109 were associated with an increase in 2° bile acids.

**Methods.** Whole metagenomic shotgun (WMS) data were generated from stool, and species were identified using a proprietary build of MetaPhlAn. Evaluation of spore-forming species richness and bile acid concentrations identified effects of SER-109 treatment. A triple stage bioreactor model of the human gut and rCDI was used to evaluate the impact of microbial therapeutics.

**Results.** Ph1b subjects who received a higher dose (>1.5 × 10<sup>8</sup> SporQ) had significantly higher spore-forming species richness than subjects who received a low dose (<1.5 × 10<sup>8</sup> SporQ) at Week 1 post-treatment (*P* = 0.017, Figure 1). Spore-forming species richness in patients receiving a low dose in Ph1b was comparable to that observed in non-recurrent patients in Ph2, who received the same mean dose (Figure 1). Ph1b subjects in the high dose group had a significantly higher concentration of 2° bile acids as compared with Ph1b low dose subjects and non-recurrent Ph2 subjects (*P* = 0.036, *P* < 0.001, respectively, Figure 2). A higher dose (3 × 10<sup>8</sup> SporQ × 3 days) suppressed recurrence in a gut model of rCDI; a single dose did not.

**Conclusion.** Higher doses of SER-109 are significantly associated with (i) higher spore-forming species richness, (ii) concentrations of secondary bile acids, and (iii) prevention of recurrence in an gut model of CDI. These results suggest that SER-109 in the Phase 2 trial was biologically active and catalyzed a functional change in the microbiome of a subset of subjects; a dose increase may optimize efficacy across a broad population. Seres has initiated a Phase 3 study of SER-109 to reduce rCDI, which includes an increase in dose titer and frequency.