

Review

Estrogen Sulfotransferase (SULT1E1): Its Molecular Regulation, Polymorphisms, and Clinical Perspectives

MyeongJin Yi ¹, Masahiko Negishi ¹ and Su-Jun Lee ^{2,*}

- ¹ Pharmacogenetics Section, Reproductive and Developmental Biology Laboratory, National Institute of Environmental Health Sciences, National Institutes of Health, Research Triangle Park, NC 27709, USA; myeongjin.yi@nih.gov (M.Y.); negishi@niehs.nih.gov (M.N.)
- ² Department of Pharmacology and Pharmacogenomics Research Center, Inje University College of Medicine, Inje University, Bokji-ro 75, Busanjin-gu, Busan 47392, Korea
- * Correspondence: 2sujun@inje.ac.kr; Tel.: +82-51-890-8665

Abstract: Estrogen sulfotransferase (SULT1E1) is a phase II enzyme that sulfates estrogens to inactivate them and regulate their homeostasis. This enzyme is also involved in the sulfation of thyroid hormones and several marketed medicines. Though the profound action of SULT1E1 in molecular/pathological biology has been extensively studied, its genetic variants and functional studies have been comparatively rarely studied. Genetic variants of this gene are associated with some diseases, especially sex-hormone-related cancers. Comprehending the role and polymorphisms of SULT1E1 is crucial to developing and integrating its clinical relevance; therefore, this study gathered and reviewed various literature studies to outline several aspects of the function, molecular regulation, and polymorphisms of SULT1E1.

Keywords: estrogen sulfotransferase; SULT1E1; estrogen; estrogen sulfate; thyroid hormones; breast cancer; endometrial cancer; polymorphism



Citation: Yi, M.; Negishi, M.; Lee, S.-J. Estrogen Sulfotransferase (SULT1E1): Its Molecular Regulation, Polymorphisms, and Clinical Perspectives. *J. Pers. Med.* **2021**, *11*, 194. <https://doi.org/10.3390/jpm11030194>

Academic Editor: Daryl Pritchard

Received: 11 January 2021
Accepted: 8 March 2021
Published: 11 March 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

The metabolism of endogenous compounds and hormones is important in physiological homeostasis. Sulfation, which occurs in many metabolic pathways, generates sulfoconjugated forms that are typically regarded as inactive metabolites. Sulfation is one of the phase II metabolizing pathways that represent the inactivation of hormones, such as estrogens, and this reaction is usually performed by an enzyme from the cytosolic enzyme group referred to as sulfotransferases (SULTs) [1]. In various mammals, such as mice and rats, SULTs play the essential roles of sulfating estrogens, thyroid hormones, bile acids, and other xenobiotics [2–6].

Thirteen cytosolic SULTs have been identified in humans (Table 1). Sulfotransferases facilitate the SN₂-like displacement/transfer reaction of a sulfonate (SO₃[−]) group from the ubiquitous donor 3'-phosphoadenosine 5'-phosphosulfate (PAPS) to hydroxyl or amino residues of acceptor substrates [7,8]. Many active sites of SULTs are conserved; they are the same in all known crystal structures of human and mouse SULTs [9–13].

Sulfotransferase isoforms sulfate not only xenobiotics (e.g., flavonoids and hydroxyl metabolites of anticancer drugs) but also endogenous compounds, such as steroid hormones. They have essential roles in the homeostasis of bile acids, thyroid hormone, androgens, and estrogens, and their expressions are influenced by substrates and pathological conditions as well [14].

Among the SULT isoforms, SULT1E1 has the lowest K_m values for estrone (E₁), estradiol (E₂), and catecholesterogen sulfation [15–19]. This enzyme had been referred to as EST (estrogen sulfotransferase) due to its substantial role in estrogen inactivation. It was discovered and cloned in other mammalian species (Table 2), and its amino acid sequence

homologies regarding rabbit, horse, pig, mouse, cow, and rat EST compared with human EST are 82.6%, 79.2%, 77.8%, 77.5%, 73.7%, and 71.3%, respectively.

Table 1. Human sulfotransferase (SULT) isoforms.

Gene ID ¹	Locus ²	Alias ¹	Number of Amino Acids ³	Number of Exons ¹
SULT1A1	Chr 16p11.2	HAST1/HAST2, P-PST, PST, ST1A1, ST1A3, STP, STP1, TSPST1	295 (isoform a) 217 (isoform b)	13
SULT1A2	Chr 16p11.2	HAST4, P-PST, P-PST 2, ST1A2, STP2, TSPST2	295 (isoform 1) 262 (isoform 2)	8
SULT1A3	Chr 16p11.2	HAST, HAST3, M-PST, ST1A3, ST1A3/ST1A4, ST1A4, ST1A5, STM, TL-PST	295	8
SULT1A4	Chr 16p11.2	HAST3, M-PST, ST1A3, ST1A3/ST1A4, ST1A4, STM, TL-PST	295	8
SULT1B1	Chr 4q13.3	ST1B1, ST1B2, SULT1B2	296	10
SULT1C2	Chr 2q12.3	ST1C1, ST1C2, SULT1C1, humSULTC2	296 (isoform a) 307 (isoform b)	9
SULT1C3	Chr 2q12.3	ST1C3	304 (isoform 1) 304 (isoform 2)	10
SULT1C4	Chr 2q12.3	SULT1C, SULT1C2	302 (isoform 1) 227 (isoform 2)	7
SULT1E1	Chr 4q13.3	EST, EST-1, ST1E1, STE	294	9
SULT2A1	Chr 19q13.33	DHEA-ST, DHEA-ST8, DHEAS, HST, ST2, ST2A1, ST2A3, STD, SULT2A3, hSTa	285	6
SULT2B1	Chr 19q13.33	ARCI14, HSST2	350 (isoform a) 365 (isoform b)	7
SULT4A1	Chr 22q13.31	BR-STL-1, BRSTL1, DJ388M5.3, NST, SULTX3, hBR-STL-1	284	11
SULT6B1	Chr 2p22.2	ST6B1	304 (isoform 1) 265 (isoform 2)	9
SUP1C2P1	Chr 2q12.3	SULT1C1P	pseudogene	4
SULT1C2P2	Chr 2q12.3		pseudogene	
SULT1D1P	Chr 4q13.3	SULT1D1	pseudogene	
SULT6B2P	Chr 12p12.1		pseudogene	

¹ Information is described according to NCBI Gene. ² All reference loci were based on the GRCh38 assembly. ³ The way to divide genes into isoform a/b or 1/2 was described in accordance with the NCBI Protein database.

Table 2. SULT1E1 expression in other mammalian species.

Species	RefSeq ¹	RefSeq mRNA ²	RefSeq Protein ³	Number of Exons ¹
<i>Homo sapiens</i> (human)	NC_000004.12	NM_005420.3	NP_005411.1	9
<i>Mus musculus</i> (mouse)	NC_000071.7	NM_023135.2	NP_075624.2	8
<i>Rattus norvegicus</i> (rat)	NC_005113.4	NM_012883.2	NP_037015.2	10
<i>Bos taurus</i> (cow)	NC_037333.1	NM_177488.3	NP_803454.2	9
<i>Oryctolagus cuniculus</i> (rabbit)	NC_013683.1	XM_002717123.2	XP_002717169.1	8
<i>Sus scrofa</i> (pig)	NC_010450.4	NM_213992.1	NP_999157.1	9
<i>Equus caballus</i> (horse)	NC_009146.3	NM_001081918.1	NP_001075387.1	8

¹ Information is described according to the NCBI Gene database. ² All reference mRNA sequences were based on the NCBI Nucleotide database. ³ All reference protein sequences were based on the NCBI Protein database.

Although the SULT1A subfamily can sulfate estrogens, their affinity for endogenous estrogens is significantly lower than the affinity of SULT1E1 for those substrates. Moreover, SULT1E1 engages in the sulfation of thyroid hormones alongside the SULT1A subfamily. Though the important roles and regulation of SULT1E1 have been identified and stressed, functional studies related to genetic variants are relatively limited.

This review concentrates on the expression, functional characterization, regulation, associations with diseases, and genetic polymorphisms of SULT1E1.

2. Expression of SULT1E1

Human SULT1E1 cDNA was first isolated, cloned, and characterized from the liver, and its localization was mapped to human chromosome 4 [20]. SULT1E1 is expressed in the human embryo, and is also highly expressed in a wide range of fetal tissues, such as the liver, lung, kidney, and hormone-dependent tissues—such as the testis or endometrium—but its expression in adults with normal status is much lower than in the fetus and placenta [21,22]. The expression of SULT1E1 varies widely in the human population, although it is not known whether this is under genetic control or not [23]. Thus, it is possible that the variability in SULT1E1 expression results from different chemical influences.

Two agonists of peroxisome-proliferator-activated receptor α (PPAR α), WY14643 and IGF-1, show different regulatory effects on the SULT1E1 promoter activity. While WY14643 suppressed SULT1E1 activity, IGF-1 upregulated it, as measured by estrogen levels in endothelial cells and smooth muscle cells [24]. Interestingly, SULT1E1 was attenuated by both transfection with PPAR γ small interfering RNA (siRNA) and exposure to GW9662, the PPAR γ antagonist [25].

SULT1E1 regulation was observed when hepatocyte nuclear factor 4 α (HNF4 α) was silenced. The significant suppression of both mRNA and protein levels of SULT1E1 occurred via Farnesoid X receptor (FXR) agonists in HepG2 cells [26]. This finding confirmed that the effect of FXR on E₂ was SULT1E1-dependent. In patients with obstructive cholestasis, the accumulation of bile acids (activator of FXR) led to reduced mRNA and protein expression of hepatic SULT1E1, increased serum E₂ levels, and decreased serum estrone sulfate concentration [27]. Phosphorylated ROR α takes a regulatory signal to HNF4 α , and then activates the *SULT1E1* promoter in human liver cells [28].

Basal expression of SULT1E1 in the liver is relatively low [29], but its expression and role could be impacted in response to ligands/substrates for nuclear receptors, such as the liver X receptor (LXR) [29], the glucocorticoid receptor (GR) [30], the constitutive androstane receptor (CAR) [31], the estrogen receptor α (ER α) [32], the pregnane X receptor (PXR) [33], and the RAR-related orphan receptor α (ROR α) [34] (Table 3).

Table 3. The nuclear receptors associated with *Sult1e1* regulation.

Gene ID	Nuclear Receptor	Species	Tissue	Reference
NR3A1	ER α	Mouse	Liver tissue	[32]
NR3C1	GR	Mouse	Liver tissue	[30]
NR1C1	PPAR α	Human	Vascular endothelial cell Smooth muscle cell	[24]
NR1C3	PPAR γ	Human	Endothelial cell	[25]
NR1H2, H3	LXR	Mouse	Uterine	[29]
NR1H4	FXR	Human	Liver cell line	[26]
		Human	Liver tissue	[27]
		Human	Liver cell line	
NR1I2	PXR	Mouse	Liver tissue	[33]
NR1I3	CAR	Mouse	Liver tissue	[31,32]
NR2A1	HNF4 α	Human	Liver tissue	[27]
		Human	Liver cell line	[28]
NR1F1	ROR α	Mouse	Liver tissue	[34]

3. Sulfation of Estrogens and Thyroid Hormones by SULT1E1

3.1. Sulfation of Estrogens

Estrogens play fundamental roles in a variety of physiological systems. It has been widely established that estradiol (E₂) exposure is one of the risk factors for breast carcinogenesis. One of the critical pathways for E₂ inactivation is sulfation by SULT1E1. Estrone (E₁) is synthesized by aromatization of androstenedione and is subsequently sulfated. After E₁ is desulfated and subsequently turned into E₂ by the 17 β -hydroxysteroid dehydrogenases (17 β -HSD), E₂ can then be sulfated through SULT1E1 [35]. As previously mentioned, SULT1E1 is a cytosolic enzyme that catalyzes estrogen sulfation at the 3-hydroxyl site while using PAPS as a sulfate donor (Figure 1). Moreover, this enzyme has high affinity for its substrate E₂, indicating its crucial role in modulating estrogen's action and homeostasis [36].

SULT1E1 has shown the distinct characteristic of having a high sulfating affinity for not only E₂, but also other estrogens, such as E₁ and ethinylestradiol (EE₂), with nanomolar K_m values (Table 4). Due to its high affinity for sulfate estrogens, SULT1E1 exhibits inhibition of substrate with increasing E₂ and E₁ concentrations. SULT1E1 is also used to sulfate other compounds, namely dehydroepiandrosterone (DHEA), pregnenolone, diethylstilbestrol (DES), and equilenin [37,38].

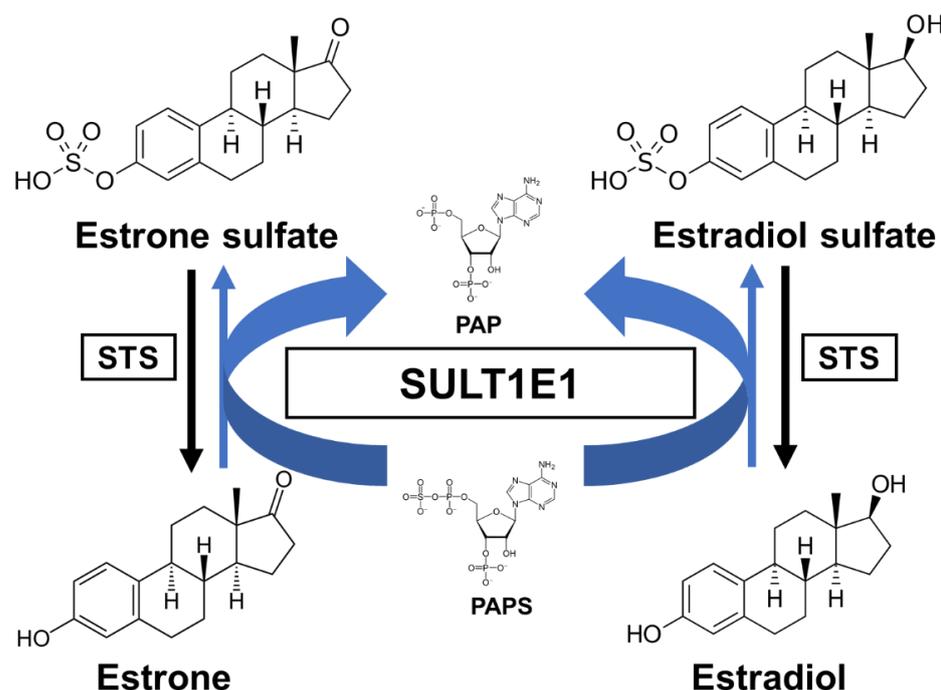


Figure 1. A schematic sulfation pathway of estrogens. STS, steroid sulfatase; PAPS, 3'-phosphoadenosine 5'-phosphosulfate; PAP, 3'-phosphoadenosine 5'-phosphate.

Table 4. Substrates of SULT1E1.

Substrate	Compound Characteristics	K_m	Reference
E ₁	Agonist of ER	~0.17 μ M	[39]
E ₂	Most active agonist of ER	5 \pm 0.8 nM	[16]
EE ₂	Agonist of GPER and ER	29 nM	[38]
EE ₂	Agonist of GPER and ER	6.7 \pm 0.1 nM	[40]
DHEA	Partial agonist of AR and ER	~0.85 μ M	[37]
T ₄	Thyroid prohormone	4.57 \pm 0.07 μ M	[40]
T ₃	Receptor active iodothyronine	22.6 \pm 1.0 μ M	
rT ₃	Receptor inactive iodothyronine	25.7 \pm 10.4 μ M	[41]
T ₂	Breakdown metabolite of triiodothyronine	2.15 \pm 1.45 μ M	
Apigenin	Common dietary flavonoid	4.75 \pm 1.25 μ M	
Epicatechin	Antioxidative flavonoid	5.3 \pm 0.65 μ M	
Resveratrol	Antioxidative flavonoid	0.96 \pm 0.17 mM	
Chrysin	Flavonoid in bee pollen or propolis	6.88 \pm 1.12 μ M	[42]
Quercetin	Flavonoid in plants or fruits	4.5 \pm 0.65 μ M	
Fulvestrant	Steroidal ER antagonist	2.0 \pm 0.34 μ M	
4-OH-TOR	Hydroxy metabolite of TOR (nonsteroidal agonist-antagonist of ER)	0.2 \pm 0.02 μ M	[43]
Troglitazone	PPAR agonist	6.4 \pm 0.09 μ M	[44]
Endoxifen	Active metabolite of Tamoxifen (nonsteroidal antagonist of ER)	8.5 \pm 0.44 μ M	[45]
4-OH TAM	Hydroxy metabolite of Tamoxifen	24 \pm 5 μ M	
N-des TAM	N-demethyl metabolite of Tamoxifen	24 \pm 5 μ M	[46]
Tibolone	Selective tissue estrogenic activity regulator	96 \pm 52 μ M	
3 α -OH-TIB	Hydroxy metabolite of TIB	19.5 \pm 2.8 μ M	
3 β -OH-TIB	Hydroxy metabolite of TIB	6.6 \pm 2.2 μ M	[47]
		2.1 \pm 0.5 μ M	

K_m , the constant value of Michaelis-Menten equation which is numerically equal to the substrate concentration at the half reaction rate of enzyme V_{max} ; E₁, estrone; E₂, estradiol; EE₂, ethinylestradiol; DHEA, dehydroepiandrosterone; T₄, thyroxine; T₃, 3,3',5-triiodothyronine; rT₃, 3,3',5'-triiodothyronine; T₂, 3,3'-diiodothyronine; TOR, toremifene; ERs, estrogen receptors; GPER, G protein-coupled receptor; AR, androgen receptor; PPARs, peroxisome proliferator-activated receptors; 4-OH TAM, 4-hydroxy tamoxifen; N-des TAM, N-desmethyltamoxifen; TIB, tibolone.

SULT1E1 is also expressed in hormone-dependent tissues, such as endometrium [22,48] and placenta [21]. SULT1E1 is specifically expressed during the secretory phase of the menstrual cycle in human endometrium [49]. Upregulated SULT1E1 activity in the endometrium may result in sulfating E_2 after ovulation [50]. In addition, SULT1E1 can be induced by progestins in human Ishikawa endometrial adenocarcinoma cells [51].

As an interesting effect, estrogens inhibit expression of the potent growth factor repressor transforming growth factor (TGF)- β 1. In addition, it was observed that MCF-7 cells expressing SULT1E1 activity did not show a decrease in ER α levels, an increase in progesterone receptor, or a decrease in transforming growth factor- β expression, suggesting the rapid sulfoconjugation of E_2 by SULT1E1.

It is possible that SULT1E1 contributes to EE $_2$ sulfation during hepatic-mediated first-pass metabolism. SULT1E1 is the high-affinity enzyme responsible for EE $_2$ sulfation at nanomolar concentrations, so SULT1E1 plays a predominant role in the sulfation of EE $_2$ in the intestine and liver.

3.2. Sulfation of Thyroid Hormones

Many factors serve as regulators for the effectiveness and bioavailability of receptor active thyroid hormone (T_3) [52,53]. The prohormone thyroxine (T_4) is predominantly secreted to regulate metabolism [54,55]. Deiodination is one of the principal and major pathways to degrade active compounds, and there are three types of deiodinase selenoproteins—iodothyronine deiodinases (D1, D2, and D3) [56]. These deiodinases are promotive of the reductive T_4 deiodination and its metabolites (Figure 2).

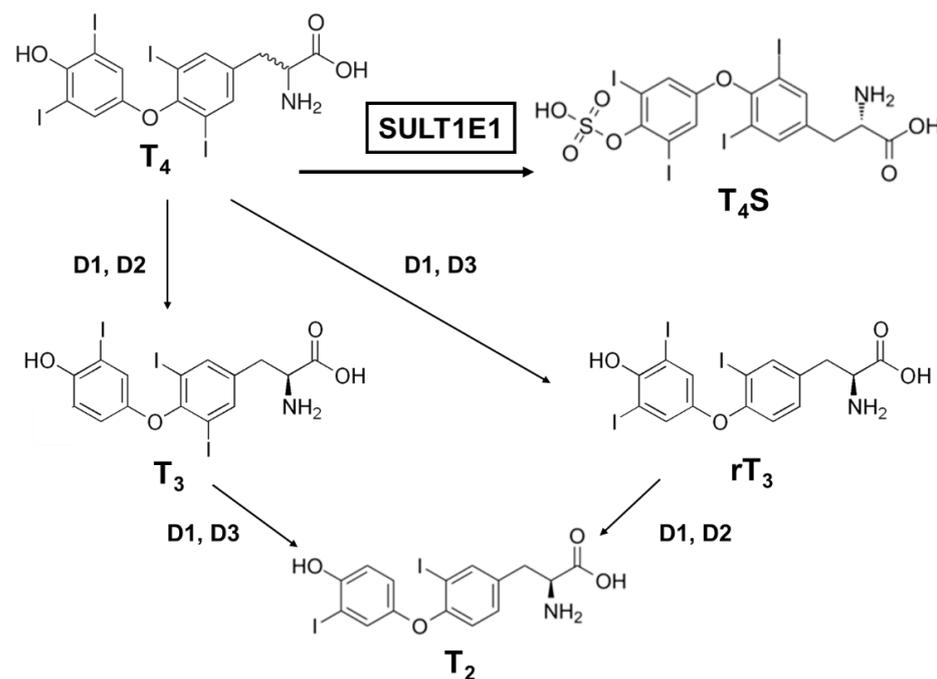


Figure 2. A schematic metabolic pathway of thyroid hormones. T_4 , thyroxine (prohormone); T_4S , thyroxine sulfate (sulfoconjugated metabolite); T_3 , 3,3',5-triiodothyronine (receptor active iodothyronine); rT_3 , 3,3',5'-triiodothyronine (receptor inactive iodothyronine); T_2 , 3,3'-diiodothyronine.

One major modification thyroid hormones receive is sulfation, which deactivates them. Thyroxine sulfate can be detected in human fetal blood and amniotic fluid, indicating that the production of sulfoconjugates is critical in utero [57]. Iodothyronine sulfates (T_4S , T_3S , rT_3S , and T_2S) are generated by SULT enzymes, which are located in a variety of different tissues, and catalyze the sulfation and substitution of the hydroxyl groups of various compounds using PAPS as the sulfate donor [58]. Interestingly, among the SULTs, most of the SULT1 enzymes catalyze the sulfation of iodothyronines [41,59–61]. SULT1E1 is

highly effective at catalyzing rT_3 sulfation and has sulfating activity for all iodothyronines (Table 4). For rT_3 sulfation especially, SULT1E1 has the highest activity among the SULT1 subfamily, even compared to SULT1A1 [59]. Moreover, SULT1E1 was reported as the most active enzyme that exhibited catalyzing activity for T_4 sulfation [62].

SULT1E1 can be detected in the human endometrium and in the mouse uterus, so it might be possible that the uterus could protect the fetus from excessive thyroid hormone by inactivating pathways via SULT1E1 or D3. It is notable that the metabolites derived from D3 (rT_3 and T_2) are also favorable substrates of SULT1E1, suggesting that T_4 and T_3 are metabolized in the uterus by consecutive sulfation. The physiological roles of each iodothyronine SULT still remain too complex to be comprehended in full. Although SULT1E1 has been proven to be a potent iodothyronine SULT along with SULT1A1, it is probable that the other SULT1 enzymes contribute to iodothyronine sulfation in a tissue- or growth-dependent way [63–65].

4. Sulfation of Other Substrates by SULT1E1

SULT1E1 has the role of sulfotransferase not only for endogenous substrates, such as estrogens or iodothyronines, but also for various other compounds (Table 4).

Flavonoids are a class of naturally occurring polyphenols in most plants, and they play diverse roles. Many of them have antioxidative influences *in vitro* and *in vivo*. Apigenin (4',5,7-trihydroxyflavone) is one of the flavonoids that usually exists in chamomile flowers, and it is a yellow compound that can dye wool [66]. Catechin enantiomers are ubiquitous constituents of herbal medicines. The active isomer (–)-epicatechin is known for its anti-inflammatory effects by the activation of the NF- κ B signaling pathway [67]. Resveratrol (3,5,4'-trihydroxy-trans-stilbene) is expressed in several plants in response to damage or attack by pathogens [68]. Chrysin (5,7-dihydroxyflavone) is typically found in honey or propolis [69]. Quercetin (5,7,3',4'-flavon-3-ol) is distributed in naturally occurring polar auxin transport inhibitor, and it is one of the most common natural dietary flavonoids [70]. Though most polyphenols are sulfated by SULT1A isoforms, many sulfoconjugated forms of polyphenols can be generated by SULT1E1 due to its phenotypic response at the cellular level [42].

Fulvestrant is a novel medicine for endocrine treatment; it is an antagonist of estrogen receptors (ERs) that provides no agonistic activity. This compound is an analog of E_2 that has a distinguishable structure from nonsteroidal medicines such as tamoxifen and other selective estrogen receptor modulators (SERMs). Fulvestrant performs as a competitive inhibitor and suppresses the binding of E_2 to the ERs, and SULT1E1 has exhibited clear sulfating activity towards fulvestrant [43].

Synthetic estrogens for oral administration are widely prescribed and given to fertile women. Various SERMs have been developed and administered to inhibit the activation of estrogen's activity in the breast. It has been revealed that SULT1E1 sulfates 4-hydroxytoremifene (4-OH TOR), an active metabolite of toremifene, alongside SULT1A1 [44]. Among the SULT isoforms, SULT1E1 has a high affinity for the tamoxifen active metabolite 4-hydroxytamoxifen (4-OH TAM) and other active tamoxifen metabolites, including endoxifen and *N*-desmethyltamoxifen (*N*-des TAM), which are substrates of SULT1E1 as well [46]. These metabolites show weak inhibitory effects on SULT1E1, suggesting that they are unlikely to interfere with the sulfation of E_2 in SULT1E1-expressing tissues.

Troglitazone acts as an agonist of PPAR α and has been used as an oral antidiabetic for the treatment of insulin-independent diabetes mellitus. SULT1E1 appropriately sulfates troglitazone and had greater activity than SULT1A1 when 10 μ M of troglitazone was treated [45].

After tibolone binds to nuclear receptors, such as ERs, progesterone receptor (PG), and androgen receptor (AR), to activate them, it is dramatically metabolized into two active hydroxylated isomers, 3 α -OH and 3 β -OH-tibolone, which can be metabolized into Δ^4 -tibolone. SULT1E1 sulfates tibolone as well as its metabolites, 3 α -OH and 3 β -OH-tibolones [47].

5. SULT1E1 and Diseases

Due to SULT1E1 being highly activated in pathophysiological conditions, such as estrogen-related diseases, the quantification of the E₁S form of estrogen during the menstrual cycle and in menopausal women has been widely used [71–73]. It has been reported that a strong association between breast cancer vulnerability and increased E₂ concentration exists [74]. Moreover, the concentrations of E₁S and E₂S are higher in patients with breast fibroadenoma [75] (Figure 3); however, in that same study, the expression of SULT1E1 decreased or was abolished in breast cancer tissues, though it was expressed in normal breast cells. In breast carcinoma cell lines, E₁ and E₂ can be sulfoconjugated by SULT1E, which appears to be expressed at low levels in breast cancer cells. The expression of SULT1E1 during the progression of tumorigenesis was characterized using an MCF-7 cell line transfected with SULT1E1, and it was observed that sulfation increased in the SULT1E1-transfected MCF-7 cells compared to the control cells [76]. A similar observation of the physiological implications of SULT1E1 expression was examined by the MCF-7 cell line as well; the response to physiological concentrations of E₂ was reduced, as determined in an estrogen-responsive reporter gene assay [77]. SULT1E1 has shown very strong affinity for the sulfation of E₂ and EE₂, so the ability of SULT1E1 to be involved in estrogen concentrations is important for regulating estrogen receptor target tissues. Estrogen-dependent breast cells with high SULT1E1 levels grow more slowly, suggesting an inhibitory role in carcinogenesis, depending on the role of SULT1E1 in creating physiologically inactive estrogen via sulfoconjugation [51,76,78,79].

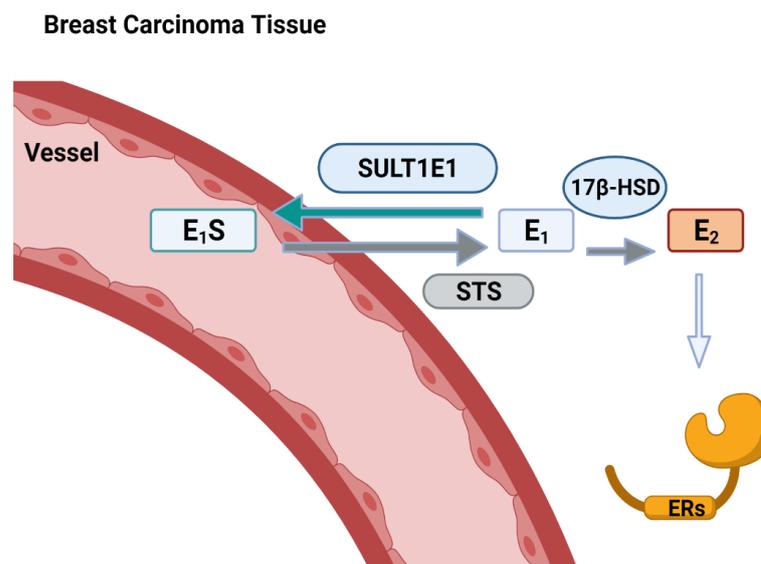


Figure 3. A schematic pathway for estrogen formation by SULT1E1 and STS in breast carcinoma tissue. E₁S, estrone sulfate; E₁, estrone; E₂, estradiol.

Due to the high homology (77.5%) between humans and mice, mouse models have been developed and studied in various approaches. Many pathological mouse models that are related to SULT1E1 have been studied, such as sepsis and diabetes. Sepsis is a lethal condition caused by physiological reactions to infections. There was an *in vivo* mouse study where hepatic SULT1E1 was upregulated via the activation of the NF-κB pathway's associated inflammatory pathways [80].

The Akita mouse was derived from C57BL/6J and inherited the mutated insulin 2 gene, so it can be used as a model of diabetes mellitus (DM) type 1. Interestingly, hepatic SULT1E1 mRNA was highly upregulated in Akita, and this pathological situation acts as a stimulus to regulate SULT1E1 expression via phosphorylated-ERα and dephosphorylated-CAR [32]. Likewise, diabetes type 2 mouse models (*db/db* and *ob/ob*) also exhibited

the hepatic overproduction of SULT1E1, representing SULT1E1's role in maintaining the balance of estrogen sulfation [81,82].

6. Functional Variants of SULT1E1 and Current Research Status

A total of 4760 single-nucleotide polymorphisms (SNPs) have been validated by frequency, cluster, and ALFA (allele frequency aggregator) out of the total of 5428 SNPs, including 214 missense variants in human *SULT1E1*, according to NCBI dbSNP. Most SNPs are intronic variants. Diverse studies have been conducted to identify *SULT1E1* polymorphisms and their effects, especially based on association cohort studies (Table 5).

Six SNPs from the introns of *SULT1E1* were associated with treatment failure of abiraterone acetate (AA) therapy in metastatic castration-resistant prostate cancer (mCRPC) patients [83]. Each DNA sample was isolated from patients with mCRPC who were treated with AA approximately three years previously, and the samples were analyzed for the study. In groups 1 (rs3775777, rs4149534, and rs10019305) and 2 (rs3775770, rs4149527, and rs3775768), it was observed that the patients carrying polymorphic alleles had the estimated hazard ratios of 3.58 and 3.12, respectively [83].

There was an association study using Korean females that included breast cancer patients and healthy subjects [84]. The patients carrying rs3775775 (TC or CC) had a hazard ratio of 3.2 (1.39–7.48) compared to that of TT carriers [84]. Regulating estrogen levels, which is especially related to SULT1E1's sulfation capacity, could facilitate the development of breast cancer or its avoidance in Korean females [84].

The most popular and broadly studied polymorphism of *SULT1E1* is rs3736599, which has a nucleotide alteration at c.-64G>A of the 5'UTR region. Though other variants were also involved, this variant influenced the DHEA sulfation, endometrial carcinogenesis risk, and bone mineral density in females [85–87]. There was a cohort study that enrolled equal numbers of African American (AA) and European-American (EA) women, and approximately 11 years after the study's inception, complete data were collected from 301 women. In the EA women, *SULT1E1* rs3736599 carriers had lower DHEA sulfate levels [85].

In a study in which 150 endometrial cancer patients in total and 165 age-matched healthy control individuals were enrolled [86], surprisingly, the odds ratios of AA and AA+GA were 3.50 and 1.76, respectively, reflecting the higher endometrial cancer risks [86].

In another study, 397 healthy Korean female subjects with menopause and without any cancer or thyroid-related disease history were genotyped to identify the differences in bone mineral density of the distal radius and calcaneus [87]. A variant of *SULT1E1*, rs3736599, was associated with bone mineral density of the distal radius and the calcaneus. Moreover, a combined effect between this polymorphism and altered estrogen consumption might exist in the calcaneus [87].

Three of the *SULT1E1* SNPs—Asp22Tyr (rs11569705), Ala32Val (rs34547148), and Pro253His (rs11569712)—were discovered, and these variants were in the encoded amino acids [88]. These alleles were transfected and expressed in COS-1 cells to discover their functional impacts on stability and activity. Among them, rs11569705 indicated the most significant decrease in enzyme activity and protein level, and rs34547148 also displayed a 50% decrease in both the enzyme and the protein [88].

Table 5. Reported human SULT1E1 functional variants.

Type	Position ¹	SNP ID ²	Effect	Reference
Intron	c.772+369T>C	rs3775777	Treatment failure on abiraterone acetate with mCRPC	[83]
	c.369+1930A>C	rs4149534		
	c.369+402T>C	rs10019305		
	c.-9-899G>A	rs3775770		
	c.-10+771C>A	rs4149527		
	c.-10+655G>A	rs3775768		
	c.-9-469G>A	rs3822172		
c.772+856G>T,C,A	rs1238574	Lower survival rate in colorectal cancer	[89]	
	c.369+1653T>C	rs3775775	Decreased survival rate from breast cancer	[84]

Table 5. Cont.

Type	Position ¹	SNP ID ²	Effect	Reference
5'UTR	c.-64G>A	rs3736599	Lower DHEA sulfate levels in the menopausal transition of European-American population	[85]
			May strongly contribute to risk for endometrial carcinogenesis in Caucasians	[86]
			Higher bone mineral density of distal radius and calcaneus in Korean women	[87]
Missense	95C>T (Ala32Val) 64G>A (Asp22Tyr)	rs34547148 rs11569705	Increased K_m value for the sulfation of E ₂	[88]

¹ All reference sequences are described according to GRCh38.p12 chromosome 4, and the accession number is NM_005420.3. ² Each single-nucleotide polymorphism (SNP) ID is described according to the NCBI dbSNP. UTR, untranslated region; mCRPC, metastatic castration-resistant prostate cancer; DHEA, dehydroepiandrosterone.

7. Future Directions for Clinical Integrations

SULT1E1 is responsible for the metabolism of active estrogens and plays crucial roles in their homeostasis. Therefore, this enzyme makes a variety of contributions to human health, including in regard to cancers and drug responses. However, the lack of genetic research on SULT1E1 needs to be enhanced by precisely designed studies in many respects. Several cohort-study-based analyses have been conducted regarding *SULT1E1* genetic variants, but relatively few compared to the number of such studies for the *SULT1A* subfamily.

Due to human SULT1E1's high nucleotide homology with several animal SULT1E1s, and their similar substrate-binding structures, animal models and in vivo studies have provided useful clues for the genetic regulation and kinetics of humans. Thus, using transgenic animal models would aid in determining gene–gene or gene–xenobiotic interactions in the study of SULT1E1 activity.

Since the substrate-binding sites and neighboring amino acids are regarded as being involved prominently in enzyme activity and structure, we suggest candidate SNPs corresponding to adjacent substrate-binding sites be investigated in genetic association studies (Table 6).

Several cohort studies have developed SULT1E1 association models, such as Predictors of Breast Cancer Recurrence (ProBeCaRE) [90] and U-statistics-based tests for identifying the pathway-based candidate genes of breast cancer and hormone metabolism pathways [91]. In addition, an intronic polymorphism (rs3775779) was discovered as a marker for analyzing the ethnic difference in the fine-scale population structure of Malays in Peninsular Malaysia and Singapore [92]. These studies suggest diverse scientific approaches to figure out the role of SULT1E1.

Many studies of SULT1E1 have highlighted aspects of its impacts on biological systems. Therefore, we encourage such studies to elucidate the related pathophysiological perspectives of human SULT1E1.

Table 6. Amino acids near to substrate-binding sites of SULT1E1.

Impacted Amino Acids	Substrate ¹	Alteration	SNP ID ²
Arg256	PAPS	Not reported	-
Phe254	E ₂ , 4-OH TCB	Phe254Cys	rs746067466
Met247	4-OH TCB	Met247Ile	rs1188553969
Ile246	4-OH TCB, TBBPA	Ile246Leu	rs1413235220
Tyr239	E ₂ , 4-OH TCB	Not reported	-
Phe228	PAPS	Not reported	-
Thr226	PAPS	Thr226Ser	rs756363002
Asn168	4-OH TCB, TBBPA	Asn168Ser	rs1265277815
Val145	4-OH TCB	Val145Leu	rs200443686
Phe141	E ₂ , 4-OH TCB, TBBPA, 3-OH BDE47	Phe141Leu	rs1220949195

Table 6. Cont.

Impacted Amino Acids	Substrate ¹	Alteration	SNP ID ²
Phe138	TBBPA	Not reported	-
Ser137	PAPS, E ₂	Ser137Pro	rs1208507410
Arg129	PAPS	Arg129Gln	rs774700339
His107	PAPS, E ₂ , 4-OH TCB, TBBPA	His107Arg	rs1316115370
Lys105	PAPS, E ₂ , 4-OH TCB, TBBPA, 3-OH BDE47	Not reported	-
Cys83	3-OH BDE47	Cys83Phe	rs1431397129
Phe80	E ₂ , 4-OH TCB, TBBPA, 3-OH BDE47	Not reported	-
Trp52	PAPS	Not reported	-
Thr51	PAPS	Thr51Ile Thr51Ala	rs1170826222 rs761632873
Thr50	PAPS	Not reported	-
Gly49	PAP	Gly49Val Gly49Ser	rs1460190031 rs1210226778
Ser48	PAP	Ser48Cys Ser48Pro	rs1336407598 rs1052854963
Lys47	PAPS, E ₂	Lys47Glu	rs1361781887
Pro46	4-OH TCB, TBBPA	Pro46Leu	rs771011878
Phe23	4-OH TCB	Phe23Cys	rs1400776691
Asp22	4-OH TCB	Asp22Asn Asp22Tyr	rs11569705
Tyr20	PAP-E ₂ , 4-OH TCB, TBBPA	Tyr20Cys	rs778407495

¹ The crystal structures and neighboring amino acids of SULT1E1 substrate-binding sites were described according to the RCSB protein data bank (PDBid: 1G3M, 1HY3, 4JVM, 4JVN, and 4JVL) [11,12,93]. ² Each SNP ID was based on NCBI dbSNP. 4-OH TCB, 4,4'-OH-3,5,3',5'-tetrachlorinated biphenyl; TBBPA, tetrabromobisphenol A; 3-OH BDE47, 3-hydroxyl bromodiphenyl ether.

Author Contributions: Literature search, M.Y.; writing, M.Y.; editing, M.Y., M.N. and S.-J.L.; funding acquisition, S.-J.L. All authors have read and agreed to the published version of the manuscript.

Funding: The article was supported by the grants from the National Research Foundation of Korea funded by the Korean government (NRF-2020R1I1A3073778) and the National Research Foundation of Korea funded by Korea (MIST) (number 2018R1A5A2021242). This article was also supported by the reviews from the National Institute of Environmental Health Sciences (NIEHS) Division of Intramural Research (DIR).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest for this article.

References

1. Gamage, N.; Barnett, A.; Hempel, N.; Duggleby, R.G.; Windmill, K.F.; Martin, J.L.; McManus, M.E. Human sulfotransferases and their role in chemical metabolism. *Toxicol. Sci.* **2006**, *90*, 5–22. [[CrossRef](#)]
2. Liu, L.; Klaassen, C.D. Regulation of hepatic sulfotransferases by steroidal chemicals in rats. *Drug Metab. Dispos.* **1996**, *24*, 854–858. [[PubMed](#)]
3. Sonoda, J.; Xie, W.; Rosenfeld, J.M.; Barwick, J.L.; Guzelian, P.S.; Evans, R.M. Regulation of a xenobiotic sulfonation cascade by nuclear pregnane X receptor (PXR). *Proc. Natl. Acad. Sci. USA* **2002**, *99*, 13801–13806. [[CrossRef](#)]
4. Song, C.S.; Echchgadda, I.; Baek, B.S.; Ahn, S.C.; Oh, T.; Roy, A.K.; Chatterjee, B. Dehydroepiandrosterone sulfotransferase gene induction by bile acid activated farnesoid X receptor. *J. Biol. Chem.* **2001**, *276*, 42549–42556. [[CrossRef](#)]
5. Qatanani, M.; Zhang, J.; Moore, D.D. Role of the constitutive androstane receptor in xenobiotic-induced thyroid hormone metabolism. *Endocrinology* **2005**, *146*, 995–1002. [[CrossRef](#)]

6. Tien, E.S.; Matsui, K.; Moore, R.; Negishi, M. The nuclear receptor constitutively active/androstane receptor regulates type 1 deiodinase and thyroid hormone activity in the regenerating mouse liver. *J. Pharmacol. Exp. Ther.* **2007**, *320*, 307–313. [[CrossRef](#)]
7. Falany, C.N. Enzymology of human cytosolic sulfotransferases. *FASEB J.* **1997**, *11*, 206–216. [[CrossRef](#)] [[PubMed](#)]
8. Klaassen, C.D.; Boles, J.W. Sulfation and sulfotransferases 5: The importance of 3'-phosphoadenosine 5'-phosphosulfate (PAPS) in the regulation of sulfation. *FASEB J.* **1997**, *11*, 404–418. [[CrossRef](#)]
9. Kakuta, Y.; Pedersen, L.G.; Carter, C.W.; Negishi, M.; Pedersen, L.C. Crystal structure of estrogen sulphotransferase. *Nat. Struct. Biol.* **1997**, *4*, 904–908. [[CrossRef](#)] [[PubMed](#)]
10. Kakuta, Y.; Petrotchenko, E.V.; Pedersen, L.C.; Negishi, M. The sulfuryl transfer mechanism. Crystal structure of a vanadate complex of estrogen sulfotransferase and mutational analysis. *J. Biol. Chem.* **1998**, *273*, 27325–27330. [[CrossRef](#)]
11. Pedersen, L.C.; Petrotchenko, E.; Shevtsov, S.; Negishi, M. Crystal structure of the human estrogen sulfotransferase-PAPS complex: Evidence for catalytic role of Ser137 in the sulfuryl transfer reaction. *J. Biol. Chem.* **2002**, *277*, 17928–17932. [[CrossRef](#)] [[PubMed](#)]
12. Shevtsov, S.; Petrotchenko, E.V.; Pedersen, L.C.; Negishi, M. Crystallographic analysis of a hydroxylated polychlorinated biphenyl (OH-PCB) bound to the catalytic estrogen binding site of human estrogen sulfotransferase. *Environ. Health Perspect.* **2003**, *111*, 884–888. [[CrossRef](#)] [[PubMed](#)]
13. Allali-Hassani, A.; Pan, P.W.; Dombrowski, L.; Najmanovich, R.; Tempel, W.; Dong, A.; Loppnau, P.; Martin, F.; Thornton, J.; Edwards, A.M.; et al. Structural and chemical profiling of the human cytosolic sulfotransferases. *PLoS Biol.* **2007**, *5*, e97.
14. Kodama, S.; Negishi, M. Sulfotransferase genes: Regulation by nuclear receptors in response to xeno/endo-biotics. *Drug Metab. Rev.* **2013**, *45*, 441–449. [[CrossRef](#)]
15. Falany, C.N.; Krasnykh, V.; Falany, J.L. Bacterial expression and characterization of a cDNA for human liver estrogen sulfotransferase. *J. Steroid Biochem. Mol. Biol.* **1995**, *52*, 529–539. [[CrossRef](#)]
16. Zhang, H.; Varlamova, O.; Vargas, F.M. Sulfuryl transfer: The catalytic mechanism of human estrogen sulfotransferase. *J. Biol. Chem.* **1998**, *273*, 10888–10892. [[CrossRef](#)]
17. Faucher, F.; Lacoste, L.; Dufort, I.; Luu-The, V. High metabolism of catecholestrogens by type 1 estrogen sulfotransferase (hEST1). *J. Steroid Biochem. Mol. Biol.* **2001**, *77*, 83–86. [[CrossRef](#)]
18. Faucher, F.; Lacoste, L.; Luu-The, V. Human type 1 estrogen sulfotransferase: Catecholesterogen metabolism and potential involvement in cancer promotion. *Ann. N. Y. Acad. Sci.* **2002**, *963*, 221–228. [[CrossRef](#)]
19. Adjei, A.A.; Weinshilboum, R.M. Catecholesterogen sulfation: Possible role in carcinogenesis. *Biochem. Biophys. Res. Commun.* **2002**, *292*, 402–408. [[CrossRef](#)]
20. Her, C.; Aksoy, I.A.; Kimura, S.; Brandriff, B.F.; Wasmuth, J.J.; Weinshilboum, R.M. Human estrogen sulfotransferase gene (STE): Cloning, structure, and chromosomal localization. *Genomics* **1995**, *29*, 16–23. [[CrossRef](#)]
21. Stanley, E.L.; Hume, R.; Visser, T.J.; Coughtrie, M.W. Differential expression of sulfotransferase enzymes involved in thyroid hormone metabolism during human placental development. *J. Clin. Endocrinol. Metab.* **2001**, *86*, 5944–5955. [[CrossRef](#)]
22. Rubin, G.L.; Harrold, A.J.; Mills, J.A.; Falany, C.N.; Coughtrie, M.W. Regulation of sulphotransferase expression in the endometrium during the menstrual cycle, by oral contraceptives and during early pregnancy. *Mol. Hum. Reprod.* **1999**, *5*, 995–1002. [[CrossRef](#)]
23. Song, W.C.; Qian, Y.; Li, A.P. Estrogen sulfotransferase expression in the human liver: Marked interindividual variation and lack of gender specificity. *J. Pharmacol. Exp. Ther.* **1998**, *284*, 1197–1202.
24. Li, Y.; Xu, Y.; Li, X.; Qin, Y.; Hu, R. Effects of PPAR- α agonist and IGF-1 on estrogen sulfotransferase in human vascular endothelial and smooth muscle cells. *Mol. Med. Rep.* **2013**, *8*, 133–139. [[CrossRef](#)]
25. Xu, Y.; Yang, X.; Wang, Z.; Li, M.; Ning, Y.; Chen, S.; Yin, L.; Li, X. Estrogen sulfotransferase (SULT1E1) regulates inflammatory response and lipid metabolism of human endothelial cells via PPAR γ . *Mol. Cell Endocrinol.* **2013**, *369*, 140–149. [[CrossRef](#)] [[PubMed](#)]
26. Wang, S.; Yuan, X.; Lu, D.; Guo, L.; Wu, B. Farnesoid X receptor regulates SULT1E1 expression through inhibition of PGC1 α binding to HNF4 α . *Biochem. Pharmacol.* **2017**, *145*, 202–209. [[CrossRef](#)]
27. Liu, X.; Xue, R.; Yang, C.; Gu, J.; Chen, S.; Zhang, S. Cholestasis-induced bile acid elevates estrogen level via farnesoid X receptor-mediated suppression of the estrogen sulfotransferase SULT1E1. *J. Biol. Chem.* **2018**, *293*, 12759–12769. [[CrossRef](#)] [[PubMed](#)]
28. Hu, H.; Negishi, M. ROR α phosphorylation by casein kinase 1 α as glucose signal to regulate estrogen sulfation in human liver cells. *Biochem. J.* **2020**, *477*, 3583–3598. [[CrossRef](#)]
29. Gong, H.; Guo, P.; Zhai, Y.; Zhou, J.; Uppal, H.; Jarzynka, M.J.; Song, W.C.; Cheng, S.Y.; Xie, W. Estrogen deprivation and inhibition of breast cancer growth in vivo through activation of the orphan nuclear receptor liver X receptor. *Mol. Endocrinol.* **2007**, *21*, 1781–1790. [[CrossRef](#)] [[PubMed](#)]
30. Gong, H.; Jarzynka, M.J.; Cole, T.J.; Lee, J.H.; Wada, T.; Zhang, B.; Gao, J.; Song, W.C.; DeFranco, D.B.; Cheng, S.Y.; et al. Glucocorticoids antagonize estrogens by glucocorticoid receptor-mediated activation of estrogen sulfotransferase. *Cancer Res.* **2008**, *68*, 7386–7393. [[CrossRef](#)]
31. Sueyoshi, T.; Green, W.D.; Vinal, K.; Woodrum, T.S.; Moore, R.; Negishi, M. Garlic extract diallyl sulfide (DAS) activates nuclear receptor CAR to induce the Sult1e1 gene in mouse liver. *PLoS ONE* **2011**, *6*, e21229. [[CrossRef](#)]
32. Yi, M.; Fashe, M.; Arakawa, S.; Moore, R.; Sueyoshi, T.; Negishi, M. Nuclear receptor CAR-ER α signaling regulates the estrogen sulfotransferase gene in the liver. *Sci. Rep.* **2020**, *10*, 5001. [[CrossRef](#)] [[PubMed](#)]

33. Hu, H.; Yokobori, K.; Negishi, M. PXR phosphorylated at Ser350 transduces a glucose signal to repress the estrogen sulfotransferase gene in human liver cells and fasting signal in mouse livers. *Biochem. Pharmacol.* **2020**, *180*, 114197. [[CrossRef](#)]
34. Fashe, M.; Hashiguchi, T.; Yi, M.; Moore, R.; Negishi, M. Phenobarbital-induced phosphorylation converts nuclear receptor RORalpha from a repressor to an activator of the estrogen sulfotransferase gene Sult1e1 in mouse livers. *FEBS Lett.* **2018**, *592*, 2760–2768. [[CrossRef](#)] [[PubMed](#)]
35. Poisson Paré, D.; Song, D.; Luu-The, V.; Han, B.; Li, S.; Liu, G.; Labrie, F.; Pelletier, G. Expression of Estrogen Sulfotransferase 1E1 and Steroid Sulfatase in Breast Cancer: A Immunohistochemical Study. *Breast Cancer* **2009**, *3*, 9–21.
36. Song, W.C.; Melner, M.H. Steroid transformation enzymes as critical regulators of steroid action in vivo. *Endocrinology* **2000**, *141*, 1587–1589. [[CrossRef](#)]
37. Falany, C.N.; Wheeler, J.; Oh, T.S.; Falany, J.L. Steroid sulfation by expressed human cytosolic sulfotransferases. *J. Steroid Biochem. Mol. Biol.* **1994**, *48*, 369–375. [[CrossRef](#)]
38. Falany, C.N.; Comer, K.A.; Dooley, T.P.; Glatt, H. Human dehydroepiandrosterone sulfotransferase. Purification, molecular cloning, and characterization. *Ann. N. Y. Acad. Sci.* **1995**, *774*, 59–72. [[CrossRef](#)]
39. Aksoy, I.A.; Wood, T.C.; Weinshilboum, R. Human liver estrogen sulfotransferase: Identification by cDNA cloning and expression. *Biochem. Biophys. Res. Commun.* **1994**, *200*, 1621–1629. [[CrossRef](#)]
40. Schrag, M.L.; Cui, D.; Rushmore, T.H.; Shou, M.; Ma, B.; Rodrigues, A.D. Sulfotransferase 1E1 is a low km isoform mediating the 3-O-sulfation of ethinyl estradiol. *Drug Metab. Dispos.* **2004**, *32*, 1299–1303. [[CrossRef](#)]
41. Kester, M.H.; van Dijk, C.H.; Tibboel, D.; Hood, A.M.; Rose, N.J.; Meinel, W.; Pabel, U.; Glatt, H.; Falany, C.N.; Coughtrie, M.W.; et al. Sulfation of thyroid hormone by estrogen sulfotransferase. *J. Clin. Endocrinol. Metab.* **1999**, *84*, 2577–2580. [[CrossRef](#)] [[PubMed](#)]
42. Ung, D.; Nagar, S. Variable sulfation of dietary polyphenols by recombinant human sulfotransferase (SULT) 1A1 genetic variants and SULT1E1. *Drug Metab. Dispos.* **2007**, *35*, 740–746. [[CrossRef](#)] [[PubMed](#)]
43. Edavana, V.K.; Yu, X.; Dhakal, I.B.; Williams, S.; Ning, B.; Cook, I.T.; Caldwell, D.; Falany, C.N.; Kadlubar, S. Sulfation of fulvestrant by human liver cytosols and recombinant SULT1A1 and SULT1E1. *Pharmgenom. Pers. Med.* **2011**, *4*, 137–145.
44. Edavana, V.K.; Dhakal, I.B.; Yu, X.; Williams, S.; Kadlubar, S. Sulfation of 4-hydroxy toremifene: Individual variability, isoform specificity, and contribution to toremifene pharmacogenomics. *Drug Metab. Dispos.* **2012**, *40*, 1210–1215. [[CrossRef](#)] [[PubMed](#)]
45. Honma, W.; Shimada, M.; Sasano, H.; Ozawa, S.; Miyata, M.; Nagata, K.; Ikeda, T.; Yamazoe, Y. Phenol sulfotransferase, ST1A3, as the main enzyme catalyzing sulfation of troglitazone in human liver. *Drug Metab. Dispos.* **2002**, *30*, 944–949. [[CrossRef](#)]
46. Squirewell, E.J.; Duffel, M.W. The effects of endoxifen and other major metabolites of tamoxifen on the sulfation of estradiol catalyzed by human cytosolic sulfotransferases hSULT1E1 and hSULT1A1*1. *Drug Metab. Dispos.* **2015**, *43*, 843–850. [[CrossRef](#)]
47. Falany, J.L.; Macrina, N.; Falany, C.N. Sulfation of tibolone and tibolone metabolites by expressed human cytosolic sulfotransferases. *J. Steroid Biochem. Mol. Biol.* **2004**, *88*, 383–391. [[CrossRef](#)]
48. Buirchell, B.J.; Hahnel, R. Metabolism of estradiol-17beta in human endometrium during the menstrual cycle. *J. Steroid Biochem.* **1975**, *6*, 1489–1494. [[CrossRef](#)]
49. Falany, J.L.; Azziz, R.; Falany, C.N. Identification and characterization of cytosolic sulfotransferases in normal human endometrium. *Chem. Biol. Interact.* **1998**, *109*, 329–339. [[CrossRef](#)]
50. Kotov, A.; Falany, J.L.; Wang, J.; Falany, C.N. Regulation of estrogen activity by sulfation in human Ishikawa endometrial adenocarcinoma cells. *J. Steroid Biochem. Mol. Biol.* **1999**, *68*, 137–144. [[CrossRef](#)]
51. Falany, J.L.; Falany, C.N. Regulation of estrogen sulfotransferase in human endometrial adenocarcinoma cells by progesterone. *Endocrinology* **1996**, *137*, 1395–1401. [[CrossRef](#)]
52. Moreau, X.; Lejeune, P.J.; Jeanningros, R. Kinetics of red blood cell T3 uptake in hypothyroidism with or without hormonal replacement, in the rat. *J. Endocrinol. Invest.* **1999**, *22*, 257–261. [[CrossRef](#)] [[PubMed](#)]
53. Kim, S.Y.; Seo, S.; Choi, K.H.; Yun, J. Evaluation of phototoxicity of tattoo pigments using the 3 T3 neutral red uptake phototoxicity test and a 3D human reconstructed skin model. *Toxicol. In Vitro* **2020**, *65*, 104813. [[CrossRef](#)] [[PubMed](#)]
54. Leiner, K.A.; Mackenzie, D.S. Central regulation of thyroidal status in a teleost fish: Nutrient stimulation of T4 secretion and negative feedback of T3. *J. Exp. Zool. Comp. Exp. Biol.* **2003**, *298*, 32–43. [[CrossRef](#)] [[PubMed](#)]
55. Mihasan, M.; Brandsch, R. A predicted T4 secretion system and conserved DNA-repeats identified in a subset of related *Arthrobacter* plasmids. *Microbiol. Res.* **2016**, *191*, 32–37. [[CrossRef](#)]
56. St Germain, D.L.; Galton, V.A.; Hernandez, A. Minireview: Defining the roles of the iodothyronine deiodinases: Current concepts and challenges. *Endocrinology* **2009**, *150*, 1097–1107. [[CrossRef](#)]
57. Wu, S.Y.; Huang, W.S.; Polk, D.; Florsheim, W.H.; Green, W.L.; Fisher, D.A. Identification of thyroxine-sulfate (T4S) in human serum and amniotic fluid by a novel T4S radioimmunoassay. *Thyroid* **1992**, *2*, 101–105. [[CrossRef](#)]
58. Chatterjee, B.; Song, C.S.; Kim, J.M.; Roy, A.K. Androgen and estrogen sulfotransferases of the rat liver: Physiological function, molecular cloning, and in vitro expression. *Chem. Biol. Interact.* **1994**, *92*, 273–279. [[CrossRef](#)]
59. Kester, M.H.; Kaptein, E.; Roest, T.J.; van Dijk, C.H.; Tibboel, D.; Meinel, W.; Glatt, H.; Coughtrie, M.W.; Visser, T.J. Characterization of human iodothyronine sulfotransferases. *J. Clin. Endocrinol. Metab.* **1999**, *84*, 1357–1364. [[CrossRef](#)]
60. Wang, J.; Falany, J.L.; Falany, C.N. Expression and characterization of a novel thyroid hormone-sulfating form of cytosolic sulfotransferase from human liver. *Mol. Pharmacol.* **1998**, *53*, 274–282. [[CrossRef](#)]
61. Li, X.; Clemens, D.L.; Anderson, R.J. Sulfation of iodothyronines by human sulfotransferase 1C1 (SULT1C1). *Biochem. Pharmacol.* **2000**, *60*, 1713–1716. [[CrossRef](#)]

62. Kester, M.; Coughtrie, M.W.; Visser, T.J. Sulfation of Thyroid Hormones. In *Human Cytosolic Sulfotransferases*; Pacifici, G.M., Coughtrie, M.W., Eds.; CRC Press/Taylor & Francis Group: Boca Raton, FL, USA, 2005; pp. 121–134.
63. Glatt, H.; Boeing, H.; Engelke, C.E.; Ma, L.; Kuhlmann, A.; Pabel, U.; Pomplun, D.; Teubner, W.; Meinel, W. Human cytosolic sulphotransferases: Genetics, characteristics, toxicological aspects. *Mutat. Res.* **2001**, *482*, 27–40. [[CrossRef](#)]
64. Weinsilboum, R.M.; Otterness, D.M.; Aksoy, I.A.; Wood, T.C.; Her, C.; Raftogianis, R.B. Sulfation and sulfotransferases 1: Sulfotransferase molecular biology: cDNAs and genes. *FASEB J.* **1997**, *11*, 3–14. [[CrossRef](#)]
65. Kiehlbauch, C.C.; Lam, Y.F.; Ringer, D.P. Homodimeric and heterodimeric aryl sulfotransferases catalyze the sulfuric acid esterification of N-hydroxy-2-acetylaminofluorene. *J. Biol. Chem.* **1995**, *270*, 18941–18947. [[CrossRef](#)] [[PubMed](#)]
66. Salehi, B.; Venditti, A.; Sharifi-Rad, M.; Kregiel, D.; Sharifi-Rad, J.; Durazzo, A.; Lucarini, M.; Santini, A.; Souto, E.B.; Novellino, E.; et al. The Therapeutic Potential of Apigenin. *Int. J. Mol. Sci.* **2019**, *20*, 1305. [[CrossRef](#)] [[PubMed](#)]
67. Tan, J.; de Bruijn, W.J.C.; van Zadelhoff, A.; Lin, Z.; Vincken, J.P. Browning of Epicatechin (EC) and Epigallocatechin (EGC) by Auto-Oxidation. *J. Agric. Food Chem.* **2020**, *68*, 13879–13887. [[CrossRef](#)] [[PubMed](#)]
68. Salehi, B.; Mishra, A.P.; Nigam, M.; Sener, B.; Kilic, M.; Sharifi-Rad, M.; Fokou, P.V.T.; Martins, N.; Sharifi-Rad, J. Resveratrol: A Double-Edged Sword in Health Benefits. *Biomedicines* **2018**, *6*, 91. [[CrossRef](#)]
69. Jana, K.; Yin, X.; Schiffer, R.B.; Chen, J.J.; Pandey, A.K.; Stocco, D.M.; Grammas, P.; Wang, X. Chrysin, a natural flavonoid enhances steroidogenesis and steroidogenic acute regulatory protein gene expression in mouse Leydig cells. *J. Endocrinol.* **2008**, *197*, 315–323. [[CrossRef](#)]
70. Li, Y.; Yao, J.; Han, C.; Yang, J.; Chaudhry, M.T.; Wang, S.; Liu, H.; Yin, Y. Quercetin, Inflammation and Immunity. *Nutrients* **2016**, *8*, 167. [[CrossRef](#)]
71. Pasqualini, J.R.; Chetrite, G.; Blacker, C.; Feinstein, M.C.; Delalonde, L.; Talbi, M.; Maloche, C. Concentrations of estrone, estradiol, and estrone sulfate and evaluation of sulfatase and aromatase activities in pre- and postmenopausal breast cancer patients. *J. Clin. Endocrinol. Metab.* **1996**, *81*, 1460–1464.
72. Mady, E.A.; Ramadan, E.E.; Ossman, A.A. Sex steroid hormones in serum and tissue of benign and malignant breast tumor patients. *Dis. Markers* **2000**, *16*, 151–157. [[CrossRef](#)] [[PubMed](#)]
73. Bonorden, M.J.; Greany, K.A.; Wangen, K.E.; Phipps, W.R.; Feirtag, J.; Adlercreutz, H.; Kurzer, M.S. Consumption of Lactobacillus acidophilus and Bifidobacterium longum do not alter urinary equol excretion and plasma reproductive hormones in premenopausal women. *Eur. J. Clin. Nutr.* **2004**, *58*, 1635–1642. [[CrossRef](#)]
74. Thomas, H.V.; Key, T.J.; Allen, D.S.; Moore, J.W.; Dowsett, M.; Fentiman, I.S.; Wang, D.Y. A prospective study of endogenous serum hormone concentrations and breast cancer risk in post-menopausal women on the island of Guernsey. *Br. J. Cancer* **1997**, *76*, 401–405. [[CrossRef](#)]
75. Pasqualini, J.R.; Cortes-Prieto, J.; Chetrite, G.; Talbi, M.; Ruiz, A. Concentrations of estrone, estradiol and their sulfates, and evaluation of sulfatase and aromatase activities in patients with breast fibroadenoma. *Int. J. Cancer* **1997**, *70*, 639–643. [[CrossRef](#)]
76. Falany, J.L.; Falany, C.N. Expression of cytosolic sulfotransferases in normal mammary epithelial cells and breast cancer cell lines. *Cancer Res.* **1996**, *56*, 1551–1555. [[PubMed](#)]
77. Chetrite, G.S.; Paris, J.; Shields-Botella, J.; Philippe, J.C.; Pasqualini, J.R. Effect of noregestrol acetate on human estrogen sulfotransferase activity in the hormone-dependent MCF-7 and T-47D breast cancer cell lines. *Anticancer Res.* **2003**, *23*, 4651–4655.
78. Qian, Y.; Deng, C.; Song, W.C. Expression of estrogen sulfotransferase in MCF-7 cells by cDNA transfection suppresses the estrogen response: Potential role of the enzyme in regulating estrogen-dependent growth of breast epithelial cells. *J. Pharmacol. Exp. Ther.* **1998**, *286*, 555–560.
79. Qian, Y.M.; Sun, X.J.; Tong, M.H.; Li, X.P.; Richa, J.; Song, W.C. Targeted disruption of the mouse estrogen sulfotransferase gene reveals a role of estrogen metabolism in intracrine and paracrine estrogen regulation. *Endocrinology* **2001**, *142*, 5342–5350. [[CrossRef](#)] [[PubMed](#)]
80. Chai, X.; Guo, Y.; Jiang, M.; Hu, B.; Li, Z.; Fan, J.; Deng, M.; Billiar, T.R.; Kucera, H.R.; Gaikwad, N.W.; et al. Oestrogen sulfotransferase ablation sensitizes mice to sepsis. *Nat. Commun.* **2015**, *6*, 7979. [[CrossRef](#)]
81. Leiter, E.H.; Chapman, H.D. Obesity-induced diabetes (diabesity) in C57BL/KsJ mice produces aberrant trans-regulation of sex steroid sulfotransferase genes. *J. Clin. Investig.* **1994**, *93*, 2007–2013. [[CrossRef](#)] [[PubMed](#)]
82. Gao, J.; He, J.; Shi, X.; Stefanovic-Racic, M.; Xu, M.; O'Doherty, R.M.; Garcia-Ocana, A.; Xie, W. Sex-specific effect of estrogen sulfotransferase on mouse models of type 2 diabetes. *Diabetes* **2012**, *61*, 1543–1551. [[CrossRef](#)]
83. Agarwal, N.; Alex, A.B.; Farnham, J.M.; Patel, S.; Gill, D.; Buckley, T.H.; Stephenson, R.A.; Cannon-Albright, L. Inherited variants in SULT1E1 and response to abiraterone acetate by men with metastatic castration refractory prostate cancer. *J. Urol.* **2016**, *196*, 1112–1116. [[CrossRef](#)]
84. Choi, J.Y.; Lee, K.M.; Park, S.K.; Noh, D.Y.; Ahn, S.H.; Chung, H.W.; Han, W.; Kim, J.S.; Shin, S.G.; Jang, I.J.; et al. Genetic polymorphisms of SULT1A1 and SULT1E1 and the risk and survival of breast cancer. *Cancer Epidemiol. Biomark. Prev.* **2005**, *14*, 1090–1095. [[CrossRef](#)] [[PubMed](#)]
85. Rebbeck, T.R.; Su, H.I.; Sammel, M.D.; Lin, H.; Tran, T.V.; Gracia, C.R.; Freeman, E.W. Effect of hormone metabolism genotypes on steroid hormone levels and menopausal symptoms in a prospective population-based cohort of women experiencing the menopausal transition. *Menopause* **2010**, *17*, 1026–1034. [[CrossRef](#)]

86. Hirata, H.; Hinoda, Y.; Okayama, N.; Suehiro, Y.; Kawamoto, K.; Kikuno, N.; Rabban, J.T.; Chen, L.M.; Dahiya, R. CYP1A1, SULT1A1, and SULT1E1 polymorphisms are risk factors for endometrial cancer susceptibility. *Cancer* **2008**, *112*, 1964–1973. [[CrossRef](#)] [[PubMed](#)]
87. Lee, S.A.; Choi, J.Y.; Shin, C.S.; Hong, Y.C.; Chung, H.; Kang, D. SULT1E1 genetic polymorphisms modified the association between phytoestrogen consumption and bone mineral density in healthy Korean women. *Calcif. Tissue Int.* **2006**, *79*, 152–159. [[CrossRef](#)] [[PubMed](#)]
88. Adjei, A.A.; Thoma, B.A.; Prondzinski, J.L.; Eckloff, B.W.; Wieben, E.D.; Weinshilboum, R.M. Human estrogen sulfotransferase (SULT1E1) pharmacogenomics: Gene resequencing and functional genomics. *Br. J. Pharmacol.* **2003**, *139*, 1373–1382. [[CrossRef](#)]
89. Li, S.; Xie, L.; Du, M.; Xu, K.; Zhu, L.; Chu, H.; Chen, J.; Wang, M.; Zhang, Z.; Gu, D. Association study of genetic variants in estrogen metabolic pathway genes and colorectal cancer risk and survival. *Arch. Toxicol.* **2018**, *92*, 1991–1999. [[CrossRef](#)]
90. Collin, L.J.; Cronin-Fenton, D.P.; Ahern, T.P.; Christiansen, P.M.; Damkier, P.; Ejlersen, B.; Hamilton-Dutoit, S.; Kjærsgaard, A.; Silliman, R.A.; Sørensen, H.T.; et al. Cohort Profile: The Predictors of Breast Cancer Recurrence (ProBe CaRE) Premenopausal Breast Cancer Cohort Study in Denmark. *BMJ Open* **2018**, *8*, e021805. [[CrossRef](#)]
91. Wei, Z.; Li, M.; Rebbeck, T.; Li, H. U-statistics-based tests for multiple genes in genetic association studies. *Ann. Hum. Genet.* **2008**, *72*, 821–833. [[CrossRef](#)]
92. Hoh, B.P.; Deng, L.; Julia-Ashazila, M.J.; Zuraihan, Z.; Nur-Hasnah, M.; Nur-Shafawati, A.R.; Hatin, W.I.; Endom, I.; Zilfalil, B.A.; Khalid, Y.; et al. Fine-scale population structure of Malays in Peninsular Malaysia and Singapore and implications for association studies. *Hum. Genom.* **2015**, *9*, 16. [[CrossRef](#)] [[PubMed](#)]
93. Gosavi, R.A.; Knudsen, G.A.; Birnbaum, L.S.; Pedersen, L.C. Mimicking of estradiol binding by flame retardants and their metabolites: A crystallographic analysis. *Environ. Health Perspect.* **2013**, *121*, 1194–1199. [[CrossRef](#)] [[PubMed](#)]